

Article

Specific Gene Duplication and Loss of Cytochrome P450 in Families 1–3 in Carnivora (Mammalia, Laurasiatheria)

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Simple Summary: In this study we investigated the specific duplication and loss events of cytochrome P450 (CYP) genes in families 1–3 in Carnivora. These genes have been recognized as essential detoxification enzymes, and, using genomic data, we demonstrated a synteny analysis of the CYP coding cluster and a phylogenetic analysis of these genes. We discovered the CYP2Cs and CYP3As expansion in omnivorous species such as the badger, the brown bear, the black bear, and the dog. Furthermore, phylogenetic analysis revealed the evolution of CYP2Cs and 3As in Carnivora. These findings are essential for the appropriate estimation of pharmacokinetics or toxicokinetic in wild carnivorans.

Abstract: Cytochrome P450s are among the most important xenobiotic metabolism enzymes that catalyze the metabolism of a wide range of chemicals. Through duplication and loss events, CYPs have created their original feature of detoxification in each mammal. We performed a comprehensive genomic analysis to reveal the evolutionary features of the main xenobiotic metabolizing family: the CYP1–3 families in Carnivora. We found specific gene expansion of CYP2Cs and CYP3As in omnivorous animals, such as the brown bear, the black bear, the dog, and the badger, revealing their daily phytochemical intake as providing the causes of their evolutionary adaptation. Further phylogenetic analysis of CYP2Cs revealed Carnivora CYP2Cs were divided into CYP2C21, 2C41, and 2C23 orthologs. Additionally, CYP3As phylogeny also revealed the 3As' evolution was completely different to that of the Caniformia and Feliformia taxa. These studies provide us with fundamental genetic and evolutionary information on CYPs in Carnivora, which is essential for the appropriate interpretation and extrapolation of pharmacokinetics or toxicokinetic data from experimental mammals to wild Carnivora.

Keywords: cytochrome P450; genome database; in silico analysis; phase I metabolism; wildlife; xenobiotic metabolism



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1. Introduction

Cytochrome P450s (CYP) catalyze major xenobiotics metabolism for a wide range of chemicals, such as drugs, phytochemicals, and environmental pollutants, and are considered the most important enzymes for detoxification [1]. The CYP genes form a superfamily and are divided into gene families based on >40% sequence similarity, but into subfamilies

when sequence similarities are >55%. Among various CYP families in mammals, the CYP1-3 families are considered one of the main families that catalyze xenobiotic metabolism [2–4], although other CYP families are generally considered important for the biosynthesis of numerous endogenous chemicals such as steroids, bile acids, cholesterol, eicosanoids, fatty acids, etc. [5,6].

These CYP genes are considered the fastest-evolving gene systems [7–9] and through gene duplication and loss events, xenobiotic-metabolizing CYPs, CYP1-3 genes, have evolved, duplicated or been lost, and diversified. Such an evolutionary history is essential for characterizing isoform-specific substrate-specificity and further characterizing species-specific metabolism in animals [10]. Reports suggest that CYP genetic duplication or loss affects the xenobiotic metabolism capacity in several animals and that these evolutionary consequences might be due to the need of some insects, birds, and herbivorous mammals to manage constant exposure to phytochemicals [11–14]. This “plant-arms race” concept may explain the necessity of CYP duplication events in herbivorous species: it hones detoxification systems in response to phytochemicals or toxins originating in plants. However, a detailed analysis of CYP duplication events in species of the Carnivora order is not available. Previous reports of CYP2D subfamily in mammals showed no clear relation with CYP2D expansion or contractions and food habitat in mammals, but suggested that some isoforms did not necessarily follow this “plant-arms race” hypothesis [15]. However, this study included only one isoform subfamily with a wide range of mammals, and evaluation for CYP expansion or contraction in various isoform subfamilies is still needed.

Carnivora is a mammalian order consisting of 297 species, including cat-like (Feli-formia) and dog-like (Caniformia) animals [16,17]. As its name suggests, several animals within this order are largely carnivorous, such as Felids, yet foraging habits of the different species are diverse in this group. Previous research has identified genetic loss of certain xenobiotic metabolism enzymes. UDP-glucuronosyltransferase (UGT) in felids and pinnipeds indicates a strong relationship between carnivory and genetic loss of xenobiotic metabolism enzymes [18–20]. However, there has been little research investigating CYP genetic loss and duplication events in this taxon, leading us to comprehensively investigate their evolutionary history in Carnivora species and the relationships between foraging habits and their genetic duplication or loss events.

Recent innovations in next generation sequencing (NGS) systems have enabled us to manage large amounts of genetic data from a wide range of wild mammals, and we can also utilize these high-quality genetic assembly data freely through an online database [21]. These whole genomics data enabled us to comprehensively analyze genetic duplication and loss events in Carnivora, and further to compare phylogenetic relationships of each gene to provide an understanding of the CYP evolutionary features in this taxon.

Firstly, we investigated and compared the synteny of CYP1-3 gene isoforms' loci in Carnivora species. We then conducted a phylogenetic analysis to further detected specific gene duplication or loss events in each studied species to reveal the evolutionary features of CYP genes in Carnivora.

2. Materials and Methods

2.1. Data Retrieval for CYP Phylogenetic Analysis

Since this study does not use samples from living organisms, no ethics committee approval review of animal experiments is required. Phylogenetic analyses were performed on the CYP genes of human (*Homo sapiens*), dog (*Canis lupus familiaris*), red fox (*Vulpes vulpes*), domestic ferret (*Mustela putorius furo*), ermine (*Mustela erminea*), mink (*Neovison vison*), badger (*Meles meles*), North American river otter (*Lontra canadensis*), Eurasian river otter (*Lutra lutra*), sea otter (*Enhydra lutris kenyoni*), polar bear (*Ursus maritimus*), giant panda (*Ailuropoda melanoleuca*), black bear (*Ursus americanus*), brown bear (*Ursus arctos*), meerkat (*Suricata suricatta*), striped hyena (*Hyaena hyaena*), domestic cat (*Felis catus*), Amur tiger (*Panthera tigris*), cheetah (*Acinonyx jubatus*), puma (*Puma concolor*), Canada lynx (*Lynx canadensis*), leopard (*Panthera pardus*), lion (*Panthera leo*), leopard cat (*Prionailurus bengalensis*).

sis), fishing cat (*Prionailurus viverrinus*), Weddell seal (*Leptonychotes weddellii*), harbor seal (*Phoca vitulina*), gray seal (*Halichoerus grypus*), Hawaiian monk seal (*Neomonachus schauinslandi*), northern elephant seal (*Mirounga angustirostris*), southern elephant seal (*Mirounga leonine*), northern fur seal (*Callorhinus ursinus*), Steller's sea lion (*Eumetopias jubatus*), California sea lion (*Zalophus californianus*), and Pacific walrus (*Odobenus rosmarus divergens*). Sequences were retrieved using National Center for Biotechnology Information (NCBI) BLAST searches using the following query sequences: human and dog CYP1A1, 1A2, and 1B1 for CYP1 searching, human CYP2A6, 2B6, 2D6, 2E1, 2F1, 2J2, 2S1, 2U1, 2W1, 2S1, rat 2T1 and 2G1, and dog 2C21, 2C41, and 2C23 were used for CYP2 investigation, and human CYP3A4, dog CYP3A12 and 3A26, and cat CYP3A131 and 132 were used for CYP3. These isoform queries were sufficiently comprehensive for detecting target genes and hitting other additional subfamily isoforms in Carnivora (e.g., the CYP2C6 BLAST search also detected CYP2Es and other subfamily genes). BLAST searches were conducted on the database Nucleotide collection (nr/nt) for each species using BLASTN (optimized for similar sequences). The gene sequences used are listed in the Supplementary Tables S1 and S2, and the protein coding region of each isozyme was analyzed. The deduced amino acid sequences were aligned using MUSCLE (Multiple Sequence Comparison by Log-Expectation) and were used for model selection (models showing minimal sets of BIC were chosen) and construction of maximum likelihood trees (bootstrapping = 100) using MEGA X (Molecular Evolutionary Genetics Analysis) [22]. The aligned sequence lengths analyzed were 1965 bp in CYP3As and 1581 bp in CYP2Cs. The JTT+G model was used. All positions containing gaps and missing data were eliminated, and the total lengths of protein-coding sequence alignments were used for phylogenetic analysis. Foraging habits of each analyzed species are also listed in the Supplementary Table S3. The phylogenetic tree was created with TimeTree 5 [23]. We also provided protein sequence files in Supplementary Information: CYP3A_protein.fas, and CYP2C_protein.fas.

2.2. Synteny Analysis of CYP Genes

Sequence data from genome projects are freely available. NCBI's genome data viewer (<https://www.ncbi.nlm.nih.gov/genome/gdv/> (accessed on 1 October 2022)) or JBrowse [24] were used to visualize the chromosomal synteny maps for each species. The following latest genome assemblies were used and listed in Supplementary Table S4. UCSC (University of California, Santa Cruz) BLAT (a BLAST-like alignment tool) (<http://genome.ucsc.edu/index.html> (accessed on 1 October 2022)) was used for additional confirmation of missing genes. The masked palm civet CYP genes were also retrieved and used to fill the gap for Feliformia species from recently assembled and annotated chromosome-level genomic data [25]

3. Results

3.1. CYP Number Counts and Isoforms in CYP1As and 2ABGFSs Clusters

Gene number counts for CYP 1–3 genes are shown in Figure 1. We found CYP gene coding loci where multiple CYPs were coded as a “cluster” of the CYP genes. CYP1As, CYP2ABGFSs, CYP2Cs, and CYP3As in each Carnivoran consist of a gene cluster. Several CYP gene clusters were conserved among Carnivorans. The CYP2ABFGSTs cluster coded CYP2As, 2Bs, 2Fs, 2Gs, and 2Ss annotated genes and was between *AXL Receptor Tyrosine Kinase* (AXL) and *Egl-9 family hypoxia inducible factor 2* (EGNL2), and the CYP1As cluster was between *C-terminal Src kinase* (CSK) and *Enhancer of mRNA decapping 3* (EDC3) when CYP1A1 and 1A2 orthologs in analyzed Carnivora were coded in this cluster. We further found specific gene duplication in brown bear CYP2As in the CYP2ABFGSTs cluster (Figure 1).

