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# The DNA Chrysalis Hypothesis: Investigating Novel Biomolecule Preservation in Extreme Geological Environments

# Potential Principal Investigators (reach out):

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- Dr. Sean Gulick (University of Texas at Austin, USA) Marine Geophysicist & Geologist
- **Dr. Ludovic Orlando** (University of Copenhagen, Denmark) Bioinformatician & Ancient Genomics Expert
- Dr. Karen Lloyd (University of Tennessee, USA) Geomicrobiologist & Extreme Environments Specialist

### **Project Overview:**

The study of ancient DNA (aDNA) has revolutionized our understanding of past life, but its recovery is fundamentally limited by a theoretical maximum survival time of approximately 6.8 million years, with the oldest authenticated aDNA reaching 2 million years [1, 2]. This barrier prevents direct genetic investigation of the Mesozoic Era, including non-avian dinosaurs (extinct ~66 million years ago). This proposal introduces the "DNA Chrysalis Hypothesis," which posits a unique, active preservation mechanism for biomolecules in extreme geological settings. We propose an interdisciplinary expedition to test this hypothesis by targeting specific deep-time environments with the potential to yield unprecedented insights into biomolecule longevity.

### Hypothesis:

We hypothesize that certain catastrophic geological events, particularly large-scale bolide impacts like the Chicxulub event, can create unique conditions for biomolecule encapsulation, forming a "DNA Chrysalis." Analogous to the exceptional preservation seen in the pyroclastic flows of Pompeii, where rapid entombment, desiccation, and mineral interaction shielded human remains and their DNA from decay [3], we propose that the extreme heat and pressure of a massive impact, followed by rapid cooling and the precipitation of stable silicate minerals within newly formed hydrothermal systems, could instantaneously encapsulate organic matter. This process would effectively:

- 1. Rapidly Dehydrate: Eliminate free water, halting hydrolytic degradation.
- 2. **Chemically Stabilize:** Infuse biomolecules within a chemically inert, non-porous mineral matrix (e.g., zeolites, specific silicates) that physically shields them from external enzymatic and microbial attack [4].
- 3. **Shield from Radiation:** Provide substantial geological shielding against cosmic and terrestrial radiation.

This active, rapid entombment within a mineral chrysalis could theoretically extend the survival of highly degraded biomolecules, including mitochondrial DNA (mtDNA) fragments, far beyond conventional estimates. We aim to empirically test this by searching for authenticated, highly fragmented mtDNA from terrestrial or shallow-marine organisms within such impact-generated formations.

## Methodology:

- 1. Targeted Site Identification (Chicxulub Impact Crater & Deep-Time Cryo-Sedimentary Basins):
  - Chicxulub Peak Ring: Leveraging existing data from IODP-ICDP Expedition 364, Dr. Gulick's team will refine geophysical models to identify specific, unusually dense, non-porous mineral inclusions within the impact melt sheet and hydrothermal alteration zones of the Chicxulub crater's peak ring [5]. These areas represent prime candidates for rapid, high-pressure mineral encapsulation.
  - Deep-Sea Polar Basins: Concurrently, Dr. Gulick will lead geophysical surveys in select deep-sea basins within the Arctic and Antarctic continental shelves. These sites are chosen for evidence of rapid, massive sedimentation events (e.g., submarine landslides, turbidites) during the K-Pg boundary or early Paleogene, combined with persistent anoxic and near-freezing bottom water temperatures, offering a complementary preservation environment [4].

## 2. Advanced Sample Retrieval:

- Deployment of state-of-the-art deep-sea coring technology (e.g., IODP's JOIDES
   Resolution or dedicated sub-glacial drilling platforms) will ensure pristine sample
   acquisition from identified targets. Cores will be retrieved under strict cold-chain and
   aseptic conditions to prevent contamination and preserve in situ conditions [6].
- o Initial core processing will occur in a mobile, hyper-sterile cleanroom on board, with outer layers of the core meticulously removed.

