Clustering Sheet for Flowcell **22KLGVLT3**

Name
Bartlomiej Gebarski

Sequencer
NovaSeqX1B

Run Type
NovaSeqX 10B XP flowcell PairedEnd 150 - single lane (r1: 150, r2: 150)

Clustering Date
2024-05-21

Sequencing Date
2024-05-21

Status
Started

1st Index Length
20

2nd Index Length
15

Comments
no comment

Reagents Checked Out
☐

num	Id	Scientist	Custom	PR	Comments
1	293535_R17174_1	[redacted]	☐	Standard	100pM 5% PhiX - 26.69µl of buffer (10mM Tris); - 6.46µl of 2.5nM library; - 0.85µl of 1nM PhiX; CRISPR_Screen
2	293628_R17179_1	[redacted]	☐	Standard	- 20.84µl of buffer (10mM Tris); - 11.63µl of 2.5nM library; - 1.53µl of 1nM PhiX; WGS
3	293629_R17179_2	[redacted]	☐	Standard	- 20.84µl of buffer (10mM Tris); - 11.63µl of 2.5nM library; - 1.53µl of 1nM PhiX; WGS
4	293630_R17179_3	[redacted]	☐	Standard	- 20.84µl of buffer (10mM Tris); - 11.63µl of 2.5nM library; - 1.53µl of 1nM PhiX; WGS
5	293632_R17179_5	[redacted]	☐	Standard	- 20.84µl of buffer (10mM Tris); - 11.63µl of 2.5nM library; - 1.53µl of 1nM PhiX; WGS
6	293633_R17179_6	[redacted]		Standard	120pM 1% PhiX - 23.88µl of buffer (10mM Tris); - 8.08µl of 2.5nM library; - 2.04µl of 0.1nM PhiX RNAseq
7	M18553_R17180_1	[redacted]	☐	Standard	120pM 1% PhiX - 23.88µl of buffer (10mM Tris); - 8.08µl of 2.5nM library; - 2.04µl of 0.1nM PhiX [various]
	⊎M18560_R17187_1	[redacted]			Solexa-Control
	⊎M18571_R17197_1				Solexa-Control
	⊎M18583_R17209_1				Solexa-Control
	⊎293927_R17194_1				DNA amplicon sequencing
8	M18562_R17189_1	[redacted]	☐	Standard	120pM 1% PhiX - 23.88µl of buffer (10mM Tris); - 8.08µl of 2.5nM library; - 2.04µl of 0.1nM PhiX ChIP-Seq
	⊎M18574_R17200_1	[redacted]			Solexa-Control
	⊎M18584_R17210_1				Solexa-Control
	⊎M18585_R17211_1				Solexa-Control

Instructions

1. Check, if instrument is idle. Check free disc space, delete old runs if necessary.
2. Thaw the reagents in water for 4h or in a 2°C to 8°C refrigerator for 48 hours. Inspect the position #12 well on the underside of the cartridge to make sure that the contents are free of ice, which indicates that the reagents are thawed. If the reagent cartridge cannot be loaded into the instrument within 24 hours, store at 2°C to 8°C for up to 72 hours or return to -25°C to -15°C storage for up to 7 days. After thawing, do not refreeze more than one time.
3. Prepare Cluster Sheet, Samples and all other reagents.
4. Take out the flowcell from the fridge to equilibrate it to room temperature (5-10min).
5. Thaw Lyo-insert and Pre-load-buffer at room temperature for 10 minutes.
6. Denature samples in Eppendorf tubes:
   - **21.35 µl** of buffer (10mM Tris);
   - **10.1 µl** of **2.5nM** library
   - **2.55 µl** of **0.1nM** PhiX
   - Add **8.5µl** of **0.2N** NaOH to each sample;
   - Incubate for 5 minutes;
   - Neutralize by ading **127.5µl** of **Pre-load Buffer**;
7. Uncap the library tube strip. Do not pierce the library tube strip foils.
8. Dispense **165µl** denatured library or denatured library with PhiX into each sample tube.
9. After dispensing libraries, cap the library tube strip. Make sure that no air gaps are present at the bottom of the tubes (if yes, spin down briefly).
10. Insert the library tube strip into the reagent cartridge and press down.
11. Insert the lyo insert into the reagent cartridge and press down.
12. Log in to the software and start new run.
13. Follow instructions on the screen.