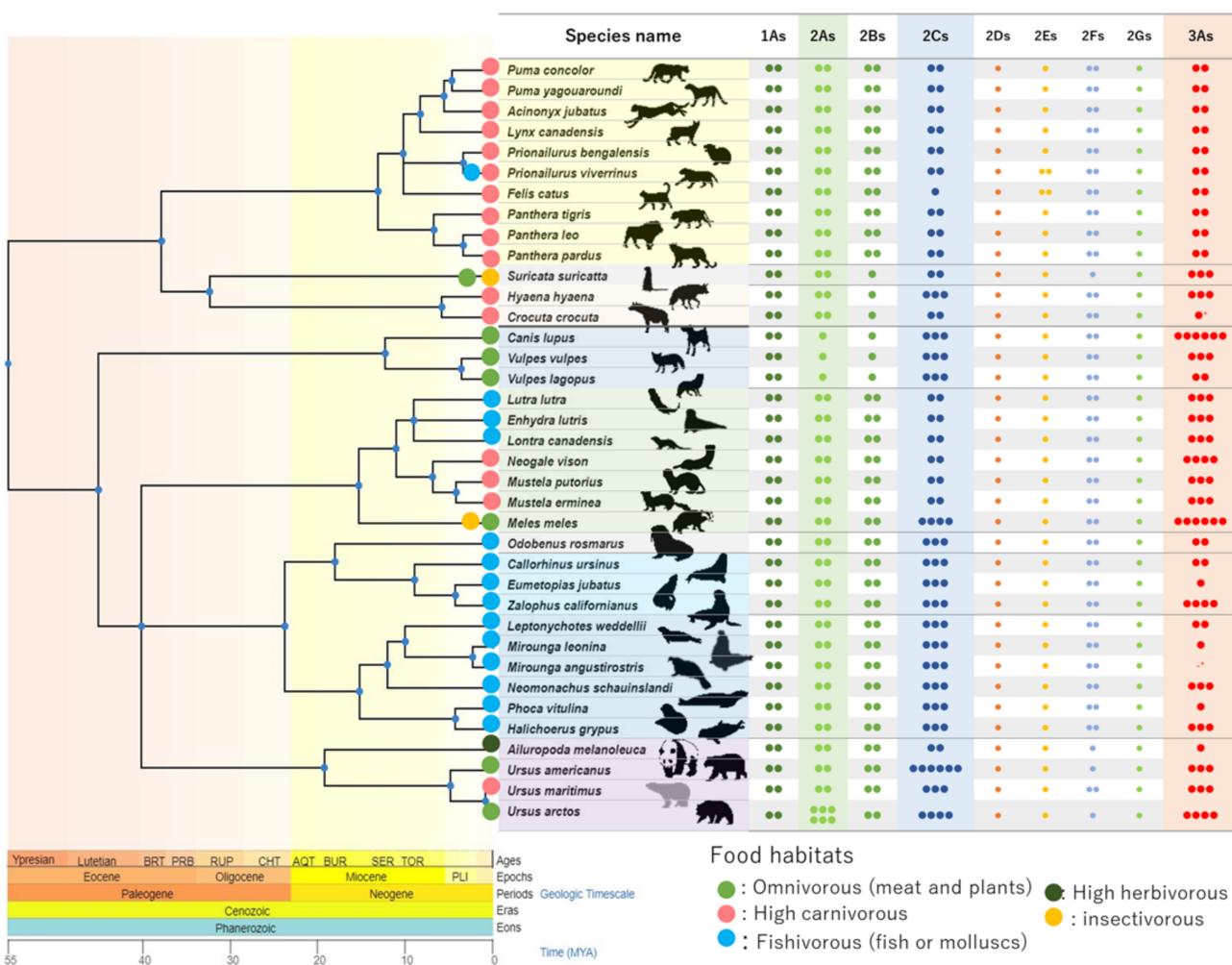


Figure 1. Isoform numbers of CYP1-3 families among Carnivora. Gene numbers for CYP1As, 2A, 2B, 2C, 2D, 2E, 2F, 2G, and 3As are shown by the number of small filled circles. Large filled circles next to the scientific name of each species are colored according to their known diet (Supplementary Table S1). Isoforms coding “low quality” or partial genes were not included in this case. The phylogenetic tree was created with TimeTree 5 [23].

3.2. CYP Isoforms in CYP2Cs and CYP 2CEs Clusters

Synteny of CYP2C coding loci are shown in Figure 2. The CYP2Cs coding loci also consisted of gene clusters, labeled the CYP2Cs clusters, and were highly conserved between *Helicase*, *lymphoid specific* (HELLS) and the *PDZ and LIM domain 1* (PDLIM1) among carnivorans, humans, and rodents. Multiple CYP2Cs were coded in the Carnivoran cluster. In almost all Mustelidae (except badger), Felidae (except the domestic cat), Pinnipedian, Canidae, and meerkat, the analyzed genome had two CYP2Cs annotated genes in this cluster, whereas CYP2Cs in badger had three isoforms (with one isoform in another un-scaffolded contig), and the domestic cat had one intact isoform and one possible dysfunctional gene. The striped hyena genome had three isoforms in this cluster, whereas the meerkat had two. Some species genomes had CYP2Cs in several un-scaffolded contigs, and we did not find completely connected cluster loci for the Pacific walrus genome although we found two possible isoforms in different contigs.

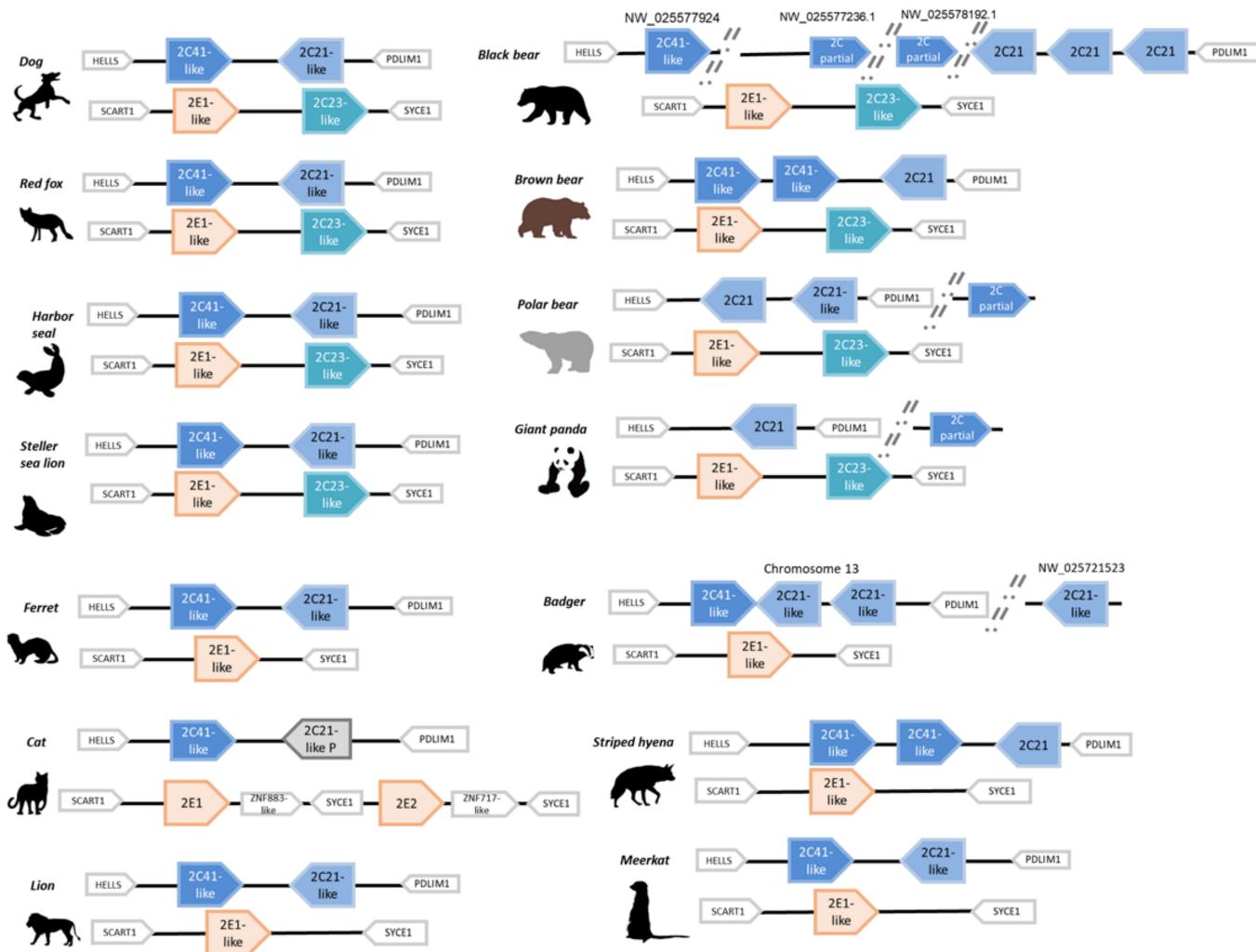


Figure 2. Synteny of CYP2Cs and CYP2CEs clusters in Carnivora. Synteny of the CYP2C cluster and the CYP2CE cluster among Carnivora are shown. Representative species for each family were selected. Phylogenetic-analysis-supported classifications were applied for each isoform and colored based on each clade. CYP2C41s are shown in blue, CYP2C21s in pale blue, CYP2C23s in brilliant blue, and CYP2Es in pale orange. Letters on each locus show coding contigs if separately coded. Pseudogenes are shown as black or gray blocks.

In contrast, within Ursidae we found huge species differences. The giant panda genome contained one possible CYP2C in this cluster and additional un-scaffolded isoforms in the contig NW_023254381.1. In contrast, the polar bear genome contained two possible CYP2Cs in this cluster with a partial additional isoform (CYP2C41-like) in the un-scaffolded contig NW_024425153.1. However, in the brown bear genome, we found three annotated CYP2Cs in this cluster. Moreover, the black bear genome had five possible isoforms and three partial isoforms, even though this cluster seemed to be on two separated contigs, and the partial isoforms on two other contigs (Figure 2).

We further found other specific CYP2Cs loci coding CYP2Cs and 2Es in Carnivora between *Synaptonemal complex central element protein 1* (SYCE1) and *Scavenger receptor family member expressed on T Cells 1* (SCART1): which were labeled the CYP2CEs cluster. Only Canids, ursids, and Pinnipeds had CYP2Cs in this cluster whereas other Carnivoran genomes (Feliformia (Felidae, striped hyena, and meerkat) and Mustelidae) had only CYP2Es in this cluster.

Other CYP2 subfamily genes such as CYP2Ds, 2Js, 2Rs, 2Us, and 2Ws did not show any duplication in Carnivora coded as isolated genes, although coding loci were highly

conserved among all analyzed Carnivorans and among other mammals (data not shown). Nevertheless CYP2J, 2R, 2S, 2T, 2U, and 2Ws are generally known as biosynthesis-type or unknown substrate isoforms, and we did not include these genes in the analyses.

3.3. Synteny Analysis of the CYP3As Cluster

We also analyzed the CYP3As gene cluster shown in Figure 3, and this cluster, which was also conserved among carnivorans, is between *Zinc finger and SCAN domain containing 25* (ZSCAN25) and *Olfactory receptor family 2 subfamily AE member 1* (OR2AE1) or *Tripartite motif containing 4* (TRIM4). All Felidae analyzed had only two possible isoforms of CYP3As, and no species-specific differences were observed. However, the Mustelidae genome had four or three possible isoforms in the CYP3As cluster and the ermine and Canadian river otter had one pseudogene-annotated gene in this cluster. In Ursidae, the brown and black bear genomes also contained four annotated CYP3As, whereas the polar bear and black bear genomes had three isoforms. The giant panda genome contained only one isoform annotated as “LOW-QUALITY PROTEIN” coding gene in this cluster, with six other very short partial un-scaffolded isoforms (CDS: CoDing Sequence length less than 515 bp) observed. Canids also have multiple CYP3As in this cluster, and the dog genome had four isoforms in this cluster (chromosome 6: NC_051810.1) with several intact and partial isoforms. Recently, two dog CYP3As were characterized and renamed, and the NCBI annotated name was different to the CYP nomenclature in dog CYP3As. The NCBI naming system was followed and genes were renamed in this paper (CYP3A4; Gene ID: 479740 as CYP3A98 and CYP3A12-like; LOC119875773 as CYP3A99) [26]. Red fox CYP3As were also on three un-scaffolded contigs (NW_020356965.1, NW_020356599.1, and NW_020356653.1) with two isoforms and three partial isoforms. However, in the Arctic fox genome, CYP3As were not coded as a cluster and these genes were coded on two different loci on the same chromosome (chromosome 3: NC_054826.1 with two intact and one short isoform). In several Pinnipedia genomes, CYP3As were also located on several un-scaffolded contigs, suggesting much higher quality assemblies are essential for clear analysis. Three Otariidae or Odobenidae genomes (from the Pacific walrus, northern fur seal, and Stellar sea lion) had two intact or partial CYP3As and one annotated pseudogene of CYP3As, whereas four isoforms were observed in the California sea lion genome (Supplementary Figure S1). Phocidae genomes also have scattered genes of CYP3As and we could not find a clear CYP3A cluster or isoform, with 1–2 intact CYP3As and several partial genes in each genome.

