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Title: Source or sink? Integrating biogeochemical, trophic and landscape processes to model lake organic carbon budgets

Source or sink? Dominant processes in organic carbon cycling in lakes revealed by dynamic mechanistic modeling

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## **ABSTRACT**

Lakes are active processors of organic carbon (OC) and are thought to play important roles in landscape and global carbon cycles. OC is produced and consumed during primary production and respiration in lakes, buried in lake sediments and exported via surface or groundwater outflows. Although these processes provide a basis for a conceptual understanding of lake OC budgets, few studies have integrated these various fluxes under a dynamic modeling framework to examine their interactions and relative magnitudes. We used our conceptual understanding of lake OC budgets to develop a flexible, dynamical mass balance model for OC, and applied the model to a heterogeneous set of five lakes. We examined the relative magnitudes of OC fluxes and found that long-term lake OC dynamics were predominantly driven by allochthonous loads in four of the five lakes, underscoring the importance of terrestrially-derived OC in lake ecosystems. Burial, the main OC storage mechanism in lakes, represented a relatively small component of the total budget and was driven largely by autochthony. Respiration, the mechanism by which lakes export carbon to the atmosphere, ranged from 14-90% of total OC loads, whereas burial accounted for 7-37% of total OC loads. Given the relatively low rates of burial compared to respiration across 4 of the 5 lakes, we concluded that these lakes consistently represented net OC sources across all modeled years; however, source capacity varied four-fold among lakes. The lone sink was Lake Monona, Wisconsin, the most productive lake in our dataset, in which average burial rates were more than twice those of respiration across modeled years. Additionally, we found that lakes generally transitioned seasonally from OC sinks to sources as water temperatures and lake productivity increased. Finally, we highlighted critical research needs, which include surface water DOC observations in paired tributary and lake settings, measurements of OC burial rates, and budgets of particulate OC. These data will help better constrain parameter estimates in future lake OC models and improve our understanding of landscape carbon cycling.

Key words: carbon cycle, carbon flux, limnology, mass balance, allochthony, autochthony, sedimentation, dissolved organic carbon, particulate organic carbon, LTER

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## **INTRODUCTION**

Lakes are dynamic components of the landscape that actively process and transport terrestrially derived organic carbon (OC; Cole et al. 2007, Tranvik et al. 2009, Tanentzap et al. 2017), as well as emit inorganic carbon to the atmosphere (Arvola et al. 2002, Raymond et al., 2013, Weyhenmeyer et al., 2015), making them important in global carbon (C) cycling. Models, whether steady-state or dynamical mass balance or statistical, are necessary to estimate the importance of lakes as either sources (i.e., emit) or sinks (i.e. retain) C to global budgets. Dynamical mass balance approaches to model the internal processing of OC in lakes that incorporate all critical OC fluxes through time, however, are still relatively uncommon. Further, existing mass balances are generally based on low frequency data, confined to single lakes, and tend to be concentrated in boreal regions (e.g., Jonsson et al. 2001, Urban et al. 2005, Andersson and Sobek 2006, Cremona et al. 2014). Here we developed and applied a dynamical mass balance model to examine whether a heterogeneous set of five lakes function as net sources or sinks of OC and identify drivers of OC source or sink status.

### *Overview of concepts of key OC fluxes in lake ecosystems*

For lakes, the term “mass balance” has been broadly used to quantify budgets (not strictly OC) as the combination of inputs, internal processes, and outputs (Likens , Weathers et al. 2013). Inputs to lake ecosystem OC budgets are the sum of allochthonous (externally derived) dissolved (DOC) and particulate OC (POC) inflows from surface or groundwater sources, precipitation, and litterfall. Internal processes consist of autochthonous (in-lake) mechanisms that produce, consume, and store OC (i.e., primary production, respiration, burial in sediments). Outputs include DOC and POC that exit a lake via surface or groundwater exports. It is the balance of inputs, internal processes, and outputs that determines *lake function* as a *net source* of C to the atmosphere, or a *net sink*, whereby C is retained by the lake ecosystem (Box 1, Fig. 1).

Lakes described as carbon sources have generally been classified based on the net flux of carbon dioxide (Kortelainen et al. 2006, Tranvik et al. 2009, Raymond et al. 2013) or methane (Bastviken et al. 2011) across the air-water interface. The contribution of lakes to OC export is less frequently considered but equally important both in terms of burial and the quality and quantity of OC ultimately exported to the ocean via tributaries (Raymond and Bauer 2001, Santoso et al. 2017). Because lakes store OC in sediments, they act as important sinks in the global carbon cycle (Mulholland and Elwood 1982, Dillon and Molot 1997, Einsele et al. 2001, Einola et al. 2011). The magnitude of burial, however, has not been adequately compared to the set of other important fluxes in overall lake OC budgets (Hanson et al. 2015), precluding determination of net lake function in terms of OC. Here, we examined the dominant processes in lake OC cycling, and provided broad definitions of net sources and net sinks for lake OC to account for the interacting nature of fluxes (Box 1).

***Box 1. Mass balance conceptual equations for organic carbon (OC) in lake ecosystems***

*OCALLOCHTHONOUS:* surface and groundwater inflows + litterfall + atmospheric deposition

*OCAUTOCHTHONOUS*: gross primary production - autotrophic respiration - heterotrophic respiration

*Net Source*: Burial < Respiration

*Net Sink*: Burial > Respiration

*Full budget*: *OCALLOCHTHONOUS* + *OCAUTOCHTHONOUS* = Respiration + Burial + Export+ ΔOC storage (in water column)

A limitation of many existing mass balance studies is the omission of key fluxes that contribute to the full budget, which underscores the importance of a unified model framework (Hanson et al. 2015). We synthesized existing knowledge of lake OC budgets into a conceptual model that integrates these important mechanisms, including both in-lake as well as external (i.e., watershed) processes (Fig. 1). Below we described these processes in three main categories 1) allochthony, 2) autochthony, and 3) storage and export.

### *Allochthony*

Allochthonous inputs include all terrestrially derived OC, including OC from surface and groundwater inflows, litterfall, and direct-fall precipitation. Although surface water inflows regularly deliver OC to lake ecosystems, the role of terrestrially derived OC is perhaps the most commonly overlooked set of processes in OC budgets, largely owing to data limitations (Hanson et al. 2015). Prior studies have included direct measurements of inflow stream concentrations of OC when available (Schindler et al. 1997, Jonsson et al. 2001 , Urban et al. 2005, Klump et al. 2009), but other approaches have included use of literature-derived input estimates (Striegl et al. 1998), equations based on watershed area (Sobek et al. 2006), watershed-wetland area ratios (O’Connor et al. 2009), precipitation (Hanson et al. 2004, Staehr et al. 2010), or GIS-based estimates based on land cover and distance-weighted hydrological flow paths (Canham et al. 2004). In lakes without surface inflows, including closed-basin and seepage lakes, groundwater can be the dominant hydrological input (e.g., Gaiser et al. 2009) and can deliver OC to lakes, especially in organic-rich soils (Schindler and Krabennhoft 1998). Empirical measurements of groundwater discharge and OC concentration, however, are rare and difficult to estimate (Hanson et al. 2014). OC inputs from litterfall, and wet and dry atmospheric deposition are typically small and are generally estimated as a function of lake size and literature- or expert-based loading coefficients (Hanson et al. 2004).

### *Autochthony*

Autochthonous OC originates within lakes through photosynthesis by primary producers. Since gross primary production (GPP) is difficult to measure at the ecosystem level, net primary production (NPP), considered the difference between GPP and autotrophic respiration, is typically measured instead (Pace and Lovett 2013). Approaches previously employed to estimate NPP include using bottle incubations (Urban 2005, Yang et al. 2008), and more recently high frequency measurements of dissolved oxygen or carbon dioxide concentrations (Cole et al. 2002, Staehr et al. 2010). Statistical relationships have also been built to estimate NPP from lake temperature and total phosphorus (TP) (Hanson et al. 2004) or chlorophyll-A (chl-*a*) (Jonsson et al. 2001, Ramlal et al. 2003), or static proportions of the overall OC pool (Aberg et al. 2004).

*Storage and export*

Long-term burial of OC in lake sediments is the only mechanism by which lakes permanently remove carbon from the global carbon cycle, and is therefore a critical flux in our understanding of source or sink dynamics of both allochthonous and autochthonous OC (Cole et al. 2002, Tranvik et al. 2009). Permanent OC burial in lakes is a product of in-lake POC concentrations, POC particle sizes and associated settling rates, sediment particle size and density that affect resuspension, lake hydrodynamics that affect settling rates and resuspension, and benthic biogeochemistry (Downing et al. 2008, Xu et al. 2013). Methods for estimating sediment accumulation rates are diverse and have commonly included functions based on lake area, particularly for studies containing multiple lakes (Canham et al. 2004, Hanson et al. 2004). Previous studies have estimated burial rates using sediment cores (Yang et al. 2008, Klump et al. 2009, Heathcote & Downing 2012), sediment traps (Jonsson et al. 2001, Ramlal et al. 2003), or estimates based on changing bathymetry (Downing 2008). A challenge associated with estimating accumulation rates is the reliance on point measurements to characterize sediment accumulation rates that can vary dramatically over both space and time. Allochthonous and autochthonous POC that is not buried is leached in the form of DOC and exported via surface or groundwater (Cole et al. 1984). Exports represent allochthonous inputs to downstream aquatic ecosystems and therefore contribute to landscape carbon cycling (Kling et al. 2000).

