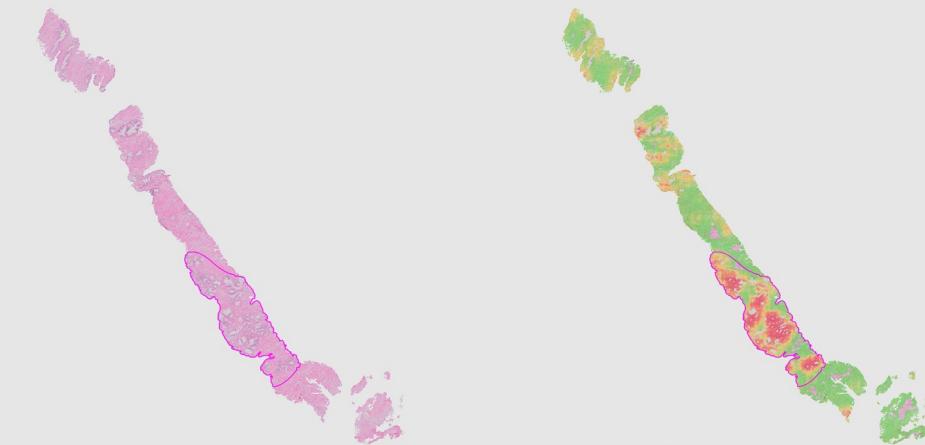


Applications of Machine Learning for Clinical Practice



Geert Litjens
Department of Pathology
Radboud University Medical Center

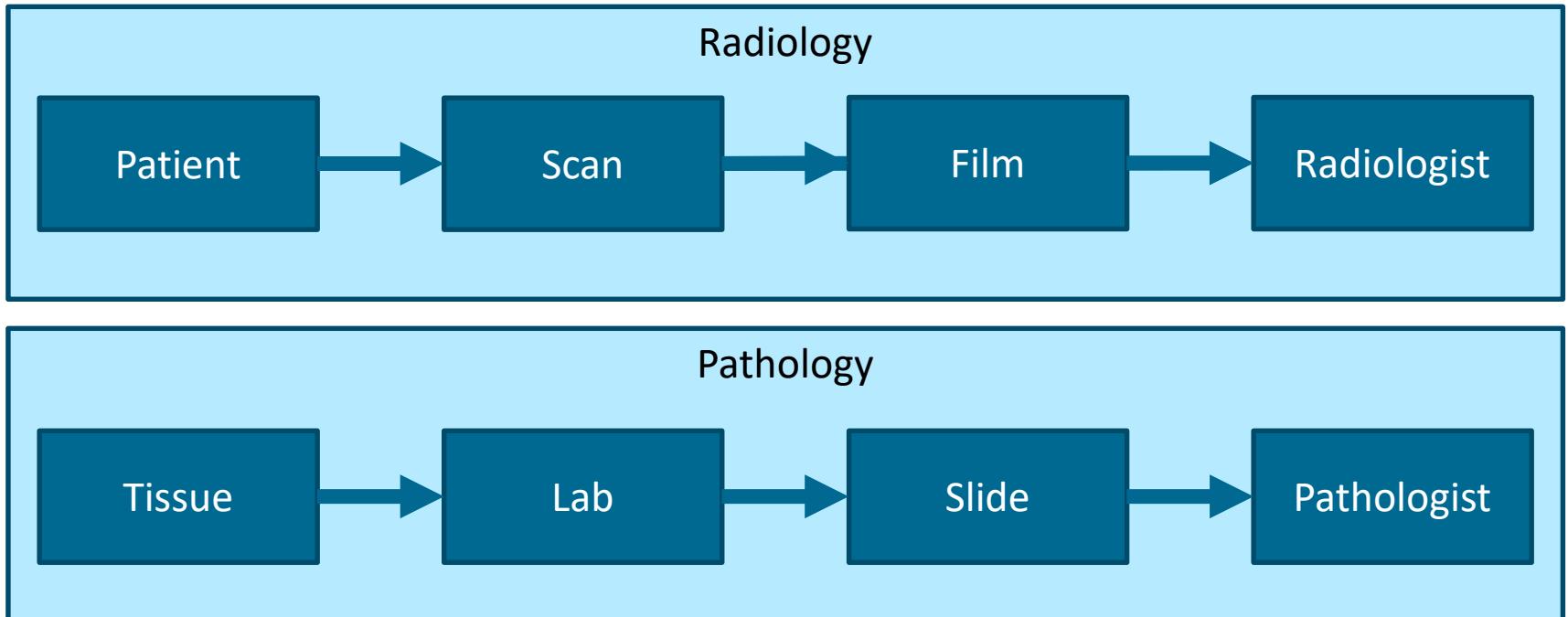
The promise of digital pathology?

5 Key criteria for evaluating Digital Pathology

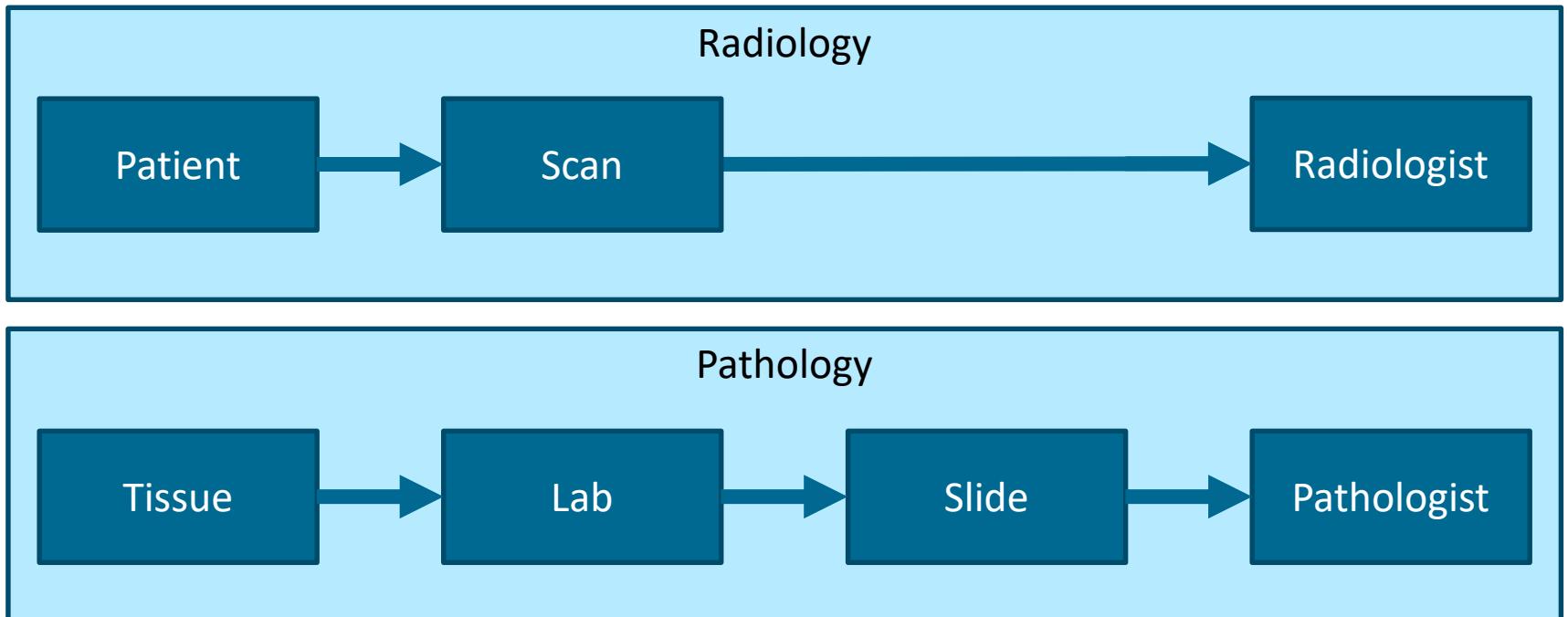
The adoption of digital pathology is evolving and offers functionality that goes far beyond the microscope. These new opportunities significantly increase workflow efficiency. They move time-consuming tasks to the computer and allow the pathologist to spend more time on reviewing cases. Here are five key criteria when evaluating a solution for digital pathology.



