

# Class10 - Structural Bioinformatics pt 1

AUTHOR

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## 1. Introduction to the RCSB Protein Data Bank (PDB)

The main repository of biomolecular structure data is called **PDB** ([Protein Data Bank](#)). It's the second oldest database, after GenBank.

### PDB Statistics

The CSV file of the PDB data distributions. It was accessed by: "Analyze" > "PDB Statistics" > "by Experimental Method and Molecular Type"

```
PDBdata <- read.csv("Data Export Summary.csv", row.names = 1)
```

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

```
xray <- gsub(",", "", PDBdata$X.ray) #substitute comma for n h  
xray <- as.integer(xray) # converting to numeric.  
sum(xray)
```

```
[1] 193952
```

Now, we turn this snippet into a function, so we can reuse it for something else.

```
comma.sum <- function(x) {  
  y <- gsub(",", "", x)  
  y <- as.integer(y)  
  return(sum(y))  
}
```

```
#Try it!  
comma.sum(PDBdata$X.ray)
```

```
[1] 193952
```

Now we answer the question!

```
xray.sum <- comma.sum(PDBdata$X.ray)
```

```
EM.sum <- comma.sum(PDBdata$EM)
total.sum <- comma.sum(PDBdata$Total)

#Division:
(xray.sum+EM.sum)/total.sum*100 #93.7% For both EM and X Ray
```

```
[1] 93.6787
```

```
xray.sum/total.sum*100 #82.4%
```

```
[1] 82.37223
```

```
EM.sum/total.sum*100 #11.3%
```

```
[1] 11.30648
```

- X-ray solved **82.4%** of PDB structures
- EM solved **11.3%** of PDB structures

Q2: What proportion of structures in the PDB are protein (only)?

```
comma.sum(PDBdata["Protein (only)","Total"])/comma.sum(PDBdata$
```

```
[1] 86.2107
```

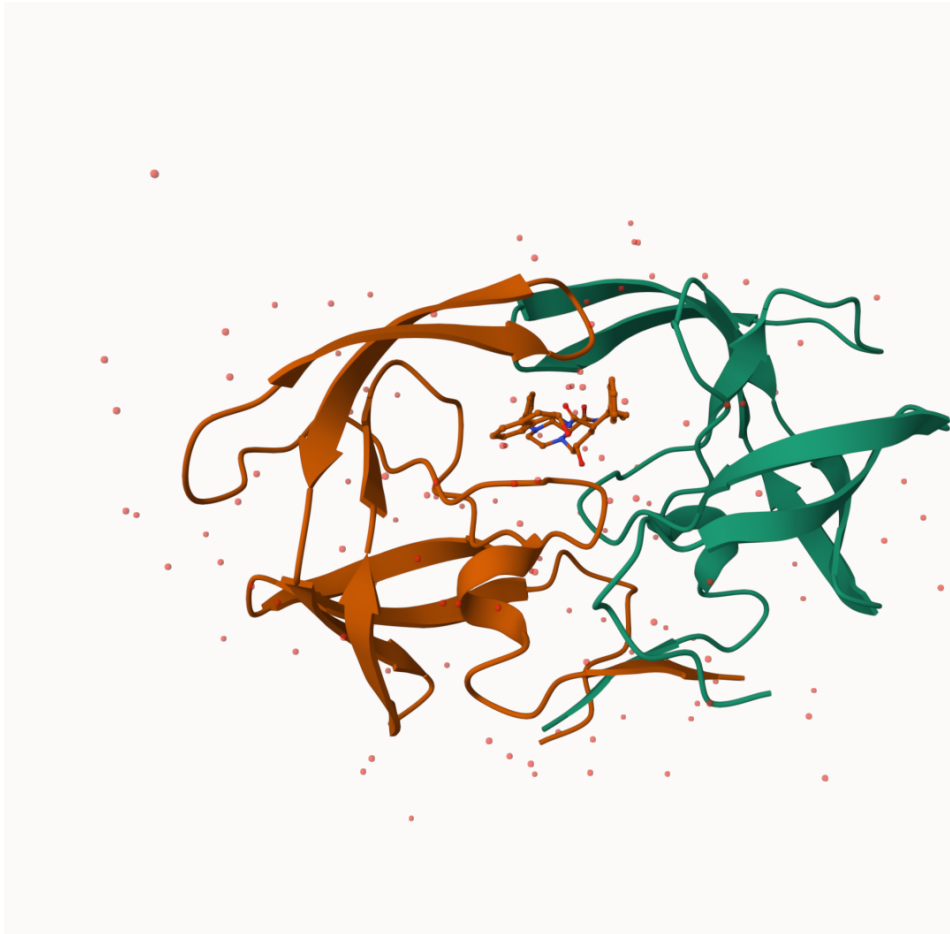
**86.2%** of the structures in PDB is just the **protein (only)**