### *Objective and research questions*

Our broad objective was to develop a flexible OC model for lakes that represented long-term dynamics and magnitudes of key OC fluxes, and to use that model both to reveal key drivers of lake function as net sources or sinks as well as uncertainties associated with these processes. Here, we described a simple dynamical mass balance model that incorporates the OC fluxes described above, parameterized for five lakes that span a gradient of morphological characteristics. We applied the model to address the following questions:

1. What are the relative magnitudes of varying fluxes that drive lake OC budgets and lake function as net sources or sinks?
2. Which mechanisms represent the greatest source of uncertainty in our understanding and modeling of carbon budgets?

## **METHODS**

### *Study lakes and data sources*

We dynamically modeled OC budgets for five lakes that span a range of environmental conditions and limnological characteristics (e.g., hydrological residence time, depth; Table 1). Lakes were selected based on the availability of observational data. Required observational data included precipitation, hydrological inflow (discharge), inflow DOC concentration and various in-lake measurements (surface temperature, chlorophyll-a (chl-*a*), and Secchi depth). All lakes had a minimum of 10 years of limnological data used for model training (Table 1) and at least four years of in-lake DOC and DO measurements for model validation. See supplemental material for detailed data descriptions and sources (S2).

*General model approach*

Given our objective to represent the relative contributions of individual fluxes to whole-system OC dynamics, an overly complex modeling approach would hinder our ability to apply the model across multiple lakes with limited observational data. We therefore specifically aimed to build a parsimonious model to maintain generalizability across lake ecosystems, using our conceptual framework as the foundation for the mass balance model (Fig. 1). We operated the model on a daily time step. Lake volume was assumed static and inflow volume was assumed equal to outflow volume. Because daily observational data of inflow DOC, in-lake chl-*a*, Secchi depth, and temperature were not always available (i.e., weekly to bi-weekly for some lakes), we gap-filled to a daily time step using linear interpolations, except for precipitation, which was assumed zero for missing data; however, precipitation data gaps were rare. Seasonal evaporative losses were assumed to be negligible and were not accounted for in the model. To account for the absence of winter data at Toolik, we set inflow DOC to 0 when the main inflow (Toolik Inlet) was frozen (S2). We summarized static and calibrated model parameters in Table 2. We developed the model using R version 3.3.2.

### *Allochthonous DOC and POC*

Allochthonous DOC load (DOCalloch) was calculated as a combined function of 1) stream load, 2) precipitation load, 3) wetland load, and 4) groundwater load. POCalloch (5) is the sum of an aerial load based on canopy cover and 10% of DOCalloch (Hanson et al. 2014).

1) Surface water inflow discharge (m3 s-1) was generally available for the main tributaries of all lakes. Inflow DOC concentration (DOC SW; g m-3) was calculated as a product of discharge volume and measured concentration (Table 3: Eq. 1a). Inflow DOC concentration was one of the least frequently measured variables for our study lakes; data were available from sub-weekly to monthly frequencies and sometimes differed in frequency across a lake’s tributaries (S2). When inflow DOC concentrations were not available for all tributaries, DOC contributions for each tributary were estimated based on the proportion of total inflow volume and the assumption that inflow DOC concentration was the same in all tributaries.

2) DOC loads from precipitation were calculated as the product of lake surface area (m2) and daily precipitation (mm) measured at the weather station nearest to each lake (Table 3: Eq. 1b-c). The concentration of DOC in precipitation was set to 2 g m-3 (Hanson et al. 2014) (Table 2: DOC\_precip\_conc).

3) Contributions of DOC from shoreline-adjacent wetlands were estimated using GIS and publicly available spatial datasets (S2). We focused on wetlands adjacent to the shoreline because they contribute the majority of wetland-derived DOC to lakes; distant wetlands contribute negligible DOC (Hanson et al. 2014). DOC contribution of wetlands was calculated by multiplying the proportion of lake shoreline covered by wetlands (Table 2: PropWetland) by lake perimeter (m), and then multiplying this value by a static parameter representing wetland DOC in g m-1 shoreline d-1 (Table 3: Eq. 1d).

4) Lake-specific groundwater inflow volume (m3) and DOC concentration (g m-3) were not available for any lake. The proportion of inflow as groundwater in our study lakes ranged from 0-19%; we used literature values when available, but assumed no groundwater inflow in the absence of data (S2). Groundwater DOC concentration was assumed to be 10 g m-3 (Table 2: DOC\_GW, Hanson et al. 2014). The contribution of groundwater to the allochthonous DOC load was therefore calculated as the product of DOC concentration and a percentage of total surface inflow (groundwater inflow rate, Table 3: Eq. 1e).

5) We calculated the contribution of all adjacent forests, including coniferous, deciduous, and mixed stands (POC canopy), to the OC budget via litterfall by multiplying the proportion of lake shoreline covered by forests (Table 2: PropCanopy) by lake perimeter (m), and multiplying this value by an aerial POC rate of 1 g m shoreline-1 day-1 (Table 3: Eq. 1f, Hanson et al. 2014).

### *Autochthonous DOC and POC: primary production*

Autochthonous OC production as GPP was modeled as a function of chl-*a* (µg L-1) and surface water temperature (°C) per Morin et al. (1999, Table 3: Eq. 2a). This empirical statistical model was based on observational temperature and chl-*a* data that ranged from 5-25 °C and 1-1000 mg m-2, respectively. Since it is difficult to predict GPP well for winter temperatures, we set GPP to zero if surface water temperatures were < 4 °C. Chl-*a* concentrations were converted from volume to areal units by multiplying by photic depth, which was estimated from Secchi depth (m; Wetzel 2001). Autotrophic respiration (Table 2: R\_autotroph) was assumed to equal 80% of total GPP; therefore, net primary production (NPP) was set equal to 20% of GPP (Quay et al. 1986, Cole et al. 2002, Hanson et al. 2004). NPP was partitioned into DOCauto and POCauto fractions using the Pace and Prairie (2005) estimate that 71.4% (as a function of chl-*a*) of NPP is respired and therefore must be converted to DOC (Table 3: Eqs. 2b-2d). The resulting rates of DOCauto and POCauto production were then applied to the whole-lake carbon balance by scaling with lake surface area (Table 3: Eqs. 2e-f).

Heterotrophic respiration was calculated as a function of DOCauto and DOCalloch concentration (g m-3) in the photic zone, epilimnion temperature (assumed to be uniform through the photic zone), and two calibrated parameters: Respiration\_autoch and Respiration\_alloch (Table 2, Table 3: Eqs. 2g-h). We determined epilimnion temperature by averaging observed temperatures throughout the photic zone when data were available, but otherwise used surface temperature (S2). Respiration\_alloch was constrained between 0.0003 and 0.03 (d-1) based on the range of OC decomposition rates for inland waters with residence times between 1-10 years presented in Catalan et al. (2016). Respiration\_autoch was constrained between 0.03 and 0.3 (d-1).

### *Burial of POC*

At each time step, a small proportion of POCauto and POCalloch was allowed to leave the lake via export, and the rest was either buried based on the Burial\_autoch and Burial\_alloch parameters (Table 2, Table 3: Eqs. 3c-h) or leached to DOCauto. Given the uncertainty associated with estimating burial, burial parameters were calibrated in the model as a proportion between 0 (no burial of POC) and 1 (all POC is buried). This simple approach allowed burial to function as the expected feedback mechanism to high POC loads from inflows and/or primary production without the need for highly uncertain gross burial and resuspension estimates.

### *Model output and calibration*

All fluxes and loads of DOC and POC were tracked at a daily time step. Net ecosystem production (NEP) was calculated as the difference between NPP and heterotrophic respiration (Table 3: Eq. 4a). Fatm (atmospheric flux rate) was calculated as k × (DOconc - DOsat)/Zmix. The mixed depth (Zmix) was set equal to half the photic depth (m) and the piston velocity (k) was set at 0.7 m d-1 (Table 3: Eq. 4b) The saturation of DO (DOsat) is temperature dependent and was determined using the Garcia-Benson method in the LakeMetabolizer R package (Winslow et al. 2016). NEP, Fatm, and DO were used to calculate the change in DO in the surface water at each time step owing to respiration (Table 3: Eq. 4c).

The four calibrated parameters in the model (Respiration\_autoch, Respiration\_alloch, Burial\_autoch and Burial\_alloch; Table 2) were fit by minimizing the sum of the squared residuals of DOC and DO modeled minus DOC and DO observed (equally weighted by number of observations). The model was fit using a pseudo-random search algorithm in the R package FME (Soetaert and Petzoldt 2010). Modeled DOC and DO were compared to corresponding observed concentrations using root mean square error (RMSE) and Nash-Sutcliffe efficiency scores for each lake to determine model goodness of fit. A sensitivity analysis of each parameter was conducted by allowing the parameter to vary at 100 different values within the set bounds while fixing the other three parameters at their calibrated values.

*Bootstrapping*

We assessed parameter uncertainty using a bootstrapping routine. Residual errors between observed and modeled DOC as well as observed and modeled DO were calculated. We created 100 pseudo-observational data sets by adding randomly selecting residuals to the original observational data. The model was then re-fit to the pseudo-observational data sets to provide 100 new parameter estimates.

