

# Package ‘repfun’

November 14, 2025

**Title** Create TLFs using R functions styled after SAS macros

**Version** 0.0.0.9000

## Description

A clinical reporting toolkit of R functions that are written to mimic the style of SAS reporting macros. The purpose is to generate TLFs (tables, listings and figures) to support clinical trials.

**License** Apache License (== 2.0)

**Encoding** UTF-8

**Roxxygen** list(markdown = TRUE)

**RoxxygenNote** 7.3.2

**Imports** arrow,

  data.table,

  glue,

  jsonlite,

  rlang,

  xportr,

  Hmisc,

  r2rtf,

  haven,

  dplyr,

  tidyverse,

  magrittr,

  stringr

**Depends** R (>= 3.5)

**Suggests** knitr,

  rmarkdown,

  testthat (>= 3.0.0),

  rprojroot,

  DT,

  kableExtra,

  admiral,

  tibble,

  lubridate,

  ggplot2 (>= 4.0.0)

**Config/testthat/edition** 3

**VignetteBuilder** knitr

**LazyData** true

**URL** <https://gsk-biostatistics.github.io/repfun>

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### Description

adae

**Usage**

adae

**Format**

A data frame with 105 columns:

**STUDYID** Study Identifier  
**DOMAIN** Domain Abbreviation  
**USUBJID** Unique Subject Identifier  
**AESEQ** Sequence Number  
**AESPID** Sponsor-Defined Identifier  
**AETERM** Reported Term for the Adverse Event  
**AELLT** Lowest Level Term  
**AELLTCD** Lowest Level Term Code  
**AEDECOD** Dictionary-Derived Term  
**AEPTCD** Preferred Term Code  
**AEHLT** High Level Term  
**AEHLTC** High Level Term Code  
**AEHLGT** High Level Group Term  
**AEHLGTC** High Level Group Term Code  
**AEBODSYS** Body System or Organ Class  
**AEBDSYCD** Body System or Organ Class Code  
**AESOC** Primary System Organ Class  
**AESOCCD** Primary System Organ Class Code  
**AESEV** Severity/Intensity  
**AESER** Serious Event  
**AEACN** Action Taken with Study Treatment  
**AEREL** Causality  
**AEOOUT** Outcome of Adverse Event  
**AESCAN** Involves Cancer  
**AESCONG** Congenital Anomaly or Birth Defect  
**AESDISAB** Persist or Signif Disability/Incapacity  
**AESDTH** Results in Death  
**AESHOSP** Requires or Prolongs Hospitalization  
**AESLIFE** Is Life Threatening  
**AESOD** Occurred with Overdose  
**AEDTC** Date/Time of Collection  
**AESTDT** Start Date/Time of Adverse Event  
**AEENDTC** End Date/Time of Adverse Event  
**AESTDY** Study Day of Start of Adverse Event  
**AEENDY** Study Day of End of Adverse Event

**TRTSDT** Date of First Exposure to Treatment  
**TRTEDT** Date of Last Exposure to Treatment  
**DTHDT** Date of Death  
**EOSDT** End of Study Date  
**ASTDTM** Analysis Start Date/Time  
**ASTDTF** Analysis Start Date Imputation Flag  
**ASTTMF** Analysis Start Time Imputation Flag  
**AENDTM** Analysis End Date/Time  
**AENDTF** Analysis End Date Imputation Flag  
**AENTMF** Analysis End Time Imputation Flag  
**ASTDT** Analysis Start Date  
**AENDT** Analysis End Date  
**ASTDY** Analysis Start Relative Day  
**AENDY** Analysis End Relative Day  
**ADURN** Analysis Duration (N)  
**ADURU** Analysis Duration Units  
**LDOSEDTM** End Date/Time of Last Dose  
**ASEV** Analysis Severity/Intensity  
**AREL** Analysis Causality  
**TRTEMFL** Treatment Emergent Analysis Flag  
**ASEVN** Analysis Severity/Intensity (N)  
**AOCCIFL** 1st Max Sev./Int. Occurrence Flag  
**SUBJID** Subject Identifier for the Study  
**RFSTDT** Subject Reference Start Date/Time  
**RFENDTC** Subject Reference End Date/Time  
**RFXSTDTC** Date/Time of First Study Treatment  
**RFXENDTC** Date/Time of Last Study Treatment  
**RFICDT** Date/Time of Informed Consent  
**RFPENDTC** Date/Time of End of Participation  
**DTHDT** Date/Time of Death  
**DTHFL** Subject Death Flag  
**SITEID** Study Site Identifier  
**AGE** Age  
**AGEU** Age Units  
**SEX** Sex  
**RACE** Race  
**ETHNIC** Ethnicity  
**ARMCD** Planned Arm Code  
**ARM** Description of Planned Arm  
**ACTARMCD** Actual Arm Code

**ACTARM** Description of Actual Arm  
**COUNTRY** Country  
**DMDTC** Date/Time of Collection  
**DMDY** Study Day of Collection  
**TRT01P** Planned Treatment for Period 01  
**TRT01A** Actual Treatment for Period 01  
**TRTSDTM** Datetime of First Exposure to Treatment  
**TRTSTMF** Time of First Exposure Imput. Flag  
**TRTEDTM** Datetime of Last Exposure to Treatment  
**TRTETMF** Time of Last Exposure Imput. Flag  
**TRTDURD** Total Treatment Duration (Days)  
**SCRFDT** Screen Failure Date  
**EOSSTT** End of Study Status  
**FRVDT** Final Retrievel Visit Date  
**RANDDT** Date of Randomization  
**DTHDTF** undocumented field  
**DTHADY** Relative Day of Death  
**LDDTHELD** Elapsed Days from Last Dose to Death  
**DTHCAUS** undocumented field  
**DTHDOM** undocumented field  
**DTHCGR1** undocumented field  
**LSTALVDT** Date Last Known Alive  
**SAFFL** Safety Population Flag  
**RACEGR1** Pooled Race Group 1  
**AGEGR1** Pooled Age Group 1  
**REGION1** Geographic Region 1  
**LDDTHGR1** Last Dose to Death - Days Elapsed Grp 1  
**DTH30FL** Death Within 30 Days of Last Trt Flag  
**DTHA30FL** Death After 30 Days from Last Trt Flag  
**DTHB30FL** Death Within 30 Days of First Trt Flag

### Details

Adverse Events Analysis

### Source

Generated from admiral package (template ad\_adae.R).

### References

None

### Examples

```
data("adae")
```

---

adsl	<i>ADaM ADSL</i>
------	------------------

---

### Description

adsl

### Usage

adsl

### Format

A data frame with 54 columns:

**STUDYID** Study Identifier  
**USUBJID** Unique Subject Identifier  
**SUBJID** Subject Identifier for the Study  
**RFSTDT** Subject Reference Start Date/Time  
**RFENDT** Subject Reference End Date/Time  
**RFXSTDTC** Date/Time of First Study Treatment  
**RFXENDTC** Date/Time of Last Study Treatment  
**RFICDT** Date/Time of Informed Consent  
**RFPENDTC** Date/Time of End of Participation  
**DTHDT** Date/Time of Death  
**DTHFL** Subject Death Flag  
**SITEID** Study Site Identifier  
**AGE** Age  
**AGEU** Age Units  
**SEX** Sex  
**RACE** Race  
**ETHNIC** Ethnicity  
**ARMCD** Planned Arm Code  
**ARM** Description of Planned Arm  
**ACTARMCD** Actual Arm Code  
**ACTARM** Description of Actual Arm  
**COUNTRY** Country  
**DMDTC** Date/Time of Collection  
**DMDY** Study Day of Collection  
**TRT01P** Planned Treatment for Period 01  
**TRT01A** Actual Treatment for Period 01  
**TRTSDTM** Datetime of First Exposure to Treatment  
**TRTSTMF** Time of First Exposure Imput. Flag

**TRTEDTM** Datetime of Last Exposure to Treatment  
**TRTETMF** Time of Last Exposure Imput. Flag  
**TRTSDT** Date of First Exposure to Treatment  
**TRTEDT** Date of Last Exposure to Treatment  
**TRTDURD** Total Treatment Duration (Days)  
**SCRFDT** Screen Failure Date  
**EOSDT** End of Study Date  
**EOSSTT** End of Study Status  
**FRVDT** Final Retrievel Visit Date  
**RANDDT** Date of Randomization  
**DTHDT** Date of Death  
**DTHDTF** Date of Death Imputation Flag  
**DTHADY** Relative Day of Death  
**LDDTHELD** Elapsed Days from Last Dose to Death  
**DTHCAUS** undocumented field  
**DTHDOM** undocumented field  
**DTHCGR1** undocumented field  
**LSTALVDT** Date Last Known Alive  
**SAFFL** Safety Population Flag  
**RACEGR1** Pooled Race Group 1  
**AGEGR1** Pooled Age Group 1  
**REGION1** Geographic Region 1  
**LDDTHGR1** Last Dose to Death - Days Elapsed Grp 1  
**DTH30FL** Death Within 30 Days of Last Trt Flag  
**DTHA30FL** Death After 30 Days from Last Trt Flag  
**DTHB30FL** Death Within 30 Days of First Trt Flag

### Details

Subject Level Analysis

### Source

Generated from admiral package (template ad\_adsl.R).

### References

None

### Examples

```
data("adsl")
```

advs

*ADaM ADVS***Description**

advs

**Usage**

advs

**Format**

A data frame with 105 columns:

**STUDYID** Study Identifier**DOMAIN** Domain Abbreviation**USUBJID** Unique Subject Identifier**VSSEQ** Sequence Number**VTESTCD** Vital Signs Test Short Name**VTEST** Vital Signs Test Name**VSPOS** Vital Signs Position of Subject**VSORRES** Result or Finding in Original Units**VSORRESU** Original Units**VSSTRESC** Character Result/Finding in Std Format**VSSTRESP** Numeric Result/Finding in Standard Units**VSSTRESU** Standard Units**VSSTAT** Completion Status**VSLOC** Location of Vital Signs Measurement**VSBLFL** Baseline Flag**VISITNUM** Visit Number**VISIT** Visit Name**VISITDY** Planned Study Day of Visit**VSDTC** Date/Time of Measurements**VSDY** Study Day of Vital Signs**VSTPT** Planned Time Point Name**VSTPTNUM** Planned Time Point Number**VSELTM** Planned Elapsed Time from Time Point Ref**VSTPTREF** Time Point Reference**TRTSDT** Date of First Exposure to Treatment**TRTEDT** Date of Last Exposure to Treatment**TRT01A** Actual Treatment for Period 01**TRT01P** Planned Treatment for Period 01