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

(SKIPPED)

## 2. Visualizing the HIV-1 protease structure

Explore the HIV-1 protease structure with PDB code: **1HSG** Molstar homepage [here](#).



This is a protease protein, that cuts off aspartic amino acid in HIV replication. The small molecule inhibitor is a Merck Drug (MK1). The hydrogen is not shown in this structure, because it's too small compared to the resolution (of 2Å). We can change the molecular views

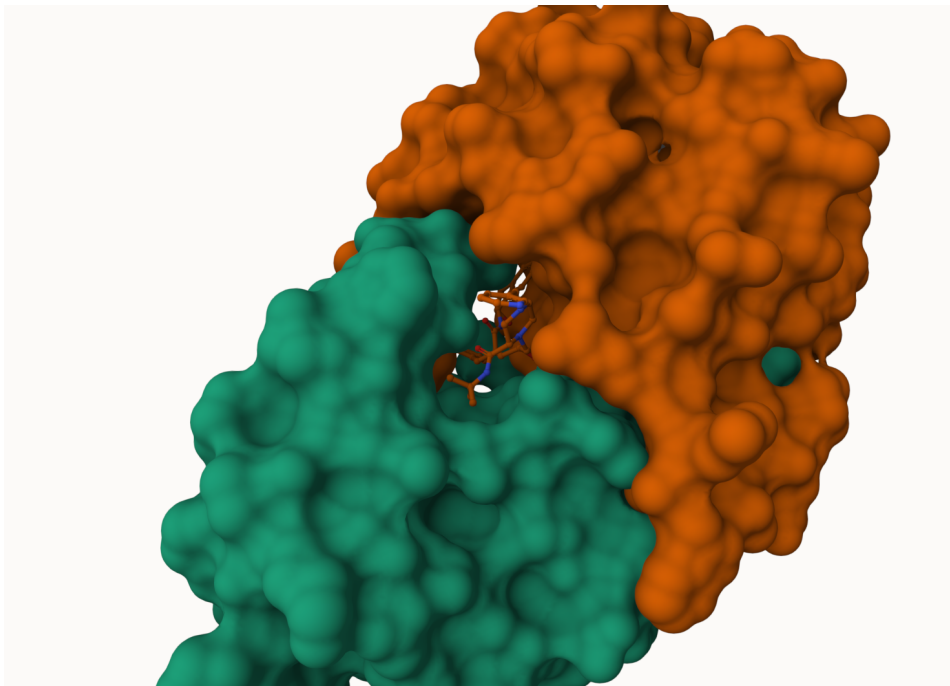


Fig 2. A molecular surface view showing binding cavity



Fig 3. The catalytically important Asp25 amino acids and drug-interacting HOH 308 water molecule

So far, we've used the MolStar website to view protein structures, but we haven't done much bioinformatics.

