Submitted to bioRxiv April 2021

Cellular and molecular mechanisms involved in LTP induced by mild theta-burst stimulation in hippocampal slices from young male rats: from weaning to adulthood.

Rodrigues NC[1], Silva-Cruz A[1], Caulino-Rocha A[3,4], Bento-Oliveira A[3,4], Ribeiro JA[1,2], Cunha-Reis D[1,3,4]

[1]Unidade de Neurociências, Instituto de Medicina Molecular e [2]Instituto de Farmacologia e Neurociências, Faculdade de Medicina, Universidade de Lisboa, Av. Prof. Egas Moniz, 1649-028 Lisboa, Portugal. [3]Departamento de Química e Bioquímica and [4]BioISI - Biosystems & Integrative Sciences Institute, Faculdade de Ciências, Universidade de Lisboa, Campo Grande, 1749-016 Lisboa, Portugal.

Abbreviated title: Transduction mechanisms involved in LTP expression in early life.

Keywords: theta-burst, LTP, CaMKII, PKA, AMPA Rs, Kv4.2, $GABA_A$ receptor, $GABA_B$ receptor, hippocampus.

*Corresponding author – current address: Departamento de Química e Bioquímica, Faculdade de Ciências da Universidade de Lisboa, Campo Grande, 1749-016 Lisboa, Portugal. dcreis@ciencias.ulisboa.pt

Number of pages: 39

Number of figures: 6

Conflict of interests: The authors have no conflict of interests to publication of this paper.

NC Rodrigues: formal analysis and methodology; ***A Silva-Cruz:*** formal analysis and methodology; ***A Caulino-Rocha:*** formal analysis and writing - review and editing; ***A Bento-Oliveira:*** methodology; ***JA Ribeiro:*** resources, supervision and writing – review and editing and ***D Cunha-Reis:*** formal analysis and methodology, resources, supervision, funding acquisition, project administration, and writing – original draft, review and editing.

Funding: This work was supported national and international funding managed by Fundação para a Ciência e a Tecnologia (FCT, IP), Portugal. **Grants:** UIDB/04046/2020 and UIDP/04046/2020 centre grants (to BioISI) and research grants PTDC/SAU-NEU/103639/2008 and FCT/POCTI (PTDC/SAUPUB/28311/2017) EPIRaft grant (to DC-R). **Fellowships:** SFRH/BPD/34661/2007 and SFRH/BPD/81358/2011 to DCR and **Researcher contract:** Norma Transitória - DL57/2016/CP1479/CT0044 to DCR. Funding sources made no contribution to the writing, research plan and decision to publish this paper.

Acknowledgements: We acknowledge the Institute of Physiology, FMUL, for animal housing facilities.

Abstract:

Long-term potentiation (LTP) is a highly studied phenomenon yet the essential vs. modulatory transduction and GABAergic pathways involved in LTP elicited by theta-burst stimulation (TBS) in the CA1 area of the hippocampus are still unclear, due to the use of different TBS intensities and patterns or of different rodent/cellular models. We now characterized the essential transduction and GABAergic pathways in mild TBS-induced LTP in the CA1 area of the rat hippocampus. LTP induced by TBS (5x4) (5 bursts of 4 pulses delivered at 100Hz) lasted for up to 3h and was increasingly greater from weaning to adulthood. Stronger TBS patterns - TBS (15x4) or three TBS (15x4) separated by 6 min induced nearly maximal LTP not being the best choice to study the value of LTP-enhancing drugs. LTP induced by TBS (5x4) was fully dependent on NMDA receptor and CaMKII activity but independent on PKA or PKC activity. In addition, it was partially dependent on $GABA_B$ receptor activation and was potentiated by $GABA_A$ receptor blockade and less by GAT-1 transporter blockade. AMPA GluA1 phosphorylation on Ser_{831} (CaMKII target) but not GluA1 Ser_{845} (PKA target) was essential for LTP expression. The phosphorylation of the Kv4.2 channel was observed at Ser438 (targeted by CaMKII) but not at Thr_{602} or Thr_{607} (ERK/MAPK pathway). This suggests that cellular kinases like PKA, PKC or kinases of the ERK/MAPK family although important modulators of TBS (5x4)-induced LTP are not essential for its expression in the CA1 area of the hippocampus.

Keywords: theta burst LTP, CaMKII, PKA, AMPA Rs, Kv4.2, $GABA_A$ receptor, $GABA_B$ receptor, hippocampus.

1. Introduction:

Synaptic plasticity generated by activity dependent changes in the strength of synaptic communication, is widely accepted as the cellular mechanism underlying memory storage (Bliss & Collingridge, 2019). Long-term potentiation (LTP) evoked by high-frequency stimulation (HFS) was first described in 1973 in the contacts of perforant path fibres to granule cells in the dentate gyrus, and later in the remaining excitatory pathways of the hippocampus and was the first synaptic plasticity mode to be associated with hippocampal dependent memory formation (Bliss & Collingridge, 1993; Lynch *et al.*, 2007). The stability or long-lasting expression of LTP is dependent on the subsequent activation of multiple intracellular cascades that are differentially activated by various patterns of electrical activity *in vivo.* The magnitude and stability of LTP evaluated *in vitro* depends on several characteristics of the stimulus used to induce LTP such as frequency, intensity and pattern (Albensi *et al.*, 2007; Baudry *et al.*, 2015). Notwithstanding, intracellular cascades are differently activated in different rodent species and strains and at different animal developmental stages, making it difficult to set the stage for studies on the action of endogenous modulators and pharmacological tools on synaptic plasticity.

LTP can be associated to three mechanistically different processes (Park *et al.*, 2018). Early-LTP can be triggered by a single episode of HFS, such as a tetanus or theta burst, requires activation of Ca^{2+}/calmodulin-dependent protein kinase II (CaMKII) but is independent of both protein kinase A (PKA) and protein synthesis. Considerable evidence implicates an increase in the number of AMPA receptors (AMPARs) inserted into the postsynaptic membrane in early-LTP expression (Park *et al.*, 2018). Late-LTP normally requires multiple episodes of HFS such as a tetani or theta burst for its induction; and these episodes need to be appropriately spaced in time (normally 10 min intervals). These protocols induce first an early-LTP but then further mechanisms are activated, and the resulting potentiation thus involves both early and late-LTP. Late-LTP lasts more than 3h *in vitro*, is likely associated with synaptic contact enlargement and requires activation of PKA and *de novo* protein

synthesis (Park *et al.*, 2018). Longer-lasting forms of LTP require additional gene transcription and will not be addressed in this paper.

Theta burst stimulation (TBS) (Larson & Lynch, 1986; Rose & Dunwiddie, 1986) is a sequence of electrical stimuli that mimics CA3 and CA1 pyramidal complex-spike cell discharges observed during the hippocampal theta rhythm (3-7Hz). This EEG pattern has been associated with hippocampal memory storage and is believed to serve as a 'tag' for short term memory processing and to activate mechanisms eliciting early LTP (Vertes, 2005; Larson & Munkácsy, 2015). Bursts repeated at the theta frequency induce maximal LTP due to the suppression of feedforward inhibition by the first burst or a priming single pulse, that allows for enough depolarization to activate NMDA receptors (Larson & Lynch, 1988; Davies *et al.*, 1990). This is mediated by activation of $GABA_B$ autoreceptors that strongly inhibit GABA release from feedforward interneurons (Cobb *et al.*, 1999) and synaptic GABA availability. For strong TBS trains the activation of presynaptic L-type Ca^{2+} channels also contributes to LTP induction (Morgan *et al.*, 2001), but such intense stimulation patterns have been argued not to be required *in vivo* to induce maximal LTP (Larson & Munkácsy, 2015).

LTP induced by TBS is dependent on the elevation of dendritic Ca^{2+} resulting in the Ca^{2+}-dependent activation and consequent auto-phosphorylation of CaMKII (Appleby *et al.*, 2011; Kim *et al.*, 2016). This results in the phosphorylation of AMPA receptor GluA1 subunits, that in turn promotes an enhancement of the channel conductance together with its recruitment to the active zone (Derkach *et al.*, 1999; Appleby *et al.*, 2011). It has been suggested that this recruitment may not always be required for the expression of hippocampal NMDA-dependent LTP (Henley & Wilkinson, 2016), and it is still controversial if it is essential to LTP stability (Henley & Wilkinson, 2016; Benke & Traynelis, 2019). Depending on the stimulation pattern and intensity used *in vitro* additional transduction pathways leading to the expression of late-LTP with involvement of PKA dependent phosphorylation of GluA1 subunits have been described (Huang Yan You & Kandel, 1994; Park *et al.*, 2016) in the rat. Yet, LTP induced by a brief TBS in hippocampal slices elicits an LTP that is PKA-dependent in mice (Nguyen & Kandel, 1997). The diversity of rodent and cell culture models as well of TBS protocols described in the

literature to elicit early and late LTP *in vitro* hinders the development of LTP modifying strategies targeting specific transduction pathways under firing conditions that are relevant for their activation *in vivo*.

The duration and extent of postsynaptic depolarization sustaining NMDA receptor activation, and ultimately leading to LTP induction, relies on the activity of both depolarizing and repolarizing postsynaptic currents and backpropagating action potentials. One channel that has been extensively studied regarding LTP expression is the Kv4.2 channel, that is largely responsible for the fast activating and deactivating A-current (I_A) in CA1 pyramidal neuron distal dendrites, where it acts to control signal propagation and compartmentalization in dendrites (Beck & Yaari, 2008; Kim & Hoffman, 2008). Interestingly, it has also been shown that Kv4.2 expression is more prominent in dendritic spines than in dendritic shafts (Kim *et al.*, 2007) and that LTP induction with strong TBS stimuli reduces the membrane levels of Kv4.2 on dendrites, which is accompanied by a shift in the voltage-dependence of I_A, suggesting that the activity of the channel, dependent on modifications such as phosphorylation and/or trafficking, could contribute to LTP expression (Frick *et al.*, 2004; Kim & Hoffman, 2008). Dendritic Ca^{2+} dynamics is also crucial to the activation of I_A during LTP induction, this being responsible for the precision of the time window allowed for LTP induction (Zhao *et al.*, 2011). As above mentioned, the diversity of LTP induction protocols, cellular and animal models used to study the relevance of Kv4.2 activation and phosphorylation to the induction and expression of LTP hardens the evaluation of its actual contribution to LTP in different physiological conditions.

Although LTP induced by TBS is canonically related to $GABA_B$-mediated suppression of feedforward inhibition and $GABA_B$ receptor activation has been implicated in hippocampal-dependent learning and memory formation (Brucato *et al.*, 1996), other GABAergic mechanisms and interneuron populations may contribute to LTP induced by mild TBS. Dendritic inhibition is ultimately mediated by synaptic $GABA_A$ receptors that have distinct subunit composition from extra-synaptic $GABA_A$ receptors (Farrant & Nusser, 2005; Brickley & Mody, 2012). The first mediate phasic inhibition, involved in the control of network activity, whereas extra-synaptic $GABA_A$ receptors mediate tonic inhibition,

occurring at a much slower time-window, shunting neuronal excitability and controlling neuronal input-output gain (Brickley & Mody, 2012). Thus, each $GABA_A$ receptor population has likely a distinct role in regulation of TBS-induced LTP at neuronal dendrites. Synaptic inhibition at pyramidal cell dendrites has been demonstrated to be crucial for selective Ca^{2+}-dependent input selectivity and precision of LTP induction (Müllner *et al.*, 2015). Although strong TBS has been demonstrated to induce long-term depression (LTD) in hippocampal interneurons *in vivo* and *in vitro* (Camiré & Topolnik, 2014; Lau *et al.*, 2017), no study to date has described long lasting synaptic plasticity responses in hippocampal interneurons in response to mild TBS.