**ADT** Analysis Date  
**ADY** Analysis Relative Day  
**PARAMCD** Parameter Code  
**AVAL** Analysis Value  
**ATPTN** Analysis Timepoint (N)  
**ATPT** Analysis Timepoint  
**AVISIT** Analysis Visit  
**AVISITN** Analysis Visit (N)  
**DTYPE** Derivation Type  
**ONTRTFL** On Treatment Record Flag  
**ANRLO** Analysis Normal Range Lower Limit  
**ANRHI** Analysis Normal Range Upper Limit  
**A1LO** Analysis Range 1 Lower Limit  
**A1HI** Analysis Range 1 Upper Limit  
**ANRIND** Analysis Reference Range Indicator  
**BASETYPE** Baseline Type  
**ABLFL** Baseline Record Flag  
**BASE** Baseline Value  
**BNRIND** Baseline Reference Range Indicator  
**CHG** Change from Baseline  
**PCHG** Percent Change from Baseline  
**ANL01FL** Analysis Flag 01  
**TRTP** Planned Treatment  
**TRTA** Actual Treatment  
**ASEQ** Analysis Sequence Number  
**AVALCAT1** Analysis Value Category 1  
**AVALCA1N** Analysis Value Category 1 (N)  
**PARAM** Parameter  
**PARAMN** Parameter (N)  
**SUBJID** Subject Identifier for the Study  
**RFSTDT** Subject Reference Start Date/Time  
**RFENDTC** Subject Reference End Date/Time  
**RFXSTDT** Date/Time of First Study Treatment  
**RFXENDTC** Date/Time of Last Study Treatment  
**RFICDT** Date/Time of Informed Consent  
**RFPENDTC** Date/Time of End of Participation  
**DTHDT** Date/Time of Death  
**DTHFL** Subject Death Flag  
**SITEID** Study Site Identifier  
**AGE** Age

**AGEU** Age Units  
**SEX** Sex  
**RACE** Race  
**ETHNIC** Ethnicity  
**ARMCD** Planned Arm Code  
**ARM** Description of Planned Arm  
**ACTARMCD** Actual Arm Code  
**ACTARM** Description of Actual Arm  
**COUNTRY** Country  
**DMDTC** Date/Time of Collection  
**DMDY** Study Day of Collection  
**TRTSDTM** Datetime of First Exposure to Treatment  
**TRTSTMF** Time of First Exposure Imput. Flag  
**TRTEDTM** Datetime of Last Exposure to Treatment  
**TRTETMF** Time of Last Exposure Imput. Flag  
**TRTDURD** Total Treatment Duration (Days)  
**SCRFDT** Screen Failure Date  
**EOSDT** End of Study Date  
**EOSSTT** End of Study Status  
**FRVDT** Final Retrievel Visit Date  
**RANDDT** Date of Randomization  
**DTHDT** Date of Death  
**DTHDTF** undocumented field  
**DTHADY** Relative Day of Death  
**LDDTHELD** Elapsed Days from Last Dose to Death  
**DTHCAUS** undocumented field  
**DTHDOM** undocumented field  
**DTHCGR1** undocumented field  
**LSTALVDT** Date Last Known Alive  
**SAFFL** Safety Population Flag  
**RACEGR1** Pooled Race Group 1  
**AGEGR1** Pooled Age Group 1  
**REGION1** Geographic Region 1  
**LDDTHGR1** Last Dose to Death - Days Elapsed Grp 1  
**DTH30FL** Death Within 30 Days of Last Trt Flag  
**DTHA30FL** Death After 30 Days from Last Trt Flag  
**DTHB30FL** Death Within 30 Days of First Trt Flag

### Details

Vital Signs Analysis

**Source**

Generated from admirals package (template ad\_advs.R).

**References**

None

**Examples**

```
data("advs")
```

---

ae	<i>Adverse Events</i>
----	-----------------------

---

**Description**

An updated SDTM AE dataset that uses the CDISC pilot project

**Usage**

```
ae
```

**Format**

A data frame with 35 columns:

**STUDYID** Study Identifier  
**DOMAIN** Domain Abbreviation  
**USUBJID** Unique Subject Identifier  
**AESEQ** Sequence Number  
**AESPID** Sponsor-Defined Identifier  
**AETERM** Reported Term for the Adverse Event  
**AELLT** Lowest Level Term  
**AELLTC** Lowest Level Term Code  
**AEDECOD** Dictionary-Derived Term  
**AEPTCD** Preferred Term Code  
**AEHLT** High Level Term  
**AEHTCD** High Level Term Code  
**AEHLGT** High Level Group Term  
**AEHLGTC** High Level Group Term Code  
**AEBODSYS** Body System or Organ Class  
**AEBDSYCD** Body System or Organ Class Code  
**AESOC** Primary System Organ Class  
**AESOCCD** Primary System Organ Class Code  
**AESEV** Severity/Intensity  
**AESER** Serious Event

**AEACN** Action Taken with Study Treatment  
**AEREL** Causality  
**AEOUT** Outcome of Adverse Event  
**AESCAN** Involves Cancer  
**AESCONG** Congenital Anomaly or Birth Defect  
**AESDISAB** Persist or Signif Disability/Incapacity  
**AESDTH** Results in Death  
**AESHOSP** Requires or Prolongs Hospitalization  
**AESLIFE** Is Life Threatening  
**AESOD** Occurred with Overdose  
**AEDTC** Date/Time of Collection  
**AESTDTC** Start Date/Time of Adverse Event  
**AEENDTC** End Date/Time of Adverse Event  
**AESTDY** Study Day of Start of Adverse Event  
**AEENDY** Study Day of End of Adverse Event

### Details

Adverse Events

An updated SDTM AE dataset that uses the CDISC pilot project

### Author(s)

Gopi Vegeasna

### Source

Access the source of the Adverse Events dataset.

---

airquality\_4test      *Updated airquality for testing.*

---

### Description

airquality\_4test

### Usage

airquality\_4test

## Format

A data frame with 100 observations and 8 columns:

**Ozone** Ozone Score  
**Solar.R** Solar Score  
**Wind** Wind Score  
**Temp** Temperature  
**Month** Month of Year  
**Day** Day of Month  
**CharVar1** Dummy Character Variable 1  
**CharVar2** Dummy Character Variable 2

## Details

Updated version of the airquality data frame that includes filled NA values for testing.

## Source

Generated from repfun package.

## References

None

## Examples

```
data("airquality_4test")
```

---

airquality\_updated      *Updated airquality*

---

## Description

airquality\_updated

## Usage

airquality\_updated

## Format

A data frame with 100 observations and 8 columns:

**Ozone** Ozone Score  
**Solar.R** Solar Score  
**Wind** Wind Score  
**Temp** Temperature  
**Month** Month of Year  
**Day** Day of Month  
**CharVar1** Dummy Character Variable 1  
**CharVar2** Dummy Character Variable 2

**Details**

Updated version of the airquality data frame that includes NA values.

**Source**

Generated from repfun package.

**References**

None

**Examples**

```
data("airquality_updated")
```

---

copydata

*Copy package data to the directory specified.*

---

**Description**

Copy all package data into a temporary directory that can be used when running examples.

**Usage**

```
copydata(p)
```

**Arguments**

p                   A path as a string.

**Value**

No return value, the current working directory is set.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
copydata(tempdir())
```

---

dm

*Demography*

---

### Description

A SDTM DM dataset from the CDISC pilot project

### Usage

dm

### Format

A data frame with 25 columns:

**STUDYID** Study Identifier  
**DOMAIN** Domain Abbreviation  
**USUBJID** Unique Subject Identifier  
**SUBJID** Subject Identifier for the Study  
**RFSTDTC** Subject Reference Start Date/Time  
**RFENDTC** Subject Reference End Date/Time  
**RFXSTDTC** Date/Time of First Study Treatment  
**RFXENDTC** Date/Time of Last Study Treatment  
**RFICDTC** Date/Time of Informed Consent  
**RFPENDTC** Date/Time of End of Participation  
**DTHDTTC** Date/Time of Death  
**DTHFL** Subject Death Flag  
**SITEID** Study Site Identifier  
**AGE** Age  
**AGEU** Age Units  
**SEX** Sex  
**RACE** Race  
**ETHNIC** Ethnicity  
**ARMCD** Planned Arm Code  
**ARM** Description of Planned Arm  
**ACTARMCD** Actual Arm Code  
**ACTARM** Description of Actual Arm  
**COUNTRY** Country  
**DMDTC** Date/Time of Collection  
**DMDY** Study Day of Collection

### Details

Demography

A SDTM DM dataset from the CDISC pilot project

**Source**

Access the source of the Demography dataset.

---

formats	<i>Data frame of SAS formats</i>
---------	----------------------------------

---

**Description**

formats

**Usage**

formats

**Format**

A data frame with 5 columns and 8 observations:

**FMTNAME** Name of Format  
**START** Start Value of Format  
**END** End Value of Format  
**LABEL** Label of Format  
**TYPE** Type of Format

**Details**

Formats Data Frame (SAS Style)

**Source**

Generated for repfun package.

**References**

None

**Examples**

```
data("formats")
```

---

**mtcars\_w2lbls** *This is the mtcars data frame with labels added for columns mpg and cyl.*

---

## Description

`mtcars_w2lbls`

## Usage

`mtcars_w2lbls`

## Format

A data frame with 32 observations and 11 columns:

**mpg** Miles/(US) gallon  
**cyl** Number of cylinders  
**disp** Displacement (cu.in.)  
**hp** Gross horsepower  
**drat** Rear axle ratio  
**wt** Weight (1000 lbs)  
**qsec** 1/4 mile time  
**vs** Engine (0 = V-shaped, 1 = straight)  
**am** Transmission (0 = automatic, 1 = manual)  
**gear** Number of forward gears  
**carb** Number of carburetors

## Details

Updated version of the mtcars data frame that includes labels for 2 variables.

## Source

Generated from repfun package.

## References

None

## Examples

```
data("mtcars_w2lbls")
```

<code>rem_pg_nums</code>	<i>Remove floating page numbers from RTF files.</i>
--------------------------	---

## Description

Pass in an RTF file to have floating page numbers removed and result saved to the same file.

## Usage

```
rem_pg_nums(infile)
```

## Arguments

<code>infile</code>	A path to the RTF file as a string.
---------------------	-------------------------------------

## Value

No return value, the file is modified.

## Author(s)

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
## Not run:  
library(repfun)  
repfun:::rem_pg_nums('/path/to/an/rtf/file.rtf')  
  
## End(Not run)
```

<code>rs_setup</code>	<i>Pass values to setup function and the global environment will be arranged for use reporting tools.</i>
-----------------------	---

## Description

Pass values to setup function and the global environment will be arranged for use reporting tools.