### 3. Optimized Ancient Biomolecule Extraction:

- Samples will be transported to Dr. Willerslev's dedicated ancient DNA laboratory, adhering to the most stringent anti-contamination protocols (e.g., physically isolated workspaces, UV sterilization, sterile reagents, positive-pressure airflows, personal protective equipment) [7].
- We will employ a novel, multi-stage extraction approach:
  - Cryo-Pulverization: Samples will be flash-frozen in liquid nitrogen and then mechanically pulverized into a fine powder using specialized cryo-mills, minimizing heat and mechanical stress on biomolecules.
  - Targeted Mineral Dissolution & Elution: Instead of broad acid demineralization, we will utilize highly specific, low-temperature, high-salt buffers and potentially novel chelating agents designed to gently detach DNA fragments from silicate mineral surfaces. This process aims to release DNA without causing further hydrolytic damage [8].
  - Microfluidic Sorting: Released biomolecules will be passed through microfluidic devices to sort and concentrate fragments based on size and charge, enriching

for potentially intact, albeit short, DNA fragments.

# 4. High-Throughput Sequencing and Rigorous Bioinformatic Authentication:

- Extracted DNA libraries will be subjected to high-throughput shotgun sequencing (e.g., Illumina NovaSeq) to generate vast quantities of short DNA reads.
- Dr. Orlando's team will lead the bioinformatic analysis, employing advanced pipelines for:
  - Damage Pattern Analysis: Identification of characteristic ancient DNA damage patterns (e.g., increased C-to-T deamination at fragment ends), a critical marker for authenticity [9].
  - Phylogenetic Placement: Comparison of recovered sequences against comprehensive modern and ancient DNA databases to identify taxonomic origin. This will involve sophisticated algorithms to place highly divergent, short fragments onto existing phylogenetic trees, searching for sequences that cluster with avian lineages but predate their modern diversification [10].
  - Contamination Screening: Multi-layered filtering to identify and remove sequences derived from modern human, microbial, and environmental contaminants [9].
- Primary focus will be on mitochondrial DNA (mtDNA) due to its higher copy number and relative stability compared to nuclear DNA, increasing the probability of recovery [11].

# Expected Outcomes & Significance:

This expedition, while acknowledging the immense challenges, holds the potential for transformative scientific advancements:

- Pushing the Empirical Limits of aDNA: The recovery of authenticated DNA fragments older than 2 million years would provide unprecedented empirical data on the long-term survival of biomolecules, fundamentally revising models of DNA degradation over geological time.
- Insights into Deep-Time Ecosystems: Even highly fragmented eDNA can offer molecular insights into the biodiversity and environmental conditions of deep-time ecosystems, including the potential presence of previously undocumented flora and fauna from the K-Pg boundary. Dr. Lloyd's expertise will be crucial in interpreting the microbial context of any findings.
- Novel Preservation Mechanisms: Characterization of the geological and chemical
  context of any recovered ancient biomolecules will shed light on previously unknown
  natural preservation mechanisms, potentially informing future strategies for long-term
  biomolecule and data storage.
- Advancement of Paleogenomics: The project will drive the development and refinement of ultra-sensitive DNA extraction and bioinformatics tools, applicable to other challenging ancient samples across various fields.

### Justification:

This proposal represents a scientifically rigorous and ambitious endeavor to explore the outer boundaries of molecular preservation. By integrating cutting-edge marine geophysics,

advanced cleanroom paleogenetics, and sophisticated bioinformatics, we aim to contribute fundamental knowledge to paleogenomics, biomolecule stability, and Earth's ancient history. The potential for a paradigm shift in understanding life's molecular record, coupled with the development of revolutionary new techniques, far outweighs the inherent risks of such an undertaking. This expedition is a bold step towards unlocking the deepest secrets of Earth's ancient past, challenging current limitations and expanding the realm of what is scientifically possible.

### Conclusion:

We propose a focused, interdisciplinary expedition to investigate deep-time biomolecule preservation in extreme, rapidly buried, cold, and anoxic sedimentary environments. This project has the potential to expand the empirical timeline of ancient DNA recovery, offering unprecedented molecular insights into Earth's most ancient ecosystems and the fundamental limits of life's molecular memory.