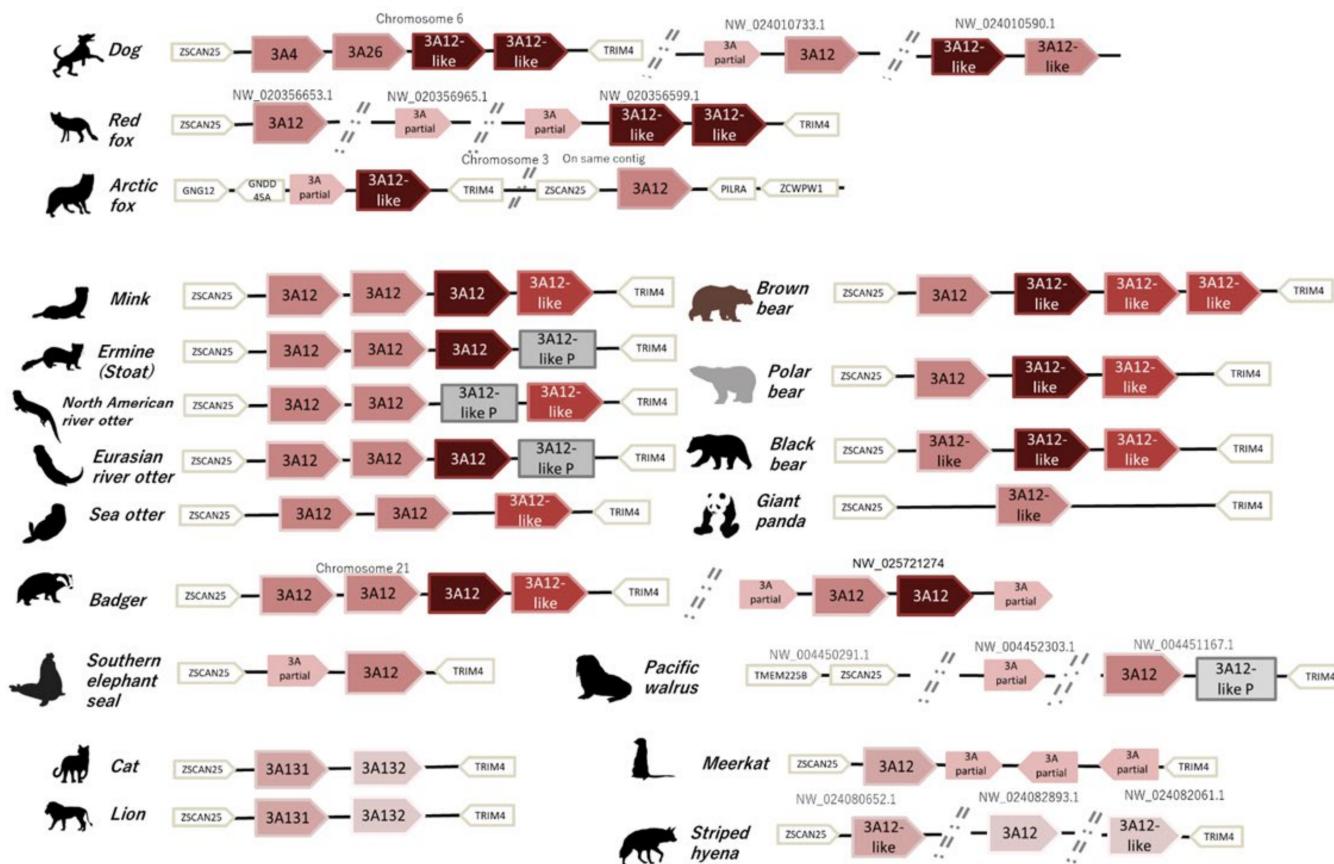


Figure 3. Synteny of the CYP3A cluster among Carnivora. Representative species of each family were selected. Phylogenetic-analysis-supported classifications were applied to each isoform and colored based on each clade. Caniformia CYP3As in clade 1, clade 2, and clade 3, and CYP3A131s and CYP3A132s in Felidae, are all differently colored. Letters on each locus show coding contigs if separately coded. Pseudogenes are shown as gray blocks.

3.4. Phylogeny of CYP2Cs in Carnivorans

We performed a phylogenetic analysis of CYP2Cs in carnivorans, and we revealed Carnivoran CYP2Cs were divided into three clades, namely CYP2C41s, CYP2C21s, and CYP2C23s (Figure 4). Based on the phylogeny, the CYP2C23s clade was located close to CYP2Es clades. Each clade contained orthologous Carnivoran genes to CYP2C41 and 2C21 in dogs and CYP2C23 in rats, respectively. Almost all CYP phylogenies in the CYP2C41s, 2C21s, and 2C23s clades followed their organisms' phylogeny order. We also found specific duplication of the CYP2C21s clade in Ursidae, suggesting these duplication events occurred after the divergence of Ursidae.

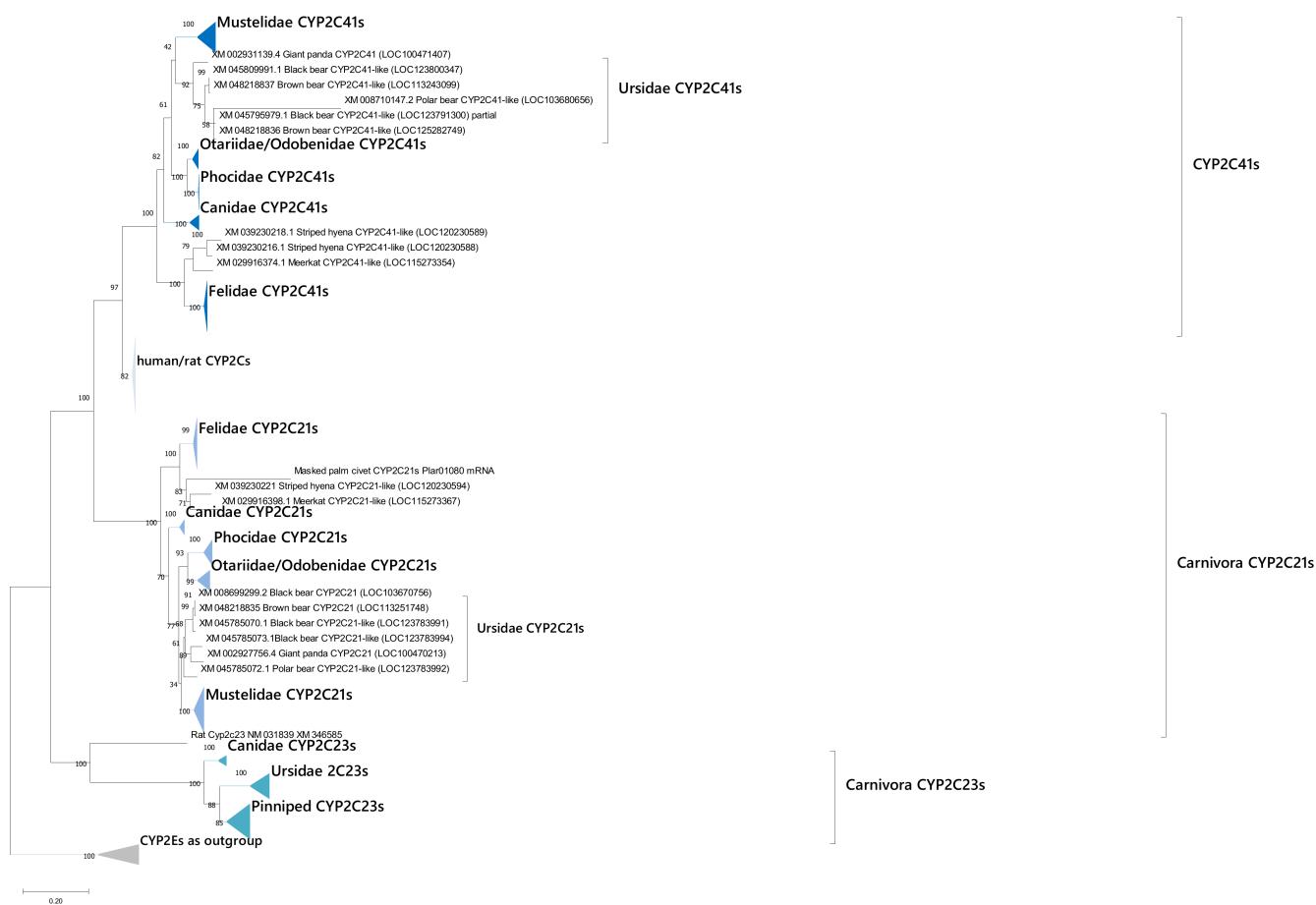


Figure 4. A phylogenetic tree of CYP2Cs sequences in humans and carnivorans. Gene sequences of protein-coding regions for each isozyme were analyzed. The numbers next to the branches indicate the number of occurrences per 100 bootstrap replicates. Genes and clades are tentatively labeled with Carnivoran CYPs examined in this article. Clades of CYP2C41s, CYP2C21s, and Carnivora 2C23s are shown as differently colored triangles. CYP2Es are shown as an outgroup.

3.5. Phylogeny of CYP3As in Carnivorans

We conducted similar phylogenetic analysis on CYP3As that revealed a Caniformia-clade and a Feliformia-unique clade in CYP3As (Figure 5), suggesting the CYP3As evolutionary history between Feliformia and Caniformia was completely different. Based on the phylogeny, the Caniformia-clade was further subdivided into three clades which were labeled Caniformia CYP3As clade 1 to clade 3. For clade 1, almost all Caniformia species possessed these genes, and Mustelidae had two each specifically duplicated clades (1-1 and 1-2), suggesting canid-specific and Mustelid-specific duplication of genes in this clade. Canidae also showed lineage specific duplication in this clade. Ursidae, Mustelidae, and some Pinniped genomes possessed CYP3As in clade 2, whereas Canidae and Feliformia did not have isoforms in this clade. In clade 3, however, Canidae, Mustelidae, and Ursidae have these genes and canids have a family-specific duplication of this clade similar to clade 1. Although some pinnipeds have multiple CYP3As, some with partial genes were removed for phylogenetic analysis to ensure clear results were produced, so there could be some pinniped CYP3As that might have been classified into clade 3.