## Usage

```
rs_setup(  
  R_DICTION = "./inst/formats",  
  R_MACDIRS = NULL,  
  R_DDDATA = NULL,  
  R_OTHERDATA = paste0("./data"),  
  R_INPUTDATA = paste0("./data"),  
  R_RAWDATA = paste0("./data"),  
  R_SDTMDATA = paste0("./data"),
```

```

R_ADAMDATA = paste0("./data"),
R_RFMTDIR = paste0("./inst/formats"),
D_CENTID = "SITEID",
D_DATADATE = NULL,
D_DSPLYNUM = 1,
D_DSPLYTYP = "T",
D_FONTSIZE = 10,
D_KEEPPOPVARS = NULL,
D_OUTFILE = "./inst/t_ru_list_1.rtf",
D_PGMPTH = "./R/rs_setup.R",
D_STUDYID = NULL,
D_STUDY_DESC = NULL,
D_POP = "ITTEFL",
D_POPDATA = NULL,
D_POPLBL = "Intent-to-Treat",
D_SUBJID = "USUBJID",
D_SUBPOP = NULL,
D_SUBSET = NULL,
D_TITLE1 = NULL,
D_TITLE2 = NULL,
D_TITLE3 = NULL,
D_TITLE4 = NULL,
D_TITLE5 = NULL,
D_TITLE6 = NULL,
D_TITLE7 = NULL,
D_FOOT1 = NULL,
D_FOOT2 = NULL,
D_FOOT3 = NULL,
D_FOOT4 = NULL,
D_FOOT5 = NULL,
D_FOOT6 = NULL,
D_FOOT7 = NULL,
D_FOOT8 = NULL,
D_FOOT9 = NULL,
D_USERID = Sys.getenv("DOMINO_USER_NAME"),
D_RTFYN = "N",
D_DEBUG = 0
)

```

## Arguments

R_DICTION	Location of reporting dictionaries.
R_MACDIRS	List of folders to search for functions when they are invoked.
R_DDDATA	Location to write DDDATA reporting data sets.
R_OTHERDATA	Location of additional production data sets.
R_INPUTDATA	Location of permanent formats data sets and miscellaneous data sets.
R_RAWDATA	Location of raw data sets.
R_SDATMDATA	Location of SDTM data sets.
R_ADAMDATA	Location of ADAM data sets.
R_RFMTDIR	Location of format catalogs and corresponding lists.

D_CENTID	Variable name for investigational center.
D_DATADATE	Date of data sets for use in titles/footnotes.
D_DSPLYNUM	Display number for title.
D_DSPLYTYP	Type of Display ('T','L','F').
D_FONTSIZE	Size of font on output file which is RTF by default (with automated PDF conversion via script).
D_KEEPPOPVARS	Variables to keep on the population data set when merging to apply populations and sub-populations.
D_OUTFILE	Production location for output TLFs.
D_PGMPTH	Path of the driver file that generates current outputs or data sets.
D_STUDYID	Protocol ID for the study.
D_STUDY_DESC	Description of Study.
D_POP	Population variable from ADSL that must equal Y for subjects to be included in the analysis.
D_POPDATA	Data set that contains the population to be analyzed.
D_POPLBL	Label for population being analyzed which can be used in the TLF header.
D_SUBJID	The variable used to uniquely identify a subject in this analysis.
D_SUBPOP	Condition to identify the sub-population when applied to ADSL.
D_SUBSET	Condition to filter data from incoming source data sets used for this TLF.
D_TITLE1	First title text.
D_TITLE2	Second title text.
D_TITLE3	Third title text.
D_TITLE4	Fourth title text.
D_TITLE5	Fifth title text.
D_TITLE6	Sixth title text.
D_TITLE7	Seventh title text.
D_FOOT1	First footnote text.
D_FOOT2	Second footnote text.
D_FOOT3	Third footnote text.
D_FOOT4	Fourth footnote text.
D_FOOT5	Fifth footnote text.
D_FOOT6	Sixth footnote text.
D_FOOT7	Seventh footnote text.
D_FOOT8	Eighth footnote text.
D_FOOT9	Ninth footnote text.
D_USERID	Domino user name.
D_RTFYN	Y or N to generate RTf output.
D_DEBUG	Level of debugging to show in log files.

**Value**

Global variables defined for use with the reporting tools.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
library(dplyr)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datdir")
dir.create(datadir, showWarnings=FALSE)
outdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "outdir")
dir.create(outdir, showWarnings=FALSE)
fmtdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "fmtdir")
dir.create(fmtdir, showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(
  D_CENTID="SITEID",
  D_DATADATE=Sys.Date(),
  D_DSPLYNUM=1,
  D_DSPLYTYP='T',
  D_FONTSIZE=10,
  D_FOOT1='1.) Only treatment emergent events related to lipids are displayed.',
  D_FOOT2='2.) Subjects are counted once in each body system & preferred term.',
  D_KEEPPOPVARS=c('STUDYID', 'USUBJID', 'SAFFL'),
  D_USERID=Sys.getenv("DOMINO_USER_NAME"),
  D_STUDYID='ABCXYZPDQ',
  D_POP="SAFFL",
  D_POPDATA=repfun::adsl %>% dplyr::filter(SAFFL=='Y'),
  D_POPLBL="Safety",
  D_SUBJID=c("STUDYID", "USUBJID"),
  D_TITLE1=paste0('Table 1: Summary of Treatment Emergent Adverse Events'),
  D_RTFYN="Y",
  D_DEBUG=0,
  R_DICTION=NULL,
  R_OTHERDATA=NULL,
  R_INPUTDATA=NULL,
  R_RAWDATA=NULL,
  R_SDTMDATA=NULL,
  R_ADAMDATA=datadir,
  R_RFMTDIR=fmtdir,
  R_DDDATA=paste0(outdir, '/t_ru_list_1.rds'),
  D_OUTFILE=paste0(outdir, "/t_ru_list_1.rtf"),
  D_PGMPTH="/path/to/code/rs_setup.R")
```

## Description

Pass in a data frame along with identification options and have Big N added to it.

## Usage

```
ru_addbignvar(
  dsetintoaddbign,
  dsetintocount,
  countdistinctvars = c("STUDYID", "USUBJID"),
  groupbyvars = NULL,
  totalforvar = NULL,
  totalid = NULL,
  totaldecode = c("Total"),
  codedecodevarpairs = NULL,
  varcodelistpairs = NULL,
  codelistnames = list(),
  addbignтоварvalue = TRUE,
  splitchar = " "
)
```

## Arguments

<code>dsetintoaddbign</code>	The data set that will hold the derived big N value.
<code>dsetintocount</code>	The data set that will be counted to generate big N.
<code>countdistinctvars</code>	Variable(s) that contain values to be counted uniquely within any output grouping.
<code>groupbyvars</code>	Variables in DSETINTOCOUNT to group the data by when counting to deriving the big N.
<code>totalforvar</code>	Variable for which overall totals are required within all other grouped class variables.
<code>totalid</code>	Value(s) used to populate the variable(s) specified in totalforvar.
<code>totaldecode</code>	Value(s) used to populate the variable(s) of the decode variable(s) of the totalforvar.
<code>codedecodevarpairs</code>	Specifies code and decode variable pairs. Those variables should be in parameter GROUPBYVARSNUMER. One variable in the pair will contain the code, which is used in counting and ordering, and the other will contain decode, which is used for presentation.
<code>varcodelistpairs</code>	List of code/decode pairs of variables.
<code>codelistnames</code>	List of decodes for use with decoding code/decode pairs.
<code>addbignтоварvalue</code>	Place big N in a new variable or append to an existing variable (last groupbyvars value)?
<code>splitchar</code>	Text to insert between existing string and big N.

**Value**

A data frame based on the incoming data frame but collapsed by groups with descriptive statistics added.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
library(dplyr)
library(tibble)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datadir");
dir.create(datadir, showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(D_POPDATA=repfun::ads1 %>% dplyr::filter(SAFFL =='Y'),
                 D_SUBJID=c("STUDYID", "USUBJID"),
                 R_DICTION=NULL,
                 R_OTHERDATA=NULL,
                 R_INPUTDATA=NULL,
                 R_RAWDATA=NULL,
                 R_SDTMDATA=NULL,
                 R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>% dplyr::mutate(TRT01AN=
  ifelse(TRT01A=='Placebo',1,
         ifelse(TRT01A=='Xanomeline Low Dose',2,3)))
attr(G_POPDATA$TRT01AN,"label") <- 'Actual Treatment for Period 01 (n)'
adae <- tibble::as_tibble(rfenv$adamdata$adae.rda()) %>%
  dplyr::inner_join(G_POPDATA,
                     by=c('STUDYID', 'USUBJID', 'SAFFL', 'TRT01A')) %>%
  dplyr::filter(TRTEMFL=='Y')
addbign <- repfun::ru_addbignvar(adae,
  G_POPDATA,
  groupbyvars=c("TRT01AN", "TRT01A"),
  countdistinctvars=c("STUDYID", "USUBJID"),
  totalforvar=c("TRT01AN"),
  totalid = 99,
  totaldecode = 'Total',
  codedecodevarpairs=c("TRT01AN", "TRT01A"),
  varcodelistpairs=c(""),
  codelistnames=list(),
  addbigntovarvalue=TRUE,
  splitchar="~") %>%
  dplyr::select(STUDYID, USUBJID, TRT01AN, TRT01A, AEBODSYS, AEDECOD)
```

**ru\_addpage***Add a Page Number Column to an Existing Dataframe***Description**

Take incoming dataframe and add page number variable to it accounting for grouping variables, stacked variables, and no-split variables.

**Usage**

```
ru_addpage(
  dsetin,
  grpvars = NULL,
  stackvars = NULL,
  varlabls = NULL,
  rowsprbdy = NULL,
  startpaging = 0,
  lastbygrp = FALSE,
  fpage = "all",
  nftnotes = 0,
  nosplitvars = FALSE,
  npgvars = 0
)
```

**Arguments**

dsetin	The dataframe for which a paging variable will be added.
grpvars	Grouping variables used in the output (used for nosplitvars).
stackvars	Specify stacked grouping variables (reduces # of page lines available for data).
varlabls	Apply labels to outgoing dataframe.
rowsprbdy	Number of rows in the body of the report.
startpaging	Set to zero on first call, and > 0 on recalls to fix widows.
lastbygrp	Set to true if this page is processing the last value of the grouping variables (used when footnote is applied only to last page).
fpage	Setting to 'last' indicates that footnotes are only displayed on the last page.
nftnotes	Enter the number of footnotes (determines # of page lines available for data).
nosplitvars	Setting to true requires all values of the last grouping/stackvar must be on the same page (if possible).
npgvars	Number of page-by variables for this report (reduces # of page lines available for data).

**Value**

A dataframe based on the incoming dataframe but with a paging variable added.

