Automatic literature mining tool to extract glycosylation information from literature

GlyGen
www.glygen.org

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. METHODS: We analyzed several human cancer cell lines as well as tissue homogenates using Western blotting and quantitative PCR for the

0,2,3,4,6,7,8,11,1

biotm.cis.udel.edu/glyco/pmid_large/28531887

Extracted text from Abstracts using the Text mining

pipeline

Processing Full-Length Articles

Pipeline: List of PMIDs → PMCID → Open Access set → xml files → parse xml

Enrichment of Cell Adhesion Molecules and Hematopoietic Cell Linage Markers in Serum Glycoproteins

Except for the component of complement and coagulation cascades, cell adhesion molecules and

retrieve results/conclusion sections and subsections --> run glycosylation extraction

hematopoietic cell linage markers (CD molecules) were also enriched in the identified serum glycoproteins

(supplementary Document S1). Most of these proteins are blood cell or neural cell surface proteins, and

they may shed or secreted from cells, such as CD44 protein. Once shedding from cell surface, the soluble

CD44 plays versatile biological functions which are different from cell adhesion (27). The N-glycosite N25

of CD44 locates at its extracellular part. One glycopeptide containing this site were identified using the 739

N-glycan masses. (supplemental Table S13). It is known that elevated levels of soluble CD44 in the serum

soluble CD44 may provide more information regarding to disease status. CD44 is also a marker for T cell

and erythrocyte lineage differentiation. It is reasonable to detect those CD markers in serum once they shed

from progenitor cells into circulation during cell differentiation. The alteration of N-glycosylation on these

Modified N-glycans Carried by Serum N-linked Glycoproteins The proposed method will achieve the highest

accuracy by using a complete database of N-glycans as well as N-linked deglycopeptides derived from

human serum. However, this database is not established to date. More than 140 N-glycans in serum was

Annotation of nine different glycan structures with varied degree of complexity and sialylation by Mascot on a

HCD MS2 spectra annotated by Mascot. The nine different glycopeptide variants included the mono-sialylated bi-

(A), tri- (B), tetra-antennary (C), and the di-sialylated bi- (D), tri- (E), tetra-antennary (F). Tri- (G,H) and tetra-

sialylated (I) glycan structures on the same glycosylation site were also annotated by Mascot.

single glycosylation site (Asn 93) of alpha-1-acid glycoprotein 1 in serum. Shown here are the representative

of patients is a marker of tumor burden and metastasis in several cancers (28), hence, the glycans on

CD markers may provide information of aberrant cell differentiation.

. Organelles of SK- Mel-28 cells were separated using continuous lodixanol gradients

11. N-glycosylation, , MT4-MMP, -, site

Output_Site_Fusion (sent_index, protein, sugar, site):

6. Glycosylation. . protein. .

11. MT4-MMP, site

11. MT4-MMP, -, Asn318



Presenter: Jeet Vora, Catherine Hayes

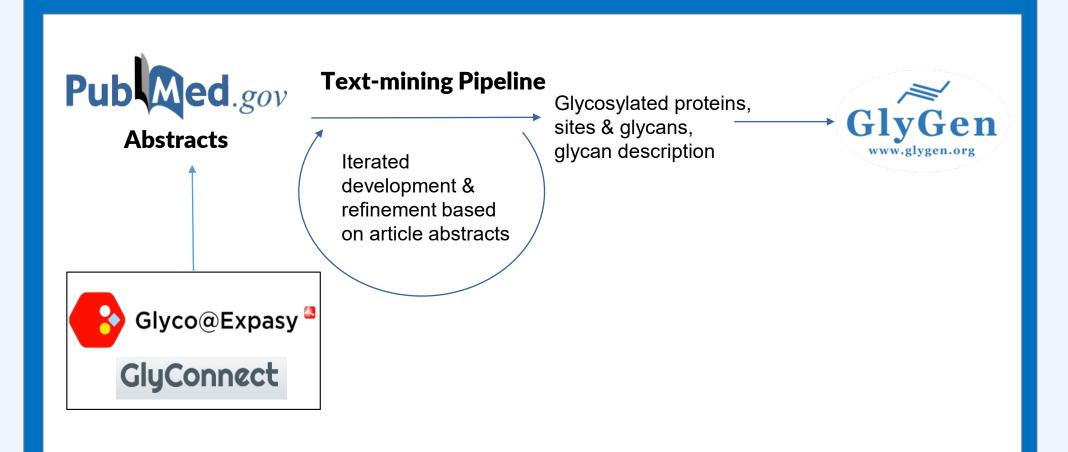
Abstract

Glycosylation is one of the most common and complex post-translational phenomena which impacts several key biological processes making it vital to study it with regard to human health and disease. Recent years have seen a great influx of data in glycosylation glycosylation in reference to disease, etc. However, transferring such data into existing bioinformatics databases for public use requires manual curation by experts, which is often time-consuming and expensive. Accelerating this process is key in facilitating biomedical research by providing the latest findings in a standardized way, ready for research use.

To facilitate Biocuration, we developed a literature mining tool that will detect glycosylated proteins, sites and glycan descriptions automatically in abstracts of publications from the MEDLINE resource. Extracted protein names are normalized to their NCBI gene ID and UniProtKB accession and the data is processed through manual and automatic quality control (QC) checks. The QC process assesses the validity of the protein accessions as well as the reported sites against fasta sequence from the UniProtKB database. The data that passes all QC checks is integrated in the GlyGen (https://www.glygen.org/) database and is publicly available.

