selscan: an efficient multi-threaded program to perform EHH-based scans for positive selection

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Abstract

Haplotype-based scans to detect natural selection are useful to identify recent or ongoing positive selection in genomes.

As both real and simulated genomic datasets grow larger, spanning thousands of samples and millions of markers, there As both real and simulated genomic datasets grow larger, spanning thousands of samples and millions of markers, there is a need for a fast and efficient implementation of these scans for general use. Here we present selscan, an efficient multi-threaded application that implements Extended Haplotype Homozygosity (EHH), Integrated Haplotype Score (iHS), and Cross-population Extended Haplotype Homozygosity (XPEHH). selscan accepts phased genotypes in multiple formats, including TPED, and performs extremely well on both simulated and real data and over an order of magnitude faster than existing available implementations. It calculates iHS on chromosome 22 (22,147 loci) across 204 CEU haplotypes in 353s on one thread (33s on 16 threads) and calculates XPEHH for the same data relative to 210 YRI haplotypes in 578s on one thread (52s on 16 threads). Source code and binaries (Windows, OSX and Linux) are available code and binaries (Windows, OSX and Linux) are available at https://github.com/szpiech/selscan.

1 ${f Introduction}$

Extended Haplotype Homozygosity (EHH) (Sabeti et al., 2002), Integrated Haplotype Score (iHS) (Voight et al., 2006), and Cross-population Extended Haplotype Homozygosity (XPEHH) (Sabeti et al., 2007) are statistics designed to use phased genotypes to identify putative regions of recent or ongoing positive selection in genomes. They are all based on the model of a hard selective sweep, where a de novo adaptive mutation arises on a haplotype that quickly sweeps toward fixation, reducing diversity around the locus. If selection is strong enough, this occurs faster than recombination or mutation can act to break up the haplotype, and thus a signal of high haplotype homozygosity can be observed extending from an adaptive locus.

As genetics data sets grow larger both in number of individuals and number of loci, there is a need for a fast and efficient publicly available implementation of these statistics. Below we introduce these statistics and provide concise definitions for their calculations. We then evaluate the performance of our implementation, selscan.

Extended Haplotype Homozygosity 1.1

In a sample of n chromosomes, let \mathcal{C} denote the set of all possible distinct haplotypes at a locus of interest (named x_0), and let $C(x_i)$ denote the set of all possible distinct haplotypes extending from the locus x_0 to the i^{th} marker either upstream or downstream from x_0 . For example, if the locus of interest x_0 is a biallelic SNP where 0 represents the ancestral allele and 1 represents the derived allele, then $\mathcal{C} := \{0,1\}$. If x_1 is an immediately adjacent marker, then the set of all possible haplotypes is $C(x_1) := \{11, 10, 00, 01\}.$

EHH of the entire sample, extending from the locus x_0 out to marker x_i , is calculated as

$$EHH(x_i) = \sum_{h \in \mathcal{C}(x_i)} \frac{\binom{n_h}{2}}{\binom{n}{2}},\tag{1}$$

where n_h is the number of observed haplotypes of type $h \in$ $\mathcal{C}(x_i)$.

In some cases, we may want to calculate the haplotype homozygosity of a sub-sample of chromosomes all carrying a 'core' haplotype at locus x_0 . Let $\mathcal{H}_c(x_i)$ be a partition of $\mathcal{C}(x_i)$ containing all distinct haplotypes carrying the core haplotype, $c \in \mathcal{C}$, at x_0 and extending to marker x_i . Note that

$$C(x_i) = \bigcup_{c \in C} \mathcal{H}_c(x_i). \tag{2}$$

Following the example above, if the derived allele (1) is chosen as the core haplotype, then $\mathcal{H}_1(x_1) := \{11, 10\}$. Similarly, if the ancestral allele is the core haplotype, then $\mathcal{H}_0(x_1) :=$ {00,01}

We calculate the EHH of the chromosomes carrying the core haplotype c to marker x_i as

$$EHH_c(x_i) = \sum_{h \in \mathcal{H}_c(x_i)} \frac{\binom{n_h}{2}}{\binom{n_c}{2}},$$
 (3)

where n_h is the number of observed haplotypes of type $h \in$ $\mathcal{H}_c(x_i)$ and n_c is the number of observed haplotypes carrying the core haplotype $(c \in \mathcal{C})$.