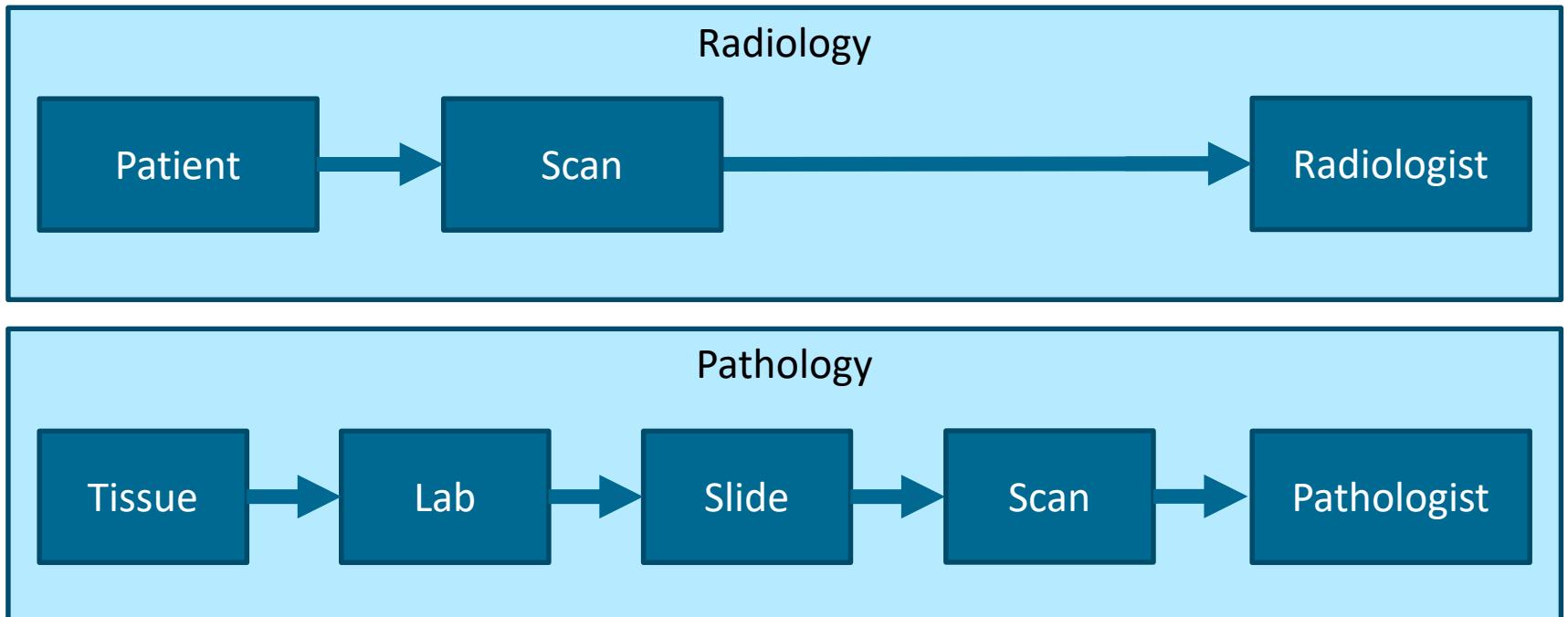
The promise of digital pathology?



The promise of digital pathology?



The promise of digital pathology?



The promise of digital pathology?



Scanners



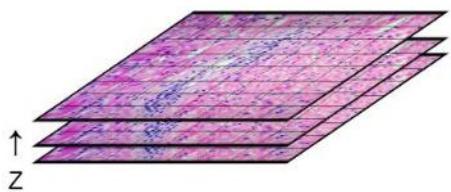
Storage



Computers



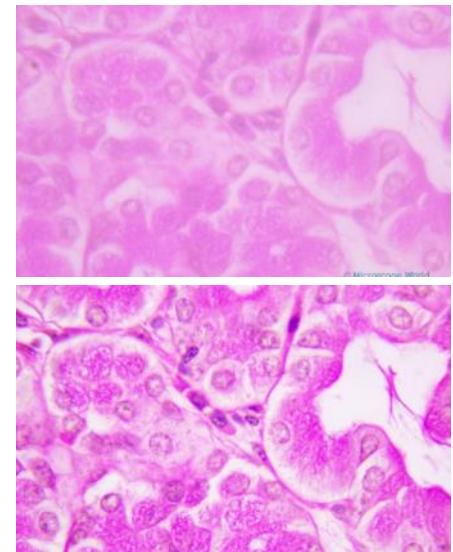
The promise of digital pathology?