## **RESULTS**

### *Model performance, parameter estimates, and sensitivity analysis*

Modeled DOC and DO generally followed observed temporal patterns across years and study lakes (Fig. 2). There was an overall strong goodness of fit for both DOC and DO based on RMSE across lakes (0.73-1.46 mg L-¹; Table 4). The model also accounted for considerably more information than long-term means of DOC based on NSE values across lakes (0.70-0.96). Although the model captured annual and seasonal DOC and DO dynamics, the model did not consistently characterize the magnitude of short-term spikes in DOC (i.e., days to weeks). Nonetheless, long-term model performance indicated the ability to account for lake variability in DOC and DO from seasonal to inter-annual time scales.

Burial\_alloch, which represented the daily proportion of POCalloch buried in sediments, was 1 (i.e., 1=100% burial, 0% leached to DOC) for all lakes (Table 2). Similarly, Burial\_autoch was near 1 for Monona, Vanern and Toolik; however, values were 0 for Harp and Trout, indicating nearly all autochthonous POC was leached to DOC in these two lakes and subsequently mineralized or exported. DOC respiration parameters were lower for DOCalloch than DOCauto across all lakes, indicating that DOCauto was more readily respired than DOCalloch. Respiration\_alloch was relatively similar across lakes, ranging 0.001-0.003, whereas Respiration\_autoch was more variable, ranging 0.015 (Trout) to 0.297 (Toolik).

Modeled DOC (mg L-1) was generally most sensitive to Respiration\_alloch, except for Monona, for which modeled DOC was most sensitive to Burial\_autoch (Fig. 3). The other 4 lakes were minimally affected by changes in this parameter (< 1 mg L-1 difference across the range of parameter values). Changes in Burial\_alloch had consistently minimal effects on the range of modeled DOC across lakes. Harp and Monona were the only lakes with considerable sensitivity to Respiration\_autoch. Overall, parameter sensitivity was greatest for Vanern, Harp and Monona, for which modeled DOC varied as much as 5-6 mg L-1 across the range of parameter values. Conversely, modeled DOC varied no more than 2 and 3 mg L-1 for Toolik and Trout, respectively.

[Yep, we need a paragraph about parameter uncertainty, which will come from the bootstrap analysis. My expectation is that the burial factors for POC will not be well-constrained, and this will be an important take-home point from the manuscript.]

### *Summary of fluxes and fates*

OC inputs to Harp averaged 71.9 g m-2 yr-1 (based on complete years only), predominantly driven by allochthony (55.5%) rather than autochthony (44.5%) (Table 5). Harp processed 75.5% of its load (24.5% exported via surface water) through more respiration (47.5 g m-2 yr-1) than burial (5.3 g m-2 yr-1). Monona had the largest OC load of the five lakes (119.0 g m-2 yr-1) and was also driven primarily by allochthony (54.5%) rather than autochthony (45.6%). Monona processed 49.6% of its load (50.4% exported) through greater burial (43.8 g m-2 yr-1) than respiration (1.68 g m-2 yr-1). Trout had the smallest OC load of the five lakes (41.5 g m-2 yr-1) and was the only lake drive primarily by autochthony (67.8%) rather than allochthony (32.2%). Trout processed 90.5% of its load, which was the greatest among all lakes (9.5% exported), through greater respiration (37.5 g m-2 yr-1) than burial (1.2 g m-2 yr-1). Vanern had the second lowest OC load (58.3 g m-2 yr-1) and was driven predominantly by allochthony (55.3%) rather than autochthony (44.7%), which were similar ratios to Harp and Monona. Vanern processed 79.2% of its load (20.8% exported) through greater respiration (29.2 g m-2 yr-1) than burial (19.9 g m-2 yr-1). Toolik had the intermediate OC load (76 g m-2 yr-1) and was driven mostly by allochthony (87.1%) rather than autochthony (12.9%), which was a larger ratio than the other lakes. Toolik processed 36.4% of its load, which was the lowest among all lakes (63.6% exported), through greater respiration (23.7 g m-2 yr-1) than burial (6.7 g m-2 yr-1). Overall, with the exception of Trout, OC loads were primarily driven by allochthony, underscoring the importance of terrestrially derived OC in overall lake budgets (Table 5).

Seasonal patterns in OC fluxes were consistent across the entire time series, with autochthony and respiration increasing to a summer maximum (Fig. 4) Respiration consistently exceeded burial in all lakes but Monona across all modeled years, indicating that these lakes were net sources of OC (Fig. 4, Table 5); on average, respiration accounted for 14.1-90.2% of total OC loads, whereas burial accounted for 7.4-36.8% of total OC loads. Therefore, the net source capacity of the four source lakes varied approximately four-fold. Trout and Harp were the largest net sources in terms of differences between respiration and burial. Monona was the only long-term net sink of OC; on average, burial rates were greater than twice respiration rates across modeled years (Fig. 4, Table 5).

### *Seasonal fates*

As water temperatures increased during the growing season (e.g., May – Aug.), the balance between allochthony and autochthony generally shifted in favor of autochthony due to increases in NPP, whereas the ratio between respiration and burial generally shifted towards respiration and therefore source function (Fig. 5). There was seasonal variability in the dominant fluxes acting in each lake. Trout remained a source for the entire year across all sampled years, but became a greater source as the growing season progressed. Harp, Toolik and Vanern were sinks early in the growing season, but became net sources as temperatures warmed. Conversely, Monona remained a sink throughout most of the year and only became a source late in the growing season. Despite this late pulse of respiration, Monona remained a net sink on an annual basis (Table 5). Monona and Vanern also showed less of an increase in autochthony as the growing season progressed compared to other lakes, suggesting the importance of continued allochthonous inputs during summer months coinciding with increases in autochthony.

## **DISCUSSION**

### *Capturing lake processes*

Our results indicate that a relatively simple, dynamical model can represent the set of key biogeochemical, trophic, and landscape processes that combine to determine the fate of OC in lake ecosystems. Our model performed well for both mesotrophic and oligotrophic lakes, in temperate and subarctic climate zones, forested and agricultural watersheds, and orders of magnitude differences in lake morphometry, which demonstrated the generalizable nature of our approach. A key inference is that a few processes are responsible for control over the OC dynamics. While postulated previously (Hanson et al. 2011), this is the first demonstration in a dynamical model applied to contrasting lake types. Further, the fluxes we modeled were within the range of other published studies for these lakes based on steady-state models. Using much of the same LTER data, Hanson et al. (2014) provided similar estimates for Trout of allochthony (Hanson estimate: 15.92 g m-2 yr-1, our model estimate: 13.35 g m-2 yr-1), burial and export, but not for respiration, because that study did not account for autochthony. Whalen and Cornwall (1985) modeled Toolik based on a different set of years from our study (1980-1981), but similarly demonstrated that the system contained high allochthony relative to autochthony, low burial and high export. Similar to Hanson et al. (2014), Dillon and Molot’s (1997) estimates for Harp of autochthony, burial (Dillon and Molot: 6.3 g m-2 yr-1, our model estimate: 5.3 g m-2 yr-1) and export (Dillon and Molot: 16.9 g m-2 yr-1, our model estimate: 17.6 g m-2 yr-1), but not respiration (based on field data from 1981-1989), were comparable to our results. Although our results generally agreed with prior studies based on steady-state models, this was not true for autochthony and respiration. We offer that dynamical models better represent these processes by accounting for seasonal changes in temperature and chl-*a* concentrations. Therefore, although steady-state models may be sufficient for recreating some key ecological processes, dynamical models are needed for determining the net source or sink function of lakes, given the importance of autochthony and respiration.

### *Under what conditions are lakes net sources or sinks of organic carbon?*

Although prior studies have identified lakes as OC sinks owing to burial in lake sediments, our study showed that burial can be a relatively small component of overall lake OC budgets. Whereas this may have been assumed previously in empirical studies (e.g., Cole et al. 2007), we demonstrated how ratios between respiration and burial can be constrained by mass balance and the inclusion of both allochthonous and autochthonous sources. Many lakes may be net OC sources because the ratio between burial and respiration is smaller than 1. A key consideration is that our model buried 100% of POCalloch (Table 2: Burial\_alloch); therefore, any increase in POCalloch would be directly proportional to increases in burial. Owing to lack of observational data, we additionally assumed POCalloch was 10% of DOCalloch. Intense precipitation can increase POC concentration disproportionately to DOC concentration in streams (Jeong et al. 2012, Dhillon and Inamdar 2013), which could temporarily increase POCalloch and thus burial. In addition, our model demonstrated DOC responses to precipitation events when we had corresponding weather and DOCalloch data, but the assumed linear changes in inflow volume and DOCalloch between observation points may also have underrepresented DOCalloch, leading indirectly to underrepresentation of POCalloch. Wet years increase DOCalloch inputs to lakes at regional scales (Rose et al. 2016) and therefore probably also increase POCalloch. Although our burial estimates were somewhat uncertain due to underrepresentation of POCalloch during precipitation events, burial would have to increase substantially over the course of the entire modeling period to switch lake function to sink rather than source, including three-fold or greater for Harp, Toolik and Trout (Table 5). Therefore, missing POCalloch may constitute noise in the overall OC budget, but is unlikely to account for enough OC to alter overall lake function due to the magnitudes of respiration.