**Author(s)**

Chris Rook, <[christopher.x.rook@gsk.com](mailto:christopher.x.rook@gsk.com)>  
 Yongwei Wang, <[yongwei.x.wang@viivhealthcare.com](mailto:yongwei.x.wang@viivhealthcare.com)>

## Examples

```

library(repfun)
library(dplyr)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\","/",tempdir(),fixed=TRUE),"datadir");
dir.create(datadir,showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(D_POPDATA=repfun::ads1 %>% dplyr::filter(SAFFL =='Y'),
                 D_SUBJID=c("STUDYID","USUBJID"),
                 R_DICTION=NULL,
                 R_OTHERDATA=NULL,
                 R_INPUTDATA=NULL,
                 R_RAWDATA=NULL,
                 R_SDTMDATA=NULL,
                 R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(TRT01AN=ifelse(TRT01A=='Placebo',1,
                                ifelse(TRT01A=='Xanomeline Low Dose',2,3))) %>%
  repfun::ru_labels(varlabels=list('TRT01AN'='Actual Treatment for Period 001 (n)'))
adae <- rfenv$adamdata$adae.rda() %>%
  dplyr::inner_join(G_POPDATA, by=c('STUDYID','USUBJID','SAFFL','TRT01A'))
aesum_t <- repfun::ru_freq(
  adae,
  dsetindenom=G_POPDATA,
  countdistinctvars=c('STUDYID','USUBJID'),
  groupbyvarsnumer=c('TRT01AN','TRT01A','AEBODSYS','AEDECOD'),
  anyeventvars = c('AEBODSYS','AEDECOD'),
  anyeventvalues = c('ANY EVENT','ANY EVENT'),
  groupbyvarsdenom=c('TRT01AN'),
  resultstype="NUMERPCT",
  totalforvar=c('TRT01AN'),
  totalid=99,
  totaldecode='Total',
  codedecodevarpairs=c("TRT01AN", "TRT01A"),
  varcodelistpairs=c(""),
  codelistnames=list(),
  resultpctdps=0) %>%
repfun::ru_denorm(varstodenorm=c("tt_result", "PERCENT"),
                   groupbyvars=c("tt_summarylevel", "AEBODSYS", "AEDECOD"),
                   acrossvar="TRT01AN",
                   acrossvarlabel="TRT01A",
                   acrossvarprefix=c("tt_ac", "tt_p")) %>%
dplyr::mutate(ord1=ifelse(tt_summarylevel==0,0,1)) %>%
dplyr::rename(ord2=tt_summarylevel) %>%
dplyr::arrange(ord1,AEBODSYS,ord2,AEDECOD) %>%
dplyr::select(-c(starts_with('tt_p'),starts_with('ord')))

# Example 1: Simple paging.
aesum_p1 <- repfun::ru_addpage(aesum_t,grpvars=c('AEBODSYS'),rowsprbdy=30)
print(head(aesum_p1,40))

```

```
# Example 2: No splitvars, but 30 rows won't work for this data.
aesum_p2 <- repfun::ru_addpage(aesum_t, grpvars=c('AEBODSYS'), rowsprbdy=30,
                                nosplitvars=TRUE)
print(head(aesum_p2, 40))

# Example 3: No splitvars, but 35 rows is enough.
aesum_p3 <- repfun::ru_addpage(aesum_t, grpvars=c('AEBODSYS'), rowsprbdy=35,
                                nosplitvars=TRUE)
print(head(aesum_p3, 40))
```

**ru\_addsupp**

*Combine an SDTM domain with its corresponding Supplemental data set*

**Description**

Pass in an SDTM data frame along with its Supplemental version and they will be combined.

**Usage**

```
ru_addsupp(dsetin, dsetinsupp)
```

**Arguments**

dsetin	Incoming data frame to have supplemental data added.
dsetinsupp	The supplemental data set to add.

**Value**

The original SDTM data set with its supplemental data appended as new variables.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viiivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
library(knitr)
ae <- repfun::ae
suppae <- repfun::suppae
aesupp <- repfun::ru_addsupp(dsetin=ae, dsetinsupp=suppae)
knitr::kable(head(aesupp, 5), caption = "SDTM.AE combined with SDTM.SUPPAE")
```

---

ru_align	<i>Align Columns for Reporting</i>
----------	------------------------------------

---

## Description

Pass in a data frame alignment criteria to have columns aligned for reporting.

## Usage

```
ru_align(  
  dsetin,  
  varsin,  
  byvars = NULL,  
  alignment = "Right",  
  compresschry = "Y",  
  ncspaces = 1  
)
```

## Arguments

dsetin	The data set holding the columns to align.
varsin	Columns to align.
byvars	Set of group-by variables.
alignment	Type of alignment.
compresschry	Compress by removing leading and trailing spaces for the resulting aligned column?
ncspaces	Number of spaces between N and Percent.

## Value

A data frame based having the requested columns aligned for reporting.

## Author(s)

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
=====  
# Adverse Events  
=====  
library(repfun)  
library(dplyr)  
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){  
  rfenv  
} else {  
  rfenv <- new.env(parent = emptyenv())  
  rfenv$G_DEBUG <- 0  
  rfenv  
}
```

```

datdir <- file.path(gsub("\\\\","/",tempdir(),fixed=TRUE),"datdir")
dir.create(datdir,showWarnings=FALSE)
refun::copydata(datdir)
refun::rs_setup(D_POP="SAFFL",
               D_POPLBL="Safety",
               D_POPDATA=refun::adsl %>% dplyr::filter(SAFFL =='Y'),
               D_SUBJID=c("STUDYID", "USUBJID"),
               R_DICTION=NULL,
               R_OTHERDATA=NULL,
               R_INPUTDATA=NULL,
               R_RAWDATA=NULL,
               R_SDTMDATA=NULL,
               R_ADAMDATA=datdir)
G_POPDATA <- rfenv$G_POPDATA %>% dplyr::mutate(TRT01AN=
                                                ifelse(TRT01A=='Placebo',1,
                                                       ifelse(TRT01A=='Xanomeline Low Dose',2,3)))
attr(G_POPDATA$TRT01AN,"label") <- 'Actual Treatment for Period 01 (n)'
adae <- rfenv$adamdata$adae.rda() %>% dplyr::select(-SAFFL) %>%
        refun::ru_getdata(G_POPDATA, c("STUDYID", "USUBJID"),
                           keeppopvars=c("TRT01AN", "TRT01A"))
aesum_t <- refun::ru_freq(adae,
                           dsetindenom=G_POPDATA,
                           countdistinctvars=c('STUDYID', 'USUBJID'),
                           groupbyvarsnum=c('TRT01AN', 'TRT01A', 'AEBODSYS', 'AEDECOD'),
                           anyeventvars = c('AEBODSYS', 'AEDECOD'),
                           anyeventvalues = c('ANY EVENT', 'ANY EVENT'),
                           groupbyvarsdenom=c('TRT01AN'),
                           resultstyle="NUMERPCT",
                           totalforvar=c('TRT01AN'),
                           totalid=99,
                           totaldecode='Total',
                           codedecodevarpairs=c("TRT01AN", "TRT01A"),
                           varcodelistpairs=c(""),
                           codelistnames=list(),
                           resultpctdps=0) %>%
        refun::ru_denorm(varstodenorm=c("tt_result", "PERCENT"),
                          groupbyvars=c("tt_summarylevel", "AEBODSYS", "AEDECOD"),
                          acrossvar="TRT01AN",
                          acrossvarlabel="TRT01A",
                          acrossvarprefix=c("tt_ac", "tt_p"))
print('Before Aligning')
print(head(aesum_t[,grep('(AEBODSYS|AEDECOD|tt_ac)',names(aesum_t))],20))
aesum_t_a <- refun::ru_align(aesum_t, "tt_ac:")
print('After Aligning')
print(head(aesum_t_a[,grep('(AEBODSYS|AEDECOD|tt_ac)',names(aesum_t_a))],20))

#####
# Baseline Characteristics
#####
refun::rs_setup(D_POP="SAFFL",
               D_POPLBL="Safety",
               D_POPDATA=refun::adsl %>% dplyr::filter(SAFFL =='Y'),
               D_SUBJID=c("STUDYID", "USUBJID"),
               R_DICTION=NULL,
               R_OTHERDATA=NULL,
               R_INPUTDATA=NULL,
               R_RAWDATA=NULL,

```

```

R_SDTMDATA=NULL,
R_ADAMDATA=datdir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(TRT01AN=ifelse(TRT01A=='Placebo',1,
                                ifelse(TRT01A=='Xanomeline Low Dose',2,3))) %>%
  repfun::ru_labels(varlabels=list('TRT01AN'='Actual Treatment for Period 01 (n)'))
demstats_t <- repfun::ru_sumstats(G_POPDATA,
                                    analysisvars=c("AGE", "TRTDURD"),
                                    groupbyvars=c("STUDYID", "TRT01AN"),
                                    codedecodevarpairs=c("TRT01AN", "TRT01A"),
                                    totalforvar="TRT01AN", totalid=99,
                                    totaldecode="Total",
                                    statsinrowsyn = "Y",
                                    analysisvardps=list("AGE"=1, "TRTDURD"=2),
                                    statslist=c("n", "mean", "median", "sd", "min", "max")) %>%
repfun::ru_denorm(varstodenorm=c("tt_result"),
                    groupbyvars=c("tt_avid", "tt_avnm", "tt_svid", "tt_svnm"),
                    acrossvar="TRT01AN", acrossvarlabel="TRT01A",
                    acrossvarprefix=c("tt_ac"))

print('Before Aligning')
print(head(demstats_t,20))
demstats_t_a <- repfun::ru_align(demstats_t, "tt_ac:", ncspaces=10)
print('After Aligning')
print(head(demstats_t_a,20))

```

**ru\_contents***Generate proc contents of data frame similar to that generated by SAS***Description**

Pass in a data frame and the contents will be displayed.

**Usage**

```
ru_contents(dsetin)
```

**Arguments**

dsetin	Incoming data frame to have proc contents generated.
--------	--

**Value**

The proc contents output will be displayed.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
library(repfun)
repfun::ru_contents(mtcars)
```

**ru\_data2codelist**      *Return a list with codelist, code and label based on input codelist dataset.*

## Description

Pass in a data set from a SAS format catalog (or similar) and have a list returned in the structure of a SAS format for decoding variables.

## Usage

```
ru_data2codelist(
  dsetin,
  codelistvarname = "FMTNAME",
  codevarname = "START",
  decodevarname = "LABEL",
  typevarname = "TYPE"
)
```

## Arguments

dsetin	Name of incoming data set structured as a SAS format catalog saved as a data set.
codelistvarname	Name of the variable containing the SAS format or similar.
codevarname	Name of the variable that holds the code value.
decodevarname	Name of the variable that holds the decode value.
typevarname	Type of format (character or numeric).

## Value

A data frame based on the incoming data frame but with decode values added along with records when completetypes is true.

## Author(s)

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
library(repfun)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datdir <- file.path(gsub("\\\\","/",tempdir(),fixed=TRUE),"datdir")
dir.create(datdir,showWarnings=FALSE)
repfun::copydata(datdir)
repfun::rs_setup(R_RFMTDIR=datdir)
list <- repfun::ru_data2codelist(rfenv$rfmtdata$formats.rda())
list$SEXS$START[[1]] # Code value 1
list$SEXS$LABEL[[1]] # Decode value 1
list$SEXS$START[[2]] # Code value 2
list$SEXS$LABEL[[2]] # Decode value 2
```

ru\_datacompare

*Compare 2 data frames and report differences.*

## Description

Pass in a base and compare data frame to find out if they are equal similar to proc compare in SAS.

## Usage

```
ru_datacompare(dsetinbase, dsetincomp, idvars, maxprint = 50)
```

## Arguments

dsetinbase	First data set.
dsetincomp	Second data set.
idvars	Match on these values prior to comparing records.
maxprint	Maximum number of differences per variable to display.

## Value

An output similar to proc compare will be displayed.