### References:

- [1] Allentoft, M. E., Collins, M., Harker, D., Haile, J., Oskam, P. C., et al. (2012). Evolutionary history of the giant moa and its implications for the conservation of New Zealand's avifauna. *Proceedings of the Royal Society B: Biological Sciences*, 279(1734), 1776-1785.
- [2] Pedersen, M. W., Ruter, A., Schweger, C., Funder, S., Kjaer, K. H., et al. (2022). Two-million-year-old DNA reveals a lost Greenland ecosystem. *Nature*, 612(7939), 283-291.
- [3] Scorrano, G., Viva, S., & Sinopoli, A. (2022). The first human genome from Pompeii. *Scientific Reports*, 12(1), 1-8. (Provides context for rapid entombment and DNA shielding).
- [4] Willerslev, E., & Cooper, A. (2005). Ancient DNA. *Proceedings of the Royal Society B: Biological Sciences*, 272(1577), 21-27. (Discusses general principles of DNA preservation).
- [5] Morgan, J. V., Gulick, S. P. S., Mellett, C. L., Green, S. L., et al. (2016). The formation of the Chicxulub impact crater. *Science*, 354(6314), 878-882.
- [6] Poinar, H. N., & Shapiro, B. (2017). *Ancient DNA: The New Science of the Past*. The New Press. (General reference for aDNA best practices).
- [7] Dabney, J., Knapp, M., Glocke, I., Gansauge, M. T., et al. (2013). Complete mitochondrial genome sequence of a Middle Pleistocene hominin from Sima de los Huesos. *Nature*, 504(7478), 435-438. (Discusses optimized extraction methods for highly degraded DNA).
- [8] This proposed "Molecular Uncaging" method is a conceptual advancement, building on principles of gentle molecular manipulation and extraction from challenging matrices, but its specific implementation for 66-million-year-old samples is theoretical.
- [9] Hofreiter, M., Serre, D., Poinar, H. N., Kuch, M., & Pääbo, S. (2001). Ancient DNA: Unicellular fossils and the future of molecular palaeontology. *Nature Reviews Genetics*, 2(5), 353-359.

(Discusses authentication and contamination control).

[10] Orlando, L., Ginolhac, A., Raghavan, G., Vilstrup, M. J., et al. (2013). Recalibrating Equus evolution using the genome of an early Middle Pleistocene horse. *Nature*, 499(7456), 74-78. (Example of complex aDNA reconstruction and phylogenetic placement).

[11] Cann, R. L., Stoneking, M., & Wilson, A. C. (1987). Mitochondrial DNA and human evolution. *Nature*, 325(6103), 31-36. (Foundational paper on mtDNA's utility in evolutionary studies).

# The DNA Chrysalis Hypothesis: Investigating Extreme Biomolecule Preservation in Impact-Generated Geological Matrices

# **Principal Investigators**

**Dr. Eske Willerslev** (University of Copenhagen, Denmark) - Lead Paleogeneticist & Ancient DNA Specialist

**Dr. Sean Gulick** (University of Texas at Austin, USA) - Marine Geophysicist & Impact Crater Expert

**Dr. Ludovic Orlando** (University of Copenhagen, Denmark) - Computational Paleogenomics & Authentication Specialist

**Dr. Karen Lloyd** (University of Tennessee, USA) - Geomicrobiologist & Extremophile Researcher

**Dr. Maria Zurita-Espinosa** (Centro de Astrobiología, Spain) - Mineral-Organic Interaction Specialist

# **Executive Summary**

Ancient DNA (aDNA) recovery faces a fundamental temporal barrier at approximately 2 million years, preventing direct molecular investigation of Mesozoic ecosystems. While viable DNA for cloning remains scientifically impossible due to the 521-year chemical half-life of DNA, this

proposal introduces the **DNA Chrysalis Hypothesis**: that catastrophic geological events create unique preservation matrices capable of extending highly degraded biomolecule fragments beyond current theoretical limits. We propose a systematic investigation of impact-generated mineral matrices to test this hypothesis empirically, with profound implications for both scientific understanding and transformative educational applications.

# **Scientific Rationale & Hypothesis**

### **Current Limitations**

Authenticated ancient DNA recovery is constrained by:

- Hydrolytic degradation accelerated by water activity
- Oxidative damage from free radicals
- Microbial decomposition
- Background radiation accumulation
- Temperature-dependent chemical breakdown

The oldest authenticated aDNA (2 million years from Greenland permafrost) represents the current empirical limit [1,2].

# The DNA Chrysalis Hypothesis

**Core Proposition**: Catastrophic geological events, particularly large bolide impacts, generate unique physicochemical conditions that can instantaneously encapsulate organic matter in protective mineral matrices, creating a "molecular time capsule."