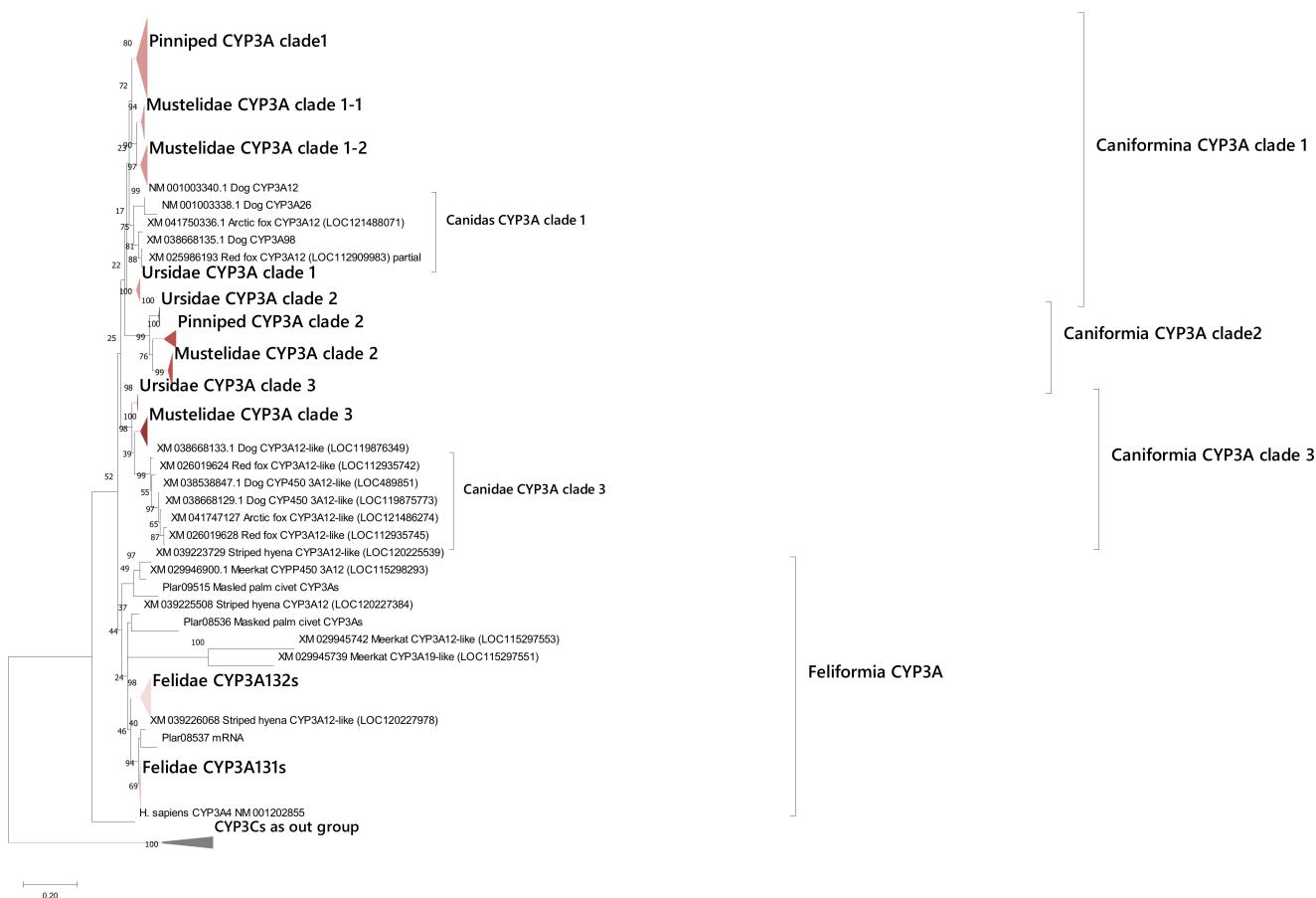


Figure 5. A phylogenetic tree of CYP3As. A phylogenetic tree of CYP3As sequences in carnivorans. Gene sequences of protein-coding regions for each isozyme were analyzed. The numbers next to the branches indicate the number of occurrences per 100 bootstrap replicates. Genes and clades are tentatively labeled with Carnivoran CYPs examined in this article. Caniformia clade 1, clade 2, clade 3, and Felidae CYP3A131s and 3A132s clades are shown with differently colored triangles. CYP3Cs are shown as an out group.

For the CYP3As Feliformia-clade, Felidae CYP3As were divided into two clades named CYP3A131s and CYP3A132s as per the domestic cat CYP3A131 and CYP3A132 [27]. In each clade, specific clades for Felidae were further established and only one isoform from each respective Felidae species was contained in these clades. However, other Feliformia CYP3As genes were not classified into these CYP3A131s and CYP3A132s clades, suggesting unique CYP3As loss or duplication events occurred in each species through the evolutionary history of Feliformia.

4. Discussion

In this analysis, we utilized the genome assembly data for multiple Carnivorans, and still these data have several limitations. Genome-assembled data only represent one individual element of genomic information and several variants could happen in all genomic assembly. Also, genome-assembly qualities should be considered. Although recent genomic assemble quality has improved, and several chromosomal-level assemblies were available even for wildlife Carnivorans, some assemblies have relatively low quality which made it difficult for us to analyze some of the CYP genes. However, we considered this genomic data analysis to have strong importance in the evaluation of the wildlife CYP genome.

4.1. CYPs Duplication and Loss in Mammals and Relationships with Foraging Habits

Several reports have revealed CYP gene duplication events in a variety of mammals, especially herbivorous mammals. Recently, a koala genome project revealed that a huge expansion of CYP2Cs was possibly an adaptation to a diet of eucalypts [11]. Further, among woodrat genomes, especially juniper-eating species, several reports have found higher gene copy numbers of CYP2As, 2Bs, and 3As compared to other rodents [12,28,29], and studies have also suggested that the woodrat's CYP2Bs gene expansion might contribute to their high metabolic capacity for terpenes from juniper plants. In this study, we found specific duplication of CYP2Cs and CYP2As in the brown bear, CYP2Cs in the black bear, CYP2Cs and 3As in the badger, and CYP3As in the dog. All these animals are omnivorous and their foraging habits include a wide variety of food types, indicating that these species have a "generalist" diet [30–34]. Therefore, these gene expansions might be the consequences of a need for detoxification of a wide variety of plant-secondary metabolites in their daily diets.

However, in other omnivorous or herbivorous animals, (e.g., the red fox, Arctic fox, and giant panda), we did not find any specific duplication of CYPs. In the red fox genome, CYP3As clusters were separated because of assembly quality, which suggested that these regions require re-assembling or target re-sequencing. Further, we found separated clusters of CYP3As for the Arctic fox, suggesting that these loci in Arctic fox genome are instable. Hence, further in-depth genomic analysis is required to clarify whether the CYP3A expansion is limited to dogs or is also applicable to other Canidae species.

Notably, we assumed giant panda genomes would show expansion of CYPs in response to various plant secondary metabolites in their exclusively bamboo diets. However, our result suggested that there was neither expansion of any CYPs, nor even a contraction trend of CYP3As in the giant panda. These trends were supported by our analysis of UGTs genomics for this species (unpublished data). These results strongly suggested that giant panda do not rely on an "enzymatic-strategy" to deal with their daily toxin intake. Their unbalanced "specialist" bamboo-exclusive diet might be the reason they have not evolved or expanded CYP or other xenobiotic metabolism enzymes, and gut microbiota is an alternative strategy they might use instead [35–37].

4.2. CYP2Cs in Carnivora

From the phylogenetic analysis of this study, we identified that Carnivora CYP2Cs are divided into three clades, which were possibly orthologs of CYP2C21s, CYP2C41 in the dog, and CYP2C23s in the rat [38–40]. These features strongly suggest that the substrate specificity of dog CYP2Cs is similar to that of CYP2C21 and CYP2C41 in other Carnivora. Previous studies have revealed that dog CYP2C21 showed substrate specificity for diclofenac, midazolam, and the 4-methyl-N-methyl analog of sulfaphenazole, O-desmethyltramadol, testosterone (16-alpha OH), and (S)-Mephenytoin [39,41–43]. CYP2C41 also showed similar substrate specificity to diclofenac and midazolam albeit with lower activity. However, comprehensive and systematic analysis of recombinant CYP2C21 and CYP2C41 in the dog is necessary for the clarification of specific substrate-specificity. Our results together with this substrate specificity suggest that Carnivora CYP2C21s and 2C41s might also show similar substrate specificity. Our analysis revealed brown bear and black bear also showed gene expansion in CYP2C21 and 2C41s. Since data regarding substrate specificity of CYP2Cs in Carnivora are limited, we need to estimate Carnivora CYP2Cs in Ursidae from other mammals. Human CYP2Cs showed metabolism of a wide variety of chemicals including endogenous eicosanoids and fatty acid [44], xenobiotic drugs such as antimalarials, oral antidiabetics, most NSAIDs, most proton pump inhibitors and warfarin [2,45], and some terpenoids [46–48]. Interestingly, brown bears, black bears, and even badger are known to consume pine nuts [33,49,50], which are from conifer trees that contain terpenoids [51,52]. Further study investigating recombinant CYP2Cs is essential to confirm whether these expanded CYP2Cs in the Ursid and badger are able to metabolize the terpenoids in their daily diets.

Among Canids, CYP2C41s show polymorphism as a complete loss in several breeds [53], suggesting some dogs have contracted CYP2Cs. This further suggests that other species might also show polymorphism or copy number variants within species, providing further justification for the importance of genomic analyses that use several individuals to conclude isoform numbers in each species.

4.3. CYP2C23s Orthologous Genes in Carnivora

From the phylogenetic analysis, we discovered a possible orthologue to rat CYP2C23 and mouse CYP2C44 in Carnivora. The coding locus of rat CYP2C23 and mouse CYP2C44 (between Erlin1 and Cpn1) was neither in the CYP2Cs cluster, nor in a similar region to that of Carnivorans. These isoforms have been cloned and characterized as arachidonic acid or eicosanoid metabolizing CYPs and are closely related to the endogenous biosynthesis of these animals [38,54,55], although humans have pseudogenes of these isoforms. We found possible orthologues to rodents CYP2C23s in Carnivora, which strongly suggests these isoforms in Carnivora also have similar eicosanoid metabolism roles. We also found specific deletion of CYP2C23s in Feliformia and Mustelidae. This could indicate that eicosanoid metabolism in these animals is different to other carnivorans. A further BLAST analysis indicated possible CYP2C23s orthologues in cattle, horse, and pangolin in loci between Erlin1 and Cpn1 (data not shown), suggesting the CYP2C23 gene translocated after Carnivora divergence.

4.4. CYP3As in Mammals

Human CYP3As catalyzes a wide range of prescribed drugs and is considered one of the most important subfamilies for drug metabolism in humans [2,56], yet they also catalyze metabolism of endogenous chemicals such as steroids, cholesterol, and bile acids [2,57]. Canine CYP3As (e.g., CYP3A12, 3A26, 3A98, and 3A99) have been cloned and characterized, showing similar substrate specificity patterns as human CYP3As [39,58,59]; however, with different expression patterns. In contrast, CYP3A12 and 3A26 are liver specific [58,60], whereas CYP3A98 is expressed in the intestine and 3A99 in the liver and intestine [26]. These features and the phylogenetic result indicate the CYP3As clade 3 could be intestinal-specific isoforms whereas the CYP3As clade 1 could be liver-specific orthologs in Carnivora. However, the CYP3As clade 2 expression patterns remain unclear. This tissue-specific expression of the CYP3As, especially in the liver and intestine, has also been observed in evolutionarily distant species, like humans.