Across all lakes in our study, warm surface temperatures were generally associated with autochthony in productive lakes (Monona, Vanern) and allochthony when the OC pool was dominated by allochthonous inputs (Harp, Trout, Toolik; Fig. 5). In lakes with primarily allochthonous carbon pools, the ratio of respiration to burial decreased as allochthony decreased. More productive systems had the opposite response, however, and revealed an increase in the ratio of respiration to burial as allochthonous inputs decreased. Across lakes, warm surface temperatures appear to drive lakes toward OC export rather than burial, though the mechanisms driving this trend likely differs between systems dominated by allochthony versus autochthony. The implications of this are that climate warming and associated increases in lake surface temperatures will not have consistent effects across all lakes, but instead may favor net heterotrophy and export of terrestrial OC from some systems, and autochthony and export of labile OC from others.

Our long-term and seasonal analyses have important implications for temporal scaling of lake function. Many previous mass balance studies were conducted over a single open-water season and therefore cannot represent longer term trends. As a post hoc analysis, we found overall consistency in lake function across all modeled years (using only complete years); however, differences between respiration and burial (lake function) varied two- or three-fold across years in some lakes (Fig. 6). Over multi-year periods, weather may influence lake function, given that differences in precipitation across years drive variation in DOCalloch and POCalloch (Jeong et al. 2012) and that warmer years increase autochthony. As such, single-year studies may not be representative of long-term conditions.

### *On-going research needs and future implications*

Our work is an important advance in terms of understanding the fates of OC across aquatic ecosystems and landscapes; however, we did encounter constraints associated with current data availability. If necessary data were collected for a larger number of lakes spanning wider environmental gradients (e.g., climate, watershed conditions), contributions of lakes to landscape carbon cycles based on lake function could be estimated at broad spatial scales. Particularly necessary are high-frequency measurements of inflow DOC concentration. These data were among the least frequently collected among lakes we considered for this study, yet without them, examining whole-lake OC budgets is prohibitively difficult. Our study generally used data collected at weekly or bi-weekly intervals; although data collection may be expensive and logistically challenging, the increasing availability of automated, high-frequency sensor equipment may alleviate long-term costs associated with sensor deployment and manual data retrieval (e.g., Porter et al. 2009). In addition, relatively little is currently known about POC budgets despite their key interactions with DOC (Einsele et al. 2001); we need more POC observational data for incorporation into dynamical models of OC, particularly in inflows for estimating POCalloch. Such studies would help constrain POC parameters and improve estimates of the fates of POC within overall OC budgets.

Thus, a key implication of our study is the need for more observational data, particularly pertaining to surface water DOC concentrations, POC cycling, and burial rates. Lake resource managers should be aware of the importance of allochthonous inputs derived from watershed sources in lake OC budgets; allochthony exceeded autochthony in 4 of the 5 lakes in this study. Lakes have become increasingly productive under recent climate warming (Kraemer et al. 2016); this increases autochthony as well as the OC source capacity of lakes. As climates continue to warm, we would therefore expect lakes to become larger sources of OC over time.

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## **LITERATURE CITED**

Åberg, J., Bergström, A. K., Algesten, G., Söderback, K., & Jansson, M. (2004). A comparison of the carbon balances of a natural lake (L. Örträsket) and a hydroelectric reservoir (L. Skinnmuddselet) in northern Sweden. *Water Research*, *38*(3), 531-538.

Algesten, G., Sobek, S., Bergström, A. K., Ågren, A., Tranvik, L. J., & Jansson, M. (2004). Role of lakes for organic carbon cycling in the boreal zone. *Global change biology*, *10*(1), 141-147.

Alin, S.R., and T.C. Johnson (2007), Carbon cycling in large lakes of the world: A synthesis of production, burial, and lake-atmosphere exchange estimates, *Global Biogeochem. Cycles, 21,* GB3002, doi:10.1029/2006GB002881.

Andersson, E., & Sobek, S. (2006). Comparison of a mass balance and an ecosystem model approach when evaluating the carbon cycling in a lake ecosystem. *AMBIO: A Journal of the Human Environment*, *35*(8), 476-483.

Arvola, L., Kortelainen, P. I. R. K. K. O., Bergström, I., Kankaala, P., Ojala, A., Pajunen, H. A. N. N. U., ... & Rantakari, M. I. I. T. T. A. (2002). Carbon pathways through boreal lakes: A multi-scale approach (CARBO). *Understanding the Global System, The Finnish Perspective, edited by Käyhkö J and Talve L*, 97-106.

Canham, C. D., Pace, M. L., Papaik, M. J., Primack, A. G., Roy, K. M., Maranger, R. J., ... & Spada, D. M. (2004). A spatially explicit watershed-scale analysis of dissolved organic carbon in Adirondack lakes. *Ecological Applications*, *14*(3), 839-854.

Cole, J. J., McDowell, W. H., & Likens, G. E. (1984). Sources and molecular weight of" dissolved" organic carbon in an oligotrophic lake. *Oikos*, 1-9.

Cole, J. J., Carpenter, S. R., Kitchell, J. F., & Pace, M. L. (2002). Pathways of organic carbon utilization in small lakes: Results from a whole‐lake 13C addition and coupled model. *Limnology and Oceanography*, *47*(6), 1664-1675.

Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., ... & Melack, J. (2007). Plumbing the global carbon cycle: integrating inland waters into the terrestrial carbon budget. *Ecosystems*, *10*(1), 172-185.

Cremona, F., Kõiv, T., Nõges, P., Pall, P., Rõõm, E. I., Feldmann, T., ... & Nõges, T. (2014). Dynamic carbon budget of a large shallow lake assessed by a mass balance approach. *Hydrobiologia*, *731*(1), 109-123.

Dhillon, G. S., & Inamdar, S. (2014). Storm event patterns of particulate organic carbon (POC) for large storms and differences with dissolved organic carbon (DOC). *Biogeochemistry*, *118*(1-3), 61-81.

Dillon, P. J., & Molot, L. A. (1997). Dissolved organic and inorganic carbon mass balances in central Ontario lakes. *Biogeochemistry*, *36*(1), 29-42.

Downing, J. A., Cole, J. J., Middelburg, J. J., Striegl, R. G., Duarte, C. M., Kortelainen, P., ... & Laube, K. A. (2008). Sediment organic carbon burial in agriculturally eutrophic impoundments over the last century. *Global Biogeochemical Cycles*, *22*(1).

Einola, E., Rantakari, M., Kankaala, P., Kortelainen, P., Ojala, A., Pajunen, H., ... & Arvola, L.

(2011). Carbon pools and fluxes in a chain of five boreal lakes: a dry and wet year comparison. *Journal of Geophysical Research: Biogeosciences*, *116*(G3).

Einsele, G., Yan, J., & Hinderer, M. (2001). Atmospheric carbon burial in modern lake basins and its significance for the global carbon budget. *Global and Planetary Change*, *30*(3), 167-195.

Gaiser, E. E., Deyrup, N. D., Bachmann, R. W., Battoe, L. D., & Swain, H. M. (2009). Multidecadal climate oscillations detected in a transparency record from a subtropical Florida lake.

Hanson, P. C., Buffam, I., Rusak, J. A., Stanley, E. H., & Watras, C. (2014). Quantifying lake allochthonous organic carbon budgets using a simple equilibrium model. *Limnol. Oceanogr*, *59*(1), 167-181.

Hanson, P. C., Hamilton, D. P., Stanley, E. H., Preston, N., Langman, O. C., & Kara, E. L. (2011). Fate of allochthonous dissolved organic carbon in lakes: a quantitative approach. *PLoS One*, *6*(7), e21884.

Hanson, P. C., Pace, M. L., Carpenter, S. R., Cole, J. J., & Stanley, E. H. (2015). Integrating landscape carbon cycling: research needs for resolving organic carbon budgets of lakes. *Ecosystems*, *18*(3), 363-375.

Hanson, P. C., Pollard, A. I., Bade, D. L., Predick, K., Carpenter, S. R., & Foley, J. A. (2004). A model of carbon evasion and sedimentation in temperate lakes. *Global Change Biology*, *10*(8), 1285-1298.

Heathcote, A. J., & Downing, J. A. (2012). Impacts of eutrophication on carbon burial in

freshwater lakes in an intensively agricultural landscape. *Ecosystems*, *15*(1), 60-70.

Jeong, J. J., Bartsch, S., Fleckenstein, J. H., Matzner, E., Tenhunen, J. D., Lee, S. D., ... & Park, J. H. (2012). Differential storm responses of dissolved and particulate organic carbon in a mountainous headwater stream, investigated by high‐frequency, in situ optical measurements. *Journal of Geophysical Research: Biogeosciences*, *117*(G3).

Jonsson, A., Meili, M., Bergström, A. K., & Jansson, M. (2001). Whole‐lake mineralization of allochthonous and autochthonous organic carbon in a large humic lake (Örträsket, N. Sweden). *Limnology and Oceanography*, *46*(7), 1691-1700.

Karlsson, J., Ask, J., & Jansson, M. (2008). Winter respiration of allochthonous and autochthonous organic carbon in a subarctic clear‐water lake. *Limnology and oceanography*, *53*(3), 948-954.

Kling, G. W., Kipphut, G. W., Miller, M. M., & O'Brien, W. J. (2000). Integration of lakes and streams in a landscape perspective: the importance of material processing on spatial patterns and temporal coherence. *Freshwater Biology*, *43*(3), 477-497.

Klump, J. V., Fitzgerald, S. A., & Waplesa, J. T. (2009). Benthic biogeochemical cycling, nutrient stoichiometry, and carbon and nitrogen mass balances in a eutrophic freshwater bay. *Limnology and Oceanography*, *54*(3), 692-712.