1.2 Integrated Haplotype Score

iHS is calculated by using Equation 3 to track the decay of haplotype homozygosity for both the ancestral and derived haplotypes extending from a query site. To calculate iHS at a site, we first calculate the integrated haplotype homozygosity (iHH) for the ancestral (0) and derived (1) haplotypes ($\mathcal{C} := \{0,1\}$) via trapezoidal quadrature.

$$iHH_{c} = \sum_{i=1}^{|\mathcal{D}|} \frac{1}{2} \left(EHH_{c}(x_{i-1}) + EHH_{c}(x_{i}) \right) g(x_{i-1}, x_{i}) + \sum_{i=1}^{|\mathcal{U}|} \frac{1}{2} \left(EHH_{c}(x_{i-1}) + EHH_{c}(x_{i}) \right) g(x_{i-1}, x_{i}), \quad (4)$$

where \mathcal{D} is the set of markers downstream from the current locus such that $x_i \in \mathcal{D}$ denotes the i^{th} closest downstream marker from the locus of interest (x_0) . \mathcal{U} and $x_i \in \mathcal{U}$ are defined similarly for upstream markers. $g(x_{i-1}, x_i)$ gives the genetic distance between two markers. The (unstandardized) iHS is then calculated as

$$\ln\left(\frac{iHH_1}{iHH_0}\right).$$
(5)

Note that this definition differs slightly from that in Voight *et al.* (2006), where unstandardized iHS is defined with iHH_1 and iHH_0 swapped.

Finally, the unstandardized scores are normalized in frequency bins across the entire genome.

$$iHS = \frac{\ln\left(\frac{iHH_1}{iHH_0}\right) - E_p\left[\ln\left(\frac{iHH_1}{iHH_0}\right)\right]}{SD_p\left[\ln\left(\frac{iHH_1}{iHH_0}\right)\right]},\tag{6}$$

where $E_p\Big[\ln\Big(\frac{iHH_1}{iHH_0}\Big)\Big]$ and $SD_p\Big[\ln\Big(\frac{iHH_1}{iHH_0}\Big)\Big]$ are the expectation and standard deviation in frequency bin p.

In practice, the summations in Equation 4 are truncated once $EHH_c(x_i) < 0.05$. Additionally with low density SNP data, if the physical distance b (in kbp) between two markers is > 20, then $g(x_{i-1}, x_i)$ is scaled by a factor of 20/b in order to reduce possible spurious signals induced by lengthy gaps. During computation if the start/end of a chromosome arm is reached before $EHH_c(x_i) < 0.05$ or if a gap of b > 200 is encountered, the iHS calculation is aborted for that locus. iHS is not reported at core sites with minor allele frequency < 0.05. In selscan, the EHH truncation value, gap scaling factor, and core site MAF cutoff value are all flexible parameters definable on the command line.

1.3 Cross-population Extended Haplotype Homozygosity

To calculate XPEHH between populations A and B at a marker x_0 , we first calculate iHH for each population separately, integrating the EHH of the entire sample in the population.

ulation (Equation 1).

$$iHH = \sum_{i=1}^{|\mathcal{D}|} \frac{1}{2} \left(EHH(x_{i-1}) + EHH(x_i) \right) g(x_{i-1}, x_i) + \sum_{i=1}^{|\mathcal{U}|} \frac{1}{2} \left(EHH(x_{i-1}) + EHH(x_i) \right) g(x_{i-1}, x_i)$$
 (7)

If iHH_A and iHH_B are the iHHs for populations A and B, then the (unstandardized) XPEHH is

$$\ln\left(\frac{iHH_A}{iHH_B}\right),$$
(8)

and after genome-wide normalization we have

$$XPEHH = \frac{\ln\left(\frac{iHH_A}{iHH_B}\right) - E\left[\ln\left(\frac{iHH_A}{iHH_B}\right)\right]}{SD\left[\ln\left(\frac{iHH_A}{iHH_B}\right)\right]}.$$
 (9)

In practice, the sums in each of iHH_A and iHH_B (Equation 7) are truncated at x_i —the marker at which the EHH of the haplotypes pooled across populations is $EHH(x_i) < 0.05$. Scaling of $g(x_{i-1}, x_i)$ and handling of gaps is done as for iHS, and these parameters are definable on the selscan command line.