Among Feline CYP3As, similar expression patterns have been characterized, which means that CYP3A132 is mainly expressed in the liver, whereas CYP3A131 is mainly expressed in the intestine with lower levels in the liver [27,61,62]. These features, together with our result, further indicate that the Feliformia CYP3A132 may be major CYPs in the liver, whereas CYP3A131 may be intestine specific in Feliformia. In other Feliformia, such as the striped hyena, meerkat, and masked palm civet, however, we found other clades of CYP3As. Unfortunately, Feliformia genomic data are limited and we could not clearly demonstrate the evolutionary history of CYP3As in this taxon. Further studies on a greater variety of Feliformia species' genomics and expression pattern are necessary.

Similar to CYP2Cs, we found CYP3As expansion in omnivorous species: the dog, badger, brown bear, and black bear. CYP3As catalyze a wide range of xenobiotics including some pyrrolizidine alkaloids [63–65] found mainly in the families Asteraceae, Boraginaceae, and Fabaceae. Thus, the omnivorous diets of bears and badgers might be an evolutionary driving force enabling these species to cope with accidental intake of these toxins.

Our phylogenetic analysis indicated that diversification of CYP3As in Caniformia and Feliformia was completely different, suggesting that substrate specificity of CYP3As among these taxa is different. However, the phylogenetically distant isoform human CYP3A4 showed similar substrate specificity to CYP3As in dogs and cats (albeit with several differences) [26,27,43], indicating that substrate specificity in Carnivora need not

range widely to cope with a variety of chemicals. However, further systemic analysis CYP3As function in Carnivora is required.

4.5. Other CYP Isoforms

In our analysis, CYP1-3 families did not show strong differences. However, in domestic cat, specific duplication and loss events were observed. For instance, CYP2C21 has been reported as a pseudogene [66], and CYP2E1, which mainly catalyzes acetaminophen or alcohol, has been specifically duplicated in this species [67]. In our study, we only identified a similar phenomenon of CYP2E duplication in the fishing cat (*Prionailurus bengalensis*) and we did not record CYP2C21 loss in any other Felidae species, suggesting that these duplication and loss events might be specific to cats or Felidae species closely related to cats. Expression of CYP2C has also been limited in cats, whereas CYP2Es are dominant in the liver compared to other isoforms [62,68], which makes it difficult to clarify which isoforms are important for xenobiotic metabolism in Felidae. Thus, a functional analysis is essential for further discussion.

5. Conclusions

In this study we comprehensively analyzed the evolutionary features of CYP1-3 families in Carnivora. We found specific expansion of CYP2C and 3As in omnivorous Carnivora such as the badger, the brown bear, the black bear and the dog genomes. Our phylogenetic analysis further revealed possible orthologs of CYP2C21s, 2C41s, and 2C23s in Carnivora. Furthermore, we found the evolution of CYP3As was completely different in Caniformia and Feliformia, and within each taxon we detected specific CYP3As duplication events. These studies provide fundamental evolutionary and genetic information for extrapolating the pharmacokinetics or toxicokinetic of experimental animals to that of wild Carnivora, which include a wide variety of top-predator, key-stone, and rare species threatened with extinction.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/ani12202821/s1>; Figure S1: Synteny of Pinniped CYP3As.; Sequence files: CYP2C_protein.fas, CYP3A_protein.fas; Table S1: Sequence information of CYP2Cs; Table S2: Sequence information of CYP3As; Table S3: Food habitat of carnivoran [69–131]; Table S4: Assemble and annotation information. We also provided protein sequence files in CYP3A_protein.fas, and CYP2C_protein.fas.

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References

- Nelson, D.R.; Kamataki, T.; Waxman, D.J.; Guengerich, F.P.; Estabrook, R.W.; Feyereisen, R.; Gonzalez, F.J.; Coon, M.J.; Gunsalus, I.C.; Gotoh, O.; et al. The P450 Superfamily: Update on New Sequences, Gene Mapping, Accession Numbers, Early Trivial Names of Enzymes, and Nomenclature. *DNA Cell Biol.* **2009**, *12*, 1–51. [[CrossRef](#)] [[PubMed](#)]
- Zanger, U.M.; Schwab, M. Cytochrome P450 Enzymes in Drug Metabolism: Regulation of Gene Expression, Enzyme Activities, and Impact of Genetic Variation. *Pharmacol. Ther.* **2013**, *138*, 103–141. [[CrossRef](#)] [[PubMed](#)]
- Verma, H.; Bahia, M.S.; Choudhary, S.; Singh, P.K.; Silakari, O. Drug Metabolizing Enzymes-Associated Chemo Resistance and Strategies to Overcome It. *Drug Metab. Rev.* **2019**, *51*, 196–223. [[CrossRef](#)] [[PubMed](#)]
- Ramaiahgari, S.C.; Waidyanatha, S.; Dixon, D.; DeVito, M.J.; Paules, R.S.; Ferguson, S.S. From the Cover: Three-Dimensional (3D) HepaRG Spheroid Model With Physiologically Relevant Xenobiotic Metabolism Competence and Hepatocyte Functionality for Liver Toxicity Screening. *Toxicol. Sci.* **2017**, *159*, 124–136. [[CrossRef](#)]
- Nelson, D.R.; Koymans, L.; Kamataki, T.; Stegeman, J.J.; Feyereisen, R.; Waxman, D.J.; Waterman, M.R.; Gotoh, O.; Coon, M.J.; Estabrook, R.W.; et al. P450 Superfamily: Update on New Sequences, Gene Mapping, Accession Numbers and Nomenclature. *Pharmacogenetics* **1996**, *6*, 1–42. [[CrossRef](#)]
- Zhao, M.; Ma, J.; Li, M.; Zhang, Y.; Jiang, B.; Zhao, X.; Huai, C.; Shen, L.; Zhang, N.; He, L.; et al. Cytochrome P450 Enzymes and Drug Metabolism in Humans. *Int. J. Mol. Sci.* **2021**, *22*, 12808. [[CrossRef](#)]
- Sabbagh, A.; Marin, J.; Veyssiére, C.; Lecompte, E.; Boukouvala, S.; Poloni, E.S.; Darlu, P.; Crouau-Roy, B. Rapid Birth-and-Death Evolution of the Xenobiotic Metabolizing NAT Gene Family in Vertebrates with Evidence of Adaptive Selection. *BMC Evol. Biol.* **2013**, *13*, 62. [[CrossRef](#)]
- Kawashima, A.; Satta, Y. Substrate-Dependent Evolution of Cytochrome P450: Rapid Turnover of the Detoxification-Type and Conservation of the Biosynthesis-Type. *PLoS ONE* **2014**, *9*, e100059. [[CrossRef](#)]
- Thomas, J.H. Rapid Birth–Death Evolution Specific to Xenobiotic Cytochrome P450 Genes in Vertebrates. *PLOS Genet.* **2007**, *3*, e67. [[CrossRef](#)]
- Bock, K.W. Vertebrate UDP-Glucuronosyltransferases: Functional and Evolutionary Aspects. *Biochem. Pharmacol.* **2003**, *66*, 691–696. [[CrossRef](#)]
- Johnson, R.N.; O’Meally, D.; Chen, Z.; Etherington, G.J.; Ho, S.Y.W.; Nash, W.J.; Grueber, C.E.; Cheng, Y.; Whittington, C.M.; Dennison, S.; et al. Adaptation and Conservation Insights from the Koala Genome. *Nat. Genet.* **2018**, *50*, 1102–1111. [[CrossRef](#)] [[PubMed](#)]
- Greenhalgh, R.; Holding, M.L.; Orr, T.J.; Henderson, J.B.; Parchman, T.L.; Matocq, M.D.; Shapiro, M.D.; Dearing, M.D. Trio-Binned Genomes of the Woodrats Neotoma Bryanti and Neotoma Lepida Reveal Novel Gene Islands and Rapid Copy Number Evolution of Xenobiotic Metabolizing Genes. *Mol. Ecol. Resour.* **2022**, *22*, 2713–2731. [[CrossRef](#)]
- Watanabe, K.P.; Kawai, Y.K.; Ikenaka, Y.; Kawata, M.; Ikushiro, S.I.; Sakaki, T.; Ishizuka, M. Avian Cytochrome P450 (CYP) 1-3 Family Genes: Isoforms, Evolutionary Relationships, and mRNA Expression in Chicken Liver. *PLoS ONE* **2013**, *8*, e75689. [[CrossRef](#)]
- Radominska-Pandya, A.; Bratton, S.; Little, J. A Historical Overview of the Heterologous Expression of Mammalian UDP-Glucuronosyltransferase Isoforms Over the Past Twenty Years. *Curr. Drug Metab.* **2005**, *6*, 141–160. [[CrossRef](#)] [[PubMed](#)]
- Feng, P.; Liu, Z. Complex Gene Expansion of the CYP2D Gene Subfamily. *Ecol. Evol.* **2018**, *8*, 11022–11030. [[CrossRef](#)]
- Hassanin, A.; Veron, G.; Ropiquet, A.; van Vuuren, B.J.; Lécu, A.; Goodman, S.M.; Haider, J.; Nguyen, T.T. Evolutionary History of Carnivora (Mammalia, Laurasiatheria) Inferred from Mitochondrial Genomes. *PLoS ONE* **2021**, *16*, e0240770. [[CrossRef](#)]
- Werdelin, L.; Dehghani, R. *Carnivora*; Springer: Berlin/Heidelberg, Germany, 2011.
- Kondo, T.; Ikenaka, Y.; Nakayama, S.M.M.; Kawai, Y.K.; Mizukawa, H.; Mitani, Y.; Nomiyama, K.; Tanabe, S.; Ishizuka, M. Uridine Diphosphate-Glucuronosyltransferase (UGT) 2B Subfamily Interspecies Differences in Carnivores. *Toxicol. Sci.* **2017**, *158*, 90–100. [[CrossRef](#)]
- Kakehi, M.; Ikenaka, Y.; Nakayama, S.M.M.; Kawai, Y.K.; Watanabe, K.P.; Mizukawa, H.; Nomiyama, K.; Tanabe, S.; Ishizuka, M. Uridine Diphosphate-Glucuronosyltransferase (UGT) Xenobiotic Metabolizing Activity and Genetic Evolution in Pinniped Species. *Toxicol. Sci.* **2015**, *147*, 360–369. [[CrossRef](#)]
- Shrestha, B.; Reed, J.M.; Starks, P.T.; Kaufman, G.E.; Goldstone, J.V.; Roelke, M.E.; O’Brien, S.J.; Koepfli, K.P.; Frank, L.G.; Court, M.H. Evolution of a Major Drug Metabolizing Enzyme Defect in the Domestic Cat and Other Felidae: Phylogenetic Timing and the Role of Hypercarnivory. *PLoS ONE* **2011**, *6*, 221–237. [[CrossRef](#)]
- Genereux, D.P.; Serres, A.; Armstrong, J.; Johnson, J.; Marinescu, V.D.; Murén, E.; Juan, D.; Bejerano, G.; Casewell, N.R.; Chemnick, L.G.; et al. A Comparative Genomics Multitool for Scientific Discovery and Conservation. *Nature* **2020**, *587*, 240–245. [[CrossRef](#)]
- Kumar, S.; Stecher, G.; Li, M.; Knyaz, C.; Tamura, K. MEGA X: Molecular Evolutionary Genetics Analysis across Computing Platforms. *Mol. Biol. Evol.* **2018**, *35*, 1547–1549. [[CrossRef](#)] [[PubMed](#)]
- Kumar, S.; Stecher, G.; Suleski, M.; Blair Hedges, S. TimeTree: A Resource for Timelines, Timetrees, and Divergence Times. *Mol. Biol. Evol.* **2017**, *34*, 1812–1819. [[CrossRef](#)] [[PubMed](#)]
- Buels, R.; Yao, E.; Diesch, C.M.; Hayes, R.D.; Munoz-Torres, M.; Helt, G.; Goodstein, D.M.; Elsik, C.G.; Lewis, S.E.; Stein, L.; et al. JBrowse: A Dynamic Web Platform for Genome Visualization and Analysis. *Genome Biol.* **2016**, *17*, 1–12. [[CrossRef](#)]