Kraemer, B. M., Chandra, S., Dell, A. I., Dix, M., Kuusisto, E., Livingstone, D. M., ... &

McIntyre, P. B. (2016). Global patterns in lake ecosystem responses to warming based on the temperature dependence of metabolism. *Global change biology*.

Lehner, B., & Döll, P. (2004). Development and validation of a global database of lakes, reservoirs and wetlands. *Journal of Hydrology* 296, 1-22.

Mulholland, P. J., & Elwood, J. W. (1982). The role of lake and reservoir sediments as sinks in

the perturbed global carbon cycle. *Tellus*, *34*(5), 490-499.

O'Connor, E. M., Dillon, P. J., Molot, L. A., & Creed, I. F. (2009). Modeling dissolved organic carbon mass balances for lakes of the Muskoka River Watershed. *Hydrology Research*, *40*(2-3), 273-290.

Pace ML & Lovett G. 2013. Primary production: the foundation of ecosystems. In: Weathers K,

Strayer D, Likens G, editors. Fundamentals of ecosystem science. Academic Press. p. 312.

Paerl, H. W., Hall, N. S., & Calandrino, E. S. (2011). Controlling harmful cyanobacterial blooms

in a world experiencing anthropogenic and climatic-induced change. *Science of the Total Environment*, *409*(10), 1739-1745.

Porter, J. H., Nagy, E., Kratz, T. K., Hanson, P., Collins, S. L., & Arzberger, P. (2009). New eyes on the world: advanced sensors for ecology. *BioScience*, *59*(5), 385-397.

Quay, P. D., Emerson, S. R., Quay, B. M., & Devol, A. H. (1986). The carbon cycle for Lake Washington-- a stable isotope study. *Limnology and Oceanography*, *31*(3), 596-611.

Ramlal, P. S., Hecky, R. E., Bootsma, H. A., Schiff, S. L., & Kingdon, M. J. (2003). Sources and fluxes of organic carbon in Lake Malawi/Nyasa. *Journal of Great Lakes Research*, *29*, 107-120.

Raymond, P. A., & Bauer, J. E. (2001). Riverine export of aged terrestrial organic matter to the

North Atlantic Ocean. *Nature*, *409*(6819), 497-500.

Raymond, P. A., Hartmann, J., Lauerwald, R., Sobek, S., McDonald, C., Hoover, M., ... &

Kortelainen, P. (2013). Global carbon dioxide emissions from inland waters. *Nature*, *503*(7476), 355-359.

Santoso, A. B., Hamilton, D. P., Hendy, C. H., & Schipper, L. A. Carbon dioxide emissions and

sediment organic carbon burials across a gradient of trophic state in eleven New Zealand lakes. *Hydrobiologia*, 1-14.

Schindler, D. W., Curtis, P. J., Bayley, S. E., Parker, B. R., Beaty, K. G., & Stainton, M. P. (1997). Climate-induced changes in the dissolved organic carbon budgets of boreal lakes. *Biogeochemistry*, *36*(1), 9-28.

Schindler, J. E., & Krabbenhoft, D. P. (1998). The hyporheic zone as a source of dissolved organic carbon and carbon gases to a temperate forested stream. *Biogeochemistry*, *43*(2), 157-174.

Sobek, S., Söderbäck, B., Karlsson, S., Andersson, E., & Brunberg, A. K. (2006). A carbon budget of a small humic lake: an example of the importance of lakes for organic matter cycling in boreal catchments. *AMBIO: A Journal of the Human Environment*, *35*(8), 469-475.

Soetaert, Karline and Petzoldt, Thomas, 2010. Inverse Modelling, Sensitivity and Monte Carlo Analysis in R Using Package FME. Journal of Statistical Software, 33(3), 1-28. DOI 10.18637/jss.v033.i03 URL http://www.jstatsoft.org/v33/i03/.

Staehr, P. A., Sand-Jensen, K., Raun, A. L., Nilsson, B., & Kidmose, J. (2010). Drivers of metabolism and net heterotrophy in contrasting lakes. *Limnology and Oceanography*, *55*(2), 817.

Striegl, R. G., & Michmerhuizen, C. M. (1998). Hydrologic influence on methane and carbon dioxide dynamics at two north-central Minnesota lakes. *Limnology and Oceanography*, *43*(7), 1519-1529.

Tanentzap, A. J., Szkokan-Emilson, E. J., Desjardins, C. M., Orland, C., Yakimovich, K.,

Dirszowsky, R., ... & Gunn, J. (2017). Bridging between litterbags and whole-ecosystem experiments: a new approach for studying lake sediments. *Journal of Limnology*.

Tranvik, L. J., Downing, J. A., Cotner, J. B., Loiselle, S. A., Striegl, R. G., Ballatore, T. J., ... & Kortelainen, P. L. (2009). Lakes and reservoirs as regulators of carbon cycling and climate. *Limnology and Oceanography*, *54*(6part2), 2298-2314.

Urban, N. R., Auer, M. T., Green, S. A., Lu, X., Apul, D. S., Powell, K. D., & Bub, L. (2005). Carbon cycling in Lake Superior. *Journal of Geophysical Research: Oceans (1978–2012)*, *110*(C6).

Verpoorter, C., Kutser, T., Seekell, D. A., & Tranvik, L.J. (2014). A global inventory of lakes based on high-resolution satellite imagery. *Geophysical Research Letters* 41(18), 6396-6402.

Wetzel, R. G. (2001). *Limnology: lake and river ecosystems*. Gulf Professional Publishing.

Weyhenmeyer, G. A., Kosten, S., Wallin, M. B., Tranvik, L. J., Jeppesen, E., & Roland, F. (2015). Significant fraction of CO2 emissions from boreal lakes derived from hydrologic inorganic carbon inputs. *Nature Geoscience*.

Whalen, S. C., & Cornwell, J. C. (1985). Nitrogen, phosphorus, and organic carbon cycling in an arctic lake. *Canadian Journal of Fisheries and Aquatic Sciences*, *42*(4), 797-808.

Winslow, L. A., Zwart, J. A., Batt, R. D., Dugan, H., Woolway, R. I., Corman, J., ... & Read, J. S. (2016). LakeMetabolizer: an R package for estimating lake metabolism from free-water oxygen using diverse statistical models. *Inland Waters*, *6*(4), 622-636.

Xenopoulos, M. A., Lodge, D. M., Frentress, J., Kreps, T. A., Bridgham, S. D., Grossman, E., &

Jackson, C. J. (2003). Regional comparisons of watershed determinants of dissolved organic carbon in temperate lakes from the Upper Great Lakes region and selected regions globally. *Limnology and Oceanography*, *48*(6), 2321-2334.

Xu, H., Lan, J., Liu, B., Sheng, E., & Yeager, K. M. (2013). Modern carbon burial in Lake Qinghai, China. *Applied geochemistry*, *39*, 150-155.

Yang, H., Xing, Y., Xie, P., Ni, L., & Rong, K. (2008). Carbon source/sink function of a subtropical, eutrophic lake determined from an overall mass balance and a gas exchange and carbon burial balance. *Environmental Pollution*, *151*(3), 559-568.

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| **TABLES**  **Table 1. Lake characteristics** | | | |  | |  | |  | |  | |  | |  | | |  | |  |  | |  |  |  |
| **Lake** | **Location** | **N lat,**  **W lon** | **Data years** | | **zmean (m)** | | **RT (yr)** | | **Trophic status** | | **Secchi (m)** | | **Chl-*a* (µg L-1)** | | **SW DOC (g m-3)** | **Lake DOC (g m-³)** | | **References** | | |
| Harp | Ontario, Canada | 45°38', 79°14' | 1991-2001 | | 12 | | 2.5 | | oligotrophic | | 4.47 | | 2.07 | | 9.35 | 4 | | Yao et al. 2011 | | |
| Monona | Wisconsin, USA | 43°11', 89°42' | 2003-2014 | | 8.3 | | 0.8 | | eutrophic | | 3.03 | | 9.21 | | 5.09 | 6 | | NTL LTER | | |
| Toolik | Alaska, USA | 68°63', 149°61' | 2001-2010 | | 7 | | 0.8 | | oligotrophic | | 4.53 | | 1.41 | | 7.72 | 5 | | Kling et al. 2000 | | |
| Trout | Wisconsin, USA | 46°02', 89°40' | 2004-2013 | | 14.6 | | 5.9 | | oligotrophic | | 5.32 | | 2.23 | | 5.11 | 3 | | Webster et al. 1996, NTL LTER | | |
| Vanern | Sweden | 59°06’,  -13°62’ | 2001-2013 | | 27 | | 6.3 | | oligotrophic | | 4.47 | | 2.07 | | 9.35 | 4 | | Kvarnäs 2001 | | |

zmean = mean depth, RT = hydrologic residence time, Secchi = Secchi depth, Chl-*a* = chlorophyll-a, SW DOC = inflow dissolved organic carbon, Lake DOC = in-lake DOC (mean water column). All values calculated from model calibration data or derived from cited references. NTL LTER = [https://lter.limnology.wisc.edu/](https://lter.limnology.wisc.edu/datacatalog/search)