Mild TBS such as a single episode of five bursts (4 stimuli, 100Hz) delivered at 5Hz is a stimulation pattern that closely resembles the naturally occurring *in vivo* firing patterns in the hippocampal CA1 area during learning and memory acquisition and induces an early-LTP that is insensitive to PKA and protein synthesis inhibitors. This typically lasts for 2-3h *in vitro* in slices obtained from naïve rats where naturally occurring reinforcement or decay, either through behavioural experience or coincident motivational inputs are not present (Aidil-Carvalho *et al.*, 2017; Çalışkan & Stork, 2018; Papaleonidopoulos & Papatheodoropoulos, 2018; Reyes-Garcia *et al.*, 2018). Since the transduction pathways implicated in the expression of LTP induced by mild TBS in the rat hippocampus are unevenly described in the literature and depend often on the stimulation paradigm, animal model (and age) or in vitro preparation used, we re-evaluated the involvement of these different mechanisms in LTP induced by mild TBS in rat hippocampal slices from young rats (6-7 weeks).

2. Material and Methods:

The experiments were performed in hippocampal slices taken from juvenile (3 weeks old), young adult (6-7 weeks old) and adult (12 weeks old) male Wistar rats (Harlan Iberica, Barcelona, Spain) essentially as previously described (Cunha-Reis *et al.*, 2014; Aidil-Carvalho *et al.*, 2017) and were in agreement with the EU Directive 2010/63/EU for animal experiments. The animals were anesthetized with halothane, decapitated, and the right hippocampus dissected free in ice-cold artificial cerebrospinal fluid (aCSF) of the following composition in mM: NaCl 124, KCl 3, NaH_2PO_4 1.25, $NaHCO_3$ 26, $MgSO_4$ 1.5, $CaCl_2$ 2, glucose 10, and gassed with a 95% O_2 - 5% CO_2 mixture.

2.1 LTP experiments

Hippocampal slices (400 µm thick) cut perpendicularly to the long axis of the hippocampus with a McIlwain tissue chopper were allowed energetic and functional recovery in a resting chamber in gassed aCSF at room temperature 22ºC–25ºC for at least 1 h. Each slice was transferred at a time to a submerged recording chamber of 1 ml capacity, where it was continuously superfused at a rate of 3 ml/min with the same gassed solution at 30.5ºC. To obtain electrophysiological recordings slices were stimulated (rectangular pulses of 0.1 ms) through a bipolar concentric wire electrode placed on the Schaffer collateral/commissural fibres in the *stratum* radiatum. Two separate sets of the Schaffer pathway (S1 and S2) were stimulated Fig. 1A. Responses were evoked every 10 s alternately on the two pathways, each pathway being stimulated every 20 s (0.05Hz). The initial intensity of the stimulus was that eliciting a field excitatory post-synaptic potential (fEPSP) of 600–1000 mV amplitude, while minimizing contamination by the population spike, and of similar magnitude in both pathways.

Evoked fEPSPs were recorded extracellularly from CA1 *stratum* radiatum (Fig. 1A) using micropipettes filled with 4 M NaCl and of 2–4 MΩ resistance. The averages of six consecutive responses from each pathway were obtained, measured, graphically plotted and recorded for further analysis with a personal computer using the LTP software (Anderson & Collingridge, 2001). The fEPSPs were quantified as the slope of the initial phase of the potential.

The independence of the two pathways was tested at the end of the experiments by studying paired-pulse facilitation (PPF) across both pathways, less than 10% facilitation being usually observed. To elicit PPF, the two Schaffer pathways were stimulated with 50 ms interpulse interval. The synaptic facilitation was quantified as the ratio P2/P1 between the slopes of the fEPSP elicited by the second P2 and the first P1 stimuli.

LTP was induced by one of three growing intensity theta-burst stimulation (***TBS***) patterns: a ***mild TBS*** (five trains of 100 Hz, 4 stimuli, separated by 200 ms), a ***moderate TBS*** (fifteen trains of 100 Hz, 4 stimuli, separated by 200 ms) and a ***strong TBS*** stimulation pattern consisting of three moderate TBS stimulation trains delivered with a 6 min interval. The stimulation protocol used to induce LTP was only applied after having a stable fEPSP slope baseline for at least 20 min. The intensity of the stimulus was not changed during these stimulation protocols. LTP was quantified as the % change in the average slope of the potentials taken from 50 to 60 min after the induction protocol (except where otherwise stated), in relation to the average slope of the fEPSP measured during the 10 min that preceded the induction protocol. Control and test conditions were tested in independent pathways in the same slice. In all experiments S1 always refers to the first pathway (left or right, randomly assigned) to which TBS was applied. Test drugs were added to the perfusion solution 20 min before TBS stimulation of the test pathway (S2) and were present until the end of the experiment except for the anti-VIP antibody that was present only until 20 min after TBS.

2.2 Western blot analysis of GluA1 and Kv4.2 phosphorylation

For western blot studies, hippocampal slices were prepared as described above, allowed for functional recovery, and then placed in the electrophysiology chambers and superfused at a flow rate of 3 ml/min with gassed aCSF at 30.5 ºC. Stimulation was delivered once every 15s in the form of rectangular pulses (0.1 ms duration) through a bipolar concentric wire electrode placed on the Schaffer collateral/commissural fibres in the *stratum radiatum* and lasted for 80 min (average duration of an electrophysiological experiment). In test chambers (but not in control chambers) TBS was applied 20

min after the beginning of basal stimulation to test but not to control slices, and basal stimulation continued until the end of the experiment. For each experimental condition, hippocampal slices were collected in sucrose solution (320mM Sucrose, 1mg/ml BSA, 10mM HEPES e 1mM EDTA, pH 7,4) containing protease (complete, mini, EDTA-free Protease Inhibitor Cocktail, Sigma) and phosphatase (1 mM PMSF, 2 mM Na_3VO_4, and 10 mM NaF) inhibitors, homogenized with a Potter-Elvejham apparatus and centrifuged at 1500g for 10 min. The supernatant was collected and further centrifuged at 14000g for 12 min. The pellet was washed twice with modified aCSF (20mM HEPES, 1mM $MgCl_2$, 1.2mM NaH_2PO_4, 2.7mM NaCl; 3mM KCl, 1.2mM $CaCl_2$, 10mM glucose, pH 7.4) also containing protease and phosphatase inhibitors and resuspended in 300µl modified aCSF *per* hippocampus. Aliquots of this suspension of hippocampal membranes were snap-frozen in liquid nitrogen and stored at -80ºC until use.

For western blot, samples incubated at 95ºC for 5 min with Laemmli buffer (125mM Tris-BASE, 4% SDS, 50% glycerol, 0,02% Bromophenol Blue, 10% β-mercaptoethanol), were run on standard 10% sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to PVDF membranes (Immobilon-P transfer membrane PVDF, pore size 0.45 µm, Immobilon). These were then blocked for 1 h with a 3% BSA solution, and incubated overnight at 4ºC with rabbit antiphospho-Ser845-GluA1 (1:2000, Chemicon), rabbit antiphospho-Ser-831-GluA1 (1:3000, Chemicon), rabbit anti-GluA1 (1:4000, Millipore), rabbit anti-Kv4.2 (1:1000, Millipore), rabbit anti-phospho-Ser438-Kv4.2 (1:100, Santa Cruz Biotech), rabbit anti-phospho-Thr607-Kv4.2 (1:100, Santa Cruz Biotech), mouse monoclonal anti-phospho-Thr602-Kv4.2 (1:2000, Santa Cruz Biotech) and rabbit anti-alpha-tubulin (1:5000, Santa Cruz Biotech) primary antibodies or rabbit anti-beta-actin (1:10000, Proteintech) primary antibodies. After washing the membranes were incubated for 1h with anti-rabbit or anti-mouse IgG secondary antibody both conjugated with horseradish peroxidase (HRP) (Proteintech) at room temperature. HRP activity was visualized by enhanced chemiluminescence with Clarity ECL Western Blotting Detection System (Bio-Rad). Intensity of the bands was evaluated with the Image J software. Beta-actin density was used as a loading control. The % phosphorylation for each target on

AMPA GluA1 subunits or Kv4.2 channels was determined by normalizing the band intensity of the phosphorylated form by the band intensity of the total GluA1 or Kv4.2. A total of six animals per condition was used to

2.3 Statistics

Values are presented as the mean ± S.E.M of n experiments for electrophysiological studies and the mean ± S.E.M of n western blot experiments performed in duplicate for western-blot studies. For some of these western blot experiments slices from more than one animal were used. The significance of the differences between the means was calculated using the paired Student's t-test when comparing two experimental groups, or with one-way analysis of variance ANOVA when comparing more than two experimental groups. P values of 0.05 or less were considered to represent statistically significant differences.

3. Results:

fEPSP obtained in hippocampal slices from *young-adult* (6-7 week-old) rats (Fig. 1.A, raw data from a single experiment) under basal stimulation conditions (40-60% of the maximal response in each slice) showed an average slope of 0.620±0.019 mV/ms (n=49). When *mild TBS* (5x4) was applied to the control pathway (S1) an LTP was observed, corresponding to a 29.6±1.2% increase in fEPSP slope (n=31, P<0.05) observed 50-60min after TBS. This potentiation progressively decayed until reaching average basal slopes 2h 30m to 3h after TBS 5x4 (n=3-6, Fig 1.B). Application of a second TBS train in the test pathway (S2) in the absence of drugs always resulted in an LTP of similar magnitude (% increase in fEPSP slope of 32.4±3.6%, n=4) of the control pathway (S1), i.e., LTP obtained under these experimental conditions was similar in S1 and S2. Increasing the number of bursts to 10 or 15 (*moderate TBS*) enhanced the resulting potentiation evaluated 50-60min after TBS to a 38.2±1.5% (n=5, P<0.05) and 51.2±3.0% (n=5, P<0.05) enhancement in fEPSP slope (Fig 1.C). By applying a *strong TBS* paradigm (TBS 3x (15x4) separated by 6 min) potentiation evaluated 50-60min after stimulation was 68.2±5.5% (n=5, P<0.05, Fig 1.C).

In *juvenile* rats (3-4 weeks old) fEPSPs had an average slope of 0.596±0.026 mV/ms (n=22). Induction of LTP with a *mild TBS (5x4)* caused a smaller potentiation of the fEPSP slope observed 50-60min after stimulation (% increase in fEPSP slope 24.5±2.6%, n=6, P<0.05, Fig 1.D). *Moderate TBS (15x4)* and *strong TBS (3x TBS 15x4)* enhanced this potentiation to a 37.8±3.2% (n=6, P<0.05) and 52.2±2.7% (n=5, P<0.05) enhancement in fEPSP slope (Fig 1.D), respectively, that were also smaller than the one obtained in young adult rats for the same *TBS* paradigm. In *adult* rats (12-13 weeks old) the average fEPSP slope was of 0.635±0.022 mV/ms (n=32). The same *mild*, *moderate* and *strong TBS* paradigms caused a larger LTP and stronger enhancement in response to protocol strength than the ones observed in young adult rats. *Mild TBS* induced a potentiation of the fEPSP slope of 33.6±5.3%, n=6, P<0.05, Fig 1.D), while *moderate* and *strong TBS* enhanced this potentiation to a 65.5±6.4% (n=6, P<0.05) and 95.1±5.8% (n=5, P<0.05) enhancement in fEPSP slope (Fig 1.D) observed 50-60 min after stimulation, respectively. Except where otherwise stated, the involvement of the different

transduction pathways in LTP induced by TBS was, as intended in this study, evaluated in young adult rats.