Multiple focal points

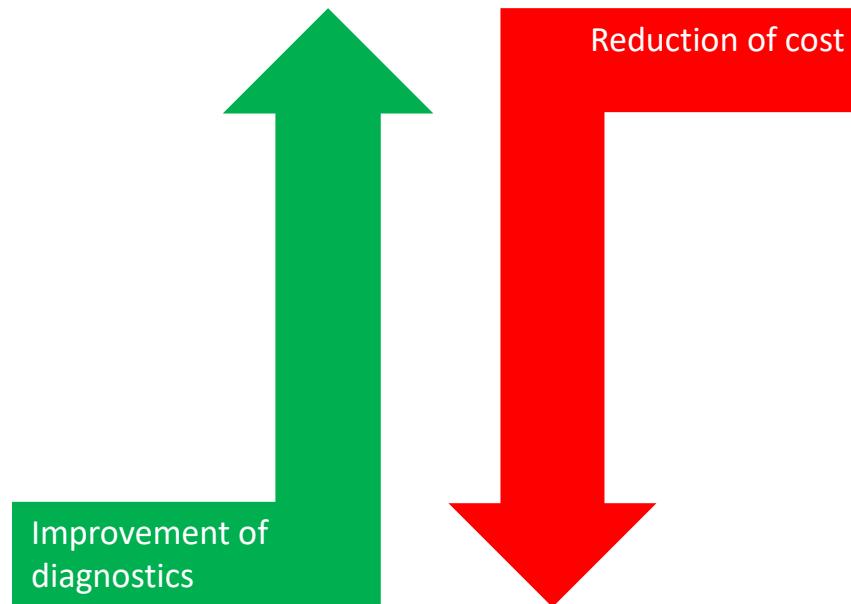


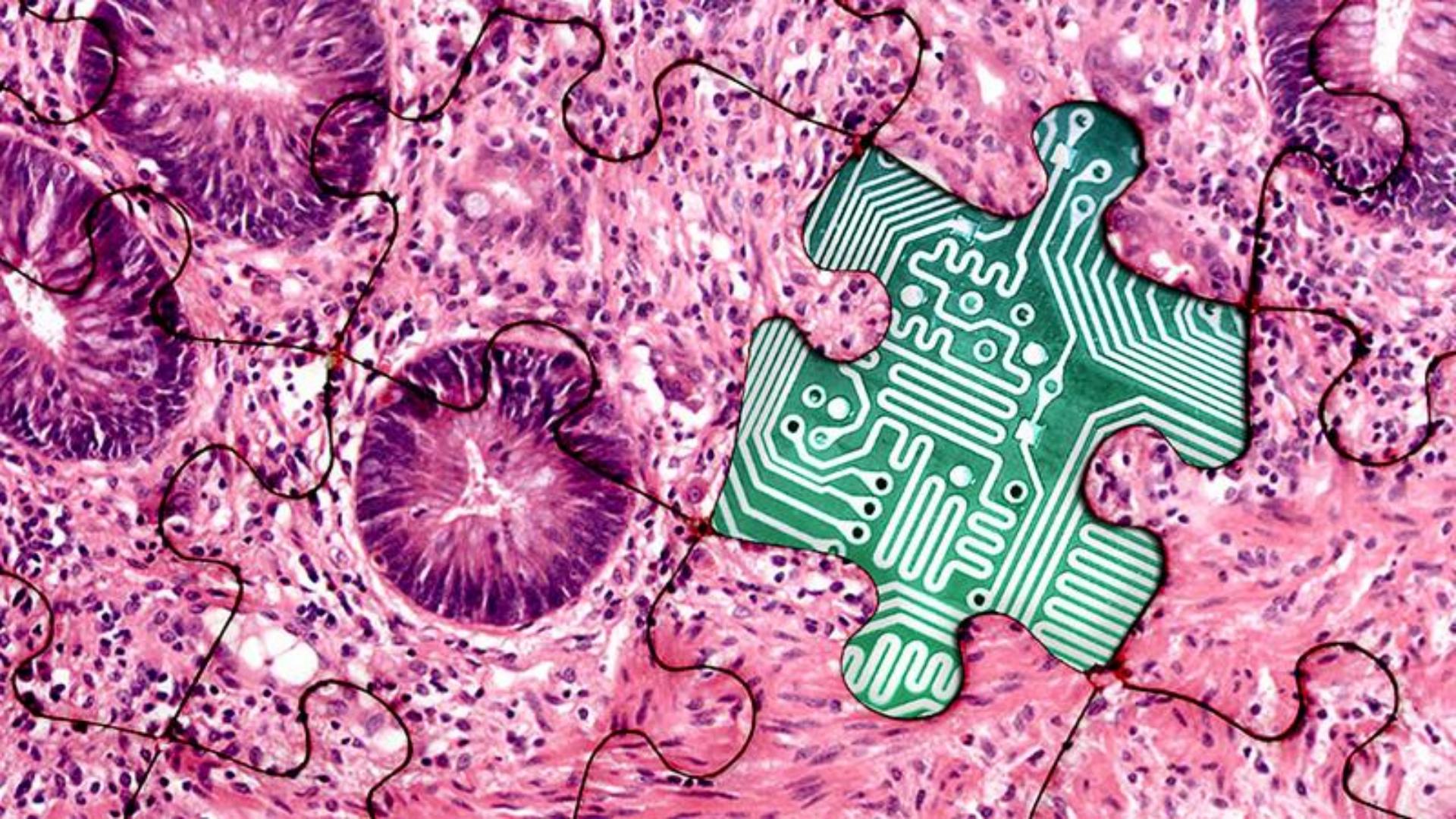
Large slides



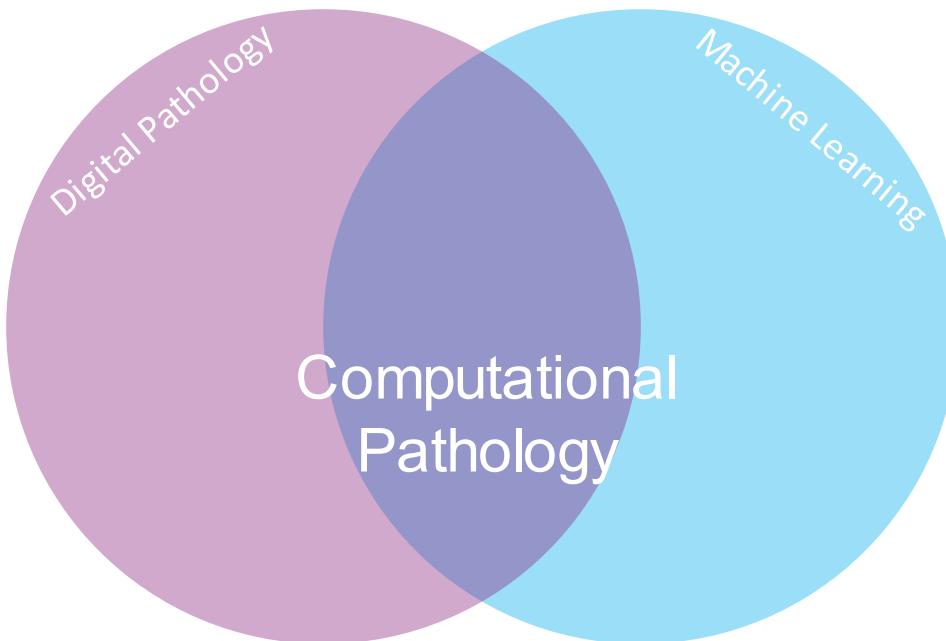
Oil immersion

The promise of digital pathology?

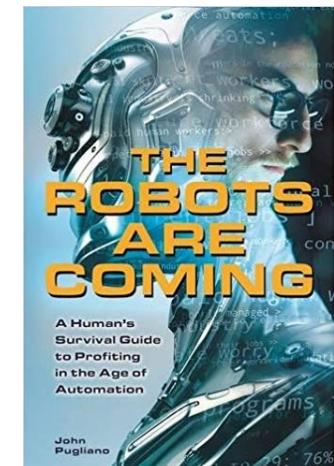
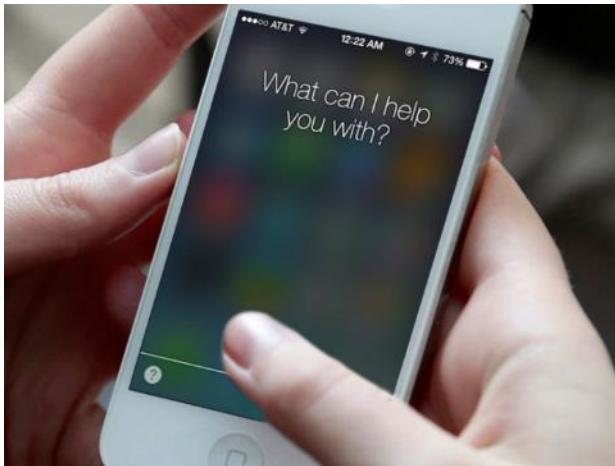