2 Performance

Here evaluate the performance of selscan (https://github.com/szpiech/selscan) for computing the iHS and XPEHH statistics. In addition, we compare performance on these statistics programs rehh (Gautier and Vitalis, 2012, with the http://cran.r-project.org/package=rehh), ihs (Voight et al., 2006) and xpehh (Pickrell et al., 2009). Both ins and xpehh are available for download at http://hgdp.uchicago.edu/Software/. All computations were run on a MacPro running OSX 10.8.5 with two 2.4 GHz 6-core Intel Xeon processors with hyperthreading enabled.

2.1 iHS

For runtime evaluation of iHS calculations, we simulated a 4 Mbp region of DNA with the program ms (Hudson, 2002) and generated four independent data sets with varying numbers of sampled haplotypes ($\theta = 1600$ and $\rho = 1600$). We sampled 250 haplotypes (9,625 SNP loci), 500 haplotypes (10,646 SNP loci), 1,000 haplotypes (11,655 SNP loci), and 2,000 haplotypes (12,724 SNP loci). We name these data sets IHS250, IHS500, IHS1000, IHS2000, respectively. These data sets represent a densely typed region similar to next-generation sequencing data. Although these data sets are generated via strictly neutral processes, they serve the purpose of runtime evaulation perfectly well. We also use data from The 1000 Genomes Project (The 1000 Genomes Project Consortium,

2012) Omni genotypes, calculating iHS scores at 22, 147 SNP loci on chromosome 22 across 102 CEU individuals (204 haplotypes). We name this data set CEU22.

Table 1 summarizes the runtimes of ihs, rehh, and selscan. We note that rehh integrates haplotype homozygosity over a physical map, whereas ihs and selscan integrate over a genetic map by default. This does not affect runtimes (data not shown), which are measured using genetic maps for ihs and selscan. Even operating on a single thread, selscan calculates iHS scores at least an order of magnitude faster than ihs and up to 1.8x faster than rehh for large data sets.

We compare unstandardized iHS scores for the CEU22 data set using ihs and selscan and find excellent agreement (Figure 1A, Pearson's r=0.9946). The slight variance in scores between the two programs is likely due to an undocumented difference in the way ihs calculates its scores (Sabeti *et al.* (2007) Supplemental Information), but the effect is negligible. We also calculate unstandardized iHS scores for the CEU22 data set using rehh and selscan (using a physical map) and again find excellent agreement (Pearson's r=0.9953).

2.2 **XPEHH**

For runtime evaluation of XPEHH calculations, we simulated a 4 Mbp region of DNA with the program ms (Hudson, 2002) with a simple two population divergence model (time to divergence $t=0.05, \theta=1600$ and $\rho=1600)$ and generated four independent data sets with varying numbers of sampled haplotypes. We sampled 250 haplotypes (125) from each population, 12,920 SNP loci), 500 haplotypes (250 from each population, 14,989 SNP loci), 1,000 haplotypes (500 from each population, 17,142 SNP loci), and 2,000 haplotypes (1,000 from each population, 19,567 SNP) loci). We name these data sets XP250, XP500, XP1000, XP2000, respectively. These data sets represent a densely typed region similar to next-generation sequencing data. Although these data sets are generated via strictly neutral processes, they serve the purpose of runtime evaulation perfectly well. We also use data from The 1000 Genomes Project (The 1000 Genomes Project Consortium, 2012) Omni genotypes, calculating XPEHH scores at 22, 147 SNP loci on chromosome 22 across 102 CEU individuals (204 haplotypes) and 105 YRI individuals (210 haplotypes). We name this data set CEUYRI22.