LTP expression induced by TBS in the hippocampus is believed to depend on suppression of feed-forward inhibition (Larson & Lynch, 1988) and to be modulated by disinhibition by additional interneuron populations in the CA1 area of the hippocampus. To confirm the involvement of fast GABAergic transmission in hippocampal CA1 LTP induced by *mild TBS* we tested the influence of blocking $GABA_A$ signaling using the selective $GABA_A$ receptor antagonist bicuculline on LTP expression. When added to the slices bicuculline (10μM) increased fEPSP slope by 43.7±5.9% (n=5), inducing also an enhancement in dendritic excitability as reflected by an enhancement in the EPSP spike superimposition in the *stratum radiatum* of CA1 area. We thus reduced stimulation intensities to make fEPSP slopes of similar magnitude to the ones obtained in the absence of bicuculline. Stimulation with *mild TBS (5x4)* in the presence of bicuculine (10μM) caused a higher LTP than the one observed in control slices, corresponding to an enhancement of 44.0±4.7% (n=5, Fig. 2.A) of fEPSP slope observed 50-60 min after TBS. This difference in the response to *mild TBS (5x4)* was evident early after LTP induction since 5 min post stimulation the enhancement in fEPSP slope was significantly higher than the one observed in control slices. The resulting potentiation was also longer-lasting and showing no significant decay ($P<0.05$) when compared to the one observed in control conditions, as evidenced by the enhancement in fEPSP slope remaining 110-120min after TBS (29.5%±4.9%, n=5, in the presence of bicuculine vs. 13.9%±2.6%, n=6, in control conditions, Fig. 2.A, $P<0.05$). This suggests that additional inhibitory pathways mediate the influence of GABAergic transmission on TBS-induced LTP induction and maintenance. $GABA_B$ autoreceptor activation by enhanced GABA release has been implicated in the suppression of feedforward inhibition during TBS (Brucato *et al.*, 1996). Synaptically released GABA is subject to reuptake by high-affinity Na^+- and Cl^--dependent GABA transporters (GATs) (Cunha-Reis *et al.*, 2017). GAT1 is predominantly expressed in presynaptic GABAergic boutons and plays a crucial role in controlling GABA spillover and, consequently, modulation of both phasic and tonic GABAergic inhibition (Gong *et al.*, 2009). As such, we tested the role of synaptic GABA availability in the induction

and expression of LTP induced by *mild TBS* upon partial inhibition of GAT1 transporters. When added to the slices, the selective GAT1 antagonist SKF89976a (5μM) did not significantly change fEPSP slope. Unlike what was previously observed in mice (Gong *et al.*, 2009), upon blockade of GAT1 transporters with SKF89976a (5μM), LTP induction with *mild TBS (5x4)* caused a long-lasting enhancement of 40.6±2.9% (n=6, Fig. 2.B) in fEPSP slope 50-60 min after stimulation, that was higher ($P<0.05$) than the one observed in control conditions 29.7±3.4% (n=6), but this difference was attenuated 110-120min after stimulation (Fig. 2.B). Finally, we tested the influence of $GABA_B$ receptor blockade with the selective antagonist CGP55845 on the induction and expression of LTP induced by *mild TBS*. When CGP55845 (1μM) was present in the bath, stimulation with *mild TBS (5x4)* elicited a weaker LTP than the one observed in control slices, corresponding to an enhancement of fEPSP slope of 17.8±0.5% (n=5, Fig. 2.C) of fEPSP slope observed 50-60 min after TBS, and that was nearly extinct (% increase in fEPSP slope: 4.7±1.5%, n=5) 110-120 min after TBS, confirming the importance of activation of $GABA_B$ receptors to the induction of LTP by TBS.

To investigate if LTP induced by *mild TBS* was dependent on the activation of NMDA receptors we tested the effect *mild TBS (5x4)* in the presence of the NMDA receptor antagonist (2R)-amino-5-phosphonopentanoate (AP-5). As expected under basal stimulation conditions, when added to the slices from the beginning of experiments AP-5 (100μM) did not significantly change fEPSP slope. In the presence of AP-5 (100μM), stimulation with *mild TBS (5x4)* did not elicit an LTP ($P>0.05$, n=4, Fig. 3.A), as previously described in the same experimental conditions (e. g. Aidil-Carvalho et al., 2017).

It is generally accepted the expression of LTP relies on the Ca^{2+}-dependent activation and consequent auto-phosphorylation of CaMKII, that then elicits the recruitment of AMPA GluA1 subunits (Appleby *et al.*, 2011). However, it has been reported that this is not required for the expression of NMDA-dependent LTP in the mouse (Cooke *et al.*, 2006). Involvement of CaMKII in LTP induced by *mild TBS (5x4)* was investigated using the CaMKII selective inhibitor KN-62. When added to the slices KN-62 (50μM) did not significantly change fEPSP slope. When *mild TBS (5x4)* was delivered in the presence of KN-62 (50μM) LTP expression was suppressed (% change in fEPSP slope of 2.2±1.4%, n=4, Fig. 3.B).

These results confirm that LTP induced by *mild TBS (5x4)* is fully dependent on CamKII activity. We further investigated the involvement of the G_s/adenylate cyclase/PKA transduction system in CA1 on LTP induced by *mild TBS (5x4)*. In the presence of the selective PKA inhibitor (Chijiwa *et al.*, 1990) H-89 (3µM), LTP induced by *mild TBS (5x4)* was not changed (% increase in fEPSP slope 50-60 min after stimulation: 27.2±1.2%, n=5, Fig. 3.C, $P<0.05$). In view of the absence of effect of the selective PKA inhibitor LTP induced by *mild TBS (5x4)*, the involvement of PKC was also investigated. Upon selective inhibition of PKC with GF 109203X (1 µM) (Toullec *et al.*, 1991), *mild TBS (5x4)* elicited an LTP that showed similarly little change (% increase in fEPSP slope 50-60 min after stimulation: 21.5±1.5%, n=5, Fig. 3.D, $P>0.05$). Expression of late-LTP induced by *moderate TBS (15x4)* in C57BL/6J mice is dependent on PKA activity and protein synthesis (Nguyen & Kandel, 1997), yet only multiple spaced TBS trains have been described to induce an LTP dependent on PKA in hippocampal slices from Sprague-Dawley rats (Park *et al.*, 2016). To confirm this, we readdressed the PKA-dependency of TBS induced LTP by using a *moderate TBS (15x4)*, as used by Nguyen and Kandel in 1997 and a *strong TBS 3x(15x4)* separated by 6 min, that would assure the conditions of three TBS bursts with sufficient spacing to mimic the spaced TBS used by Park et al. (2016), and in rats of similar age (12 weeks). Upon *moderate TBS (15x4)*, the resulting potentiation in fEPSP slope evaluated 50-60min after TBS was 65.5±6.4% (n=6, $P<0.05$, Fig 4.A) and 110-120 min after stimulation showed very little decay 61.2±9.3% (n=6, $P<0.05$, Fig 4.A). In the presence of the selective PKA inhibitor H-89 (3µM), the potentiation of fEPSP slope induced by *moderate TBS (15x4)* was not significantly changed either 50-60 min post stimulation (62.5±2.1%, n=5, Fig. 4.A) or 110-120 min post stimulation (63.4±4.2%, n=5, Fig. 4.A).

H-89 is a competitive antagonist at the ATP binding site of the PKA catalytic subunit and its EC_{50} is known to depend strongly on intracellular ATP concentrations. Since the absence of effect of PKA on late-LTP could be due its ineffective inhibition of PKA, we used another inhibitor, PKI 14-22 amide, a PKA inhibitor that binds with high-affinity the catalytic subunit of PKA, similarly to the binding of the PKA regulatory subunit, and that promotes inhibition of the kinase activity a K_i in the nanomolar range (Glass *et al.*, 1989). The presence of the selective PKA inhibitor PKI 14-22 amide (1µM), again did

not significantly change (P>0.05) the potentiation of fEPSP slope induced by *moderate TBS (15x4)* either 50-60 min post stimulation (63.0±3.4%, n=4, Fig. 4.B) or 110-120 min post stimulation (58.0±3.0%, n=4, Fig. 4.B).

Upon *strong TBS 3x (15x4)*, the resulting potentiation in fEPSP slope evaluated 50-60min after TBS was 95.1±5.8% (n=6, P<0.05, Fig. 4.C) and the enhancement in fEPSP slope observed 110-120 min after stimulation was 83.8±10.7% (n=6, P<0.05, Fig. 4.C). The selective PKA inhibitor H-89 (3µM) also did not significantly change (P>0.05) the potentiation of fEPSP slope induced by *strong TBS 3x (15x4)* either 50-60 min post stimulation (90.3±10.1%, n=5, Fig. 4.C) or 110-120 min post stimulation (78.2±8.6%, n=5, Fig. 4.C).

To further investigate the transduction pathways involved in the expression of LTP induced *mild TBS (5x4),* we investigated the downstream targets of different intracellular kinases on AMPA GluA1 receptor subunits, previously reported to mediate expression of hippocampal LTP by promoting traffic or modifying opening probability of AMPA receptors (Lee *et al.*, 2003). When GluA1 phosphorylation was inspected 50 min after *mild TBS (5x4),* no significant changes in Ser_{845} phosphorylation (a site targeted by PKA) or in the total expression of GluA1 subunits (P>0.05, n=3-4, Fig 5A-B, E-F) were observed. Yet, an enhancement of 62.1±11.0%, n=4 (Fig. 5.C and D) in GluA1 phosphorylation in Ser_{831} residue, a phosphorylation site targeted by both CaMKII and PKC, was observed under these experimental conditions. Increasing the TBS stimulus to a *moderate TBS (15x4)* did not change the pattern of GluA1 phosphorylation but increased, although not significantly compared to *TBS(5x4)* (P>0.05), the level of GluA1 phosphorylation at Ser_{831} by 91.3±10.4%, n=3 (Fig. 5.C and D).

Finally, we investigated the downstream targets of different intracellular kinases on Kv4.2 dendritic K^+ channels, previously reported to contribute to the expression of hippocampal LTP by suppressing the A-current and facilitating action potential backpropagation (Frick *et al.*, 2004; Rosenkranz *et al.*, 2009). When Kv4.2 phosphorylation was inspected 50 min after *mild TBS (5x4),* a significant enhancement (119.8±43.8%, P<0.05, n=4, Fig 6.A-B) was observed in Ser_{438} phosphorylation

(a site targeted by CaMKII) but not in the total expression of Kv4.2 subunits (P>0.05, n=4, Fig 6.C-D). No significant changes (P>0.05, n=3) were observed in Kv4.2 phosphorylation in two ERK (extracellular signal-regulated kinases) phosphorylation sites Thr_{607} (Fig 6.E) and Thr_{602} (Fig 6.F) and basal phosphorylation (control conditions) at these two sites was low.

4. Discussion:

The main findings of the present work are that: 1) *mild TBS* stimulation (5 x 4 (100Hz) stimuli, separated by 200ms), a stimulation paradigm mimicking the duration and pattern of complex-spike discharges under theta-rhythm modulation that are observed during exploration and memory acquisition, induces growing long-lasting potentiation of synaptic transmission from weaning (3 week-old) through adulthood (12-week-old) in the CA1 area of hippocampal slices obtained from Wistar rats; 2) Increasing the duration of the train to 15 x 4 (100Hz) stimuli, separated by 200ms (*moderate TBS*) or concomitantly increasing the duration and number of these trains to three (*strong TBS*) resulted in an increased potentiation and stability of the elicited LTP at three developmental stages tested (3-, 7- and 12 week-old); 3) The LTP elicited by *mild TBS* in young (7-week-old) rats was dependent on $GABA_B$ receptor activation and is endogenously inhibited by $GABA_A$ receptor activation and by GAT-1 activity; 4) *mild TBS* induced LTP in young rats was fully dependent on NMDA receptor and CaMKII activity but independent of PKA activity for all ages and TBS paradigms; 5) Furthermore, LTP induced by *mild TBS* in young rats results in putative CaMKII-mediated phosphorylation of GluA1 AMPA receptor subunits at Ser845 and of Kv4.2 K^+ channels at Ser438.