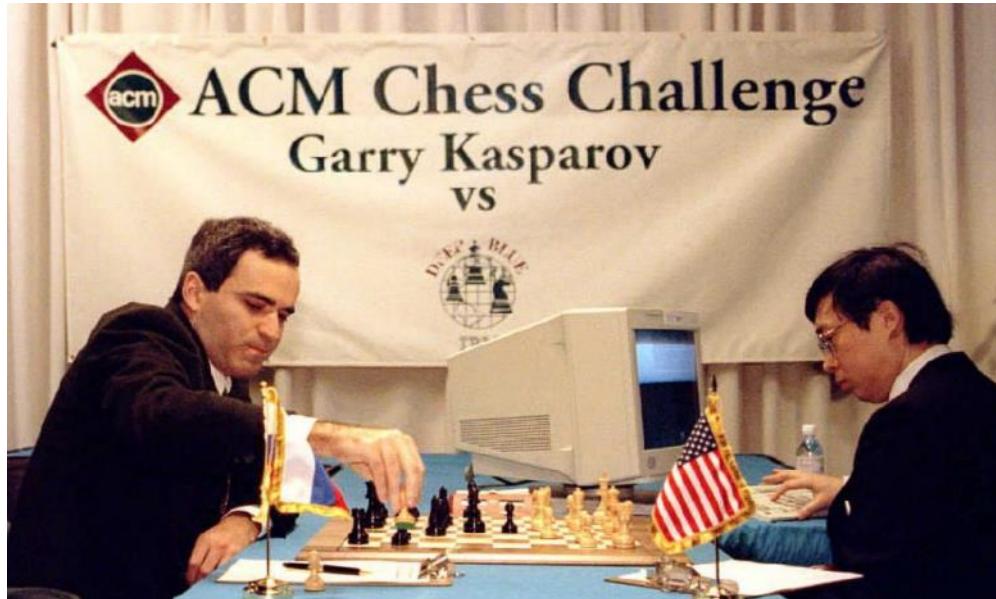
Computational Pathology



Machine learning



ML: a bit of history



nature

THE INTERNATIONAL WEEKLY JOURNAL OF SCIENCE



*At last – a computer program that
can beat a champion Go player* PAGE 484

ALL SYSTEMS GO

CONSERVATION

**SONGBIRDS
À LA CARTE**
*Illegal harvest of millions
of Mediterranean birds*

PAGE 452

RESEARCH ETHICS

**SAFEGUARD
TRANSPARENCY**
*Don't let openness backfire
on individuals*

PAGE 459

POPULAR SCIENCE

**WHEN GENES
GOT 'SELFISH'**
*Dawkins's culling
card forty years on*

PAGE 462

NATURE.COM/NATURE

26 February 2016 450

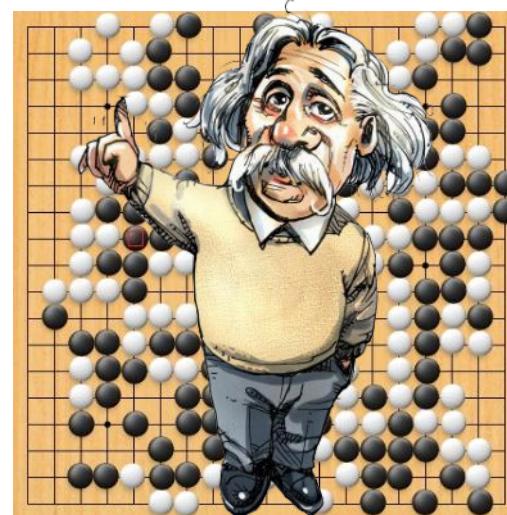
Vol. 529 No. 7607



ML: a bit of history

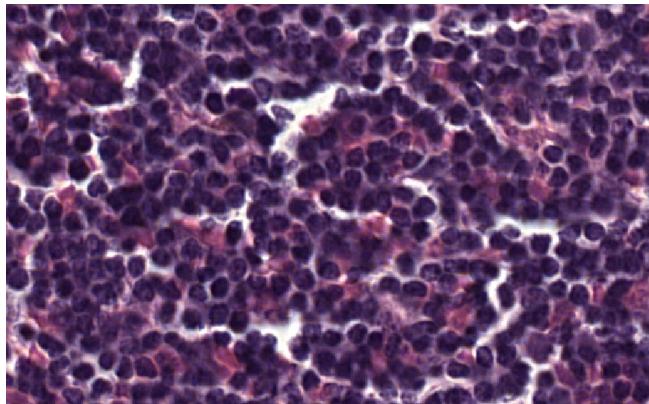


30 possible moves per turn
40 turns per game

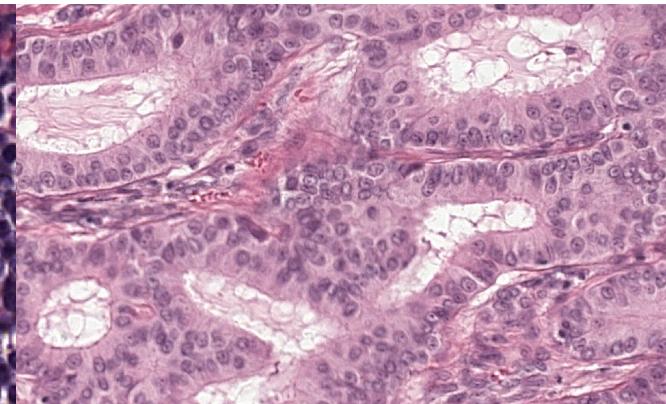


250 possible moves per turn
150 turns per game

How to build an ML system?



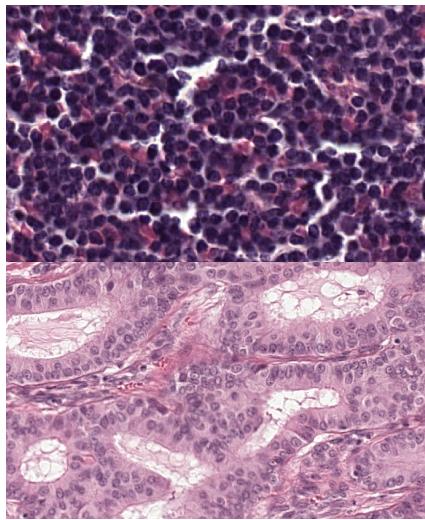
Normal lymph node



Breast cancer metastasis

How to build an ML system?