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| **Table 2. Lake model parameters (calibrated parameters italicized, n= 4)** | | |  |  |  |  |  |
| **Parameter** | **Description** | **Harp** | | **Monona** | **Toolik** | **Trout** | **Vanern** |
| **General** |  |  | |  |  |  |  |
| Perimeter (m) | Lake perimeter | 4000 | | 35200 | 8104 | 25900 | 2007000 |
| Mean Depth (m) | Mean water depth | 12 | | 8.3 | 7 | 14.6 | 27 |
| Area (m²) | Lake surface area | 7.14E+05 | | 13.3E+07 | 1.49E+06 | 1.61E+07 | 5.65E+09 |
| Volume (m³) | Lake volume | 8.32E+06 | | 1.10E+08 | 1.06E+07 | 2.35E+08 | 1.53E+11 |
| DOC\_init (g m-3) | Initial in-lake DOC concentration | 3.70 | | 7.00 | 5.00 | 2.90 | 4.40 |
| POC\_init (g m-3) | Initial in-lake POC concentration | 0.37 | | 0.10 | 0.50 | 0.29 | 0.40 |
| **Allochthony** |  |  | |  |  |  |  |
| PropCanopy | Proportion of shoreline with forest | 1.000 | | 0.167 | 0.000 | 0.780 | 0.615 |
| PropWetlands | Proportion of shoreline with wetlands | 0.000 | | 0.026 | 0.133 | 0.011 | 0.037 |
| WetlandLoad (g d-1) | Loading rate of POC from wetlands | 1 | | 1 | 1 | 1 | 1 |
| DOC\_gw (g m-3) | DOC concentration of groundwater | 10 | | 10 | 10 | 10 | 10 |
| PropGW | Proportion of lake inflow as groundwater | 0.00 | | 0.00 | 0.00 | 0.19 | 0.00 |
| DOC\_precip\_conc (g m-3) | DOC concentration of precipitation | 2 | | 2 | 2 | 2 | 2 |
| AerialLoad (g d-1) | Influx of aerial POC (i.e., leaflitter) | 1 | | 1 | 1 | 1 | 1 |
| **Autochthony** |  |  | |  |  |  |  |
| *Respiration\_alloch (d*-*1)* | *Decomposition rate of allochthonous DOC in heterotrophic respiration* | *0.002* | | *0.001* | *0.003* | *0.001* | *0.001* |
| *Respiration\_autoch (d-1)* | *Decomposition rate of autochthonous DOC in heterotrophic respiration* | *0.092* | | *0.003* | *0.297* | *0.015* | *0.069* |
| R\_autotroph | Proportion of GPP autotrophically respired | 0.8 | | 0.8 | 0.8 | 0.8 | 0.8 |
| **Burial** |  |  | |  |  |  |  |
| *Burial alloch* | *Proportion of allochthonous POC buried in sediments* | *1.000* | | *1.000* | *1.000* | *1.000* | *1.000* |
| *Burial\_autoch* | *Proportion of autochthonous POC buried in sediments* | *0.000* | | *1.000* | *0.968* | *0.000* | *1.000* |
| Observed\_MAR (g m² yr-1) | Observed sediment mass accumulation rate | 78 | | 249 | 153 | 27 | 186 |
| POClc\_alloch | Proportion of allochthonous POC leached to DOC (1-Burial\_alloch) | 0.000 | | 0.000 | 0.000 | 0.000 | 0.000 |
| POClc\_autoch | Proportion of autochthonous POC leached to DOC (1-Burial\_autoch) | 1.000 | | 0.000 | 0.032 | 1.000 | 0.000 |

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| --- | --- | --- | --- | --- |
| **Table 3. Built model with equations** | | |  |  |
| **Allochthonous DOC and POC** | **Units** | **Reference** | | |
| 1a) DOC SW = surface water concentration \* surface water inflow rate \* 86400 | g d-1 | Hanson et al. 2014 | | |
| 1b) Daily precipitation = rainfall \* 0.001 \* Area | m3 d-1 | Hanson et al. 2014 | | |
| 1c) DOC precipitation = DOC\_precip \* Daily precipitation | g d-1 | Hanson et al. 2014 | | |
| 1d) DOC Wetland = PropWetland \* WetlandLoad \* Perimeter | g d-1 | Hanson et al. 2014 | | |
| 1e) DOC GW = groundwater concentration \* groundwater inflow rate \* 86400 | g d-1 | Hanson et al. 2014 | | |
| 1f) POC canopy = PropCanopy \* AerialLoad \* Perimeter | g d-1 | Hanson et al. 2014 | | |
| 1g) Inflow load DOC = DOC Wetland + DOC GW + DOC SW + DOC precipitation | g d-1 | Hanson et al. 2014 | | |
| 1h) Internal load POC = (DOC Wetland + DOC SW) \* 0.1 | g d-1 | Hanson et al. 2014 | | |
| 1i) Inflow load POC = POC Aerial + Internal load POC | g d-1 | Hanson et al. 2014 | | |
| 1j) POC Aerial = AerialLoad \* Perimeter | g m-1 d-1 | Hanson et al. 2014 | | |
| **Autochthonous DOC and POC: primary production** |  |  | | |
| 2a) GPP rate = 10^(1.18 + (0.92 \* log10(chl-*a*\* photic depth)) + (0.014 \* epilimnion temperature)) | mg m-2 d-1 | Morin et al. 1999 | | |
| 2b) GPP percent DOC = 71.4 \* (chl-*a* \* photic depth)^(-0.22) | % | Pace and Prairie 2005 | | |
| 2c) GPP DOC rate = GPP rate \* (GPP percent DOC/100) | mg m-2 d-1 |  | | |
| 2d) GPP POC rate = GPP rate \* (1-(GPP percent DOC/100)) | mg m-2 d-1 |  | | |
| 2e) NPP DOC\_autoch = GPP DOC\_autoch \* 0.2 \* Area / 1000 | g d-1 |  | | |
| 2f) NPP POC\_autoch = GPP POC\_autoch \* 0.2 \* Area / 1000 | g d-1 |  | | |
| 2g) Autochthonous Respiration = GPP DOC rate \* Respiration\_autoch(1.08^(epilimnion temp - 20)) | g m-3 |  | | |
| 2h) Allochthonous Respiration = GPP DOC rate \* Respiration\_alloch(1.08^(epilimnion temp - 20)) | g m-3 |  | | |
| **Burial** |  |  | | |
| 3a) MAR\_alloch = POC mass \* Burial\_alloch \* 365/Area | g m-2 yr-1 |  | | |
| 3b) MAR\_autoch = POC mass \* Burial\_autoch \* 365/Area | g m-2 yr-1 |  | | |
| 3c) POC Burial\_alloch = MAR\_alloch \* (1/365) \* Area | g d-1 |  | | |
| 3d) POC Burial\_autoch = MAR\_autoch \* (1/365) \* Area | g d-1 |  | | |
| 3e) POC\_alloch leached out = POC\_alloch concentration \* POClc\_alloch \* Volume | g d-1 |  | | |
| 3f) POC\_autoch leached out = POC\_autoch concentration \* POClc\_autoch \* Volume | g d-1 |  | | |
| 3g) DOC\_alloch leached in = POC\_alloch leached out | g d-1 |  | | |
| 3h) DOC\_autoch leached in = POC\_autoch leached out | g d-1 |  | | |
| **NEP and Oxygen Flux** |  |  | | |
| 4a) NEP (as O2) = (NPP - DOCrespired) \* 32/12 | g m-3 d-1 |  | | |
| 4b) Fatm = 0.7 \* (DOconc - DOsat)/Zmix | g m-3 d-1 |  | | |
| 4c) DO(t+1) = DOconc + NEP - Fatm | g m-3 d-1 |  | | |

MAR = mass accumulation rate, Fatm = atmospheric flux rate, Zmix = mixed depth (m)

**Table 4. Model goodness of fit**

|  |  |  |
| --- | --- | --- |
| **Lake** | **RMSE\*** | **NSE\*\*** |
| Harp | 1.22 | 0.86 |
| Monona | 1.46 | 0.70 |
| Toolik | 1.19 | 0.79 |
| Trout | 0.80 | 0.95 |
| Vanern | 0.73 | 0.96 |
| \* Root mean square error (mg L-1) | | | |
| \*\* Nash-Sutcliffe efficiency | | | |

Table 5. Summary of annual mass balances (g m-2 y-1)

|  |  |  |  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- | --- | --- | --- |
| **Lake** | **Alloch** | **Autoch** | **Total Load** | **Processed** | **Resp** | **Burial** | **Net** | **Export** |
| **Annual means** | |  |  |  |  |  |  |  |
| Harp | 39.88 | 32.03 | 71.91 | 54.31 | -47.46 | -5.30 | 42.16 | -17.60 |
| Monona | 64.74 | 54.23 | 118.97 | 59.02 | -16.76 | -43.81 | -27.05 | -59.95 |
| Toolik | 76.19 | 11.27 | 87.45 | 31.86 | -23.68 | -6.67 | 17.01 | -55.59 |
| Trout | 13.35 | 28.17 | 41.52 | 37.56 | -37.45 | -1.19 | 36.25 | -3.95 |
| Vanern | 32.25 | 26.08 | 58.32 | 46.20 | -29.20 | -19.85 | 9.35 | -12.12 |
| **SD of annual means** | | |  |  |  |  |  |  |
| Harp | 10.71 | 7.99 | 11.42 | 9.89 | 9.42 | 0.95 | 9.66 | 4.86 |
| Monona | 24.16 | 18.05 | 21.81 | 15.38 | 1.99 | 12.85 | 12.26 | 20.58 |
| Toolik | 64.78 | 3.36 | 62.91 | 11.08 | 3.52 | 5.55 | 8.14 | 53.53 |
| Trout | 2.68 | 4.59 | 4.41 | 3.88 | 5.98 | 0.18 | 6.04 | 0.88 |
| Vanern | 5.81 | 4.91 | 5.86 | 4.97 | 2.35 | 3.14 | 1.83 | 1.84 |
| **Proportion of total load** | |  |  |  |  |  |  |  |
| Harp | 0.55 | 0.45 | 1.00 | 0.76 | -0.66 | -0.07 | 0.59 | -0.24 |
| Monona | 0.54 | 0.46 | 1.00 | 0.50 | -0.14 | -0.37 | -0.23 | -0.50 |
| Toolik | 0.87 | 0.13 | 1.00 | 0.36 | -0.27 | -0.08 | 0.19 | -0.64 |
| Trout | 0.32 | 0.68 | 1.00 | 0.90 | -0.90 | -0.03 | 0.87 | -0.10 |
| Vanern | 0.55 | 0.45 | 1.00 | 0.79 | -0.50 | -0.34 | 0.16 | -0.21 |

Positive fluxes represent OC inputs to lakes, whereas negative fluxes represent OC removal. Processed OC is the amount of the total load not exported. Positive net values represent OC sources. Proportions of allochthony and autochthony sum to 1. Proportions of respiration, burial, and export also sum to 1. SD = standard deviation.