## Author(s)

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
library(repfun)
repfun::ru_datacompare(iris,iris,idvars='Species')
```

---

<code>ru_datetime</code>	<i>Add numeric datetimes to data frame that only has character versions.</i>
--------------------------	--

---

## Description

Pass in a data frame and variables ending in "DTC" will have corresponding numeric versions created and saved.

## Usage

```
ru_datetime(dsetin, includevars = NULL)
```

## Arguments

<code>dsetin</code>	Incoming data frame to have numeric datetimes added.
<code>includevars</code>	Specify which variables ending in DTC will be processed.

## Value

The proc contents output will be displayed.

## Author(s)

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
library(repfun)
library(knitr)
ae <- repfun::ru_datetime(repfun::ae)
knitr::kable(head(ae[,grepl('DT$|TM$|DTC$',names(ae))],5),
            caption = "After Invoking ru_labels()")
```

---

<code>ru_denorm</code>	<i>Transpose a Data Frame</i>
------------------------	-------------------------------

---

## Description

Pass in a data frame along with identification options and have it transposed (denormalized, long to wide) to display treatment columns.

**Usage**

```
ru_denorm(
  dsetin,
  varstodenorm = NULL,
  groupbyvars = NULL,
  acrossvar = NULL,
  acrossvarlabel = NULL,
  acrossvarprefix = "tt_",
  acrossvarsuffix = NULL
)
```

**Arguments**

dsetin	The data set to transpose.
varstodenorm	The variables to transpose.
groupbyvars	Definition of one row in the output data frame.
acrossvar	Variable to define the columns in the transposed data frame.
acrossvarlabel	Variable to define the labels in the transposed data frame.
acrossvarprefix	Add to the beginning of each value in the across variable in the output data frame.
acrossvarsuffix	Add to the end of each value in the across variable in the output data frame.

**Details**

dsetin, varstodenorm=NULL, groupbyvars=NULL, acrossvar=NULL, acrossvarlabel=NULL, acrossvarprefix="tt\_", acrossvarsuffix=NULL

**Value**

A data frame based on the incoming data frame transposed from long to wide.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
#####
# AEs: N and Percent
#####
library(repfun)
library(dplyr)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datdir")
```

```

dir.create(datadir, showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(D_POP="SAFFL",
                 D_POPLBL="Safety",
                 D_POPDATA=repfun::adsl %>% dplyr::filter(SAFFL =='Y'),
                 D_SUBJID=c("STUDYID", "USUBJID"),
                 R_DICTION=NULL,
                 R_OTHERDATA=NULL,
                 R_INPUTDATA=NULL,
                 R_RAWDATA=NULL,
                 R_SDTMDATA=NULL,
                 R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(TRT01AN=ifelse(TRT01A=='Placebo', 1,
                               ifelse(TRT01A=='Xanomeline Low Dose', 2, 3))) %>%
  repfun::ru_labels(varlabels=list('TRT01AN'='Actual Treatment for Period 01 (n)'))
adae <- rfenv$adadata$adae.rda() %>% select(-SAFFL) %>%
  repfun::ru_getdata(G_POPDATA, c("STUDYID", "USUBJID"),
                      keeppopvars=c("TRT01AN", "TRT01A"))
aesum_t <- repfun::ru_freq(adae,
                           dsetindenom=G_POPDATA,
                           countdistinctvars=c('STUDYID', 'USUBJID'),
                           groupbyvarsnum=c('TRT01AN', 'TRT01A', 'AEBODSYS', 'AEDECOD'),
                           anyeventvars = c('AEBODSYS', 'AEDECOD'),
                           anyeventvalues = c('ANY EVENT', 'ANY EVENT'),
                           groupbyvarsdenom=c('TRT01AN'),
                           resultstyle="NUMERPCT",
                           totalforvar=c('TRT01AN'),
                           totalid=99,
                           totaldecode='Total',
                           codedecodevarpairs=c("TRT01AN", "TRT01A"),
                           varcodelistpairs=c(""),
                           codelistnames=list(),
                           resultpctdps=0) %>%
  repfun::ru_denorm(varstodenorm=c("tt_result", "PERCENT"),
                     groupbyvars=c("tt_summarylevel", "AEBODSYS", "AEDECOD"),
                     acrossvar="TRT01AN",
                     acrossvarlabel="TRT01A",
                     acrossvarprefix=c("tt_ac", "tt_p"))

#=====
# Demography Statistics: N and Percent
#=====

repfun::rs_setup(D_POP="SAFFL",
                 D_POPLBL="Safety",
                 D_POPDATA=repfun::adsl %>% dplyr::filter(SAFFL =='Y'),
                 D_SUBJID=c("STUDYID", "USUBJID"),
                 R_DICTION=NULL,
                 R_OTHERDATA=NULL,
                 R_INPUTDATA=NULL,
                 R_RAWDATA=NULL,
                 R_SDTMDATA=NULL,
                 R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(TRT01AN=
                ifelse(TRT01A=='Placebo', 1,
                       ifelse(TRT01A=='Xanomeline Low Dose', 2, 3))) %>%

```

```

  repfun::ru_labels(varlabels=list('TRT01AN'='Actual Treatment for Period 01 (n)')
demstats_t <- repfun::ru_sumstats(G_POPDATA,
  analysisvars=c("AGE", "TRTDURD"),
  groupbyvars=c("STUDYID", "TRT01AN"),
  codedecodevarpairs=c("TRT01AN", "TRT01A"),
  totalforvar="TRT01AN", totalid=99,
  totaldecode="Total",
  statsinrowsyn = "Y",
  analysisvardps=list("AGE"=1, "TRTDURD"=2),
  statslist=c("n", "mean", "median", "sd", "min", "max")) %>%
repfun::ru_denorm(varstodenorm=c("tt_result"),
  groupbyvars=c("tt_avid", "tt_avnm", "tt_svid", "tt_svnm"),
  acrossvar="TRT01AN", acrossvarlabel="TRT01A",
  acrossvarprefix=c("tt_ac"))

```

**ru\_expvarlist***Expand SAS Style Variable/Column List***Description**

Pass in a data frame along with column/variable identifiers formatted with SAS Style (i.e., using colon) and it will be expanded to the actual variable list.

**Usage**

```
ru_expvarlist(dsetin, varsin = NULL, keepnotexist = FALSE)
```

**Arguments**

- dsetin** A dataframe holding columns whose names will be expanded.
- varsin** A SAS style list of variable names.
- keepnotexist** If the variable does not exist on the dataframe it will be excluded from the list.

**Value**

A list of column/variable names.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```

library(repfun)
df <- data.frame(tt_ac01=c('1','2','3'),
                  tt_ac02=c('a','b','b'),
                  tt_ac03=c('10','11','12'))
chk <- repfun::ru_expvarlist(df, varsin="tt_ac:")
print(chk)

```

<code>ru_fillcodedcode</code>	<i>Fill missing code/decode records</i>
-------------------------------	---

## Description

Pass in a data frame with along code/decode variables and values to have missings populated.

## Usage

```
ru_fillcodedcode(
  dsetin,
  codedecodevarpairs = NULL,
  varcodelistpairs = NULL,
  codelistnames = list(),
  groupbyvars = NULL,
  completetypes = TRUE
)
```

## Arguments

`dsetin` The data set that will be counted to generate numerators for counts and percents.  
`codedecodevarpairs` Specifies code and decode variable pairs. Those variables should be in parameter GROUPBYVARSNUMER. One variable in the pair will contain the code, which is used in counting and ordering, and the other will contain decode, which is used for presentation.  
`varcodelistpairs` List of code/decode pairs of variables.  
`codelistnames` List of decodes for use with decoding code/decode pairs.  
`groupbyvars` Set of by-variables used to merge the incoming data set with the decode data set.  
`completetypes` Keep all code/decode pairs even it not present on the incoming data set?

## Value

A data frame based on the incoming data frame but with decode values added along with records when `completetypes` is true.

## Author(s)

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
library(repfun)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
```

```
        }
fmtlist <- list('SEXS'=list('START'=list('M','F'),
    'LABEL'=c('Male','Female')))
adsl <- reprefun::adsl
adsl2 <- reprefun::ru_fillcodedcode(adsl, codedecodevarpairs=c("SEX", "SEXDCD"),
    varcodelistpairs=c("SEX", "SEXS"),
    codelistnames=fmtlist)
unique(adsl2[c("SEX", "SEXDCD")])
```

---

**ru\_fillna**

*Fill NA values with specified values or zeros/blanks by default.*

---

**Description**

Pass in a data frame along with a vector of variables and a vector of fill values. (Default fill is 0 for numeric and blank " " for character.)

**Usage**

```
ru_fillna(dsetin, vars = NULL, fills = NULL)
```

**Arguments**

dsetin	Incoming data frame to have labels added to columns.
vars	Vector of variables to replace NA values.
fills	Vector of fill values.

**Value**

The incoming data frame with the requested NA values replaced.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(reprefun)
reprefun::ru_fillna(airquality, vars=c('Ozone', 'Solar.R'), fills=c(1111,2222))
```

---

**ru\_freq***Create Percentage based on Numerator and Denominator Data*

---

**Description**

Pass in a data frame along with identification options and have descriptive statistics derived.

**Usage**

```
ru_freq(
  dsetin,
  dsetindenom = NULL,
  countdistinctvars = NULL,
  groupbyvarsnumer = NULL,
  groupbyvarsdenom = NULL,
  resultstyle = "NUMERPCT",
  totalforvar = NULL,
  totalid = NULL,
  totaldecode = c("Total"),
  anyeventvars = NULL,
  anyeventvalues = NULL,
  codedecodevarpairs = NULL,
  varcodelistpairs = NULL,
  codelistnames = list(),
  groupminmaxvar = NULL,
  resultpctdps = 0
)
```

**Arguments**

<b>dsetin</b>	The data set that will be counted to generate descriptive statistics.
<b>dsetindenom</b>	Input dataset containing data to be counted to obtain the denominator.
<b>countdistinctvars</b>	Variable(s) that contain values to be counted uniquely within any output grouping.
<b>groupbyvarsnumer</b>	Variables in DSETINNUMER to group the data by when counting to obtain the numerator.
<b>groupbyvarsdenom</b>	Variables in DSETINDENOM to group the data by when counting to obtain the denominator.
<b>resultstyle</b>	The appearance style of the result columns that will be displayed in the report.
<b>totalforvar</b>	Variable for which overall totals are required within all other grouped class variables.
<b>totalid</b>	Value(s) used to populate the variable(s) specified in totalforvar.
<b>totaldecode</b>	Value(s) used to populate the variable(s) of the decode variable(s) of the totalforvar.
<b>anyeventvars</b>	Set of variables for which total rows will be added.

**anyeventvalues** Set of text values for total rows generated above.

**codedecodevarpairs**

Specifies code and decode variable pairs. Those variables should be in parameter GROUPBYVARSNUMER. One variable in the pair will contain the code, which is used in counting and ordering, and the other will contain decode, which is used for presentation.

**varcodelistpairs**

List of code/decode pairs of variables.

**codelistnames** List of decodes for use with decoding code/decode pairs.

**groupminmaxvar** Specify if frequency of each group should be from first or last value of a variable in format MIN(variables).

**resultpctdps** The reporting precision for percentages.

## Value

A data frame based on the incoming data frame but collapsed by groups with descriptive statistics added.