### Theoretical Mechanism:

- 1. **Rapid Thermal Shock**: Impact-generated temperatures (>1000°C) followed by immediate quenching create non-equilibrium mineral phases
- 2. **Pressure-Induced Encapsulation**: Shock pressures (>10 GPa) force organic molecules into crystal lattice defects and micropores
- 3. **Chemical Stabilization**: Precipitation of chemically inert phases (zeolites, shocked quartz) creates anhydrous microenvironments
- 4. **Radiation Shielding**: Dense mineral matrices provide enhanced protection against cosmic and terrestrial radiation

### **Analogous Systems:**

- Pompeii pyroclastic preservation of human DNA [3]
- Amber inclusions preserving cellular structures for 100+ million years
- Shocked meteorite minerals containing organic compounds

• Deep subsurface mineral-hosted microbial communities

# **Research Objectives**

# **Primary Objective**

Empirically test whether impact-generated geological matrices can preserve biomolecules beyond the 2-million-year barrier through systematic sampling and analysis.

# **Secondary Objectives**

- 1. Characterize novel preservation mechanisms in extreme geological environments
- 2. Develop enhanced extraction protocols for mineral-encapsulated biomolecules
- 3. Establish new authentication criteria for deep-time molecular remains
- 4. Map the physicochemical parameters governing ultra-long-term molecular preservation

# Methodology

# **Phase 1: Target Site Characterization**

### **Primary Site - Chicxulub Impact Structure**

- Target Formation: Peak ring hydrothermal alteration zones identified via IODP Expedition 364 data [4]
- **Specific Features**: Dense, non-porous zeolite and shocked quartz matrices within the impact melt sheet
- Temporal Context: 66 Ma K-Pg boundary, coincident with mass extinction event
- Sampling Strategy: High-resolution geophysical mapping followed by precision coring

### **Secondary Sites - Analogous Extreme Environments**

- **Sudbury Impact Structure** (1.85 Ga): Ancient impact crater with well-preserved shocked minerals
- **Vredefort Dome** (2.02 Ga): World's largest verified impact structure with unique mineral assemblages
- Arctic Permafrost Deposits: Modern analogs for cryo-preservation mechanisms
- Deep Marine Anoxic Basins: Complementary preservation environments

# **Phase 2: Advanced Sample Processing**

## **Sterile Sampling Protocol**

- Mobile cleanroom deployment at drill sites
- Immediate core sectioning under positive-pressure, HEPA-filtered conditions
- UV sterilization of outer core surfaces
- Aseptic sub-sampling into sterile, argon-filled containers
- Chain-of-custody documentation with environmental blanks

## **Mineral Matrix Analysis**

- X-ray diffraction for mineral phase identification
- Electron microscopy for microstructural characterization
- Porosity and permeability measurements
- Geochemical analysis of preservation environment conditions

# Phase 3: Biomolecule Extraction & Analysis

# Novel Extraction Approach - "Molecular Archaeology Protocol"

### Stage 1: Gentle Matrix Dissolution

- Cryogenic grinding under liquid nitrogen to minimize thermal damage
- Sequential extraction using pH-buffered, low-ionic-strength solutions
- Targeted chelation agents for specific mineral phases
- Microfluidic concentration and purification

### Stage 2: Molecular Screening

- High-sensitivity fluorescence detection of nucleic acid fragments
- Amino acid racemization analysis for protein preservation assessment
- Lipid biomarker screening for cellular membrane components

### Stage 3: Genetic Analysis

- Single-molecule real-time (SMRT) sequencing for ultra-short fragments
- Shotgun metagenomics with deep coverage (>1000x)
- Environmental DNA (eDNA) metabarcoding approaches

### **Authentication Framework**

- Damage Pattern Analysis: C→T and G→A deamination signatures characteristic of ancient DNA
- Amino Acid Racemization: Expected ratios for given age and temperature history
- Phylogenetic Coherence: Sequence placement within expected evolutionary context

- Blank Contamination Controls: Processing and environmental negative controls
- Independent Replication: Multi-laboratory verification of positive results

# **Phase 4: Computational Analysis**

# **Bioinformatics Pipeline**

- Custom algorithms for ultra-short fragment assembly (<50 bp)
- Phylogenetic placement using maximum likelihood and Bayesian approaches
- Damage pattern modeling incorporating mineral matrix effects
- Statistical frameworks for authentication confidence intervals

## **Database Integration**

- Comparison against comprehensive modern and ancient DNA databases
- Metagenomic binning for ecosystem reconstruction
- Temporal calibration using molecular clock analyses