### 3. Using the Bio3D in R

First, install & load the bio3D package. It is focused on structural bioinformatics analysis and allows us to read and analyze PB (and related) data.

```
library(bio3d)
```

Read in some data from PDB

```
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
pdb
```

Call: `read.pdb(file = "1hsg")`

```

Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A
B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]

Protein sequence:

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD

QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE

ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

+ attr: atom, xyz, seqres, helix, sheet,
        calpha, remark, call

```

```

#lookng at the attributes in the pdb data:
attributes(pdb)

```

```

$names
[1] "atom" "xyz" "seqres" "helix" "sheet" "calpha"
"remark" "call"

$class
[1] "pdb" "sse"

```

```

#open one of the attributes.
head(pdb$atom)

```

```

type eleno elety alt resid chain resno insert      x      y
z o      b
1 ATOM      1      N <NA>  PR0      A      1      <NA> 29.361 39.686
5.862 1 38.10
2 ATOM      2      CA <NA>  PR0      A      1      <NA> 30.307 38.663
5.319 1 40.62
3 ATOM      3      C  <NA>  PR0      A      1      <NA> 29.760 38.071
4.022 1 42.64
4 ATOM      4      O <NA>  PR0      A      1      <NA> 28.600 38.302
3.676 1 43.40
5 ATOM      5      CB <NA>  PR0      A      1      <NA> 30.508 37.541
6.342 1 37.87
6 ATOM      6      CG <NA>  PR0      A      1      <NA> 29.296 37.591
7.162 1 38.40
      segid elesy charge
1 <NA>      N  <NA>

```

2	<NA>	C	<NA>
3	<NA>	C	<NA>
4	<NA>	O	<NA>
5	<NA>	C	<NA>
6	<NA>	C	<NA>

```
head(pdbseq(pdb))
```

```
  1    2    3    4    5    6
"P" "Q" "I" "T" "L" "W"
```

Q7: How many amino acid residues are there in this pdb object?

There are **198 amino acid** (198 alpha carbons, each is an amino acid)

Q8: Name one of the two non-protein residues?

**Water** (HOH) is one of them.

Q9: How many protein chains are in this structure?

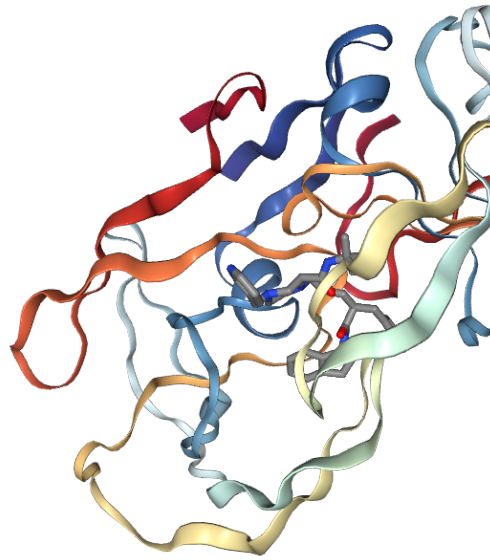
There are **2** protein chains, A and B.

## Molecular Visualization in R (interactive!)

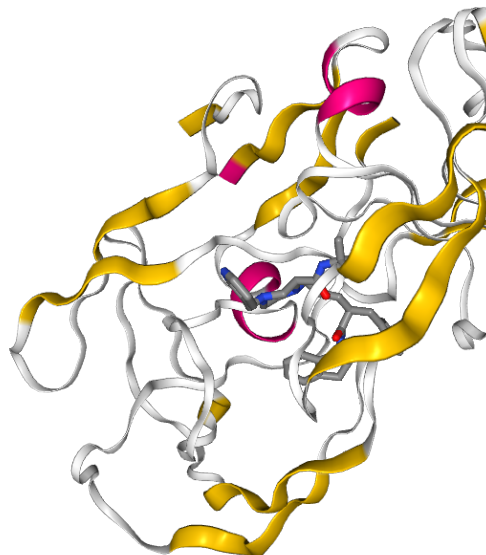
We can make a quick 3D vizualisation with the `bio3dview` package. Use the following to install.

```
{r} install.packages("pak") pak::pak("bioboot/bio3dview")
install.packages("NGLViewer")
```