In the present study *mild TBS* (5 bursts, 4 pulses delivered at 100Hz), the LTP-inducing stimulation pattern object of this study, elicited a long-lasting potentiation of fEPSP slope in the CA1 area of rat hippocampal slices that lasted at least 2h. This potentiation was increasingly larger in 3-, 7- , and 12-week-old rats. Previous studies showed that during postnatal development there is a reinforcement of the cellular mechanisms leading to LTP expression and stability (Kramár & Lynch, 2003; Cao & Harris, 2012; Larson & Munkácsy, 2015) but it was never unequivocally show in the same study that this enhancement in potentiation continues during postweaning development until full adulthood at 12-week-old rats. In fact, some studies include in the same group animals aged 4-12 weeks (Park *et al.*, 2016), and do not allow for this to be evaluated. Stronger theta burst stimulation, achieved by increasing the number of bursts to 10-15 (*moderate TBS*) or delivery of three of these trains separated by 6 min (*strong TBS*), enhanced proportionally the resulting potentiation for all age

groups studied. *Mild TBS* constitutes an ideal intensity of TBS for pharmacological studies aiming to improve LTP outcome, since increasing the number of pulses as in *moderate TBS* (e.g. 10 bursts, 4 pulses) elicits a maximal LTP for this stimulation pattern. This ceiling effect can only be overcome by delivering several TBS trains (Larson & Munkácsy, 2015). Weaker TBS stimulation (3 bursts, 4 pulses) did not elicit a long-lasting potentiation of fEPSP slope at the developmental ages tested in this study similar to what was previously observed (Costenla *et al.*, 1999). Also, as previously reported, the potentiation elicited by *mild TBS* in the CA1 area of the rat hippocampus was fully dependent on NMDA receptor activity (Fig. 3.A), unlike the potentiation elicited by other stronger and slightly differently patterned TBS (200Hz tetanus frequency) for which a concomitant dependency on voltage-gated calcium channels (VGCCs) was observed (Grover & Teyler, 1990; Grover, 1998).

Suppression of feed-forward phasic inhibition, mediated by $GABA_B$ autoreceptor suppression of GABA release, is believed to mediate induction of LTP by TBS in excitatory synapses of the hippocampus (Pacelli *et al.*, 1989; Grover & Yan, 1999; Larson & Munkácsy, 2015), allowing for enough temporal summation of excitation and prolonged depolarization that ultimately lead to NMDA receptor activation. In addition, *mild* TBS also activates cooperative postsynaptic mechanisms dependent on $GABA_B$ and metabotropic glutamate receptors, leading to a sustained potentiation of fast $GABA_A$ synaptic transmission in GABAergic synapses to pyramidal cells (Patenaude *et al.*, 2003). Despite this, the net result is a potentiation of synaptic transmission as evaluated by the long-lasting changes in fEPSP slope. In this study, blocking $GABA_A$ signaling using supramaximal concentrations of the selective $GABA_A$ receptor antagonist bicuculline resulted in an enhancement of the magnitude of LTP induced by *mild* TBS in the CA1 area. This is consistent with previous studies in Sprague-Dawley rats using primed-burst protocols (Pacelli *et al.*, 1989) and much stronger TBS protocols using either 100Hz or 200Hz tetanus frequency (Chapman *et al.*, 1998; Grover & Yan, 1999). The resulting potentiation lasted longer than the one observed in control conditions and showed little decay 2h after induction, unlike the one obtained in control conditions that was run down to one third of the potentiation observed 1h after induction. Since sustained changes in $GABA_A$ mediated currents in

response to *mild* TBS do not contribute to the fEPSP slope in the presence of bicuculine, this result likely reflects the potentiation (and its stability) elicited by TBS at glutamatergic synapses and suggests that additional inhibitory pathways (other than suppression of phasic feed-forward inhibition during induction) mediate the influence of GABAergic transmission on the expression and maintenance of TBS-induced LTP. One hypothesis, is that transient changes in tonic inhibition during LTP induction may also modulate the outcome of LTP induced by TBS under different physiological conditions, since both synaptic and non-synaptic inhibition can hinder spike backpropagation and Ca^{2+} spikes in CA1 pyramidal cell dendrites (Tsubokawa & Ross, 1996; Larson & Munkácsy, 2015; Müllner *et al.*, 2015), an essential mechanism in TBS-induced LTP. In fact, developmental changes in tonic GABAergic inhibition at pyramidal cell dendrites have been associated with developmental changes in LTP expression (Groen *et al.*, 2014), suggesting that such changes may contribute to the differences in the magnitude of potentiation induced by *mild* TBS at the three developmental stages tested in this study.

In this study, LTP induced by TBS was, as expected, dependent on endogenous activation of $GABA_B$ receptors, as previously described, thus confirming an essential role of these receptors in CA1 LTP induction by TBS (Larson *et al.*, 1986; Larson & Lynch, 1988). However, blockade of $GABA_B$ receptors with supramaximal concentrations of the antagonist CGP 55845 did not prevent the expression of TBS-induced LTP in the CA1 area of the hippocampus in this study. This suggests that additional $GABA_B$-independent mechanisms contribute to the suppression of feed-forward inhibition to induce LTP by TBS. In addition, blockade of GAT1 transporters, that are predominantly expressed in presynaptic GABAergic boutons and to a lesser extent in astrocytes, enhanced LTP induced by *mild TBS* in juvenile rats, rebutting what was previously reported in mice using the same stimulation paradigm and intensity (Gong *et al.*, 2009). GABA released to the synaptic cleft is subject to reuptake by high-affinity Na^+- and Cl^--dependent GABA transporters (GATs). GAT-1 plays a crucial role in controlling synaptic GABA spillover and, consequently, modulation of both phasic and tonic GABAergic inhibition (Gong *et al.*, 2009). In light of the results in this study, it is suggested that GAT-1 blockade, by enhancing synaptic GABAergic tone at synapses mediating disinhibition, might contribute to enhanced

depolarization of pyramidal cell dendrites during TBS, leading to enhanced potentiation of glutamatergic transmission (Cunha-Reis *et al.*, 2010, 2014; Francavilla *et al.*, 2018; Cunha-Reis & Caulino-Rocha, 2020). In contrast, enhanced synaptic GABA by GAT-1 inhibition at feedforward inhibitory contacts might enhance suppression of inhibition in mice, leading to decreased potentiation by TBS. Altogether, this suggests that theta-patterned neuronal activity may be differently regulated in mice and in rats during exploration and spatial learning, the behavioral correlates of the *mild TBS* LTP-inducing paradigm characterized in this study. In fact, mice and rats behave very differently in exploratory tasks and its modulation by sensory cues and previous experience are very distinct (Tanaka *et al.*, 2012; Thompson *et al.*, 2018). Interestingly, Reyes-Garcia et al (2018) described that an enhancement of TBS-induced LTP by $GABA_A$ receptor antagonists is not observed in rat wildlife population (Reyes-Garcia *et al.*, 2018). The enhancement in *mild* TBS-induced LTP expression caused by environmental enrichment (Malik & Chattarji, 2012) may involve changes in number and distribution of GABAergic synapses onto pyramidal cell dendrites (Foggetti *et al.*, 2019). Furthermore, the enhancement of mild TBS-induced LTP caused by mismatch exploration training (Aidil-Carvalho *et al.*, 2017) is also associated with a decrease in VIP endogenous disinhibition enhancing LTP (Cunha-Reis *et al.*, 2008). Altogether, these observations and the fact that variations in LTP across dendritic subfields of hippocampus pyramidal cells reflect a differential distribution of GABAergic interneurons responsible for feed-forward inhibition, that in turn differentially affect the local dynamics of Ca^{2+} transients (Arai *et al.*, 1994; Kanemoto *et al.*, 2011; Francavilla *et al.*, 2019), suggest that a dynamic modulation of GABAergic input to different CA1 pyramidal cell dendritic subfields mediates changes in *mild* TBS-induced LTP (Müllner *et al.*, 2015).

In this study, the long-lasting potentiation of synaptic transmission caused by mild TBS in young adult rats was fully dependent on NMDA receptor activation and CaMKII activity. This was likely a result from enhanced Ca^{2+} influx (Fig. 3. A-B), as previously described (Larson & Munkácsy, 2015), and does not suggest that additional endogenous channel activation is required to induce potentiation under these experimental conditions. Furthermore, as previously described by us and others (Cruz, 2010;

Park *et al.*, 2016), this potentiation was fully independent of PKA activity (Fig. 3.C). Even when reproducing experimental conditions and using specific inhibitors of PKA activity, as previously used in mice when observing a PKA-dependent expression of theta-burst LTP, this independence of PKA activity is confirmed (Fig. 4). This again suggests that TBS-induced LTP is differently regulated in rats and in mice, and that these differences should not be disregarded in the context of learning and memory tasks in the two species. This also implies that modulators acting through PKA-dependent mechanisms might have substantially different impact in the expression of TBS-induced LTP in mice and in rats, and that care should be taken in generalizing the interpretation of data from a single rodent species when considering substances acting through PKA-dependent transduction pathways. Finally, the expression of LTP induced by *mild* TBS was virtually independent of PKC activity. PKC has previously been reported to be involved in the expression of LTP induced by weak TBS stimulation 3x 5 (100Hz) pulses in the CA1 area of mice hippocampal slices (Miura *et al.*, 2002), but not in the rat (Hasegawa *et al.*, 2014). These results suggest once more that regulation of LTP expression in the mouse and rat hippocampus is differently regulated by intracellular protein kinases.

The results in this study showed that *mild* TBS promotes phosphorylation of AMPA receptor GluA1 subunits at Ser_{831}, a phosphorylation site targeted by both CaMKII and PKC, but not at Ser845, targeted by PKA. Phosphorylation of AMPA receptor GluA1 subunits by CaMKII, promotes enhancement of channel conductance and its recruitment of to the active zone (Derkach *et al.*, 1999; Appleby *et al.*, 2011). This has been argued not to be essential to the expression of hippocampal NMDA-dependent LTP (Henley & Wilkinson, 2016) or required for LTP stability (Henley & Wilkinson, 2016; Benke & Traynelis, 2019), yet the results in this study indicate than GluA1 phosphorylation at Ser_{831} occurs even under *mild* TBS stimulation, suggesting that it is important for LTP expression under these stimulation conditions. Phosphorylation of GluA1 at Ser_{845} has been argued to be dependent on the TBS stimulation pattern and intensity used *in vitro* in the rat (Huang Yan You & Kandel, 1994; Park *et al.*, 2016). Furthermore, since LTP induced by *moderate* TBS in mice hippocampal slices is PKA-dependent (Nguyen & Kandel, 1997), it is also dependent on the rodent species used. This again

evidences a major variability in the role of GluA1 phosphorylation on *mild* to *moderate* TBS induced LTP in different rodent models. Furthermore, it is dependent on the use of maximal and submaximal TBS stimulation, hindering the establishment of an universal rule for the role of GluA1 phosphorylation in the induction, expression and maintenance of TBS-induced LTP.

A role for Kv4.2 channels in regulation of hippocampal synaptic plasticity has for long been debated in the literature (Kim & Hoffman, 2008). The function and properties of the Kv4.2 channel can be regulated by post-translational phosphorylation at several highly conserved sites. Kv4.2 can be phosphorylated by ERK and other kinases of the MAPK (mitogen-activated protein kinases) family at Thr_{602} and Thr_{607} residues, which leads to a decrease in I_A, and internalization of the channel (Adams *et al.*, 2000; Varga *et al.*, 2000; Schrader *et al.*, 2006). Yet kinases of the ERK family can also catalyze the phosphorylation of Ser_{616}, which inhibits the channel's activity (Hu *et al.*, 2006). PKA phosphorylates the Kv4.2 channel, at Thr_{38} or Ser_{552}. Although consequences of phosphorylation on Thr_{38} are unknown, phosphorylation of Ser_{552} causes internalization of the channel (Hammond *et al.*, 2008). Finally, CaMKII can modulate Kv4.2 channel expression and upregulate neuronal I_A currents through phosphorylation at Ser_{438} (Varga *et al.*, 2004).

One important observation in this study was that *mild* TBS stimulation enhances the phosphorylation of Kv4.2 channels at Ser_{438}. As mentioned above, NMDA receptor activation during *mild* TBS mediates Ca^{2+} influx that activates CaMKII. This enhancement of CaMKII activity can be the cause for the enhanced Kv4.2 channel phosphorylation at Ser_{438} found in this work since this is a major target for CaMKII phosphorylation. This post-translational modification leads to an increase in the cell-surface expression of the channel (Varga *et al.*, 2004), which can explain the mild increase of the channel levels also found in this work. Since the increase of the channel expression leads to an increase of I_A this likely does not contribute to expression of *mild* TBS-induced LTP since it promotes an NMDA receptor increased NR2B/NR2A subunit ratio (Jung *et al.*, 2008, 2011). Rather it may be counteracting the effects of NMDA receptor activation to keep dendritic excitability under control. Furthermore, since Kv4.2 channels are differentially expressed in different dendritic segments in relation to the

distance to the soma of pyramidal cells (Hoffman *et al.*, 1997; Beck & Yaari, 2008) it is possible that these results do not reflect the modulation of the I_A in all dendritic segments.