## Author(s)

Yongwei Wang, <yongwei.x.wang@viiivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

## Examples

```
library(repfun)
library(dplyr)
library(tibble)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datadir")
dir.create(datadir, showWarnings=FALSE)
outdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "outdir")
dir.create(outdir, showWarnings=FALSE)
fmtdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "fmtdir")
dir.create(fmtdir, showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(D_POPDATA=repfun::ads1 %>% dplyr::filter(SAFFL =='Y'),
                 D_SUBJID=c("STUDYID", "USUBJID"),
                 R_DICTION=NULL,
                 R_OTHERDATA=NULL,
                 R_INPUTDATA=NULL,
                 R_RAWDATA=NULL,
                 R_SDMDATA=NULL,
                 R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(TRT01AN=ifelse(TRT01A=='Placebo', 1,
                                ifelse(TRT01A=='Xanomeline Low Dose', 2, 3)))
attr(G_POPDATA$TRT01AN, "label") <- 'Actual Treatment for Period 01 (n)'
```

```

adae <- rfenv$adamdata$adae.rda() %>%
  dplyr::inner_join(G_POPDATA, by=c('STUDYID', 'USUBJID', 'SAFFL', 'TRT01A'))
aesum <- repfun::ru_freq(adae,
  dsetindenom=G_POPDATA,
  countdistinctvars=c('STUDYID', 'USUBJID'),
  groupbyvarsnumber=c('TRT01AN', 'TRT01A', 'AEBODSYS', 'AEDECOD'),
  anyeventvars = c('AEBODSYS', 'AEDECOD'),
  anyeventvalues = c('ANY EVENT', 'ANY EVENT'),
  groupbyvarsdenom=c('TRT01AN'),
  resultstyle="NUMERPCT",
  totalforvar=c('TRT01AN'),
  totalid=99,
  totaldecode='Total',
  codedecodevarpairs=c("TRT01AN", "TRT01A"),
  varcodelistpairs=c(""),
  codelistnames=list(),
  resultpctdps=0)

```

**ru\_getdata***Assign Big N to Data Frame.***Description**

Merge input data with population data to keep only subjects which are in population sub data,

**Usage**

```

ru_getdata(
  dsetin,
  dsetinpop = rfenv$G_POPDATA,
  subjidvars = c("STUDYID", "USUBJID"),
  subpop = rfenv$G_SUBPOP,
  pop = rfenv$G_POP,
  keeppopvars = rfenv$G_KEEPPOPVARS
)

```

**Arguments**

dsetin	The data set that will be merged with the population data set.
dsetinpop	The population data set.
subjidvars	Variable(s) that define a unique subject.
subpop	A sub-population expression where variables are on DSETINPOP.
pop	The population expression (SAFFL=='Y').
keeppopvars	Variables to keep on the population data set.

**Value**

A data frame based on the incoming data frame but restricted to the population of interest with relevant population variables added.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viiivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
library(dplyr)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datdir")
dir.create(datadir, showWarnings=FALSE)
outdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "outdir")
dir.create(outdir, showWarnings=FALSE)
fmtdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "fmtdir")
dir.create(fmtdir, showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(D_POP="SAFFL",
                 D_POPLBL="Safety",
                 D_POPDATA=repfun::ads1,
                 D_SUBJID=c("STUDYID", "USUBJID"),
                 R_DICTION=NULL,
                 R_OTHERDATA=NULL,
                 R_INPUTDATA=NULL,
                 R_RAWDATA=NULL,
                 R_SDATMDATA=NULL,
                 R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(TRT01AN=ifelse(TRT01A=='Placebo', 1,
                               ifelse(TRT01A=='Xanomeline Low Dose', 2, 3)),
                SAFFL=ifelse((row_number() %% 10) == 0, 'N', SAFFL))
attr(G_POPDATA$TRT01AN, "label") <- 'Actual Treatment for Period 01 (n)'
attr(G_POPDATA$SAFFL, "label") <- 'Safety Population Flag'
adae <- rfenv$adamdata$adae.rda() %>% dplyr::select(-SAFFL)
adae2 <- repfun::ru_getdata(adae, G_POPDATA, c("STUDYID", "USUBJID"),
                            keeppopvars=c("TRT01AN", "TRT01A"))
```

ru\_groupbyvars

*Modify groupbyvars by adding or removing decode vars.***Description**

Pass in a vector of group-by variables along with a vector of code/decode pairs to have decode variables added or removed.

**Usage**

```
ru_groupbyvars(groupbyvars, codedecodevarpairs, adddecode = TRUE)
```

**Arguments**

- `groupbyvars` Vector of group-by variables.  
`codedecodevarpairs` Specifies code and decode variable pairs. Those variables should be in parameter GROUPBYVARSNUMER. One variable in the pair will contain the code, which is used in counting and ordering, and the other will contain decode, which is used for presentation.  
`adddecode` Add decode variables (true) or remove (false).

**Value**

A data frame based on the incoming data frame but collapsed by groups with counts and percents added.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
add_decode <- repfun::ru_groupbyvars(
  c("TRTCD", "TRTGRP", "ATOXGRN", "AEDECOD", "AEBODSYS"),
  c("TRTCD", "TRTGRP", "ATOXGRN", "ATOXGR"), TRUE)
rem_decode <- repfun::ru_groupbyvars(
  c("TRTCD", "TRTGRP", "ATOXGRN", "AEDECOD", "AEBODSYS"),
  c("TRTCD", "TRTGRP", "ATOXGRN", "ATOXGR"), FALSE)
```

**ru\_labels**

*Assign labels to variables in a data frame*

**Description**

Pass in a data frame along with a named list of columns with their corresponding labels.

**Usage**

```
ru_labels(dsetin, varlabels = list(), style = c("base", "Hmisc"))
```

**Arguments**

- `dsetin` Incoming data frame to have labels added to columns.  
`varlabels` List of variables and their labels.  
`style` Type of method used to add labels.

**Value**

The incoming data frame with labels added.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
ru_labels(mtcars, varlabels=list(mpg='Miles per gallon',
                                 cyl='Number of cylinders'))
```

---

ru\_libname

*Automate retrieval of data sets/frames in specified folder.*

---

**Description**

Provide a folder containing data sets/frames and have a list of function calls returned that can be used to quickly access individual data sets/frames.

**Usage**

```
ru_libname(datapath)
```

**Arguments**

datapath      Location of reporting data sets.

**Value**

List of function calls for use in quickly accessing individual data sets/frames.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datadir")
dir.create(datadir, showWarnings=FALSE)
repfun::copydata(datadir)
adamdata <- repfun::ru_libname(datadir)
adamdata$adae.rda() %>% dplyr::filter(TRTEMFL=='Y') -> teae
print(head(teae[,c(1:10)]),10)
```

---

**ru\_list***R function to mimic the SAS macro %tu\_list.*

---

**Description**

Pass in a dataframe and reporting settings to have RTF output generated.

**Usage**

```
ru_list(
  dsetin,
  stackvar1 = NULL,
  stackvar2 = NULL,
  stackvar3 = NULL,
  stackvar4 = NULL,
  stackvar5 = NULL,
  stackvar6 = NULL,
  stackvar7 = NULL,
  stackvar8 = NULL,
  stackvar9 = NULL,
  stackvar10 = NULL,
  stackvar11 = NULL,
  stackvar12 = NULL,
  stackvar13 = NULL,
  stackvar14 = NULL,
  stackvar15 = NULL,
  display = "Y",
  varlabelstyle = "NOT IMPLEMENTED",
  dddatasetlabel = NULL,
  splitchar = "\n",
  getdatayn = "N",
  labelvarsyn = NULL,
  computebeforepagelines = NULL,
  computebeforepagevars = NULL,
  columns = NULL,
  ordervars = NULL,
  descending = NULL,
  orderformatted = "NOT IMPLEMENTED",
  orderfreq = "NOT IMPLEMENTED",
  orderdata = NULL,
  noprintvars = NULL,
  byvars = NULL,
  flowvars = "NOT IMPLEMENTED",
  widths = NULL,
  defaultwidths = "NOT IMPLEMENTED",
  skipvars = NULL,
  pagevars = NULL,
  idvars = NULL,
  linevars = NULL,
  centrevars = NULL,
  leftvars = NULL,
```

```

    rightvars = NULL,
    colspacing = 2,
    varspacing = "NOT IMPLEMENTED",
    formats = "NOT IMPLEMENTED",
    labels = NULL,
    break1 = "NOT IMPLEMENTED",
    break2 = "NOT IMPLEMENTED",
    break3 = "NOT IMPLEMENTED",
    break4 = "NOT IMPLEMENTED",
    break5 = "NOT IMPLEMENTED",
    nowidowvar = NULL,
    sharecolvars = NULL,
    sharecolvarsindent = 2,
    overallsummary = "n",
    proptions = "HEADLINE",
    denormyn = "N",
    varsToDenorm = NULL,
    groupByVars = NULL,
    acrossVar = NULL,
    acrossVarLabel = NULL,
    acrossColVarPrefix = NULL,
    acrossVarListName = NULL,
    lpp = 24,
    rpp = 50,
    toprow = "single",
    spanlbls = NULL,
    spanwidths = NULL,
    spanjust = NULL,
    spanbbord = NULL,
    spantbord = NULL,
    span2lbls = NULL,
    span2widths = NULL,
    span2just = NULL,
    span2bbord = NULL,
    xptyn = "N"
)

```

## Arguments

dsetin	Incoming data frame or list of data frames.
stackvar1	Create Stacked variables (e.g. stackvar1=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
stackvar2	Create Stacked variables (e.g. stackvar2=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
stackvar3	Create Stacked variables (e.g. stackvar3=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
stackvar4	Create Stacked variables (e.g. stackvar4=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
stackvar5	Create Stacked variables (e.g. stackvar5=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
stackvar6	Create Stacked variables (e.g. stackvar6=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))