25. Lan, T.; Fang, D.; Li, H.; Sahu, S.K.; Wang, Q.; Yuan, H.; Zhu, Y.; Yang, Z.; Zhang, L.; Yang, S.; et al. Chromosome-Scale Genome of Masked Palm Civet (*Paguma Larvata*) Shows Genomic Signatures of Its Biological Characteristics and Evolution. *Front. Genet.* **2022**, *12*, 2699. [[CrossRef](#)]
26. Uno, Y.; Jikuya, S.; Noda, Y.; Murayama, N.; Yamazaki, H. A Comprehensive Investigation of Dog Cytochromes P450 3A (CYP3A) Reveals A Functional Role of Newly Identified CYP3A98 in Small Intestine. *Drug Metab. Dispos.* **2022**, *50*, DMD-AR-2021-000749. [[CrossRef](#)]
27. Honda, K.; Komatsu, T.; Koyama, F.; Kubota, A.; Kawakami, K.; Asakura, H.; Uno, Y.; Kitazawa, T.; Hiraga, T.; Teraoka, H. Expression of Two Novel Cytochrome P450 3A131 and 3A132 in Liver and Small Intestine of Domestic Cats. *J. Vet. Med. Sci.* **2011**, *73*, 1489–1492. [[CrossRef](#)] [[PubMed](#)]
28. Haley, S.L.; Lamb, J.G.; Franklin, M.R.; Constance, J.E.; Denise Dearing, M. Xenobiotic Metabolism of Plant Secondary Compounds in Juniper (*Juniperus Monosperma*) by Specialist and Generalist Woodrat Herbivores, Genus *Neotoma*. *Comp. Biochem. Physiol. Part. C Toxicol. Pharmacol.* **2007**, *146*, 552–560. [[CrossRef](#)]
29. Kitanovic, S.; Orr, T.J.; Spalink, D.; Cocke, G.B.; Schramm, K.; Wilderman, P.R.; Halpert, J.R.; Dearing, M.D. Role of Cytochrome P450 2B Sequence Variation and Gene Copy Number in Facilitating Dietary Specialization in Mammalian Herbivores. *Mol. Ecol.* **2018**, *27*, 723–736. [[CrossRef](#)]
30. Mangipane, L.S.; Belant, J.L.; Lafferty, D.J.R.; Gustine, D.D.; Hiller, T.L.; Colvin, M.E.; Mangipane, B.A.; Hilderbrand, G.V. Dietary Plasticity in a Nutrient-Rich System Does Not Influence Brown Bear (*Ursus Arctos*) Body Condition or Denning. *Polar Biol.* **2018**, *41*, 763–772. [[CrossRef](#)]
31. Costello, C.M.; Cain, S.L.; Pils, S.; Frattaroli, L.; Haroldson, M.A.; Van Manen, F.T. Diet and Macronutrient Optimization in Wild Ursids: A Comparison of Grizzly Bears with Sympatric and Allopatric Black Bears. *PLoS ONE* **2016**, *11*, e0153702. [[CrossRef](#)]
32. Kruuk, H.; Parish, T. Feeding Specialization of the European Badger (*Meles Meles*) in Scotland. *J. Anim. Ecol.* **1981**, *50*, 773–788. [[CrossRef](#)]
33. Balestrieri, A.; Remonti, L.; Saino, N.; Raubenheimer, D. The ‘Omnivorous Badger Dilemma’: Towards an Integration of Nutrition with the Dietary Niche in Wild Mammals. *Mamm. Rev.* **2019**, *49*, 324–339. [[CrossRef](#)]
34. Davies, M. Veterinary Clinical Nutrition: Success Stories: An Overview. *Proc. Nutr. Soc.* **2016**, *75*, 392–397. [[CrossRef](#)]
35. Huang, G.; Wang, X.; Hu, Y.; Wu, Q.; Nie, Y.; Dong, J.; Ding, Y.; Yan, L.; Wei, F. Diet Drives Convergent Evolution of Gut Microbiomes in Bamboo-Eating Species. *Sci. China Life Sci.* **2021**, *64*, 88–95. [[CrossRef](#)]
36. Wei, F.; Hu, Y.; Yan, L.; Nie, Y.; Wu, Q.; Zhang, Z. Giant Pandas Are Not an Evolutionary Cul-de-Sac: Evidence from Multidisciplinary Research. *Mol. Biol. Evol.* **2015**, *32*, 4–12. [[CrossRef](#)] [[PubMed](#)]
37. Huang, G.; Wang, L.; Li, J.; Hou, R.; Wang, M.; Wang, Z.; Qu, Q.; Zhou, W.; Nie, Y.; Hu, Y.; et al. Seasonal Shift of the Gut Microbiome Synchronizes Host Peripheral Circadian Rhythm for Physiological Adaptation to a Low-Fat Diet in the Giant Panda. *Cell Rep.* **2022**, *38*, 110203. [[CrossRef](#)]
38. Imaoka, S.; Wedlund, P.J.; Ogawa, H.; Kimura, S.; Gonzalez, F.J.; Kim, H.Y. Identification of CYP2C23 Expressed in Rat Kidney as an Arachidonic Acid Epoxygenase. *J. Pharmacol. Exp. Ther.* **1993**, *267*, 1012–1016.
39. Shou, M.; Norcross, R.; Sandig, G.; Lu, P.; Li, Y.; Lin, Y.; Mei, Q.; Rodrigues, A.D.; Rushmore, T.H. Substrate Specificity and Kinetic Properties of Seven Heterologously Expressed Dog Cytochromes P450. *Drug Metab. Dispos.* **2003**, *31*, 1161–1169. [[CrossRef](#)]
40. Blaisdell, J.; Goldstein, J.A.; Bai, S.A. Isolation of a New Canine Cytochrome P450 CDNA from the Cytochrome P450 2C Subfamily (CYP2C41) and Evidence for Polymorphic Differences in Its Expression. *Drug Metab. Dispos.* **1998**, *26*, 278–283.
41. Zielinski, J.; Mevissen, M. Inhibition of in Vitro Metabolism of Testosterone in Human, Dog and Horse Liver Microsomes to Investigate Species Differences. *Toxicol. Vitr.* **2015**, *29*, 468–478. [[CrossRef](#)]
42. Perez Jimenez, T.E.; Mealey, K.L.; Schnider, D.; Grubb, T.L.; Greene, S.A.; Court, M.H. Identification of Canine Cytochrome P-450s (CYPs) Metabolizing the Tramadol (+)-M1 and (+)-M2 Metabolites to the Tramadol (+)-M5 Metabolite in Dog Liver Microsomes. *J. Vet. Pharmacol. Ther.* **2018**, *41*, 815–824. [[CrossRef](#)] [[PubMed](#)]
43. Court, M.H. Canine Cytochrome P-450 Pharmacogenetics. *Vet. Clin. North. Am. Small Anim. Pract.* **2013**, *43*, 1027–1038. [[CrossRef](#)] [[PubMed](#)]
44. Capdevila, J.H.; Falck, J.R.; Harris, R.C. Cytochrome P450 and Arachidonic Acid Bioactivation: Molecular and Functional Properties of the Arachidonate Monooxygenase. *J. Lipid Res.* **2000**, *41*, 163–181. [[CrossRef](#)]
45. Esteves, F.; Rueff, J.; Kranendonk, M. The Central Role of Cytochrome P450 in Xenobiotic Metabolism—A Brief Review on a Fascinating Enzyme Family. *J. Xenobiotics* **2021**, *11*, 7. [[CrossRef](#)]
46. Miyazawa, M.; Shindo, M.; Shimada, T. Metabolism of (+)- and (−)-Limonenes to Respective Carveols and Perillyl Alcohols by CYP2C9 and CYP2C19 in Human Liver Microsomes. *Drug Metab. Dispos.* **2002**, *30*, 602–607. [[CrossRef](#)]
47. Jiang, R.; Yamaori, S.; Takeda, S.; Yamamoto, I.; Watanabe, K. Identification of Cytochrome P450 Enzymes Responsible for Metabolism of Cannabidiol by Human Liver Microsomes. *Life Sci.* **2011**, *89*, 165–170. [[CrossRef](#)]
48. Miyazawa, M.; Sugie, A.; Shimada, T. Roles of Human Cyp2a6 And 2b6 and Rat Cyp2c11 and 2b1 in the 10-Hydroxylation of (−)-Verbenone by Liver Microsomes. *Drug Metab. Dispos.* **2003**, *31*, 1049–1053. [[CrossRef](#)]
49. Kendall, K.C. Use of Pine Nuts by Grizzly and Black Bears in the Yellowstone Area. *Bears Biol. Manag.* **1983**, *5*, 166. [[CrossRef](#)]
50. Felicetti, L.A.; Schwartz, C.C.; Rye, R.O.; Haroldson, M.A.; Gunther, K.A.; Phillips, D.L.; Robbins, C.T. Use of Sulfur and Nitrogen Stable Isotopes to Determine the Importance of Whitebark Pine Nuts to Yellowstone Grizzly Bears. *Can. J. Zool.* **2011**, *81*, 763–770. [[CrossRef](#)]