In summary, the expression of LTP induced by *mild* TBS in the CA1 area of rat hippocampal slices involves several signaling mechanisms that are different than what is currently believed in the literature to be the general mechanisms for LTP expression. Determining the more physiologically relevant stimulation patterns and intensities to the study this widely cellular phenomenon that is widely accepted as the basis for learning and memory mechanisms represents still a challenge in current research. Nevertheless, it is our believe that *mild* TBS constitutes the ideal pattern and intensity to study the value of pharmacological tools aiming to modulate LTP expression and stability. The results discussed in this paper also raise several concerns on which is the best animal/rodent model to study the cellular mechanisms of LTP and evidences several differences in the endogenous regulation of TBS-induced LTP by $GABA_A$ receptors and GAT-1 transporters which suggest that the GABAergic tone in the mouse and rat hippocampus during learning and memory may be quite different. This may in turn be the cause for the high differences in exploratory and learning behavior found between several mice and rat strains. In addition, we demonstrate that a few protein kinases widely accepted to be essential to LTP expression in mice do not significantly contribute to LTP expression by *mild* to *moderate* TBS in the rat, suggesting that they are more likely involved in regulatory mechanisms or are activated by endogenous modulators of LTP in well-defined physiological situations. In addition, we show that GluA1 phosphorylation at Ser_{831} is the only GluA1 post-translational modification that is likely essential for expression of mild TBS induced LTP in the CA1 area of the rat hippocampus. Other phosphorylation sites in GluA1 subunits may then be preferential targets for endogenous modulators of synaptic plasticity. Finally, we show that Kv4.2 is more likely involved in suppressing or controlling the extent rather than contributing to the expression of LTP induced by mild TBS. Nevertheless, its multiple phosphorylation sites suggest this channel is a major target for the regulation of LTP by endogenous modulators and several intracellular kinases.

In conclusion, this paper reveals that there are several differences in the essential mechanisms involved in TBS-induced LTP in rats. This supports the need to establish the best model as well as standardized methodologies, stimulation patterns and intensities to induce LTP by TBS to determine which are the most appropriate to study the action of pharmacological tools aiming to improve synaptic plasticity and ultimately memory.

5. References:

Adams, J.P., Anderson, A.E., Varga, A.W., Dineley, K.T., Cook, R.G., Pfaffinger, P.J., & Sweatt, J.D. (2000) The A-type potassium channel Kv4.2 is a substrate for the mitogen-activated protein kinase ERK. *J. Neurochem.*, **75**, 2277–2287.

Aidil-Carvalho, M.F., Carmo, A.J.S., Ribeiro, J.A., & Cunha-Reis, D. (2017) Mismatch novelty exploration training enhances hippocampal synaptic plasticity: A tool for cognitive stimulation? *Neurobiol. Learn. Mem.*, **145**, 240–250.

Albensi, B.C., Oliver, D.R., Toupin, J., & Odero, G. (2007) Electrical stimulation protocols for hippocampal synaptic plasticity and neuronal hyper-excitability: Are they effective or relevant? *Exp. Neurol.*,.

Anderson, W.W. & Collingridge, G.L. (2001) The LTP Program: a data acquisition program for on-line analysis of long-term potentiation and other synaptic events. *J Neurosci Methods*, **108**, 71–83.

Appleby, V.J., Corrêa, S.A.L., Duckworth, J.K., Nash, J.E., Noël, J., Fitzjohn, S.M., Collingridge, G.L., & Molnár, E. (2011) LTP in hippocampal neurons is associated with a CaMKII-mediated increase in GluA1 surface expression. *J Neurochem*, **116**, 530–543.

Arai, A., Black, J., & Lynch, G. (1994) Origins of the variations in long-term potentiation between synapses in the basal versus apical dendrites of hippocampal neurons. *Hippocampus*, **4**, 1–9.

Baudry, M., Zhu, G., Liu, Y., Wang, Y., Briz, V., & Bi, X. (2015) Multiple cellular cascades participate in long-term potentiation and in hippocampus-dependent learning. *Brain Res*, **1621**, 73–81.

Beck, H. & Yaari, Y. (2008) Plasticity of intrinsic neuronal properties in CNS disorders. *Nat. Rev. Neurosci.*, **9**, 357–369.

Benke, T. & Traynelis, S.F. (2019) AMPA-Type Glutamate Receptor Conductance Changes and Plasticity: Still a Lot of Noise. *Neurochem. Res.*, **44**, 539–548.

Bliss, T. & Collingridge, G. (2019) Persistent memories of long-term potentiation and the N -methyl-d-aspartate receptor. *Brain Neurosci. Adv.*, **3**, 1–10.

Bliss, T.V.P. & Collingridge, G.L. (1993) A synaptic model of memory: Long-term potentiation in the hippocampus. *Nature*, **361**, 31–39.

Brickley, S.G. & Mody, I. (2012) Extrasynaptic GABA A Receptors: Their Function in the CNS and Implications for Disease. *Neuron*,.

Brucato, F.H., Levin, E.D., Mott, D.D., Lewis, D. V., Wilson, W.A., Swartzelder, H.S., & Swartzwelder, H.S. (1996)

Hippocampal long-term potentiation and spatial learning in the rat: effects of GABAB receptor blockade. *Neuroscience*, **74**, 331–339.

Çalışkan, G. & Stork, O. (2018) Hippocampal network oscillations as mediators of behavioural metaplasticity: Insights from emotional learning. *Neurobiol. Learn. Mem.*, **154**, 37–53.

Camiré, O. & Topolnik, L. (2014) Dendritic calcium nonlinearities switch the direction of synaptic plasticity in fast-spiking interneurons. *J. Neurosci.*, **34**, 3864–3877.

Cao, G. & Harris, K.M. (2012) Developmental regulation of the late phase of long-term potentiation (L-LTP) and metaplasticity in hippocampal area CA1 of the rat. *J. Neurophysiol.*, **107**, 902–912.

Chapman, C.A., Perez, Y., & Lacaille, J.C. (1998) Effects of GABA(A) inhibition on the expression of long-term potentiation in CA1 pyramidal cells are dependent on tetanization parameters. *Hippocampus*, **8**, 289–298.

Chijiwa, T., Mishima, A., Hagiwara, M., Sano, M., Hayashi, K., Inoue, T., Naito, K., Toshioka, T., & Hidaka, H. (1990) Inhibition of forskolin-induced neurite outgrowth and protein phosphorylation by a newly synthesized selective inhibitor of cyclic AMP-dependent protein kinase, N-[2-(p-bromocinnamylamino)ethyl]-5-isoquinolinesulfonamide (H-89), of PC12D pheochromocytoma . *J Biol Chem*, **265**, 5267–5272.

Cobb, S.R., Manuel, N.A., Morton, R.A., Gill, C.H., Collingridge, G.L., & Davies, C.H. (1999) Regulation of depolarizing GABA(A) receptor-mediated synaptic potentials by synaptic activation of GABA(B) autoreceptors in the rat hippocampus. *Neuropharmacology*, **38**, 1723–1732.

Cooke, S.F., Wu, J., Plattner, F., Errington, M., Rowan, M., Peters, M., Hirano, A., Bradshaw, K.D., Anwyl, R., Bliss, T.V.P., & Giese, K.P. (2006) Autophosphorylation of αCaMKII is not a general requirement for NMDA receptor-dependent LTP in the adult mouse. *J. Physiol.*, **574**, 805–818.

Costenla, A.R., de Mendonça, A., & Ribeiro, J.A. (1999) Adenosine modulates synaptic plasticity in hippocampal slices from aged rats. *Brain Res*, **851**, 228–234.

Cruz, A.D. da S. (2010) The influence of epilepsy in synaptic plasticity in the hippocampus: neuroprotective role of VIP and its receptors.

Cunha-Reis, D., Aidil-Carvalho, F., & Ribeiro, J.A. (2014) Endogenous inhibition of hippocampal LTD and depotentiation by vasoactive intestinal peptide VPAC1 receptors. *Hippocampus*, **24**, 1353–1363.

Cunha-Reis, D., Aidil-Carvalho, F., Sebastião, A.M., & Ribeiro, J.A. (2008) Modulation of hippocampal synaptic

plasticity by endogenous VIP: influence of novelty exploration training. In *6º Encontro Do Fórum Internacional de Investigadores Portugueses*.

Cunha-Reis, D. & Caulino-Rocha, A. (2020) VIP Modulation of Hippocampal Synaptic Plasticity: A Role for VIP Receptors as Therapeutic Targets in Cognitive Decline and Mesial Temporal Lobe Epilepsy. *Front. Cell. Neurosci.*, **14**, 153.

Cunha-Reis, D., Ribeiro, J.A., de Almeida, R.F.M., & Sebastião, A.M. (2017) VPAC1 and VPAC2 receptor activation on GABA release from hippocampal nerve terminals involve several different signalling pathways. *Br. J. Pharmacol.*, **174**, 4725–4737.

Cunha-Reis, D., Rodrigues, N.C., & Ribeiro, J.A. (2010) On the cellular and molecular pathways involved in the inhibition of LTP in the CA1 area of the hippocampus. *J. Mol. Neurosci.*, **42**, 278.

Davies, C.H., Davies, S.N., & Collingridge, G.L. (1990) Paired-pulse depression of monosynaptic GABA-mediated inhibitory postsynaptic responses in rat hippocampus. *J Physiol*, **424**, 513–531.

Derkach, V., Barria, A., & Soderling, T.R. (1999) Ca2+/calmodulin-kinase II enhances channel conductance of alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionate type glutamate receptors. *Proc. Natl. Acad. Sci. U. S. A.*, **96**, 3269–3274.

Farrant, M. & Nusser, Z. (2005) Variations on an inhibitory theme: Phasic and tonic activation of GABA A receptors. *Nat. Rev. Neurosci.*,.

Foggetti, A., Baccini, G., Arnold, P., Schiffelholz, T., & Wulff, P. (2019) Spiny and Non-spiny Parvalbumin-Positive Hippocampal Interneurons Show Different Plastic Properties. *Cell Rep.*, **27**, 3725-3732.e5.

Francavilla, R., Villette, V., Luo, X., Chamberland, S., Muñoz-Pino, E., Camiré, O., Wagner, K., Kis, V., Somogyi, P., & Topolnik, L. (2018) Connectivity and network state-dependent recruitment of long-range VIP-GABAergic neurons in the mouse hippocampus. *Nat. Commun.*, **9**.

Francavilla, R., Villette, V., Martel, O., & Topolnik, L. (2019) Calcium Dynamics in Dendrites of Hippocampal CA1 Interneurons in Awake Mice. *Front. Cell. Neurosci.*, **13**, 98.

Frick, A., Magee, J., & Johnston, D. (2004) LTP is accompanied by an enhanced local excitability of pyramidal neuron dendrites. *Nat. Neurosci.*, **7**, 126–135.

Glass, D.B., Cheng, H.C., Mende-Mueller, L., Reed, J., & Walsh, D.A. (1989) Primary structural determinants essential for potent inhibition of cAMP-dependent protein kinase by inhibitory peptides corresponding to the active portion of the heat-stable inhibitor protein. *J. Biol. Chem.*, **264**, 8802–8810.

Gong, N., Li, Y., Cai, G.Q., Niu, R.F., Fang, Q., Wu, K., Chen, Z., Lin, L.N., Xu, L., Fei, J., & Xu, T. Le (2009) GABA transporter-1 activity modulates hippocampal theta oscillation and theta burst stimulation-induced long-term potentiation. *J. Neurosci.*, **29**, 15836–15845.

Groen, M.R., Paulsen, O., Pérez-Garci, E., Nevian, T., Wortel, J., Dekker, M.P., Mansvelder, H.D., van Ooyen, A., & Meredith, R.M. (2014) Development of dendritic tonic GABAergic inhibition regulates excitability and plasticity in CA1 pyramidal neurons. *J. Neurophysiol.*, **112**, 287–299.