Examples

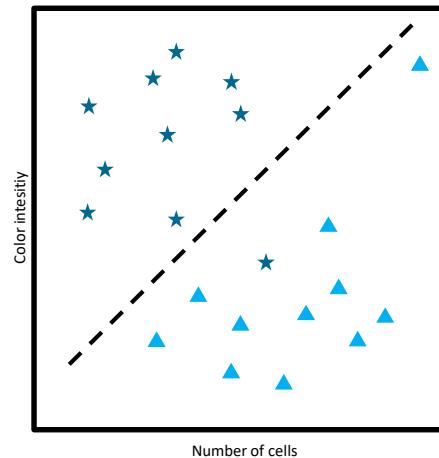


Features

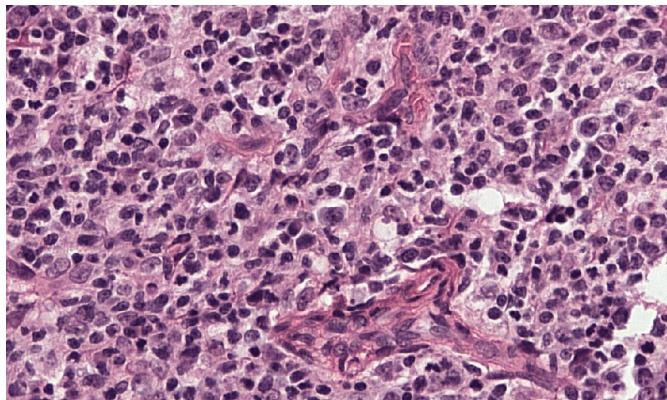
Color intensity

Number of cells

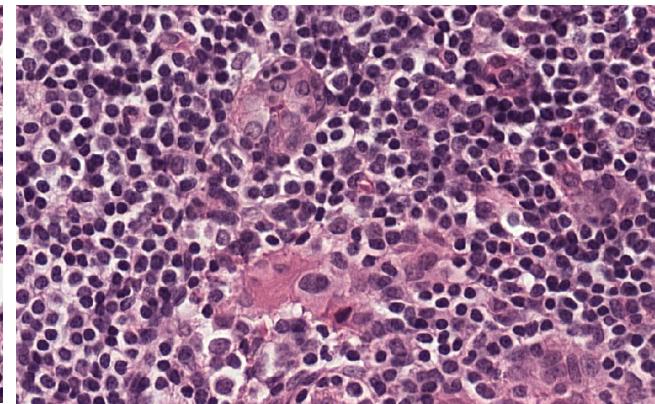
Classification



How to build an ML system?



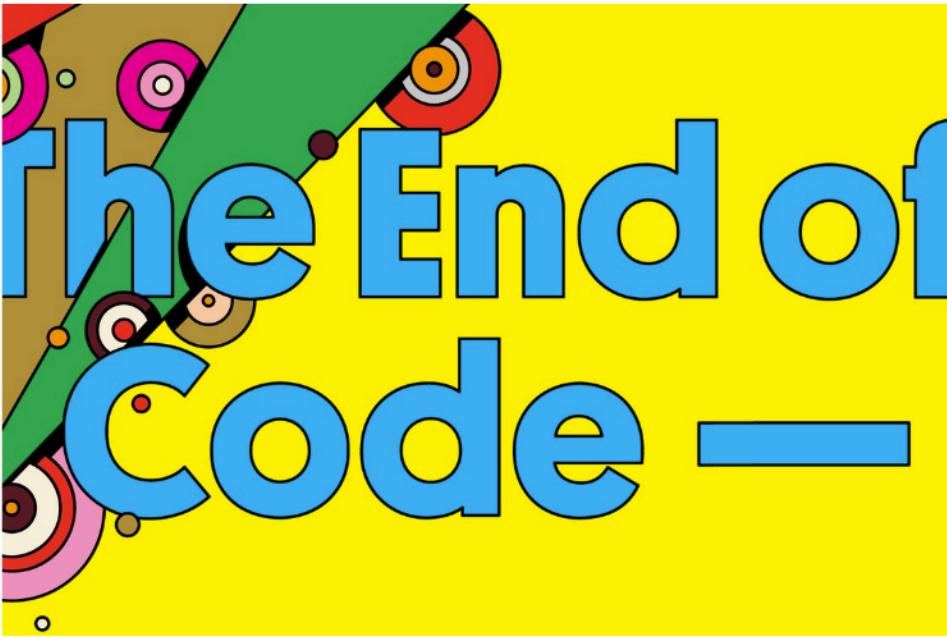
Normal lymph node



Breast cancer metastasis

JASON TANZ IDEAS 05.17.16 6:00 AM

SOON WE WON'T PROGRAM COMPUTERS. WE'LL TRAIN THEM LIKE DOGS



EDWARD C. MONAGHAN

SHARE



SHARE
13,193



TWEET

BEFORE THE INVENTION of the computer, most experimental psychologists thought the brain was an unknowable black box. You could analyze a subject's behavior—*ring bell, dog salivates*—but thoughts, memories, emotions? That stuff was obscure and inscrutable, beyond the reach of science. So these behaviorists, as they called themselves, confined their work to the study of stimulus and response, feedback and reinforcement, bells and saliva. They gave up trying to

MOST POPULAR



BUSINESS
SpaceX's President is Thinking Even Bigger Than Elon Musk
ERIN GRIFFITH

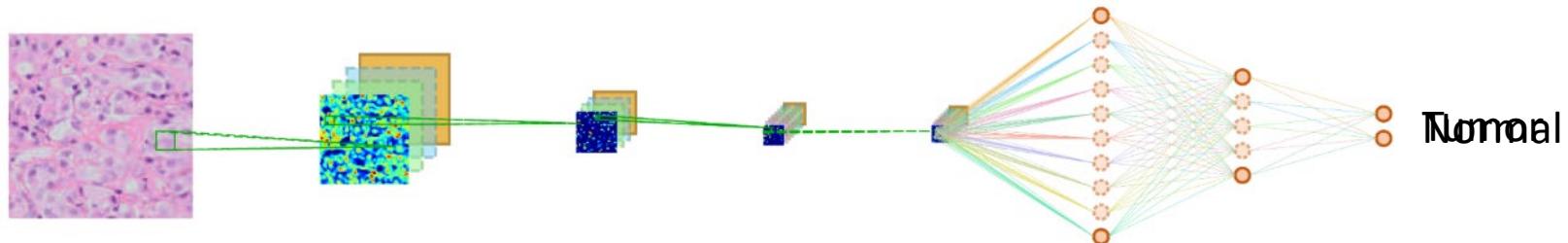
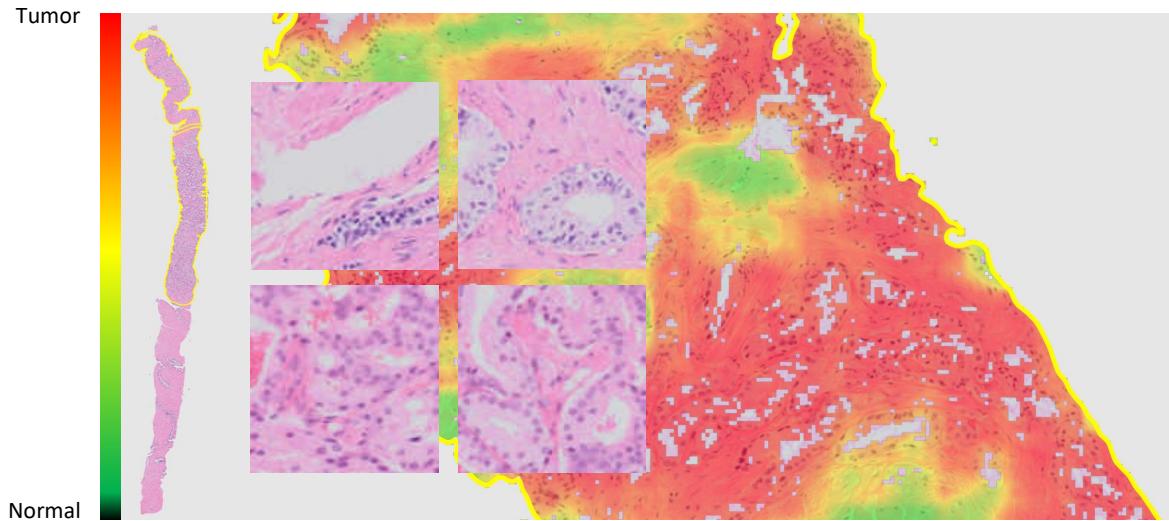
TRANSPORTATION

JASON TANZ IDEAS 05.17.16 06:50 AM

SOON WE WON'T PROGRAM COMPUTERS. WE'LL TRAIN THEM LIKE DOGS

How to build an ML system?