51. Zulak, K.G.; Bohlmann, J. Terpenoid Biosynthesis and Specialized Vascular Cells of Conifer Defense. *J. Integr. Plant. Biol.* **2010**, *52*, 86–97. [[CrossRef](#)]
52. Kopacyk, J.M.; Warguła, J.; Jelonek, T. The Variability of Terpenes in Conifers under Developmental and Environmental Stimuli. *Environ. Exp. Bot.* **2020**, *180*, 104197. [[CrossRef](#)]
53. Karakus, E.; Prinzing, C.; Leiting, S.; Geyer, J. Sequencing of the Canine Cytochrome P450 CYP2C41 Gene and Genotyping of Its Polymorphic Occurrence in 36 Dog Breeds. *Front. Vet. Sci.* **2021**, *8*, 663175. [[CrossRef](#)]
54. DeLozier, T.C.; Tsao, C.C.; Coulter, S.J.; Foley, J.; Bradbury, J.A.; Zeldin, D.C.; Goldstein, J.A. CYP2C44, a New Murine CYP2C That Metabolizes Arachidonic Acid to Unique Stereospecific Products. *J. Pharmacol. Exp. Ther.* **2004**, *310*, 845–854. [[CrossRef](#)] [[PubMed](#)]
55. Capdevila, J.; Wang, W. Role of Cytochrome P450 Epoxidase in Regulating Renal Membrane Transport and Hypertension. *Curr. Opin. Nephrol. Hypertens.* **2013**, *22*, 163–169. [[CrossRef](#)] [[PubMed](#)]
56. Ingelman-Sundberg, M. Human Drug Metabolising Cytochrome P450 Enzymes: Properties and Polymorphisms. *Naunyn. Schmiedebergs. Arch. Pharmacol.* **2004**, *369*, 89–104. [[CrossRef](#)]
57. Wright, W.C.; Chenge, J.; Chen, T. Structural Perspectives of the CYP3A Family and Their Small Molecule Modulators in Drug Metabolism. *Liver Res.* **2019**, *3*, 132–142. [[CrossRef](#)]
58. Fraser, D.J.; Feyereisen, R.; Harlow, G.R.; Halpert, J.R. Isolation, Heterologous Expression and Functional Characterization of a Novel Cytochrome P450 3A Enzyme from a Canine Liver cDNA Library. *J. Pharmacol. Exp. Ther.* **1997**, *283*, 1425–1432.
59. Locuson, C.W.; Ethell, B.T.; Voice, M.; Lee, D.; Feenstra, K.L. Evaluation of Escherichia Coli Membrane Preparations of Canine CYP1A1, 2B11, 2C21, 2C41, 2D15, 3A12, and 3A26 with Coexpressed Canine Cytochrome P450 Reductase. *Drug Metab. Dispos.* **2009**, *37*, 457–461. [[CrossRef](#)]
60. Martinez, S.E.; Shi, J.; Zhu, H.J.; Jimenez, T.E.P.; Zhu, Z.; Court, M.H. Absolute Quantitation of Drug-Metabolizing Cytochrome P450 Enzymes and Accessory Proteins in Dog Liver Microsomes Using Label-Free Standard-Free Analysis Reveals Interbreed Variability. *Drug Metab. Dispos.* **2019**, *47*, 1314–1324. [[CrossRef](#)]
61. Sugiyama, S.; Uno, Y.; Amano, T.; Kitazawa, T.; Teraoka, H. Genetic Diversity of Cytochrome P450 3A with Different Metabolic Activity in Domestic Cats. *J. Vet. Med. Sci.* **2019**, *81*, 598–600. [[CrossRef](#)]
62. Visser, M.; Weber, K.L.; Lyons, L.A.; Rincon, G.; Boothe, D.M.; Merritt, D.A. Identification and Quantification of Domestic Feline Cytochrome P450 Transcriptome across Multiple Tissues. *J. Vet. Pharmacol. Ther.* **2019**, *42*, 7–15. [[CrossRef](#)] [[PubMed](#)]
63. Fu, P.P. Pyrrolizidine Alkaloids: Metabolic Activation Pathways Leading to Liver Tumor Initiation. *Chem. Res. Toxicol.* **2017**, *30*, 81–93. [[CrossRef](#)] [[PubMed](#)]
64. Liu, Y. Incorporation of Absorption and Metabolism into Liver Toxicity Prediction for Phytochemicals: A Tiered in Silico QSAR Approach. *Food Chem. Toxicol.* **2018**, *118*, 409–415. [[CrossRef](#)] [[PubMed](#)]
65. Huan, J.Y.; Miranda, C.L.; Buhler, D.R.; Cheeke, P.R. The Roles of CYP3A and CYP2B Isoforms in Hepatic Bioactivation and Detoxification of the Pyrrolizidine Alkaloid Senecionine in Sheep and Hamsters. *Toxicol. Appl. Pharmacol.* **1998**, *151*, 229–235. [[CrossRef](#)] [[PubMed](#)]
66. Ono, Y.; Sugiyama, S.; Matsushita, M.; Kitazawa, T.; Amano, T.; Uno, Y.; Ikushiro, S.; Teraoka, H. Limited Expression of Functional Cytochrome P450 2c Subtypes in the Liver and Small Intestine of Domestic Cats. *Xenobiotica* **2018**, *49*, 627–635. [[CrossRef](#)] [[PubMed](#)]
67. Tanaka, N.; Shinkyo, R.; Sakaki, T.; Kasamatsu, M.; Imaoka, S.; Funae, Y.; Yokota, H. Cytochrome P450 2E Polymorphism in Feline Liver. *Biochim. Biophys. Acta Gen. Subj.* **2005**, *1726*, 194–205. [[CrossRef](#)]
68. Khidkhan, K.; Mizukawa, H.; Ikenaka, Y.; Nakayama, S.M.M.; Nomiyama, K.; Yokoyama, N.; Ichii, O.; Darwish, W.S.; Takiguchi, M.; Tanabe, S.; et al. Tissue Distribution and Characterization of Feline Cytochrome P450 Genes Related to Polychlorinated Biphenyl Exposure. *Comp. Biochem. Physiol. Part. C Toxicol. Pharmacol.* **2019**, *226*, 108613. [[CrossRef](#)]
69. Rezaei Khozani, A.; Kaboli, M.; Ashrafi, S.; Akbari, H. Survey Diet of Asiatic Cheetah Acinonyx Jubatus Venaticus by Scat Analysis Method in Bafgh Protected Area, Central Iran. *J. Anim. Environ.* **2016**, *8*, 1–8.
70. Mills, M.G.L.; Broomhall, L.S.; Du Toit, J.T. Cheetah Acinonyx Jubatus Feeding Ecology in the Kruger National Park and a Comparison across African Savanna Habitats: Is the Cheetah Only a Successful Hunter on Open Grassland Plains? *Wildlife Biol.* **2004**, *10*, 177–186. [[CrossRef](#)]
71. Schaller, G.B. *The Giant Pandas of Wolong*; University of Chicago Press: Chicago, IL, USA, 1985; p. 298.
72. Gudmundson, C.J.; Zeppelin, T.K.; Ream, R.R. Application of Two Methods for Determining Diet of Northern Fur Seals (*Callorhinus Ursinus*). *Fish Bull.* **2006**, *104*, 445–456.
73. Berta, A.; Churchill, M.; Boessenecker, R.W. The Origin and Evolutionary Biology of Pinnipeds: Seals, Sea Lions, and Walruses. *Annu. Rev. Earth Planet. Sci.* **2018**, *46*, 203–228. [[CrossRef](#)]
74. Costa, D.P.; McHuron, E.A. (Eds.) *Ethology and Behavioral Ecology of Phocids*; Springer: Cham, Switzerland, 2022; p. 645. [[CrossRef](#)]
75. Pauly, D.; Trites, A.W.; Capuli, E.; Christensen, V. Diet Composition and Trophic Levels of Marine Mammals. *ICES J. Mar. Sci.* **1998**, *55*, 467–481. [[CrossRef](#)]
76. Bosch, G.; Hagen-Plantinga, E.A.; Hendriks, W.H. Dietary Nutrient Profiles of Wild Wolves: Insights for Optimal Dog Nutrition? *Br. J. Nutr.* **2015**, *113*, S40–S54. [[CrossRef](#)] [[PubMed](#)]
77. Soars, M.G.; Riley, R.J.; Findlay, K.A.; Coffey, M.J.; Burchell, B. Evidence for Significant Differences in Microsomal Drug Glucuronidation by Canine and Human Liver and Kidney. *Drug Metab. Dispos.* **2001**, *29*, 121–126.

78. Baker, D.H.; Czarnecki-Maulden, G.L. Comparative Nutrition of Cats and Dogs. *Annu. Rev. Nutr.* **1991**, *11*, 239–263. [[CrossRef](#)]
79. Montanari, S. Discrimination Factors of Carbon and Nitrogen Stable Isotopes in Meerkat Feces. *PeerJ* **2017**, *2017*, e3436. [[CrossRef](#)]
80. Brox, B.W.; Edwards, K.; Buist, N.A.; Macaskill, A.C. Investigating Food Preference in Zoo-Housed Meerkats. *Zoo Biol.* **2021**, *40*, 517–526. [[CrossRef](#)]
81. Johnson, C.K.; Tinker, M.T.; Estes, J.A.; Conrad, P.A.; Staedler, M.; Miller, M.A.; Jessup, D.A.; Mazet, J.A.K. Prey Choice and Habitat Use Drive Sea Otter Pathogen Exposure in a Resource-Limited Coastal System. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 2242–2247. [[CrossRef](#)]
82. Rand, K.; McDermott, S.; Logerwell, E.; Matta, M.E.; Levine, M.; Bryan, D.R.; Spies, I.B.; Loomis, T. Higher Aggregation of Key Prey Species Associated with Diet and Abundance of the Steller Sea Lion *Eumetopias Jubatus* across the Aleutian Islands. *Mar. Coast. Fish.* **2019**, *11*, 472–486. [[CrossRef](#)]
83. Edinboro, C.H.; Scott-Moncrieff, J.C.; Janovitz, E.; Thacker, H.L.; Glickman, L.T. Epidemiologic Study of Relationships between Consumption of Commercial Canned Food and Risk of Hyperthyroidism in Cats. *J. Am. Vet. Med. Assoc.* **2004**, *224*, 879–886. [[CrossRef](#)]
84. Hammill, M.O.; Stenson, G.B. Estimated Prey Consumption by Harp Seals (*Phoca Groenlandica*) & Hooded Seals (*Cystophora Cristata*) & Grey Seals (*Halichoerus Grypus*) and Harbour Seals (*Phoca Vitulina*) in Atlantic Canada. *J. Northw. Atl. Fish. Sci.* **2000**, *26*, 1–23.
85. Alam, M.S.; Khan, J.A. Food Habits of Striped Hyena (*Hyaena Hyaena*) in a Semi-Arid Conservation Area of India. *J. Arid. Land* **2015**, *7*, 860–866. [[CrossRef](#)]
86. Bhandari, S.; Morley, C.; Aryal, A.; Shrestha, U.B. The Diet of the Striped Hyena in Nepal’s Lowland Regions. *Ecol. Evol.* **2020**, *10*, 7953–7962. [[CrossRef](#)] [[PubMed](#)]
87. Goetz, K.T.; Burns, J.M.; Hückstädt, L.A.; Sher, M.R.; Costa, D.P. Temporal Variation in Isotopic Composition and Diet of Weddell Seals in the Western Ross Sea. *Deep Res. Part II Top. Stud. Oceanogr.* **2017**, *140*, 36–44. [[CrossRef](#)]
88. Cote, D.; Stewart, H.M.J.; Gregory, R.S.; Gosse, J.; Reynolds, J.J.; Stenson, G.B.; Miller, E.H. Prey Selection by Marine-Coastal River Otters (*Lontra Canadensis*) in Newfoundland, Canada. *J. Mammal.* **2008**, *89*, 1001–1011. [[CrossRef](#)]
89. Day, C.C.; Westover, M.D.; McMillan, B.R. Seasonal Diet of the Northern River Otter (*Lontra Canadensis*): What Drives Prey Selection? *Can. J. Zool.* **2015**, *93*, 197–205. [[CrossRef](#)]
90. Krawczyk, A.J.; Bogdziewicz, M.; Majkowska, K.; Glazaczow, A. Diet Composition of the Eurasian Otter *Lutra Lutra* in Different Freshwater Habitats of Temperate Europe: A Review and Meta-Analysis. *Mamm. Rev.* **2016**, *46*, 106–113. [[CrossRef](#)]
91. Burstahler, C.M.; Terwissen, C.V.; Roth, J.D. Latitudinal Gradient in Cortisol Concentrations in Canada Lynx (*Lynx Canadensis*) Is Not Explained by Diet. *Can. J. Zool.* **2019**, *97*, 748–753. [[CrossRef](#)]
92. Parker, G.R.; Maxwell, J.W.; Morton, L.D.; Smith, G.E.J. The Ecology of the Lynx (*Lynx Canadensis*) on Cape Breton Island. *Can. J. Zool.* **2011**, *61*, 770–786. [[CrossRef](#)]
93. Cleary, G.P.; Corner, L.A.L.; O’Keeffe, J.; Marples, N.M. The Diet of the Badger *Meles Meles* in the Republic of Ireland. *Mamm. Biol.* **2009**, *74*, 438–447. [[CrossRef](#)]
94. Rita, D.; Drago, M.; Galimberti, F.; Cardona, L. Temporal Consistency of Individual Trophic Specialization in Southern Elephant Seals *Mirounga Leonina*. *Mar. Ecol. Prog. Ser.* **2017**, *585*, 229–242. [[CrossRef](#)]
95. Spurlin, S.M.; Peterson, S.H.; Crocker, D.E.; Costa, D.P. Nitrogen and Carbon Stable-Isotope Ratios Change in Adult Northern Elephant Seals (*Mirounga Angustirostris*) during the Breeding and Molting Fasts. *Mar. Mammal Sci.* **2019**, *35*, 707–717. [[CrossRef](#)]
96. Condit, R.; Le Boeuf, B.J. Feeding Habits and Feeding Grounds of the Northern Elephant Seal. *J. Mammal.* **1984**, *65*, 281–290. [[CrossRef](#)]
97. Dell’Arte, G.L.; Laaksonen, T.; Norrdahl, K.; Korpimäki, E. Variation in the Diet Composition of a Generalist Predator, the Red Fox, in Relation to Season and Density of Main Prey. *Acta Oecologica* **2007**, *31*, 276–281. [[CrossRef](#)]
98. McDonald, R.A.; Webbon, C.; Harris, S. The Diet of Stoats (*Mustela Erminea*) and Weasels (*Mustela Nivalis*) in Great Britain. *J. Zool.* **2000**, *252*, 363–371. [[CrossRef](#)]
99. Smith, G.P.; Ragg, J.R.; Waldrup, K.A.; Moller, H. Diet of Feral Ferrets (*Mustela Furo*) from Pastoral Habitats in Otago and Southland, New Zealand. *N. Z. J. Zool.* **1995**, *22*, 363–369. [[CrossRef](#)]
100. Vinke, C.M.; Schoemaker, N.J. The Welfare of Ferrets (*Mustela Putorius Furo* T): A Review on the Housing and Management of Pet Ferrets. *Appl. Anim. Behav. Sci.* **2012**, *139*, 155–168. [[CrossRef](#)]
101. Huang, A.C.; Nelson, C.; Elliott, J.E.; Guertin, D.A.; Ritland, C.; Drouillard, K.; Cheng, K.M.; Schwantje, H.M. River Otters (*Lontra Canadensis*) “Trapped” in a Coastal Environment Contaminated with Persistent Organic Pollutants: Demographic and Physiological Consequences. *Environ. Pollut.* **2018**, *238*, 306–316. [[CrossRef](#)]
102. Krawczyk, A.J.; Bogdziewicz, M.; Czyz, M.J. Diet of the American Mink *Neovison Vison* in an Agricultural Landscape in Western Poland. *Folia Zool.* **2013**, *62*, 304–310. [[CrossRef](#)]
103. Magnusdottir, R.; von Schmalensee, M.; Stefansson, R.A.; Macdonald, D.W.; Hersteinsson, P. A Foe in Woe: American Mink (*Neovison Vison*) Diet Changes during a Population Decrease. *Mamm. Biol.* **2014** *79* **2013**, *79*, 58–63. [[CrossRef](#)]
104. Robinson, S.; Barbieri, M.; Johanos, T. The Hawaiian Monk Seal: Ethology Applied to Endangered Species Conservation and Recovery. In *Ethology and Behavioral Ecology of Phocids*; Springer: Cham, Switzerland, 2022; pp. 599–635. [[CrossRef](#)]
105. Sheffield, G.; Grebmeier, J.M. Pacific Walrus (*Odobenus Rosmarus Divergens*): Differential Prey Digestion and Diet. *Mar. Mammal Sci.* **2009**, *25*, 761–777. [[CrossRef](#)]