Text mining workflow and pipeline Protein name, GenBank Acc (Nucleotide/Protein) Expression system: Cell type, Tissue Residue-Site Information Automatic UniProtKB Acc, GenBank Acc. Literature Mining Predominant glycan species Mass spectrometry data Cellosaurus ID, MeSH ID PubMed ID Align Residue-Site Information from Associated disease name paper to UniProtKB canonical Negative Results Inferred Data Disease Ontology ID (DOID) Glycan to Site specificityexplicit/fuzzy. QA/QC Mass spec data/glycan structure GlyTouCan Acc. Creating Data Objects in GlyGen GlyGen-Front End

Overview of the process shows how new abstracts and existing ones which have already been collected from UniProtKB, UniCarbKB and GlyTouCan are being mined for annotations.

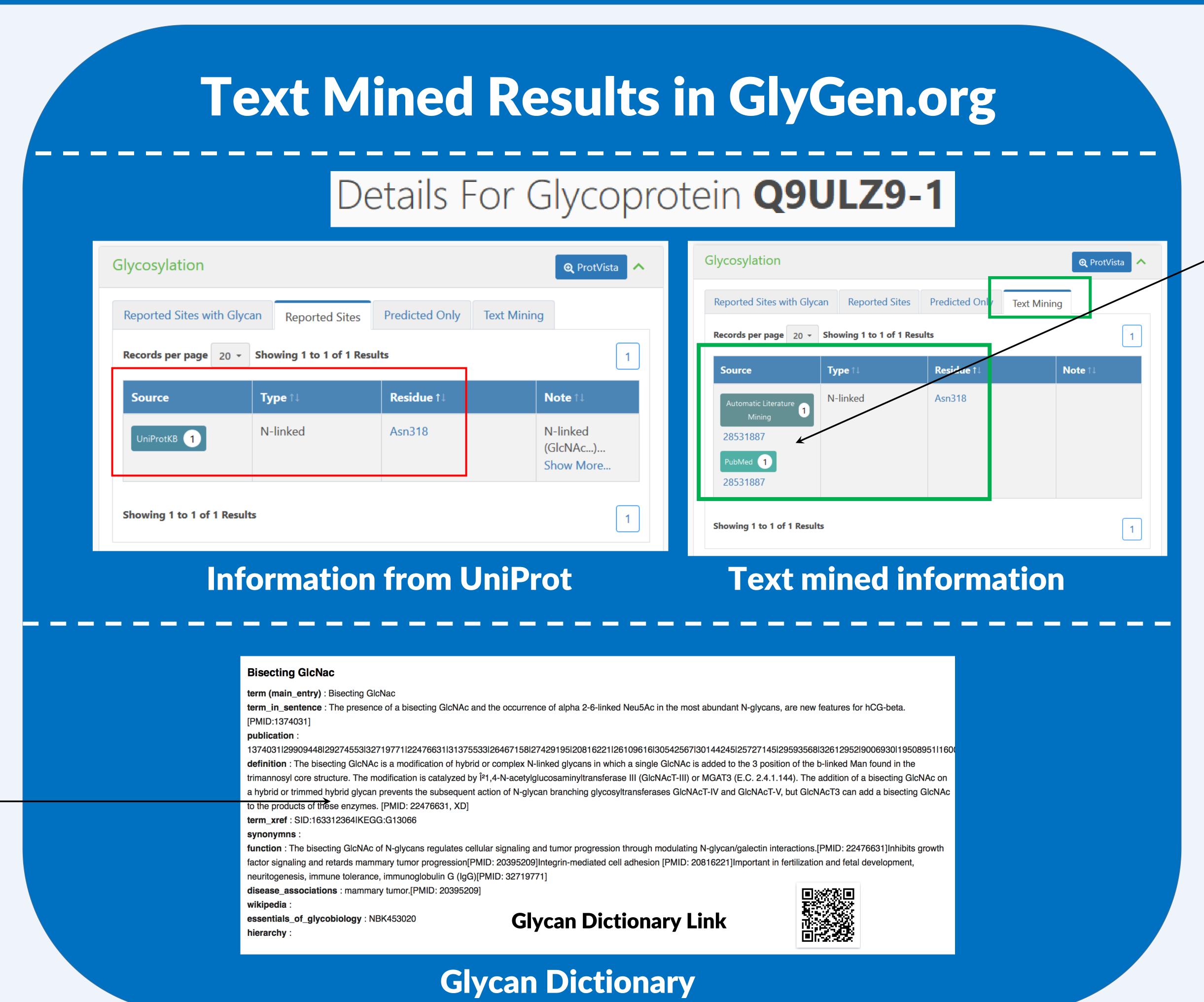


A multi-step process that involves iterations of refinement, with each step adding to the accuracy of the search parameters and algorithms

Term	PMID
Bisecting GlcNAc	1374031
Bisialo-biantennary complex	2386787
type	
core- fucosylated biantennary	10731668
complex-type oligosaccharide	

Extracted Glycan terms

Using chemical and glycobiological descriptions of glycans, a glycan detector was developed – allowed the extraction of motif names, glycan types, sequences etc. This was used to build a glycan dictionary.



The Human Glycosylation Sites [Automatic Literature Mining]

The Human Glycosylation of Sites Automatic Literature Mining (https://data.glygen.org/GLY_000481)

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contains 133 proteins, 241 sites from 160 abstracts.

Development of Protein Name and Detection and Normalization Tool

- Normalization of protein names by in house tool not normalised by PubTator
- Mapping of NCBI ids to UniProt accessions
- Created dictionary from UniProt with special processing for chains and subunits