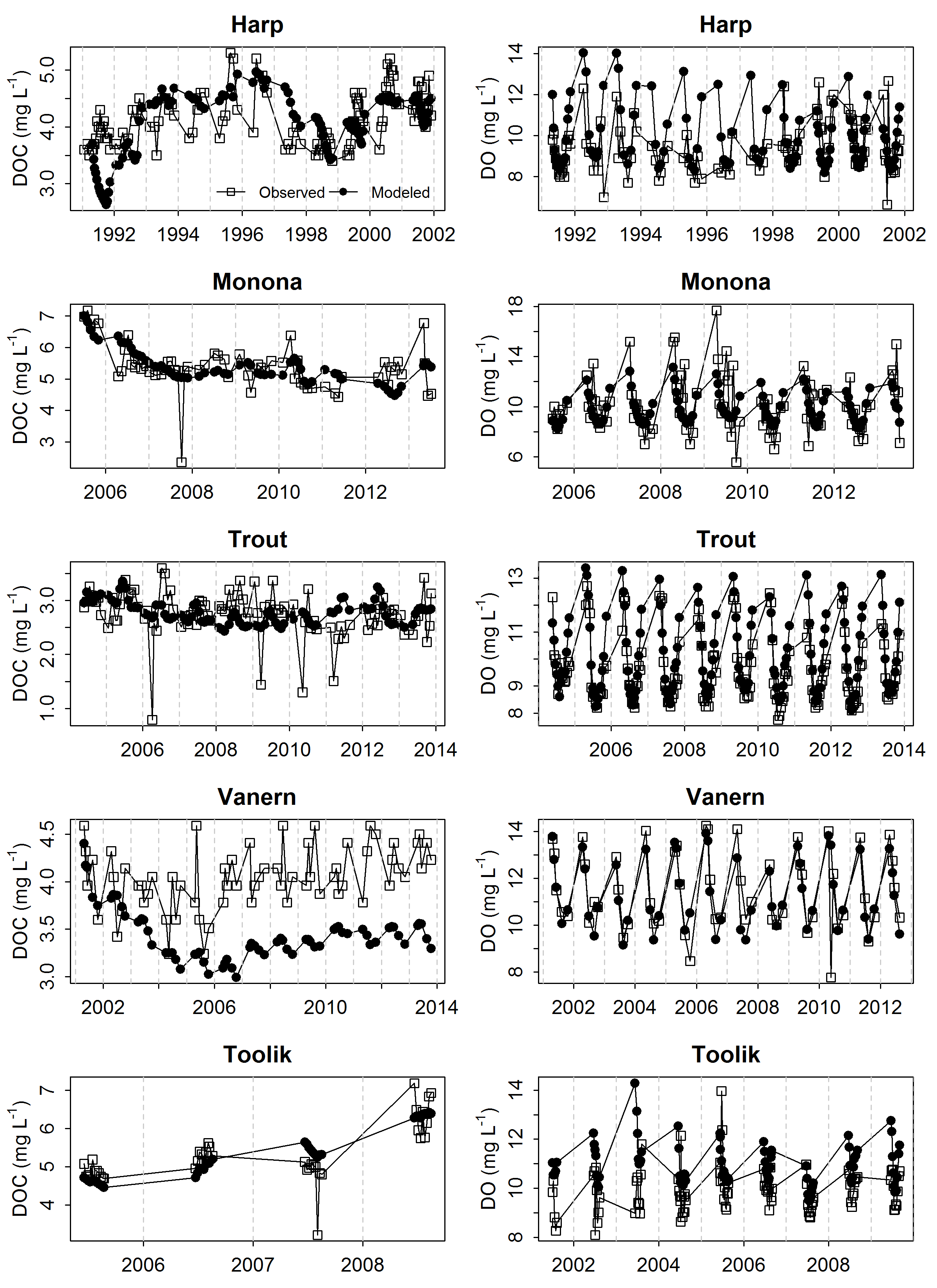
**FIGURE CAPTIONS**

**FIGURES**



**Figure 1.**

Figure 1. Conceptual diagram of the organic carbon lake model depicting fluxes based on allocthonous (externally derived) and autochthonous (NPP) inputs of organic carbon, long-term burial, leaching of particulate organic carbon (POC) to dissolved organic carbon (DOC), respiration of DOC to CO2, and export via outflow. Four parameters (Respiration\_auto, Respiration\_alloch, Burial\_auto, and Burial\_alloch) are treated as free parameters and calibrated for each individual lake. Parameters and equations are defined in Tables 2 and 3.



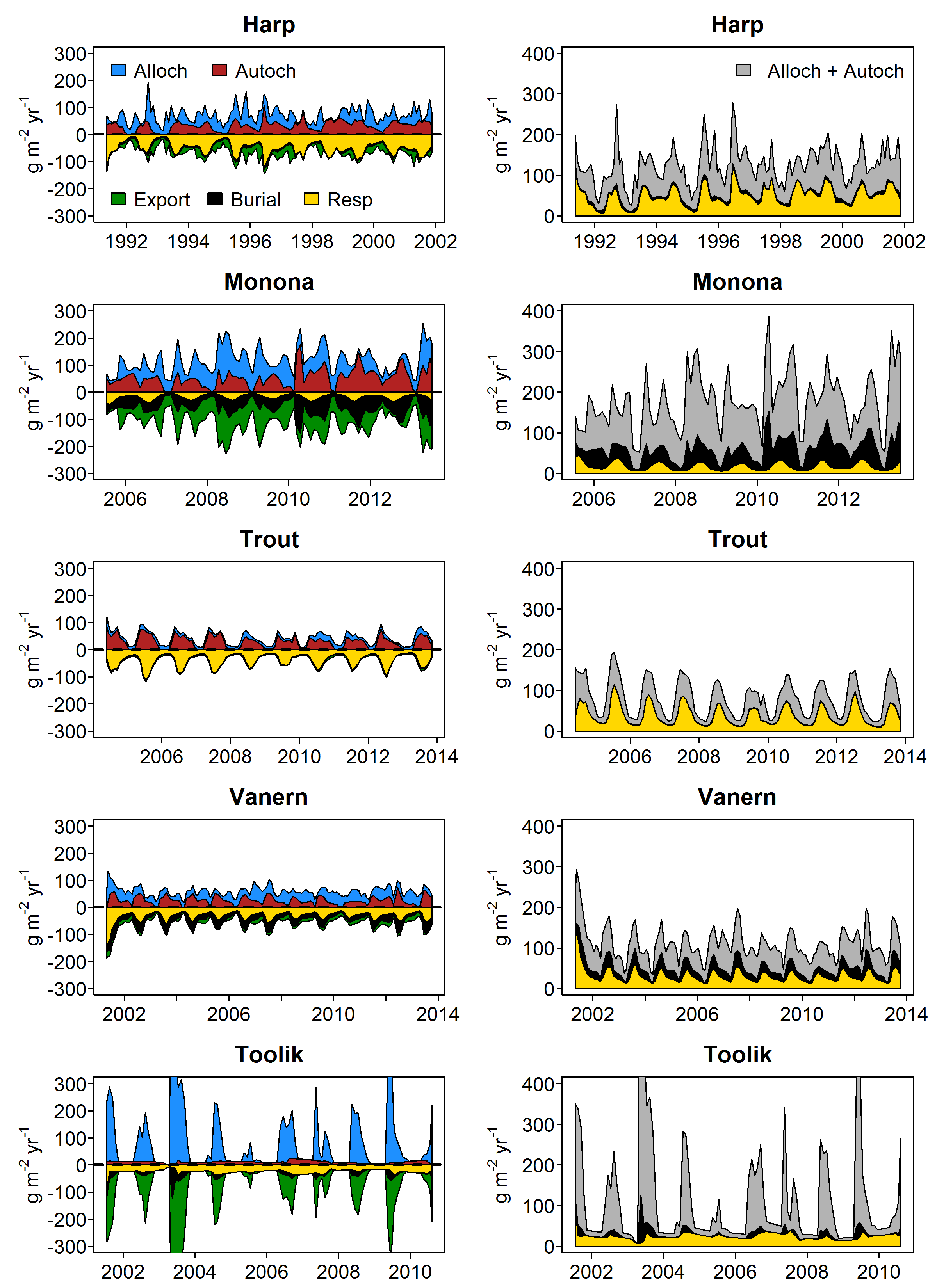
**Figure 2.**

Figure 2. Observed dissolved organic carbon (DOC) and dissolved oxygen (DO) concentrations in all lakes (blue circles) compared with modeled concentrations (red squares) on the same date. For some lakes, years differed between DOC and DO based on availability of observed data (S2).