Practical applications of computation pathology

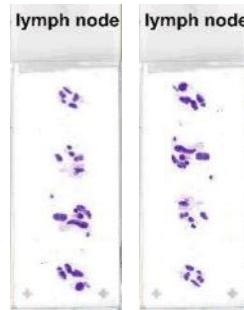
Detection of
metastases in lymph
nodes

Automatic mitotic
counts

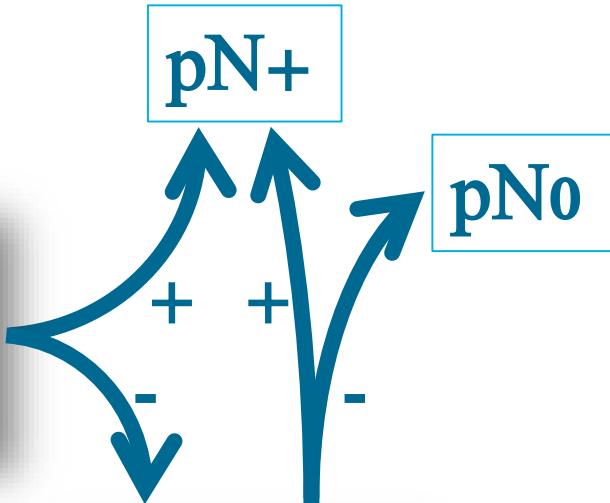
Tumor
qua

Detection of metastases in lymph nodes

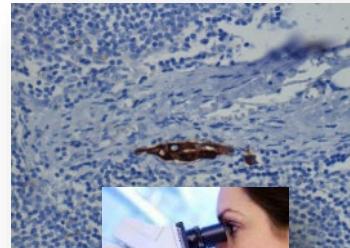




H&E



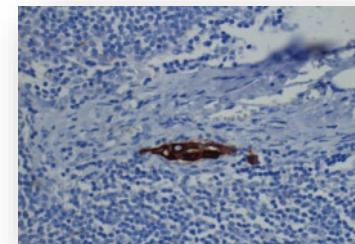
IHC



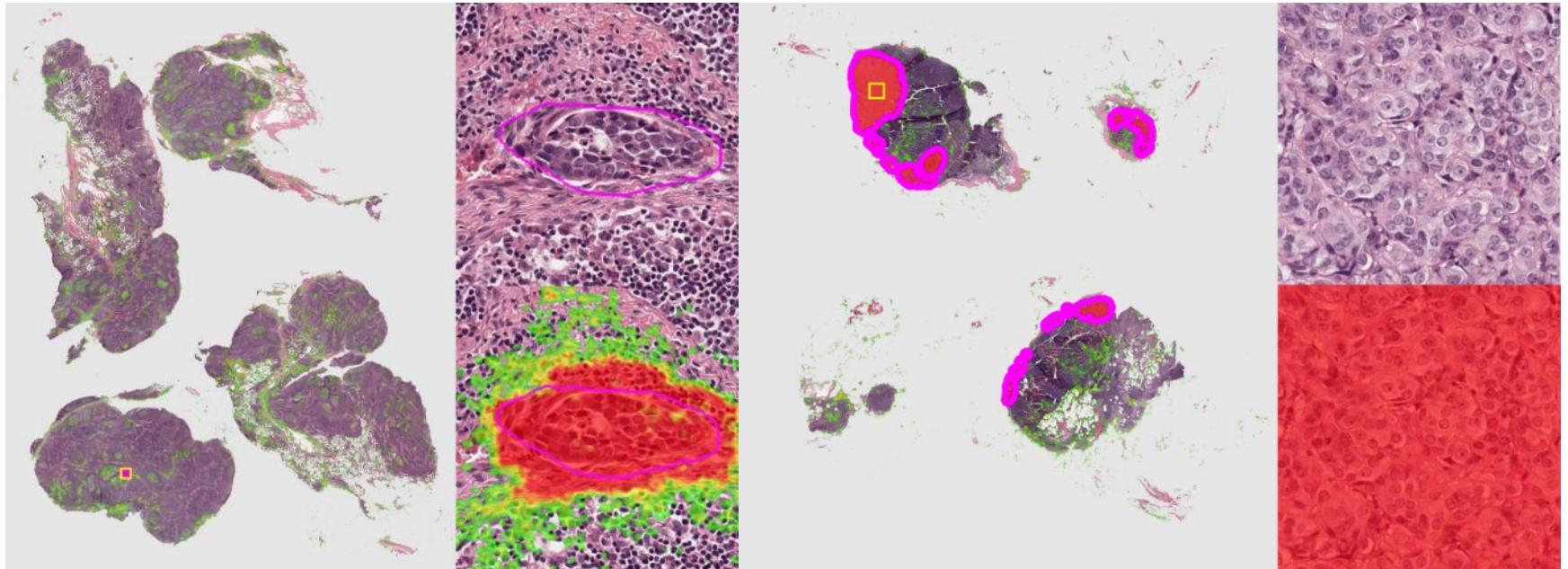


H&E

IHC



Detection of metastases in lymph nodes



Breast cancer metastasis detection



Data

Centrum	Number of slides
CWZ (Nijmegen)	200
LabPON (Hengelo)	200
Rijnstate (Arnhem)	200
Radboudumc (Nijmegen)	439
UMCU (Utrecht)	350
Total	1399



CAMELYON16



CAMELYON17

Why challenges?