Table 2 summarizes the runtimes of xpehh and selscan. Even operating on a single thread, selscan tends to calculate XPEHH scores at least an order of magnitude faster than xpehh. Figure 1B shows the correlation (Pearson's r=0.9999) of CEUYRI22 unstandardized XPEHH scores between the two programs.

3 Conclusions

selscan achieves a speed up of at least an order or magnitude over both ihs and xpehh and a speed up of nearly 2x over rehh for large data sets through general optimizations of the calculations. We also implement shared memory parallelism with multithreading to further speed up calculations on computers with multiple cores. Since iHS and XPEHH attempt to calculate a score for each site in the data and each score can be calculated indpendently of the others, selscan partitions the workload (sites at which to calculate a score) across threads, while maintaining each thread's access to the entire data set required to make the calculation.

Additional empirical testing (data not shown) suggests that rehh, ihs, and selscan (for both iHS and XPEHH calculations) are $O(ND^2)$, and xpehh is $O(N^2D^2)$, where N is the number of haploid samples and D is the SNP locus density.

Each of these statistics require phased haplotypes and a genetic or physical map as input data (TPED format) and missing genotypes must either be dropped or imputed. Because of the speed improvements we have implented, we expect that selscan will be a valuable tool for calculating EHH-based genome-wide scans for positive selection in very large genetic data sets, including whole genome sequencing and GWAS data, currently being generated for humans and other organisms. selscan will also allow for in-depth examination of the performance of these statistics under a wide range of parameters in large scale simulation studies.

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Table 1: Runtime performance (in seconds) of ihs, rehh, and selscan for calculating unstandardized iHS for various data sets. Calculations running over 100,000 seconds were aborted. *rehh integrates over a physical map instead of a genetic map. Using a physical map does not affect selscan's runtime (data not shown).

| Data Cat | ihs | rehh* | selscan | | | | |
|----------|-----------|--------|-------------|-------|-------|-------|-----|
| Data Set | | | threads = 1 | 2 | 4 | 8 | 16 |
| IHS250 | 19,275 | 563 | 618 | 306 | 162 | 84 | 58 |
| IHS500 | 45,547 | 1,652 | 1,554 | 782 | 399 | 220 | 150 |
| IHS1000 | > 100,000 | 4,834 | 4,018 | 2,019 | 1,040 | 566 | 380 |
| IHS2000 | > 100,000 | 12,652 | 7,054 | 3,633 | 1,869 | 1,046 | 752 |
| CEU22 | 19,434 | 588 | 353 | 182 | 93 | 50 | 33 |

Table 2: Runtime performance (in seconds) of xpehh and selscan for calculating unstandardized XPEHH for various data sets. Calculations running over 100,000 seconds were aborted.

| Γ | Data Set | xpehh | selscan | | | | |
|---|----------|-----------|-------------|-------|-------|-----|-----|
| | | | threads = 1 | 2 | 4 | 8 | 16 |
| Γ | XP250 | 11,113 | 287 | 141 | 71 | 38 | 25 |
| | XP500 | 57,006 | 766 | 403 | 194 | 104 | 67 |
| | XP1000 | > 100,000 | 2,037 | 1,018 | 515 | 274 | 180 |
| | XP2000 | > 100,000 | 5,683 | 2,798 | 1,471 | 763 | 493 |
| | CEUYRI22 | 37,271 | 578 | 291 | 150 | 78 | 52 |

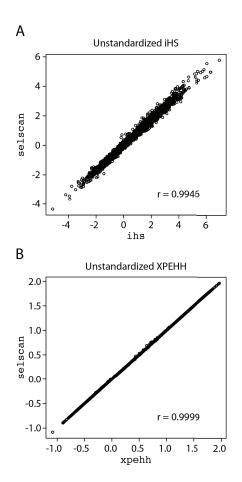


Figure 1: (A) Unstandardized iHS scores calculated on the CEU22 data set for selscan and ihs (Pearson's r=0.9946) and (B) Unstandardized XPEHH scores calculated on the CEUYRI22 data set for selscan and xpehh (Pearson's r=0.9999)