Grover, L.M. (1998) Evidence for postsynaptic induction and expression of NMDA receptor independent LTP. *J. Neurophysiol.*, **79**, 1167–1182.

Grover, L.M. & Teyler, T.J. (1990) Two components of long-term potentiation induced by different patterns of afferent activation. *Nature*, **347**, 477–479.

Grover, L.M. & Yan, C. (1999) Blockade of GABA(A) receptors facilitates induction of NMDA receptor-independent long-term potentiation. *J. Neurophysiol.*, **81**, 2814–2822.

Hammond, R.S., Lin, L., Sidorov, M.S., Wikenheiser, A.M., & Hoffman, D.A. (2008) Protein kinase a mediates activity-dependent Kv4.2 channel trafficking. *J. Neurosci.*, **28**, 7513–7519.

Hasegawa, Y., Mukai, H., Asashima, M., Hojo, Y., Ikeda, M., Komatsuzaki, Y., Ooishi, Y., & Kawato, S. (2014) Acute modulation of synaptic plasticity of pyramidal neurons by activin in adult hippocampus. *Front. Neural Circuits*, **8**, 56.

Henley, J.M. & Wilkinson, K.A. (2016) Synaptic AMPA receptor composition in development, plasticity and disease. *Nat. Rev. Neurosci.*,.

Hoffman, D.A., Magee, J.C., Colbert, C.M., & Johnston, D. (1997) K+ channel regulation of signal propagation in dendrites of hippocampal pyramidal neurons. *Nature*, **387**, 869–875.

Hu, H.J., Carrasquillo, Y., Karim, F., Jung, W.E., Nerbonne, J.M., Schwarz, T.L., & Gereau IV, R.W. (2006) The Kv4.2 Potassium Channel Subunit Is Required for Pain Plasticity. *Neuron*, **50**, 89–100.

Huang Yan You & Kandel, E.R. (1994) Recruitment of long-lasting and protein kinase A-dependent long-term potentiation in the CA1 region of hippocampus requires repeated tetanization. *Learn. Mem.*, **1**, 74–82.

Jung, S.-C.C., Kim, J., & Hoffman, D.A. (2008) Rapid, Bidirectional Remodeling of Synaptic NMDA Receptor Subunit Composition by A-type K+ Channel Activity in Hippocampal CA1 Pyramidal Neurons. *Neuron*, **60**, 657–671.

Jung, S.C., Eun, S.Y., Kim, J., & Hoffman, D.A. (2011) Kv4.2 block of long-term potentiation is partially dependent

on synaptic NMDA receptor remodeling. *Brain Res. Bull.*, **84**, 17–21.

Kanemoto, Y., Matsuzaki, M., Morita, S., Hayama, T., Noguchi, J., Senda, N., Momotake, A., Arai, T., & Kasai, H. (2011) Spatial distributions of GABA receptors and local inhibition of ca 2+ transients studied with GABA uncaging in the dendrites of CA1 pyramidal neurons. *PLoS One*, **6**, e22652.

Kim, J. & Hoffman, D.A. (2008) Potassium Channels: Newly Found Players in Synaptic Plasticity. *Neurosci.*, **14**, 276–286.

Kim, J., Jung, S.-C.C., Clemens, A.M., Petralia, R.S., & Hoffman, D.A. (2007) Regulation of Dendritic Excitability by Activity-Dependent Trafficking of the A-Type K+ Channel Subunit Kv4.2 in Hippocampal Neurons. *Neuron*, **54**, 933–947.

Kim, K., Saneyoshi, T., Hosokawa, T., Okamoto, K., & Hayashi, Y. (2016) Interplay of enzymatic and structural functions of CaMKII in long-term potentiation. *J. Neurochem.,*.

Kramár, E.A. & Lynch, G. (2003) Developmental and regional differences in the consolidation of long-term potentiation. *Neuroscience*, **118**, 387–398.

Larson, J. & Lynch, G. (1986) Induction of synaptic potentiation in hippocampus by patterned stimulation involves two events. *Science (80-.).*, **232**, 985–988.

Larson, J. & Lynch, G. (1988) Role of N-methyl-D-aspartate receptors in the induction of synaptic potentiation by burst stimulation patterned after the hippocampal theta-rhythm. *Brain Res*, **441**, 111–118.

Larson, J. & Munkácsy, E. (2015) Theta-burst LTP. *Brain Res.*, **1621**, 38–50.

Larson, J., Wong, D., & Lynch, G. (1986) Patterned stimulation at the theta frequency is optimal for the induction of hippocampal long-term potentiation. *Brain Res*, **368**, 347–350.

Lau, P.Y.P., Katona, L., Saghy, P., Newton, K., Somogyi, P., & Lamsa, K.P. (2017) Long-term plasticity in identified hippocampal GABAergic interneurons in the CA1 area in vivo. *Brain Struct. Funct.*, **222**, 1809–1827.

Lee, H.K., Takamiya, K., Han, J.S., Man, H., Kim, C.H., Rumbaugh, G., Yu, S., Ding, L., He, C., Petralia, R.S., Wenthold, R.J., Gallagher, M., & Huganir, R.L. (2003) Phosphorylation of the AMPA receptor GluR1 subunit is required for synaptic plasticity and retention of spatial memory. *Cell*, **112**, 631–643.

Lynch, G., Rex, C.S., & Gall, C.M. (2007) LTP consolidation: Substrates, explanatory power, and functional significance. *Neuropharmacology*, **52**, 12–23.

Malik, R. & Chattarji, S. (2012) Enhanced intrinsic excitability and EPSP-spike coupling accompany enriched environment-induced facilitation of LTP in hippocampal CA1 pyramidal neurons. *J. Neurophysiol.*, **107**,

1366–1378.

Miura, M., Watanabe, M., Offermanns, S., Simon, M.I., & Kano, M. (2002) Group I metabotropic glutamate receptor signaling via Gαq/Gα11 secures the induction of long-term potentiation in the hippocampal area CA1. *J. Neurosci.*, **22**, 8379–8390.

Morgan, S.L., Coussens, C.M., & Teyler, T.J. (2001) Depotentiation of vdccLTP requires NMDAR activation. *Neurobiol Learn Mem*, **76**, 229–238.

Müllner, F.E., Wierenga, C.J., & Bonhoeffer, T. (2015) Precision of Inhibition: Dendritic Inhibition by Individual GABAergic Synapses on Hippocampal Pyramidal Cells Is Confined in Space and Time. *Neuron*, **87**, 576–589.

Nguyen, P. V. & Kandel, E.R. (1997) Brief Θ-burst stimulation induces a transcription-dependent late phase of LTP requiring cAMP in area CA1 of the mouse hippocampus. *Learn. Mem.*,.

Pacelli, G.J., Su, W., & Kelso, S.R. (1989) Activity-induced depression of synaptic inhibition during LTP-inducing patterned stimulation. *Brain Res.*, **486**, 26–32.

Papaleonidopoulos, V. & Papatheodoropoulos, C. (2018) β-adrenergic receptors reduce the threshold for induction and stabilization of LTP and enhance its magnitude via multiple mechanisms in the ventral but not the dorsal hippocampus. *Neurobiol. Learn. Mem.*, **151**, 71–84.

Park, P., Kang, H., Sanderson, T.M., Bortolotto, Z.A., Georgiou, J., Zhuo, M., Kaang, B.-K., & Collingridge, G.L. (2018) The Role of Calcium-Permeable AMPARs in Long-Term Potentiation at Principal Neurons in the Rodent Hippocampus. *Front. Synaptic Neurosci.*, **10**, 42.

Park, P., Sanderson, T.M., Amici, M., Choi, S.-L.L., Bortolotto, Z.A., Zhuo, M., Kaang, B.-K.K., & Collingridge, G.L. (2016) Calcium-Permeable AMPA Receptors Mediate the Induction of the Protein Kinase A-Dependent Component of Long-Term Potentiation in the Hippocampus. *J Neurosci*, **36**, 622–631.

Patenaude, C., Chapman, C.A., Bertrand, S., Congar, P., & Lacaille, J.C. (2003) GABAB receptor- and metabotropic glutamate receptor-dependent cooperative long-term potentiation of rat hippocampal GABAA synaptic transmission. *J. Physiol.*, **553**, 155–167.

Reyes-Garcia, S.Z., de Almeida, A.C.G., Ortiz-Villatoro, N.N., Scorza, F.A., Cavalheiro, E.A., & Scorza, C.A. (2018) Robust Network Inhibition and Decay of Early-Phase LTP in the Hippocampal CA1 Subfield of the Amazon Rodent Proechimys. *Front. Neural Circuits*, **12**, 81.

Rose, G.M. & Dunwiddie, T. V (1986) Induction of hippocampal long-term potentiation using physiologically

patterned stimulation. *Neurosci Lett*, **69**, 244–248.

Rosenkranz, J.A., Frick, A., & Johnston, D. (2009) Kinase-dependent modification of dendritic excitability after long-term potentiation. *J. Physiol.*, **587**, 115–125.

Schrader, L.A., Birnbaum, S.G., Nadin, B.M., Ren, Y., Bui, D., Anderson, A.E., & Sweatt, J.D. (2006) ERK/MAPK regulates the Kv4.2 potassium channel by direct phosphorylation of the pore-forming subunit. *Am. J. Physiol. Physiol.*, **290**, C852–C861.

Tanaka, S., Young, J.W., Halberstadt, A.L., Masten, V.L., & Geyer, M.A. (2012) Four factors underlying mouse behavior in an open field. *Behav. Brain Res.*, **233**, 55–61.

Thompson, S.M., Berkowitz, L.E., & Clark, B.J. (2018) Behavioral and neural subsystems of rodent exploration. *Learn. Motiv.*, **61**, 3–15.

Toullec, D., Pianetti, P., Coste, H., Bellevergue, P., Grand-Perret, T., Ajakane, M., Baudet, V., Boissin, P., Boursier, E., & Loriolle, F. (1991) The bisindolylmaleimide GF 109203X is a potent and selective inhibitor of protein kinase C. *J Biol Chem*, **266**, 15771–15781.

Tsubokawa, H. & Ross, W.N. (1996) IPSPs modulate spike backpropagation and associated [Ca2+](i) changes in the dendrites of hippocampal CA1 pyramidal neurons. *J. Neurophysiol.*, **76**, 2896–2906.

Varga, A.W., Anderson, A.E., Adams, J.P., Vogel, H., & Sweatt, J.D. (2000) Input-specific immunolocalization of differentially phosphorylated Kv4.2 in the mouse brain. *Learn. Mem.*, **7**, 321–332.

Varga, A.W., Yuan, L.-L., Anderson, A.E., Schrader, L.A., Wu, G.-Y., Gatchel, J.R., Johnston, D., & Sweatt, J.D. (2004) Calcium-Calmodulin-Dependent Kinase II Modulates Kv4.2 Channel Expression and Upregulates Neuronal A-Type Potassium Currents. *J. Neurosci.*, **24**, 3643–3654.

Vertes, R.P. (2005) Hippocampal theta rhythm: a tag for short-term memory. *Hippocampus*, **15**, 923–935.

Zhao, C., Wang, L., Netoff, T., & Yuan, L.L. (2011) Dendritic mechanisms controlling the threshold and timing requirement of synaptic plasticity. *Hippocampus*, **21**, 288–297.