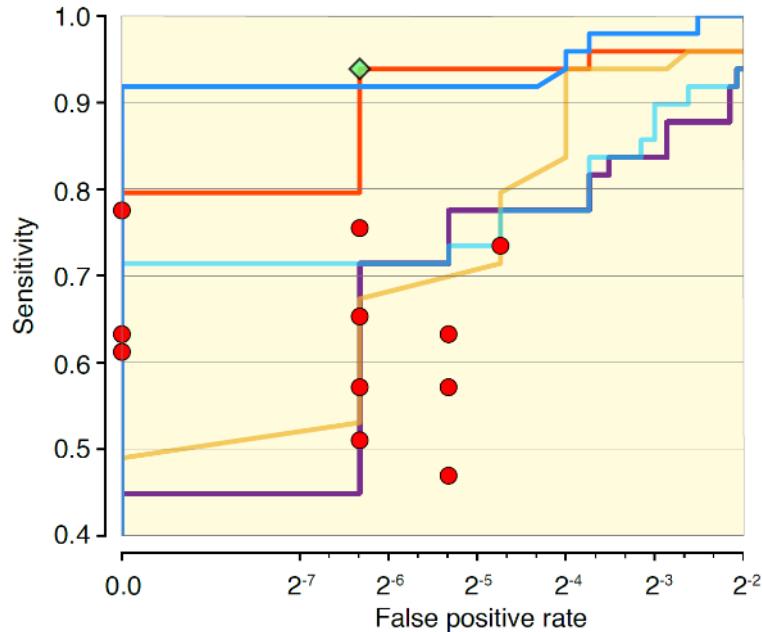
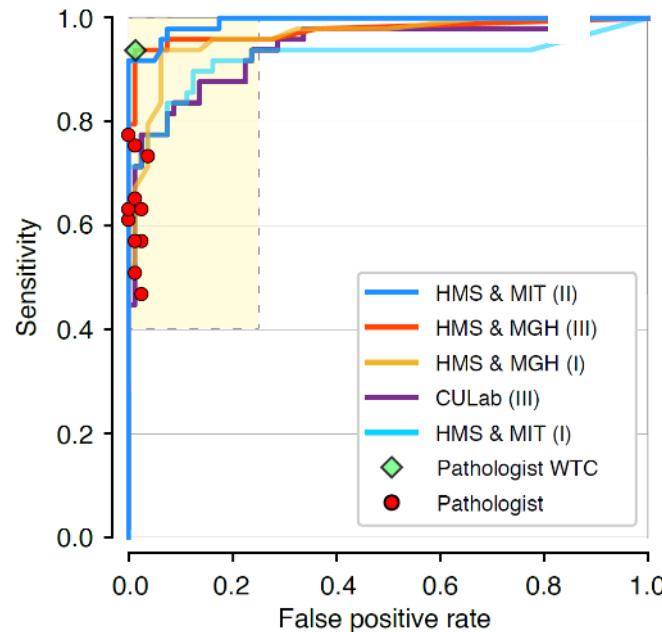
Great way to collect and compare solutions for a problem

Fair comparison of algorithms

- Same evaluation metric
- Same ground truth definition
- Same training and test datasets

Rank	Team	AUC	Description
01	Harvard Medical School (BIDMC) and Massachusetts Institute of Technology (CSAIL), USA	0.9250	  
02	ExB Research and Development co., Germany	0.9173	  
03	Independent participant, Germany	0.8680	  
04	Health Sciences Middle East Technical University, Turkey	0.8669	  
05	NLP LOGIX co., USA	0.8332	  
06	University of Toronto, Electrical and Computer Engineering, Canada	0.8181	  
07	The Warwick-QU Team, United Kingdom	0.7999	  
08	Radboud University Medical Center, Diagnostic Image Analysis Group, Netherlands	0.7828	  
09	HTW-BERLIN, Germany	0.7717	 
10	University of Toronto, Electrical and Computer Engineering, Canada	0.7666	  