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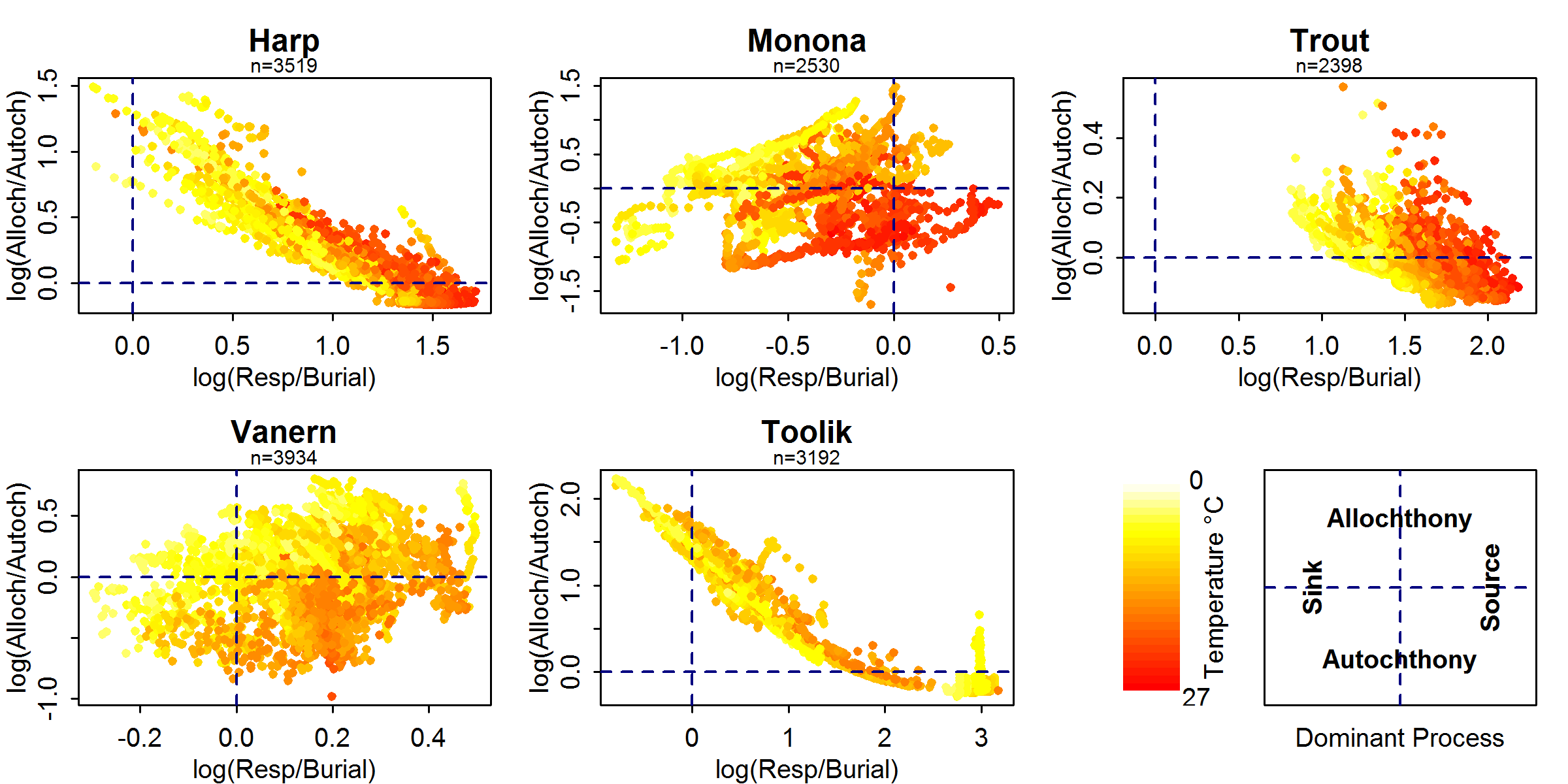
**Figure 3.**

Figure 3. Sensitivity of calibrated model parameters. Shaded areas represent the range of modeled dissolved organic carbon (DOC) concentrations as a given parameter was varied across a specified range (see legend) while the three other parameters remained fixed at their calibrated value. Black circles represent the observed in-lake DOC concentrations.



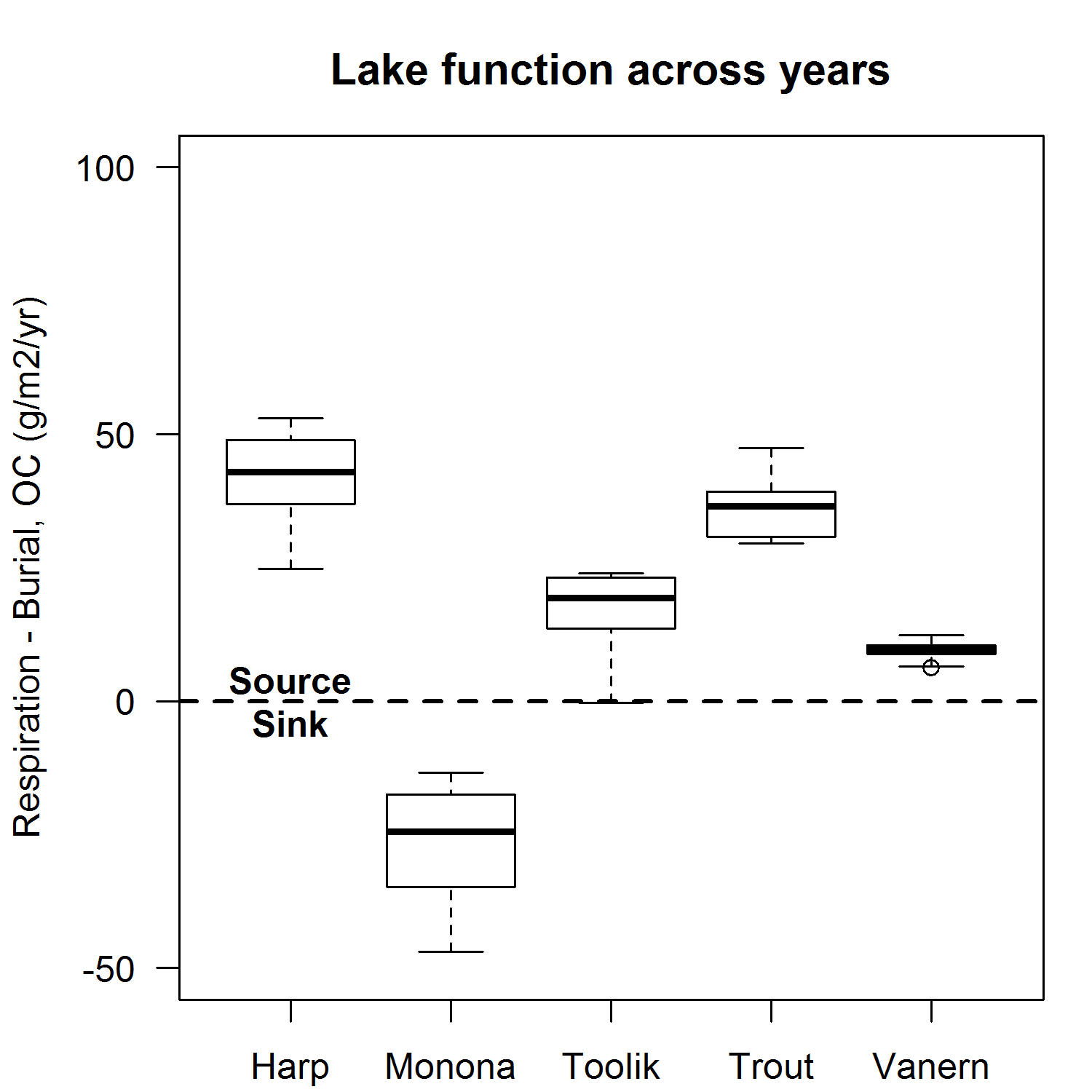
**Figure 4.**

Figure 4. Time series of organic carbon fluxes and fates. A) Colored areas represent relative magnitudes of input fluxes (allochthonous and autochthonous) and output fluxes (export, burial, and respiration). All lines are stacked to show cumulative magnitudes. b) Absolute values of burial and respiration, and input fluxes. A lake is a net source when respiration exceeds burial.



**Figure 5.**

Figure 5. Relationship between log10-transformed allochthony/autochthony and respiration/burial (g m-2 yr-1) of organic carbon, colored by water temperature. The four quadrants in each figure represent the dominant processes (either predominantly a source or sink, and either predominantly allochthonous driven or autochthonous driven) associated with each lake.



**Figure 6.**

Figure 6. Lake function (source or sink; respiration minus burial) across full modeled years and A lake functions as a source when the difference between respiration and burial exceeds zero.

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