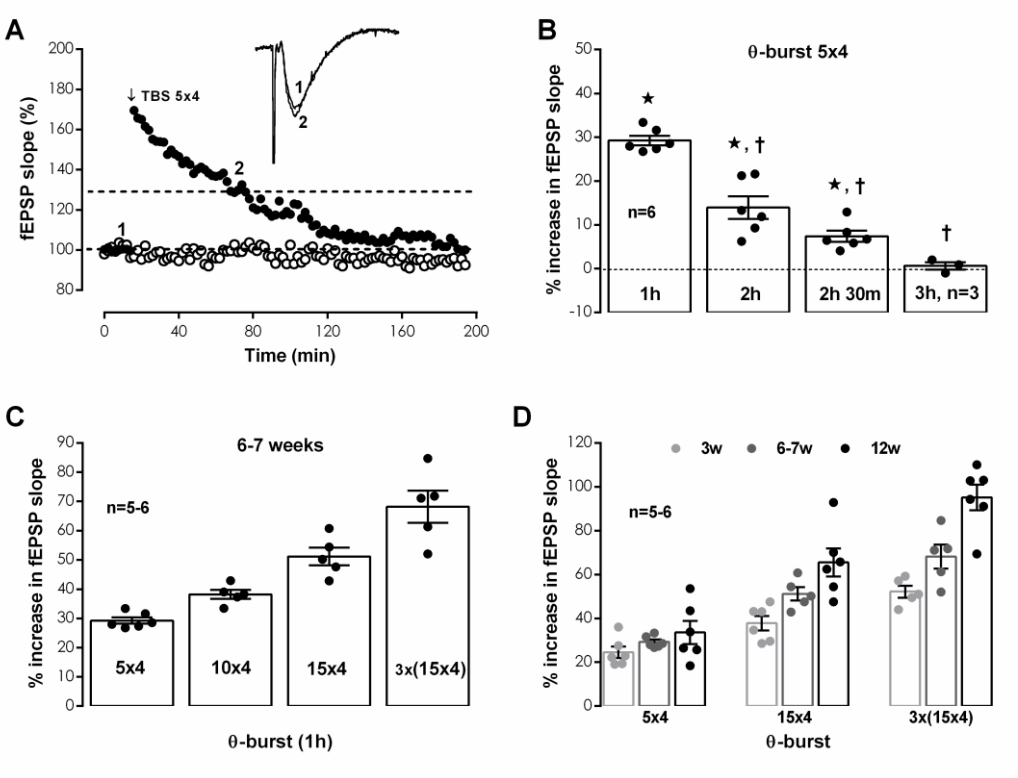

Figure 1 – Hippocampal CA1 long-term potentiation of synaptic transmission elicited by different theta-burst stimulation protocols from weaning to adulthood. A. Time-course of changes in fEPSP slope caused by theta-burst stimulation (5 bursts at 5 Hz, each composed of four pulses at 100 Hz, *mild TBS(5x4)*) in a typical experiment for which a test pathway (-●-) was stimulated in the absence of added drugs and a control pathway (-○-) was used as internal control for TBS stimulation in the same hippocampal slice obtained from a young adult rat (6-7 weeks). ***Inset:*** Traces of fEPSPs obtained in the same experiment before (time point **1**) and 50-60 min after (time point **2**) theta burst stimulation. Traces are the average of eight consecutive responses and are composed of the stimulus artifact, the presynaptic volley and the fEPSP. **B.** LTP magnitude estimated from the averaged enhancement of fEPSP slope observed 50-60 min, 110-120 min, 140-150 min and 170-180 min after *mild TBS (5x4)* in the absence of added drugs in young adult rats (6-7 weeks). **C.** Average LTP magnitude obtained 50-60 min theta-burst stimulation with increasingly stronger TBS stimulation paradigms: *mild TBS (5x4), TBS (10x4), moderate TBS (15x4)* and *strong TBS 3x (15x4)* bursts separated by 6 min in the absence of added drugs in young adult rats (6-7 weeks). **D.** Comparison of the average LTP magnitude obtained 50-60 min theta-burst stimulation with increasingly stronger TBS stimulation paradigms: *mild TBS (5x4), moderate TBS (15x4)* and *strong TBS 3x (15x4)* in *juvenile* (3 weeks), *young adult* (6-7 weeks) and *adult* (12 weeks) rats. Individual values and the mean ± S.E.M are depicted (**B.** - **D.**). *$p < 0.05$ (Student's t test) as compared to the fEPSP slope before LTP induction; †$p < 0.05$ (one-way *ANOVA*, Tukey's multiple comparison test) as compared with the potentiation obtained 50-60 min after *mild TBS (5x4)*.

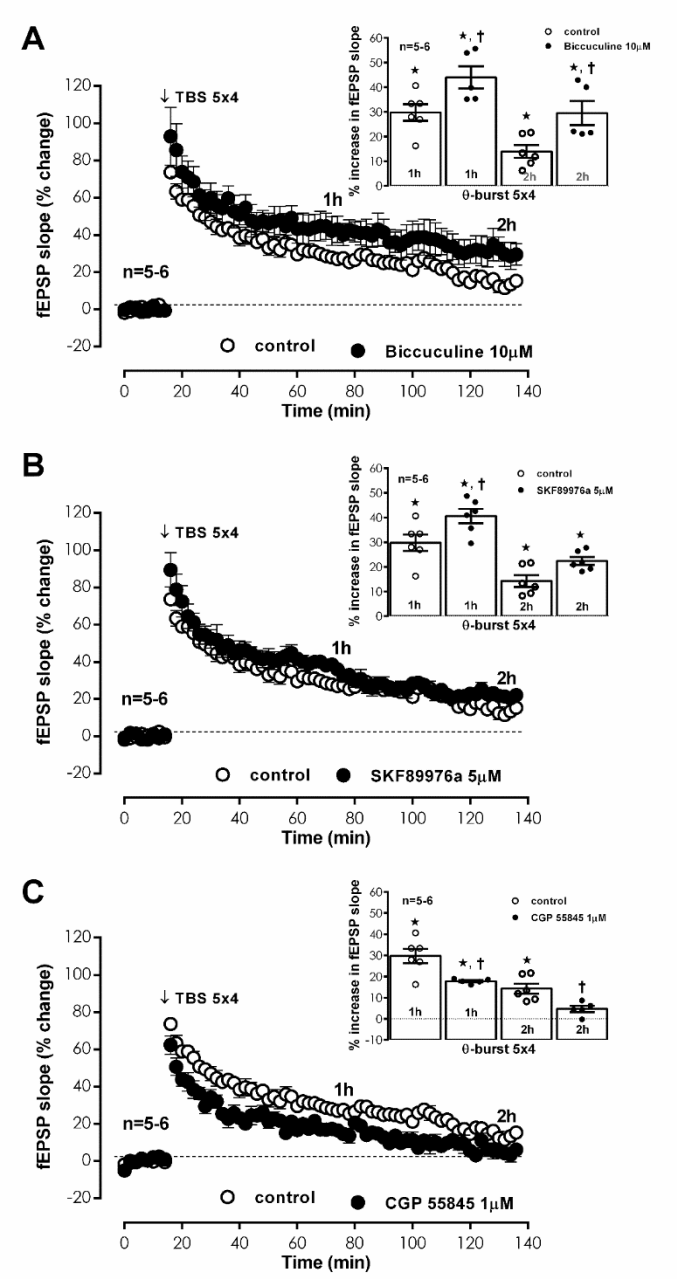

Figure 2 – Hippocampal CA1 LTP induced by *mild TBS (5x4)* is dependent on GABAergic transmission. A., B. and **C.** Averaged time-course of changes in fEPSP slope caused by theta-burst stimulation (*mild TBS (5x4)*) in the absence (-○-) and in the presence (-●-) of the selective $GABA_A$ receptor antagonist Bicuculline (10μM, **A.**), the selective GAT_1 GABA transporter inhibitor SKF8997A (5μM, **B.**) or the selective $GABA_B$ receptor antagonist CGP 55845 (1μM, **C.**). Control and test conditions (absence and presence added drugs) were tested in independent pathways in the same slice. ***Inset:*** Magnitude of LTP estimated from the averaged enhancement of fEPSP slope observed 50-60 and 110-120 min after *mild TBS (5x4)* in the absence (-○-) or in the presence (-●-) the selective $GABA_A$ receptor antagonist Bicuculline (10μM, **A.**), the selective GAT_1 GABA transporter inhibitor SKF8997A (5μM, **B.**) or the selective $GABA_B$ receptor antagonist CGP 55845 (1μM, **C.**). Individual values and the mean ± S.E.M are depicted (**A.** - **C.**). *$p < 0.05$ (Student's t test) as compared to the

fEPSP slope before LTP induction; †$p < 0.05$ (one-way *ANOVA*, Tukey's multiple comparison test) as compared with the potentiation obtained in different conditions and time points in the same slices.

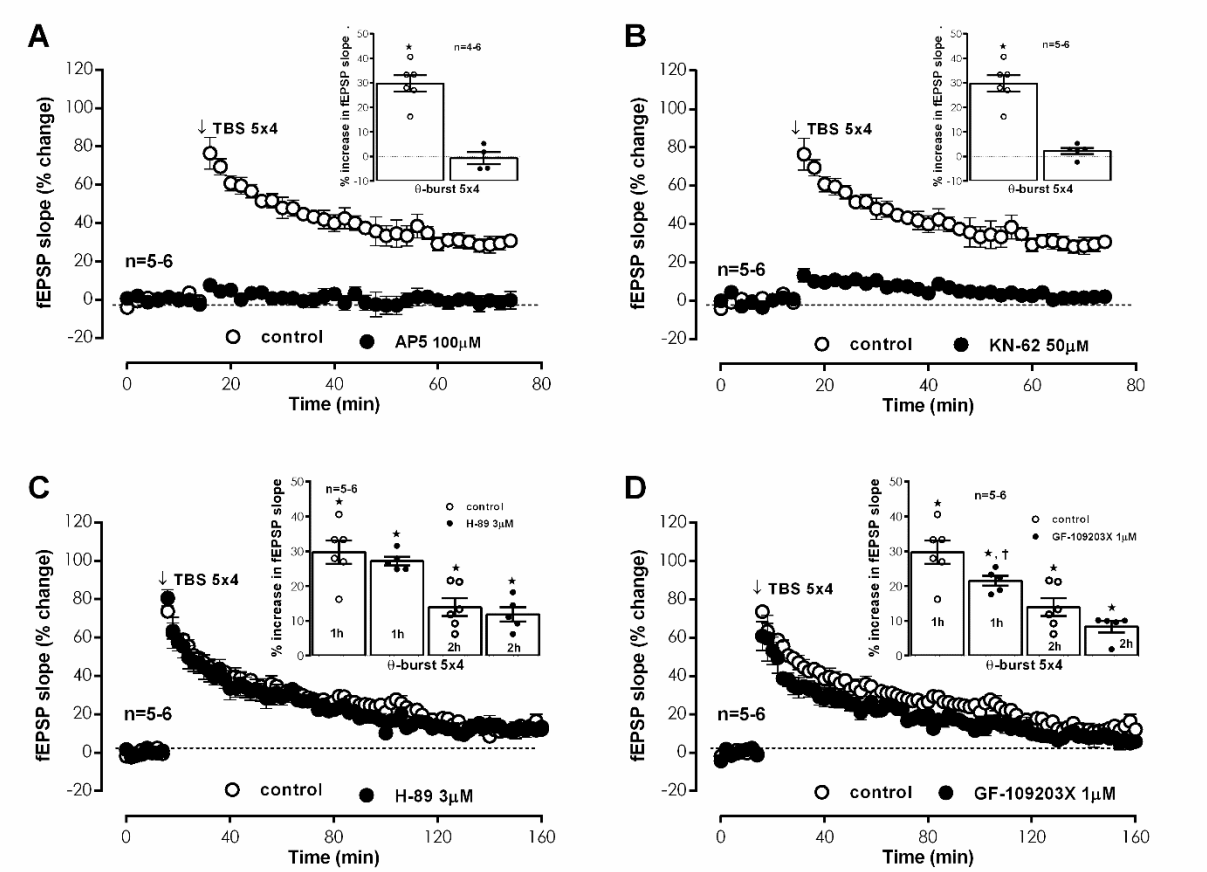

Figure 3 – Hippocampal CA1 LTP induced by *mild TBS (5x4)* in young adult rats is dependent on NMDA activation and CaMKII activity but not PKA or PKC activity. A.-D. Averaged time-course of changes in fEPSP slope induced by *mild TBS (5x4)* in the absence (-○-) and in the presence (-●-) of either the selective NMDA receptor antagonist AP5 (100µM, **A.**), the selective CaMKII inhibitor KN-62 (50µM, **B.**), the selective PKA inhibitor H-89 (3µM, **C.**) or the selective PKC inhibitor GF109203x (1µM, **D.**). Control and test conditions (absence and presence of added drugs) were tested in independent pathways in the same slice. ***Inset:*** Magnitude of LTP estimated from the averaged enhancement of fEPSP slope observed 50-60 min after theta-burst stimulation in the absence (-○-) or in the presence (-●-) of the NMDA receptor antagonist AP5 (100µM, **A.**), the selective CaMKII inhibitor KN-62 (50µM, **B.**), the selective PKA inhibitor H-89 (3µM, **C.**) or the selective PKC inhibitor GF109203x (1µM, **D.**). Individual values and the mean ± S.E.M are depicted (**A.** - **D.**).*$p < 0.05$ (Student's t test) as compared to the fEPSP slope before LTP induction; †$p < 0.05$ (one-way *ANOVA*, Tukey's multiple comparison test) as compared with the potentiation obtained in control conditions for the same time point in the same slices.