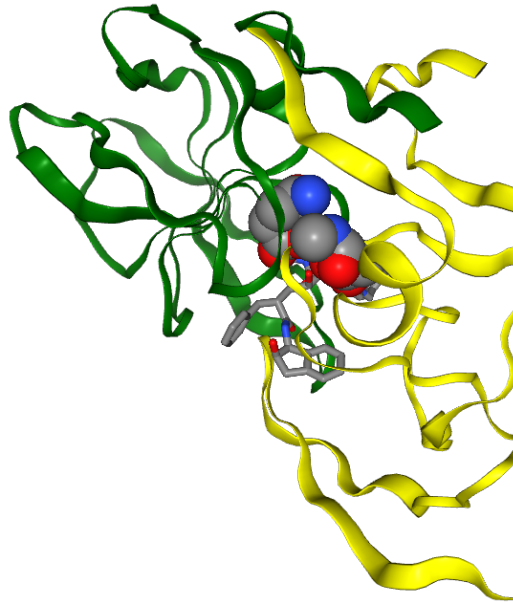
```
library(bio3dview)
library(NGLViewer)
view.pdb(pdb) |>
  setSpin()
```



```
view.pdb(pdb, backgroundColor = "black", colorScheme = "sse") |:
```



```
sel <- atom.select(pdb, resno=25)
view.pdb(pdb, cols = c("green", "yellow"),
         highlight= sel, highlight.style= "spacefill") |> setRo
```



## Predicting Functional motions of a single structure

We can finish off today with a bioinformatics prediction of functional motions of a protein.

We will run a Normal Mode Analysis (NMA) – it inserts an energy into the prediction

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file  
PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

Total Models#: 1



Total Atoms#: 1898, XYZs#: 5694 Chains#: 1 (values: A)

Protein Atoms#: 1654 (residues/Calpha atoms#: 214)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244 (residues: 244)

Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

Protein sequence:

MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGDMRLAAVKSGSELGKQAKDIMDAGKLV

DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVDYVLEFDVPDELIVDKI

VGRRVHAPSGRVYHVKNPPKVEGKDDVTGEELTRKDDQEETVRKRLVEYHQM TAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG

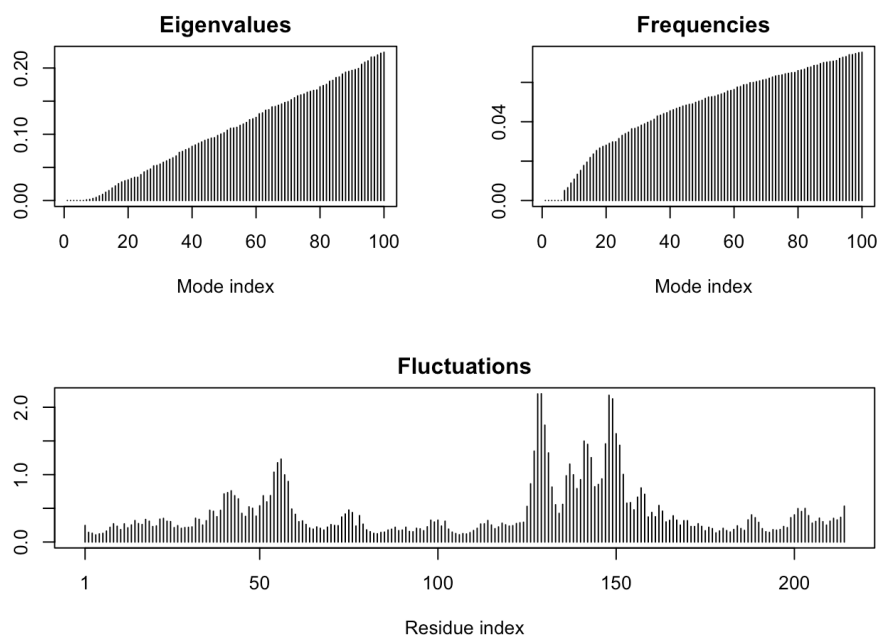
+ attr: atom, xyz, seqres, helix, sheet,  
calpha, remark, call

```
m <- nma(adk)
```

Building Hessian... Done in 0.044 seconds.

Diagonalizing Hessian... Done in 0.516 seconds.

```
plot(m)
```



We can write our a trajectory of the predicted dynamics and view in molstar.

```
mktrj(m, file="adk_m7.pdb")
```

This makes it into a playable animation in Molstar if we import it from files.