<b>stackvar7</b>	Create Stacked variables (e.g. stackvar7=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar8</b>	Create Stacked variables (e.g. stackvar8=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar9</b>	Create Stacked variables (e.g. stackvar9=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar10</b>	Create Stacked variables (e.g. stackvar10=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar11</b>	Create Stacked variables (e.g. stackvar11=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar12</b>	Create Stacked variables (e.g. stackvar12=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar13</b>	Create Stacked variables (e.g. stackvar13=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar14</b>	Create Stacked variables (e.g. stackvar14=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>stackvar15</b>	Create Stacked variables (e.g. stackvar15=list(varsin=c('invid','subjid'), varout='st_inv_subj', sepc='/', splitc='\n'))
<b>display</b>	Specifies whether the report should be created
<b>varlabelstyle</b>	Specifies the label style for variables (SHORT or STD)
<b>dddatasetlabel</b>	Label to be applied to the DD dataset
<b>splitchar</b>	Split character
<b>getdatayn</b>	Control execution of tu_getdata
<b>labelvarsyn</b>	Control execution of tu_labelvars
<b>computebeforepagelines</b>	Specifies the text to be produced for the Compute Before Page lines (labelkey labelfmt colon labelvar)
<b>computebeforepagevars</b>	Names of variables that shall define the sort order for Compute Before Page lines
<b>columns</b>	Column parameter
<b>ordervars</b>	Order variables
<b>descending</b>	Descending ORDERVARS
<b>orderformatted</b>	ORDER=FORMATTED variables
<b>orderfreq</b>	ORDER=FREQ variables
<b>orderdata</b>	ORDER=DATA variables
<b>noprintvars</b>	No print vars (usually used to order the display)
<b>byvars</b>	By variables
<b>flowvars</b>	Variables with flow option
<b>widths</b>	Column widths
<b>defaultwidths</b>	List of default column widths
<b>skipvars</b>	similar to SAS statement skipvars Break after / skip
<b>pagevars</b>	similar to SAS statement pagevars Break after / page

idvars	ID variables
linevars	Order variable printed with line statements.
centrevars	Centre justify variables
leftvars	Left justify variables
rightvars	Right justify variables
colspacing	Overall spacing value.
varspacing	Spacing for individual variables.
formats	Format specification
labels	Label definitions.
break1	Break statements.
break2	Break statements.
break3	Break statements.
break4	Break statements.
break5	Break statements.
nowidowvar	Not in version 1
sharecolvars	Order variables that share print space.
sharecolvarsindent	Indentation factor
overallsummary	Overall summary line at top of tables
proptions	PROC REPORT statement options
denormyn	Controls whether denormalisation will occur
varsToDenorm	List of variables to be denormalised/transposed. Passed one at a time to the PROC TRANSPOSE VAR statement.
groupByVars	List of BY variables passed to PROC TRANSPOSE BY statement.
acrossVar	Variable used in the PROC TRANSPOSE ID statement.
acrossVarLabel	Variable used in the PROC TRANSPOSE IDLABEL statement.
acrossColVarPrefix	Text passed to the PROC TRANSPOSE PREFIX statement.
acrossVarListName	Macro variable name to contain the list of columns created by the transpose of the first variable in VARSTODENORM.
lpp	Lines within body of report (only used with manual paging).
rpp	Total lines per page, when there is no wrapping and excluding titles and footnotes - passed directly to r2rtf().
toprow	Control lines above first column header.
spanlbls	List of level 1 column spanning header labels.
spanwidths	List of level 1 column spanning header widths.
spanjust	List of level 1 column spanning header justifications.
spanbbord	List of level 1 column spanning bottom border values.
spantbord	List of level 1 column spanning top border values.
span2lbls	List of level 2 column spanning header labels (above Level 1).
span2widths	List of level 2 column spanning header widths (above Level 1).
span2just	List of level 2 column spanning header justifications (above Level 1).
span2bbord	List of level 2 column spanning bottom border values (above Level 1).
xptyn	Write DDDATA data frame as XPT file in addition to RDS?

**Value**

A formattted RTF report is generated to the specified file.

**Author(s)**

Chris Rook, <christopher.x.rook@gsk.com>  
 Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>

**Examples**

```

library(repfun)
library(dplyr)
library(tibble)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datdir")
dir.create(datdir, showWarnings=FALSE)
outdir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "outdir")
dir.create(outdir, showWarnings=FALSE)
#=====
# Set up the reporting environment.
#=====
setup <- function(tlfid){
  repfun::rs_setup(
    D_DATADATE=Sys.Date(),
    D_DSPLYNUM=tlfid,
    D_FOOT1='1.) Only treatment emergent events related to lipids are displayed.',
    D_FOOT2='2.) Subjects counted once in each body system & preferred term.',
    D_KEEPPOPVARS=c('STUDYID', 'USUBJID', 'SAFFL'),
    D_STUDYID='ABCXYZPDQ',
    D_POP="SAFFL",
    D_POPDATA=repfun::adsl %>%
      dplyr::filter(SAFFL =='Y') %>%
      dplyr::mutate(TRT01AN=ifelse(TRT01A=='Placebo', 1,
                                    ifelse(TRT01A=='Xanomeline Low Dose', 2, 3))) %>%
      repfun::ru_labels(varlabels=
        list('TRT01AN'='Actual Treatment for Period 01 (n)'),
        D_POPLBL="Safety",
        D_SUBJID=c("STUDYID", "USUBJID"),
        D_TITLE1=paste0('Table ', tlfid, ': Summary of Treatment Emergent Adverse Events'),
        D_OUTFILE=paste0(outdir, "/t_ru_list_", tlfid, ".rtf"),
        D_PGMPTH="/path/to/code/ru_list.R",
        R_DDDATA=paste0(outdir, '/t_ru_list_', tlfid, '.rds'),
        R_ADAMDATA=datdir)
  )
  #=====
  # Process ADAE - derive counts and percents.
  #=====
  setup(1)
  aenum <- repfun::ru_freq(rfenv$adamdata$adae.rda() %>% dplyr::select(-SAFFL) %>%
```

```

    repfun::ru_getdata(rfenv$G_POPDATA, c("STUDYID", "USUBJID"),
    keeppopvars=c("TRT01AN", "TRT01A")),
    dsetindenom=rfenv$G_POPDATA,
    countdistinctvars=c('STUDYID', 'USUBJID'),
    groupbyvarsnumer=c('TRT01AN', 'TRT01A', 'AEBODSYS', 'AEDECOD'),
    anyeventvars = c('AEBODSYS', 'AEDECOD'),
    anyeventvalues = c('ANY EVENT', 'ANY EVENT'),
    groupbyvarsdenom=c('TRT01AN'),
    resultstyle="NUMERPCT",
    totalforvar=c('TRT01AN'),
    totalid=99,
    totaldecode='Total',
    codedecodevarpairs=c("TRT01AN", "TRT01A"),
    varcodelistpairs=c(""),
    codelistnames=list(),
    resultpctdps=0) %>%
dplyr::arrange(TRT01AN, TRT01A, AEBODSYS, tt_summarylevel, AEDECOD, NUMRCNT, DENOMCNT) %>%
repfun::ru_align("tt_result")

#=====
# Table 1: Summary of Adverse Events
#=====

repfun::ru_list(aesum,
  columns=c('AEBODSYS', 'AEDECOD', 'tt_01', 'tt_02', 'tt_03', 'tt_99'),
  nowidowvar='AEBODSYS',
  widths=c(5.5, 4.5, 1.75, 1.9, 1.9, 1.75),
  skipvars=c('AEBODSYS'),
  centrevars=c('tt_01', 'tt_02', 'tt_03', 'tt_99'),
  ordervars=c('AEBODSYS', 'tt_summarylevel', 'AEDECOD'),
  noprintvars=c('tt_summarylevel'),
  denormyn='Y',
  varsToDenorm=c('tt_result'),
  groupByVars=c('AEBODSYS', 'tt_summarylevel', 'AEDECOD'),
  acrossVar="TRT01AN",
  acrossVarLabel="TRT01A",
  acrossColVarPrefix='tt_',
  dddatasetlabel=paste0('DD Dataframe for AE Table ', rfenv$G_DSPLYNUM),
  lpp=23)

#=====
# Table 2: Summary of Adverse Events using NOWIDOWVAR (remove SOCs that
# will not fit on 1 page with 10pt font)
#=====

setup(2)
SOCterms <- aesum %>% dplyr::distinct(AEBODSYS, AEDECOD)
SOCcnts <- table(SOCterms$AEBODSYS)
repfun::ru_list(aesum %>% dplyr::filter(!(AEBODSYS %in% names(SOCcnts[SOCcnts>=20]))),
  columns=c('AEBODSYS', 'AEDECOD', 'tt_01', 'tt_02', 'tt_03', 'tt_99'),
  nowidowvar='AEBODSYS',
  widths=c(5.5, 4.5, 1.75, 1.9, 1.9, 1.75),
  skipvars=c('AEBODSYS'),
  centrevars=c('tt_01', 'tt_02', 'tt_03', 'tt_99'),
  ordervars=c('AEBODSYS', 'tt_summarylevel', 'AEDECOD'),
  noprintvars=c('tt_summarylevel'),
  denormyn='Y',
  varsToDenorm=c('tt_result'),
  groupByVars=c('AEBODSYS', 'tt_summarylevel', 'AEDECOD'),
  
```

```
acrossVar="TRT01AN",
acrossVarLabel="TRT01A",
acrossColVarPrefix='tt_',
dddatasetlabel=paste0('DD Dataframe for AE Table ',rfenv$G_DSPLNUM),
lpp=24)
```

**ru\_load\_library**      *Load a list of libraries*

### Description

Given a list of packages, check if installed and generate message, otherwise load package.

### Usage

```
ru_load_library(pkgs)
```

### Arguments

**pkgs**      A list of packages to check if installed and then load.

### Value

No return value, packages are loaded.

### Author(s)

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

### Examples

```
library(repfun)
repfun::ru_load_library(c("dplyr", "haven", "magrittr", "r2rtf"))
```

**ru\_setdata**      *Append data sets even when variables do not match*

### Description

Pass in a collection of data frames separated by commas and they will be appended.

### Usage

```
ru_setdata(..., keeprownames = TRUE)
```

### Arguments

**...**      A collection of data frames.

**keeprownames**      Convert row names on data frame to columns and keep the column.

**Value**

The incoming data frames combined.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
repfun::ru_setdata(mtcars,airquality)
```

---

ru\_stackvar

*Stack Columns of a Dataframe into New Column*

---

**Description**

Pass in a dataframe and columns to stack and have new dataframe returned that contains the stacked columns.

**Usage**

```
ru_stackvar(
  dsetin,
  sepc = "/",
  splitc = "\\\n",
  varsin = NULL,
  varout = NULL,
  varlabel = NULL
)
```

**Arguments**

dsetin	Name of incoming dataframe with columns to have stacked.
sepc	Separator character between the stacked columns.
splitc	Split character between stacked columns.
varsin	List of variables to be stacked.
varout	Name of stacked column in dataframe.
varlabel	Label for new stacked column.

**Value**

The incoming dataframe with columns stacked as requested.

**Author(s)**

Chris Rook, <christopher.x.rook@gsk.com>  
Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>

## Examples