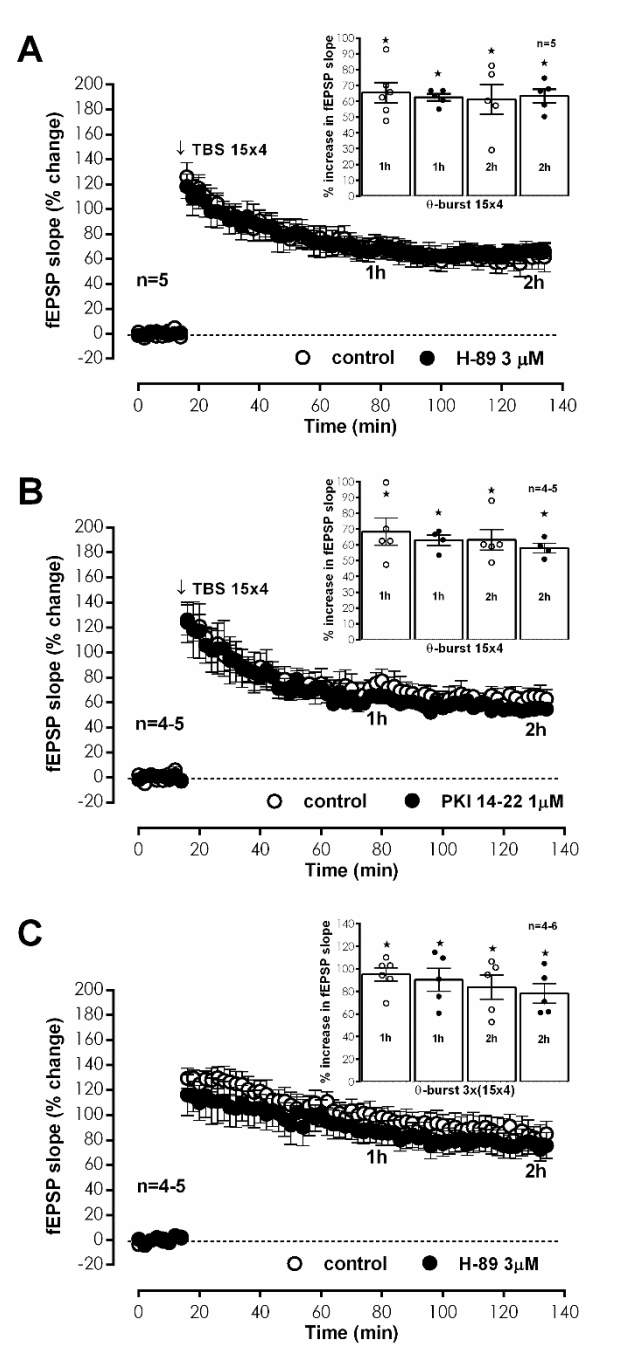

Figure 4 – Hippocampal CA1 LTP induced by *moderate TBS (15x4)* and *strong TBS 3x (15x4)* in adult rats is independent of PKA activity. A.-B. Averaged time-course of changes in fEPSP slope induced by *moderate TBS (15x4)* in the absence (-○-) and in the presence (-●-) of either the selective PKA inhibitor H-89 (3µM, **A.**) or another selective PKA inhibitor PKI 14-22 (1µM, **B.**). Control and test conditions (absence and presence of added drugs) were tested in independent pathways in the same slice obtained from adult rats. ***Inset:*** Magnitude of LTP estimated from the averaged enhancement of fEPSP slope observed 50-60 or 110-120 min after *moderate TBS (15x4)* in the absence (-○-) and in the presence (-●-) of either the selective PKA inhibitor H-89 (3µM, **A.**) or the selective PKA inhibitor PKI 14-

22 (1µM, **B.**).**C.** Averaged time-course of changes in fEPSP slope induced by *strong TBS 3x (15x4)* in the absence (-○-) and in the presence (-●-) of either the selective PKA inhibitor H-89 (3µM, **A.**) ***Inset:*** Magnitude of LTP estimated from the averaged enhancement of fEPSP slope observed 50-60 or 110-120 min after *strong TBS 3x (15x4)* in the absence (-○-) and in the presence (-●-) of the selective PKA inhibitor H-89 (3µM). Individual values and the mean ± S.E.M are depicted (**A.** - **C.**). *$p < 0.05$ (Student's t test) as compared to the fEPSP slope before LTP induction; †$p < 0.05$ (one-way *ANOVA*, Tukey's multiple comparison test) as compared with the potentiation obtained in control conditions for the same time point in the same slices.

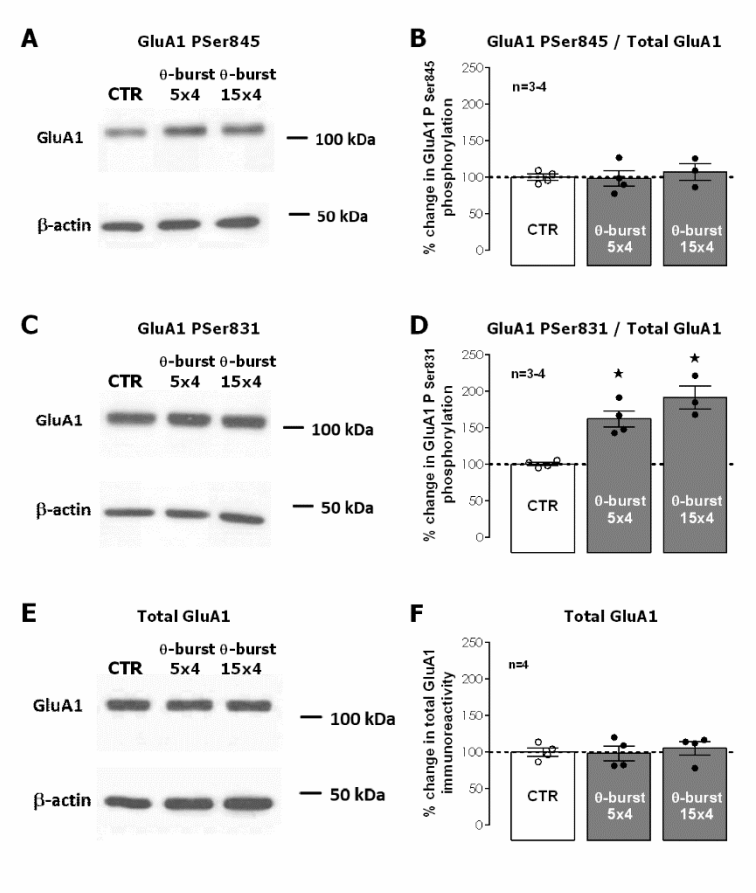

Figure 5 – Impact of theta-burst stimulation on phosphorylation of hippocampal AMPA GluA1 subunits on Ser_{845} and Ser_{831}. A., C. and **E.** Western-blot immunodetection of AMPA GluA1 phosphorylated forms in Ser_{845} and Ser_{831} and of total GluA1 subunits obtained in one individual experiment in which hippocampal slices were subjected to Schaffer collateral basal, *mild TBS (5x4)* or *moderate TBS (15x4)* stimulation. Slices were monitored for 50 min after TBS (or equivalent time for controls) before WB analysis. **B.** Averaged total GluA1 immunoreactivity and **D.** and **F.** % GluA1 phosphorylation on Ser_{845} and Ser_{831} residues of GluA1 subunits. Individual values and the mean ± S.E.M of four independent experiments performed in duplicate are depicted (**B., D.,** and **F**). 100% - averaged GluA1 immunoreactivity or GluA1 phosphorylation obtained in control conditions (absence of TBS). ***** represents $p < 0.05$ (*ANOVA*, Tukey's multiple comparison test) as compared to absence of TBS.

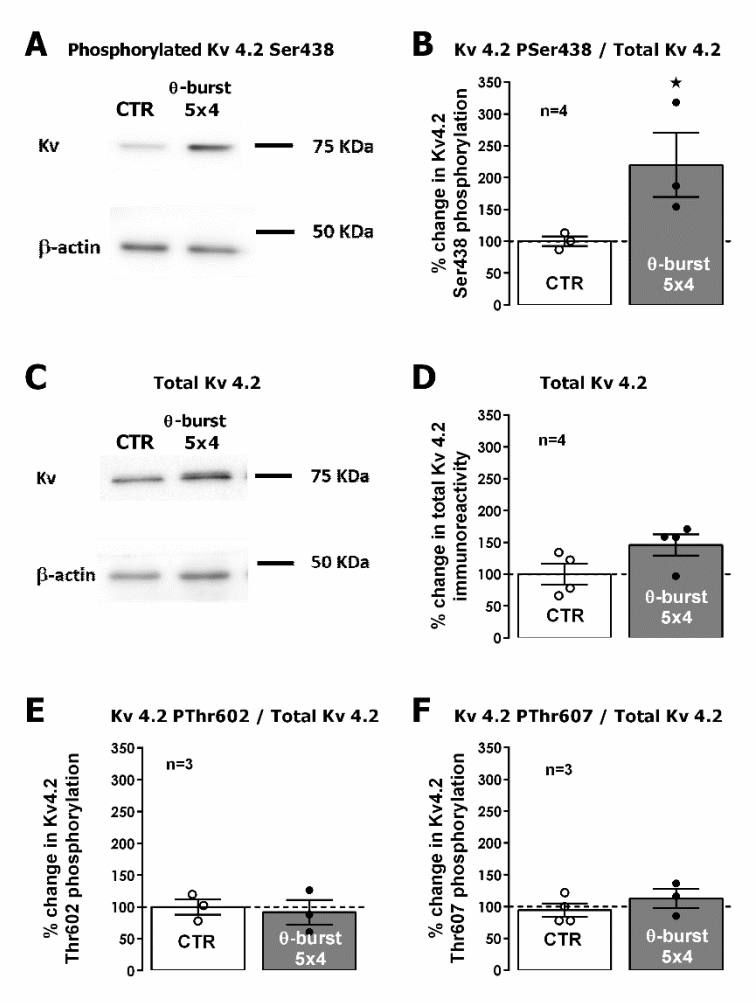

Figure 6 – Impact of theta-burst stimulation on phosphorylation of hippocampal Kv4.2 on Ser_{438} and Thr_{602} and $_{607}$. A. and **C.** Western blot immunodetection of Kv4.2 phosphorylated forms in Ser438 and total Kv4.2 obtained in one individual experiment in which hippocampal slices were subjected to Schaffer collateral basal or *mild TBS (5x4)* stimulation. Slices were monitored for 50 min after TBS (or equivalent time for controls) before WB analysis. **B., D., E,** and **F.** Averaged total Kv4.2 immunoreactivity and immunoreactivity of its phosphorylated forms in Ser_{438}, Thr_{602} or Thr_{607}, normalized to the total variation in Kv4.2. Individual values and the mean ± S.E.M of 3-4 independent experiments performed in duplicate are depicted (**B., D., E,** and **F**). 100% - averaged Kv4.2 immunoreactivity obtained in control conditions (absence of TBS). * represents $p < 0.05$ (*Student's t* test) as compared to absence of TBS.