106. Clark, C.T.; Horstmann, L.; de Vernal, A.; Jensen, A.M.; Misarti, N. Pacific Walrus Diet across 4000 Years of Changing Sea Ice Conditions. *Quat. Res.* **2022**, *108*, 26–42. [CrossRef]
107. Beukes, M.; Radloff, F.G.T.; Ferreira, S.M. Estimating Lion’s Prey Species Profile in an Arid Environment. *J. Zool.* **2017**, *303*, 136–144. [CrossRef]
108. Bothma, J.d.P.; Walker, C. The African Lion. In *Larger Carnivores of the African Savannas*; Springer: Berlin/Heidelberg, Germany, 1999; pp. 22–59. [CrossRef]
109. Kshettry, A.; Vaidyanathan, S.; Athreya, V. Diet Selection of Leopards (*Panthera Pardus*) in a Human-Use Landscape in North-Eastern India. *Trop. Conserv. Sci.* **2018**, *11*, 1–8. [CrossRef]
110. Andheria, A.P.; Karanth, K.U.; Kumar, N.S. Diet and Prey Profiles of Three Sympatric Large Carnivores in Bandipur Tiger Reserve, India. *J. Zool.* **2007**, *273*, 169–175. [CrossRef]
111. Hayward, M.W.; Jedrzejewski, W.; Jedrzejewska, B. Prey Preferences of the Tiger *P. Anthera Tigris*. *J. Zool.* **2012**, *286*, 221–231. [CrossRef]
112. Biswas, S.; Sankar, K. Prey Abundance and Food Habit of Tigers (*Panthera Tigris Tigris*) in Pench National Park, Madhya Pradesh, India. *J. Zool.* **2002**, *256*, 411–420. [CrossRef]
113. Toth, J.; Evert, S.; Zimmermann, E.; Sullivan, M.; Dotts, L.; Able, K.W.; Hagan, R.; Slocum, C. Annual Residency Patterns and Diet of *Phoca Vitulina Concolor* (Western Atlantic Harbor Seal) in a Southern New Jersey Estuary. *Northeast. Nat.* **2018**, *25*, 611–626. [CrossRef]
114. Ganguly, D.; Adhya, T. How Fishing Cats *Prionailurus Viverrinus Bennett*, 1833 Fish: Describing a Felid’s Strategy to Hunt Aquatic Prey. *Mammalia* **2022**, *86*, 182–189. [CrossRef]
115. Grassman, L.I.; Tewes, M.E.; Silvy, N.J.; Kreetiyutanont, K. Spatial Organization and Diet of the Leopard Cat (*Prionailurus Bengalensis*) in North-Central Thailand. *J. Zool.* **2005**, *266*, 45–54. [CrossRef]
116. Rau, J.R.; Jiménez, J.E. Diet of Puma (*Puma Concolor*, *Carnivora: Felidae*) in Coastal and Andean Ranges of Southern Chile. *Stud. Neotrop. Fauna Environ.* **2002**, *37*, 201–205. [CrossRef]
117. Aranda, M.; Sánchez-Cordero, V. Prey Spectra of Jaguar (*Panthera Onca*) and Puma (*Puma Concolor*) in Tropical Forests of Mexico. *Stud. Neotrop. Fauna Environ.* **1996**, *31*, 65–67. [CrossRef]
118. Tófoli, C.F.; Rohe, F.; Setz, E.Z.F. Jaguarundi (*Puma Yagouaroundi*) (Geoffroy, 1803) (*Carnivora, Felidae*) Food Habits in a Mosaic of Atlantic Rainforest and Eucalypt Plantations of Southeastern Brazil. *Braz. J. Biol.* **2009**, *69*, 871–877. [CrossRef]
119. De Cassia Bianchi, R.; Rosa, A.F.; Gatti, A.; Mendes, S.L. Diet of Margay, *Leopardus Wiedii*, and Jaguarundi, *Puma Yagouaroundi*, (*Carnivora: Felidae*) in Atlantic Rainforest, Brazil. *Zoologia* **2011**, *28*, 127–132. [CrossRef]
120. Merkle, J.A.; Polfus, J.L.; Derbridge, J.J.; Heinemeyer, K.S. Dietary Niche Partitioning among Black Bears, Grizzly Bears, and Wolves in a Multiprey Ecosystem. *Can. J. Zool.* **2017**, *95*, 663–671. [CrossRef]
121. Kirby, R.; Alldredge, M.W.; Pauli, J.N. The Diet of Black Bears Tracks the Human Footprint across a Rapidly Developing Landscape. *Biol. Conserv.* **2016**, *200*, 51–59. [CrossRef]
122. Hatch, K.A.; Kester, K.A.; Auger, J.; Roeder, B.L.; Bunnell, K.; Black, H.L. The Effect of Sex, Age, and Location on Carnivory in Utah Black Bears (*Ursus Americanus*). *Oecologia* **2019**, *189*, 931–937. [CrossRef]
123. Matsubayashi, J.; Otsubo, K.; Morimoto, J.O.; Nakamura, F.; Nose, T.; Tayasu, I. Feeding Habits May Explain the Morphological Uniqueness of Brown Bears on Etorofu Island, Southern Kuril Islands in East Asia. *Biol. J. Linn. Soc.* **2016**, *119*, 99–105. [CrossRef]
124. Mangipane, L.S.; Lafferty, D.J.R.; Joly, K.; Sorum, M.S.; Cameron, M.D.; Belant, J.L.; Hilderbrand, G.V.; Gustine, D.D. Dietary Plasticity and the Importance of Salmon to Brown Bear (*Ursus Arctos*) Body Size and Condition in a Low Arctic Ecosystem. *Polar Biol.* **2020**, *43*, 825–833. [CrossRef]
125. Johnson, A.C.; Hobson, K.A.; Lunn, N.J.; McGeachy, D.; Richardson, E.S.; Derocher, A.E. Temporal and Intra-Population Patterns in Polar Bear Foraging Ecology in Western Hudson Bay. *Mar. Ecol. Prog. Ser.* **2019**, *619*, 187–199. [CrossRef]
126. Elmhausen, B.; Tannerfeldt, M.; Verucci, P.; Angerbjörn, A. The Arctic Fox (*Alopex Lagopus*): An Opportunistic Specialist. *J. Zool.* **2000**, *251*, 139–149. [CrossRef]
127. Food Habits of Arctic Foxes (*Alopex Lagopus*) on the Western Coast of Svalbard on JSTOR. Available online: <https://www.jstor.org/stable/40511362> (accessed on 31 July 2022).
128. Baltrūnaitė, L. Diet and Winter Habitat Use of the Red Fox, Pine Marten and Raccoon Dog in Dzūkija National Park, Lithuania. *Acta Zool. Litu.* **2006**, *16*, 46–53. [CrossRef]
129. Castañeda, I.; Doherty, T.S.; Fleming, P.A.; Stobo-Wilson, A.M.; Woinarski, J.C.Z.; Newsome, T.M. Variation in Red Fox *Vulpes Vulpes* Diet in Five Continents. *Mamm. Rev.* **2022**, *52*, 328–342. [CrossRef]
130. Weise, M.J.; Harvey, J.T. Temporal Variability in Ocean Climate and California Sea Lion Diet and Biomass Consumption: Implications for Fisheries Management. *Mar. Ecol. Prog. Ser.* **2008**, *373*, 157–172. [CrossRef]
131. Bailey, K.M.; Ainley, D.G. The Dynamics of California Sea Lion Predation on Pacific Hake. *Fish. Res.* **1981**, *1*, 163–176. [CrossRef]