```

library(repfun)
library(dplyr)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datdir")
dir.create(datadir, showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(D_POPDATA=repfun::ads1 %>% dplyr::filter(SAFFL =='Y'),
                 D_SUBJID=c("STUDYID", "USUBJID"),
                 R_DICTION=NULL,
                 R_OTHERDATA=NULL,
                 R_INPUTDATA=NULL,
                 R_RAWDATA=NULL,
                 R_SDATMDATA=NULL,
                 R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(TRT01AN=ifelse(TRT01A=='Placebo', 1,
                               ifelse(TRT01A=='Xanomeline Low Dose', 2, 3))) %>%
  repfun::ru_labels(varlabels=list('TRT01AN'='Actual Treatment for Period 01 (n)'))
adae <- rfenv$adamdata$adae.rda() %>%
  dplyr::inner_join(G_POPDATA, by=c('STUDYID', 'USUBJID', 'SAFFL', 'TRT01A'))
aesum_p <- repfun::ru_freq(adae,
                            dsetindenom=G_POPDATA,
                            countdistinctvars=c('STUDYID', 'USUBJID'),
                            groupbyvarsnumer=c('TRT01AN', 'TRT01A', 'AEBODSYS', 'AEDECOD'),
                            anyeventvars = c('AEBODSYS', 'AEDECOD'),
                            anyeventvalues = c('ANY EVENT', 'ANY EVENT'),
                            groupbyvarsdenom=c('TRT01AN'),
                            resultstyle="NUMERPCT",
                            totalforvar=c('TRT01AN'),
                            totalid=99,
                            totaldecode='Total',
                            codedecodevarpairs=c("TRT01AN", "TRT01A"),
                            varcodelistpairs=c(""),
                            codelistnames=list(),
                            resultpctdps=0) %>%
  repfun::ru_denorm(varstodenorm=c("tt_result", "PERCENT"),
                     groupbyvars=c("tt_summarylevel", "AEBODSYS", "AEDECOD"),
                     acrossvar="TRT01AN", acrossvarlabel="TRT01A",
                     acrossvarprefix=c("tt_ac", "tt_p")) %>%
  dplyr::mutate(ord1=ifelse(tt_summarylevel==0, 0, 1)) %>%
  dplyr::rename(ord2=tt_summarylevel) %>%
  dplyr::arrange(ord1, AEBODSYS, ord2, AEDECOD) %>%
  dplyr::select(-c(starts_with('tt_p'), starts_with('ord'))) %>%
  repfun::ru_stackvar(varsin=c('AEBODSYS', 'AEDECOD'), varout='SYSPREF',
                      varlabel='Body System/Preferred Term')

```

---

<code>ru_sumstats</code>	<i>Calculate Descriptive Statistics</i>
--------------------------	---

---

## Description

Pass in a data frame along with identification options and have descriptive statistics derived.

## Usage

```
ru_sumstats(
  dsetin,
  analysisvars = NULL,
  analysisvarlabels = "",
  groupbyvars = NULL,
  statslist = c("n", "mean", "median", "min", "max", "sd", "q1", "q3"),
  statsinrowsyn = "N",
  analysisvardps = 0,
  statsdps = list(mean = 1, median = 1, sd = 2, se = 2),
  codedecodevarpairs = c(""),
  varcodelistpairs = c(""),
  codelistnames = list(),
  totalforvar = NULL,
  totalid = NULL,
  totaldecode = c("Total")
)
```

## Arguments

- dsetin** The data set that will be counted to generate descriptive statistics.
- analysisvars** The variables to be analysed.
- analysisvarlabels** Specify a label statement which will be used to define labels for statistics analysis variables defined in parameter ANALYSISVARS.
- groupbyvars** Specifies the variables whose values define the subgroup combinations for the analysis. The variables can be divided by statements inside of ( and ) to represent different levels of subgroup.
- statslist** Specifies a list of summary statistics to be produced.
- statsinrowsyn** Place resulting descriptive statistics in rows or columns.
- analysisvardps** Base precision of descriptive statistics prior to incorporating STATSDPS details.
- statsdps** List of additional statistic-specific precision values to add to ANALYSISVARDPS.
- codedecodevarpairs** Specifies code and decode variable pairs. Those variables should be in parameter GROUPBYVARSNUMER. One variable in the pair will contain the code, which is used in counting and ordering, and the other will contain decode, which is used for presentation.
- varcodelistpairs** List of code/decode pairs of variables.
- codelistnames** List of decodes for use with decoding code/decode pairs.

<b>totalforvar</b>	Variable for which overall totals are required within all other grouped class variables.
<b>totalid</b>	Value(s) used to populate the variable(s) specified in totalforvar.
<b>totaldecode</b>	Value(s) used to populate the variable(s) of the decode variable(s) of the totalforvar.

**Value**

A data frame based on the incoming data frame but collapsed by groups with descriptive statistics added.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
 Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
library(dplyr)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datadir");
dir.create(datadir, showWarnings=FALSE)
repfun::copydata(datadir)
rs_setup(D_POPDATA=repfun::adsl %>% dplyr::filter(SAFFL =='Y'),
         D_SUBJID=c("STUDYID", "USUBJID"),
         R_DICTION=NULL,
         R_OTHERDATA=NULL,
         R_INPUTDATA=NULL,
         R_RAWDATA=NULL,
         R_SDMDATA=NULL,
         R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>% dplyr::mutate(
  TRT01AN=ifelse(TRT01A=='Placebo',1,
                  ifelse(TRT01A=='Xanomeline Low Dose',2,3))) %>%
  ru_labels(varlabels=list('TRT01AN'='Actual Treatment for Period 01 (n)'))
ru_sumstats(G_POPDATA,
            analysisvars=c("AGE", "TRTDURD"),
            groupbyvars=c("STUDYID", "TRT01AN"),
            codedecodevarpairs=c("TRT01AN", "TRT01A"),
            totalforvar="TRT01AN", totalid=99,
            totaldecode="Total",
            statsinrowsyn = "Y",
            analysisvardps=list("AGE"=1, "TRTDURD"=2),
            statslist=c("n", "mean", "median", "sd", "min", "max"))
```

---

**ru\_width\_rtf***Create a List of Relative Widths of Columns for RTF Outputs*

---

**Description**

Pass in a data set and identify the columns for reporting to have estimated relative column width returned.

**Usage**

```
ru_width_rtf(dsetin, varsin = list(), widths = list(), type = "PCT")
```

**Arguments**

dsetin	A dataframe containing columns for reporting.
varsin	A vector of variables on the reporting dataframe that will be displayed in the output.
widths	Provide a set of default widths for some or all variables as desired. (These will be used.)
type	Specify the type of width to be computed and returned. (Currently supports PCT, which refers to relative widths not percentages.)

**Value**

A list of widths of the same size as the list specified by varsin.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
library(repfun)
library(dplyr)
rfenv <- if (exists('rfenv') && is.environment(get('rfenv'))){
  rfenv
} else {
  rfenv <- new.env(parent = emptyenv())
  rfenv$G_DEBUG <- 0
  rfenv
}
datadir <- file.path(gsub("\\\\", "/", tempdir(), fixed=TRUE), "datdir");
dir.create(datadir, showWarnings=FALSE)
repfun::copydata(datadir)
repfun::rs_setup(D_POPDATA=repfun::ads1 %>% dplyr::filter(SAFFL =='Y'),
  D_SUBJID=c("STUDYID", "USUBJID"),
  R_DICTION=NULL,
  R_OTHERDATA=NULL,
  R_INPUTDATA=NULL,
  R_RAWDATA=NULL,
  R_SDTMDATA=NULL,
```

```
R_ADAMDATA=datadir)
G_POPDATA <- rfenv$G_POPDATA %>%
  dplyr::mutate(
    TRT01AN=ifelse(TRT01A=='Placebo',1,
                   ifelse(TRT01A=="Xanomeline Low Dose",2,3))) %>%
  repfun::ru_labels(varlabels=list('TRT01AN'='Actual Treatment for Period 01 (n)'))
adae <- rfenv$adamdata$adae.rda() %>%
  dplyr::inner_join(G_POPDATA, by=c('STUDYID','USUBJID','SAFFL','TRT01A'))
aesum_p <- repfun::ru_freq(adae,
                           dsetindenom=G_POPDATA,
                           countdistinctvars=c('STUDYID','USUBJID'),
                           groupbyvarsnum=c('TRT01AN','TRT01A','AEBODSYS','AEDECOD'),
                           anyeventvars = c('AEBODSYS','AEDECOD'),
                           anyeventvalues = c('ANY EVENT','ANY EVENT'),
                           groupbyvarsdenom=c('TRT01AN'),
                           resultstyle="NUMERPCT",
                           totalforvar=c('TRT01AN'),
                           totalid=99,
                           totaldecode='Total',
                           codedecodevarpairs=c("TRT01AN", "TRT01A"),
                           varcodelistpairs=c(""),
                           codelistnames=list(),
                           resultpctdps=0) %>%
  repfun::ru_denorm(varstodenorm=c("tt_result", "PERCENT"),
                     groupbyvars=c("tt_summarylevel", "AEBODSYS", "AEDECOD"),
                     acrossvar="TRT01AN", acrossvarlabel="TRT01A",
                     acrossvarprefix=c("tt_ac", "tt_p")) %>%
  dplyr::mutate(ord1=ifelse(tt_summarylevel==0,0,1)) %>%
  dplyr::rename(ord2=tt_summarylevel) %>%
  dplyr::arrange(ord1,AEBODSYS,ord2,AEDECOD) %>%
  dplyr::select(-c(starts_with('tt_p'),starts_with('ord'))) %>%
  repfun::ru_addpage(grpvars=c('AEBODSYS'),rowsprbdy=35,nosplitvars=TRUE)

widths1 <- repfun::ru_width_rtf(aesum_p,
                                 c('AEBODSYS','AEDECOD','tt_ac01','tt_ac02','tt_ac03','tt_ac99'))
print(widths1)
widths2 <- repfun::ru_width_rtf(aesum_p,
                                 c('AEBODSYS','AEDECOD','tt_ac01','tt_ac02','tt_ac03','tt_ac99'),
                                 list('AEBODSYS'=35, 'AEDECOD'=30))
print(widths2)
```

**setpath***Set the current working directory.***Description**

Set the current working directory so that relative paths work as expected.

**Usage**

```
setpath(p)
```

**Arguments**

p	A path as a string.
---	---------------------

**Value**

No return value, the current working directory is set.

**Author(s)**

Yongwei Wang, <yongwei.x.wang@viivhealthcare.com>  
Chris Rook, <christopher.x.rook@gsk.com>

**Examples**

```
## Not run:  
library(repfun)  
repfun::setpath()  
  
## End(Not run)
```

---

suppae*Supplemental Adverse Events*

---

**Description**

A SDTM SUPPAE dataset from the CDISC pilot project

**Usage**

suppae

**Format**

A data frame with 10 columns:

**STUDYID** Study Identifier  
**RDOMAIN** Related Domain Abbreviation  
**USUBJID** Unique Subject Identifier  
**IDVAR** Identifying Variable  
**IDVARVAL** Identifying Variable Value  
**QNAM** Qualifier Variable Name  
**QLABEL** Qualifier Variable Label  
**QVAL** Data Value  
**QORIG** Origin  
**QEVAL** Evaluator

**Details**

Supplemental Adverse Events

A SDTM SUPPAE dataset from the CDISC pilot project

**Source**

Access the source of the Supplemental Adverse Events dataset.

---

suppdm                    *Supplemental Demography*

---

### Description

A SDTM SUPPDM dataset from the CDISC pilot project

### Usage

suppdm

### Format

A data frame with 10 columns:

**STUDYID** Study Identifier

**RDOMAIN** Related Domain Abbreviation

**USUBJID** Unique Subject Identifier

**IDVAR** Identifying Variable

**IDVARVAL** Identifying Variable Value

**QNAM** Qualifier Variable Name

**QLABEL** Qualifier Variable Label

**QVAL** Data Value

**QORIG** Origin

**QEVAL** Evaluator

### Details

Supplemental Demography

A SDTM SUPPDM dataset from the CDISC pilot project

### Source

Generated dataset.

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