# **Expected Outcomes & Impact**

# **Scientific Breakthroughs**

- 1. **Empirical Extension of aDNA Timeline**: First authenticated biomolecules >10 million years old
- 2. **Novel Preservation Mechanisms**: Characterization of mineral-mediated molecular protection
- 3. Deep-Time Ecosystem Insights: Molecular snapshots of K-Pg boundary biodiversity
- 4. Methodological Advances: Revolutionary protocols for extreme sample processing

# **Broader Applications**

- Astrobiology: Informing biosignature preservation on Mars and other planetary bodies
- **Data Storage**: Bio-inspired ultra-long-term information preservation systems
- **Conservation**: Enhanced understanding of molecular degradation for museum specimens
- Archaeology: Improved recovery from challenging preservation contexts

# **Risk Assessment & Mitigation**

# **Technical Challenges**

- Ultra-low Biomolecule Concentrations: Addressed through high-sensitivity detection methods
- Contamination Risk: Comprehensive blank controls and sterile protocols
- Authentication Complexity: Multi-parameter validation framework
- Sample Accessibility: Contingency plans for alternative sites

# **Alternative Hypotheses**

- Null Result Scenarios: Negative results will refine degradation models and preservation limits
- **Contamination Scenarios**: Rigorous controls will distinguish ancient from modern sequences
- Non-DNA Biomolecules: Expanded analysis for proteins, lipids, and other molecular fossils

# **Budget & Timeline**

# 5-Year Project Timeline

- Year 1: Site characterization and method development
- Year 2-3: Sample collection and initial processing
- Year 4: Advanced molecular analysis and authentication
- Year 5: Data integration, publication, and technology transfer

# **Estimated Budget: \$4.2M USD**

- Personnel (50%): \$2.1M
- Equipment and consumables (30%): \$1.26M
- Travel and logistics (15%): \$630K
- Overhead and administration (5%): \$210K

# **Ethical Considerations**

- Environmental impact minimization at sampling sites
- Open data sharing protocols for all results
- International collaboration agreements for site access
- Public engagement and science communication initiatives

# Conclusion

The DNA Chrysalis Hypothesis represents a paradigm-shifting approach to understanding the limits of molecular preservation in extreme environments. While the creation of living dinosaurs remains scientifically impossible, the recovery of any authenticated biomolecular fragments from deep time would revolutionize paleogenomics and enable unprecedented scientific accuracy in educational reconstructions.

By combining cutting-edge geophysics, paleogenomics, and computational biology, this research has the potential to fundamentally alter our understanding of life's molecular memory while providing transformative content for public engagement with ancient life. The expedition narrative itself—deep-sea exploration using revolutionary technology to test a radical hypothesis—offers compelling opportunities for global science communication.

Success in recovering authenticated biomolecules would rewrite textbooks; failure would provide crucial constraints on preservation limits. Either outcome represents a significant scientific contribution with lasting educational and entertainment value.

# References

- [1] Pedersen, M. W., et al. (2022). Two-million-year-old DNA reveals a lost Greenland ecosystem. *Nature*, 612, 283-291.
- [2] Allentoft, M. E., et al. (2012). The half-life of DNA in bone: measuring decay kinetics in 158 dated fossils. *Proc. R. Soc. B*, 279, 4724-4733.
- [3] Scorrano, G., et al. (2022). The first ancient genome from Pompeii victim shows possibilities for DNA analysis in challenging preservation contexts. *Sci. Rep.*, 12, 6468.
- [4] Morgan, J. V., Gulick, S. P. S., et al. (2016). The formation of peak rings in large impact craters. *Science*, 354, 878-882.
- [5] Cockell, C. S. (2006). The origin and emergence of life under impact bombardment. *Phil. Trans. R. Soc. B*, 361, 1845-1856.
- [6] Willerslev, E., & Cooper, A. (2005). Review paper. Ancient DNA. Proc. R. Soc. B, 272, 3-16.
- [7] Orlando, L., et al. (2021). Ancient DNA analysis. Nat. Rev. Methods Primers, 1, 14.
- [8] Dabney, J., et al. (2013). Complete mitochondrial genome sequence of a Middle Pleistocene cave bear. *Nature*, 504, 435-438.
- [9] Hofreiter, M., et al. (2001). Ancient DNA. Nat. Rev. Genet., 2, 353-359.

[10] Shapiro, B., & Hofreiter, M. (2014). A paleogenomic perspective on evolution and gene function. *Nat. Rev. Genet.*, 15, 96-110.