Comparing to pathologists



Pathologist

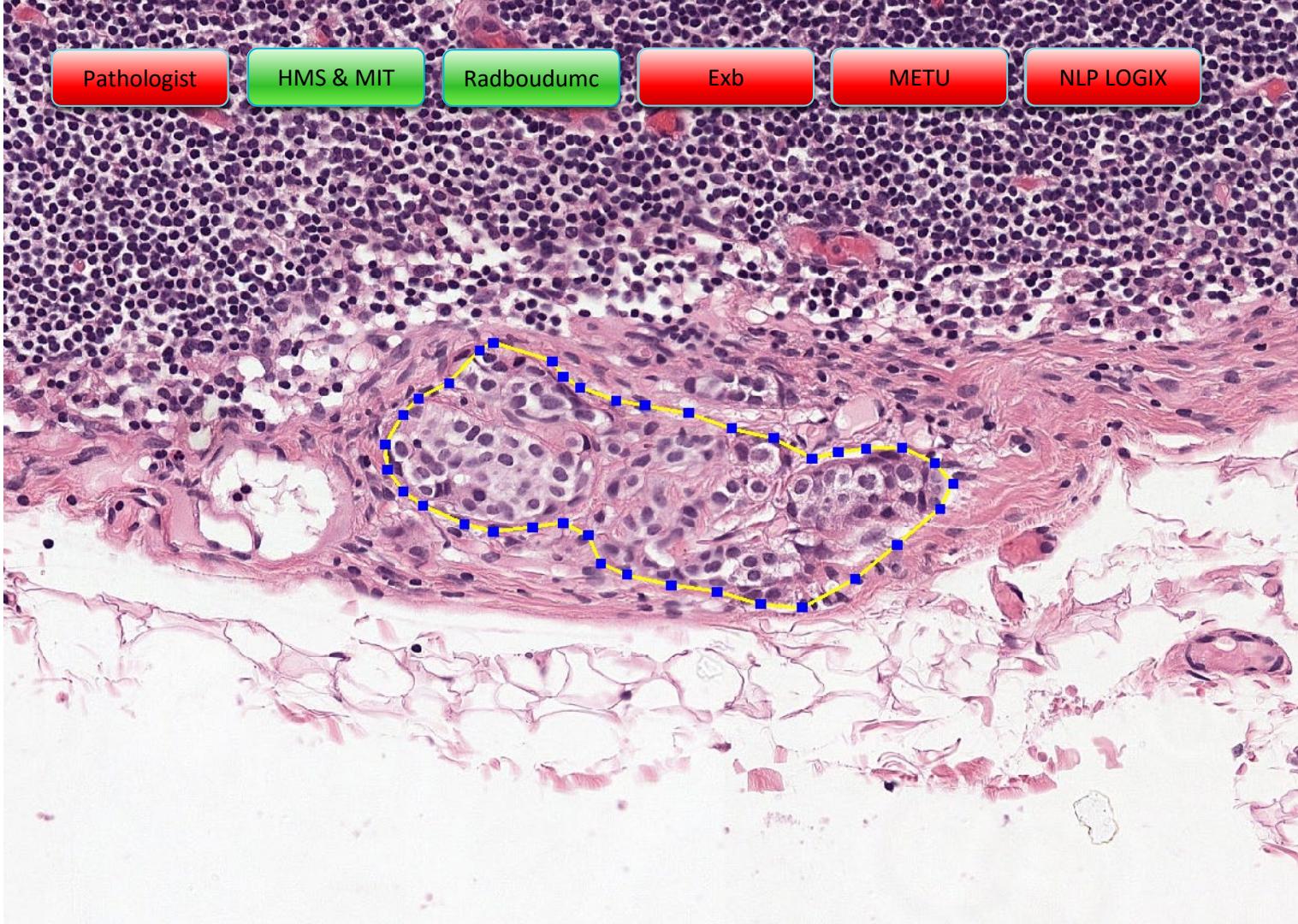
HMS & MIT

Radboudumc

Exb

METU

NLP LOGIX



Pathologist

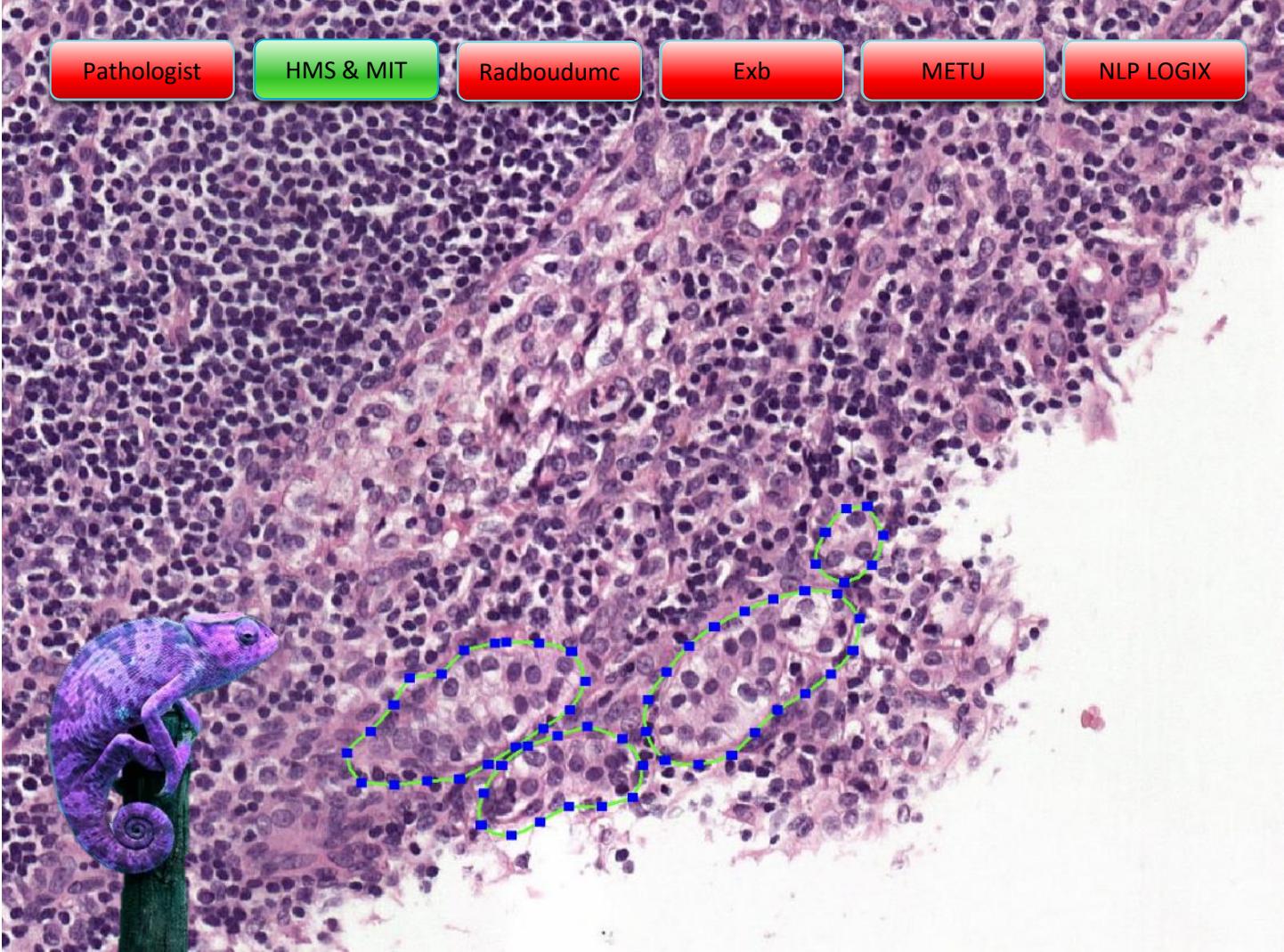
HMS & MIT

Radboudumc

Exb

METU

NLP LOGIX



Pathologist

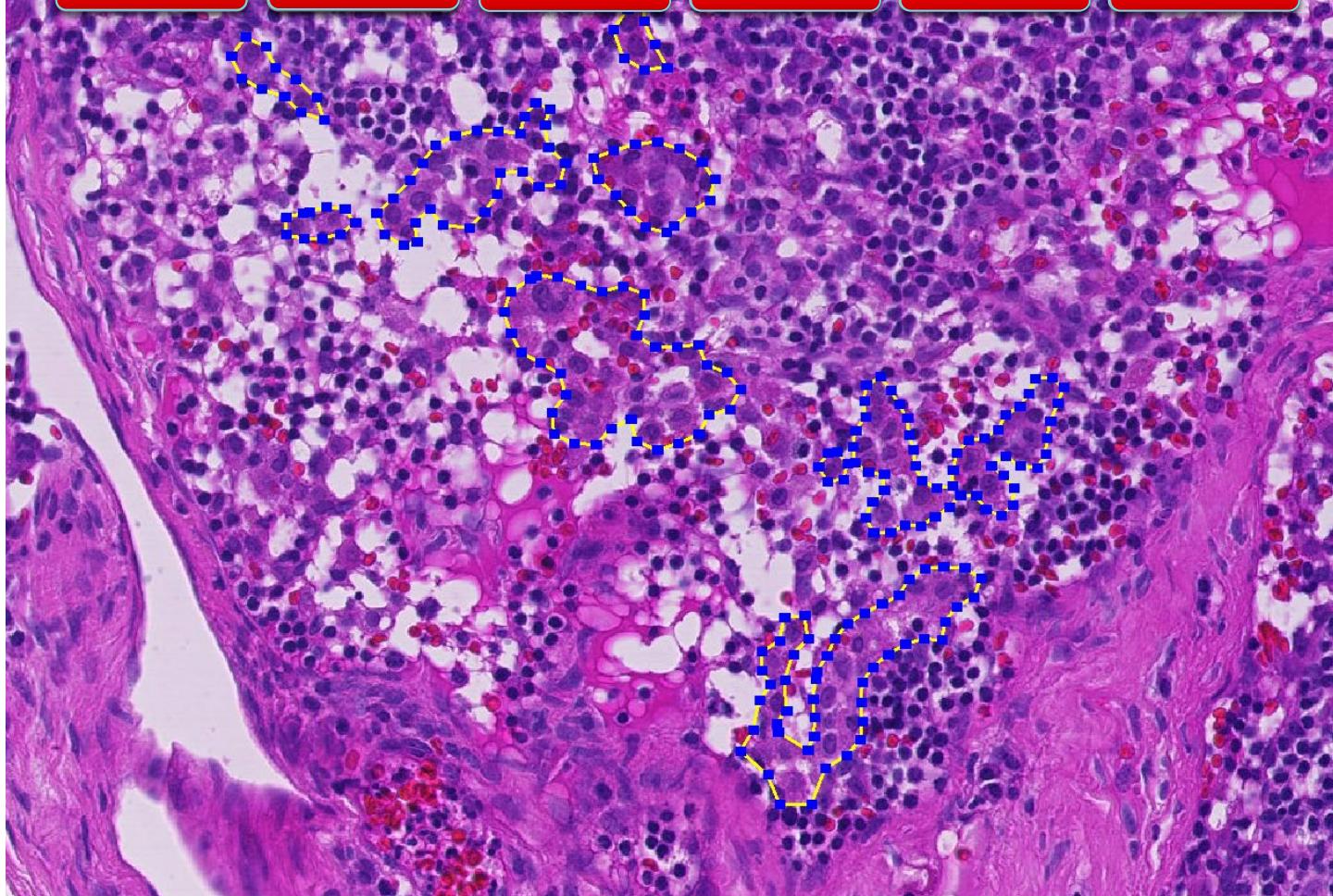
HMS & MIT

Radboudumc

Exb

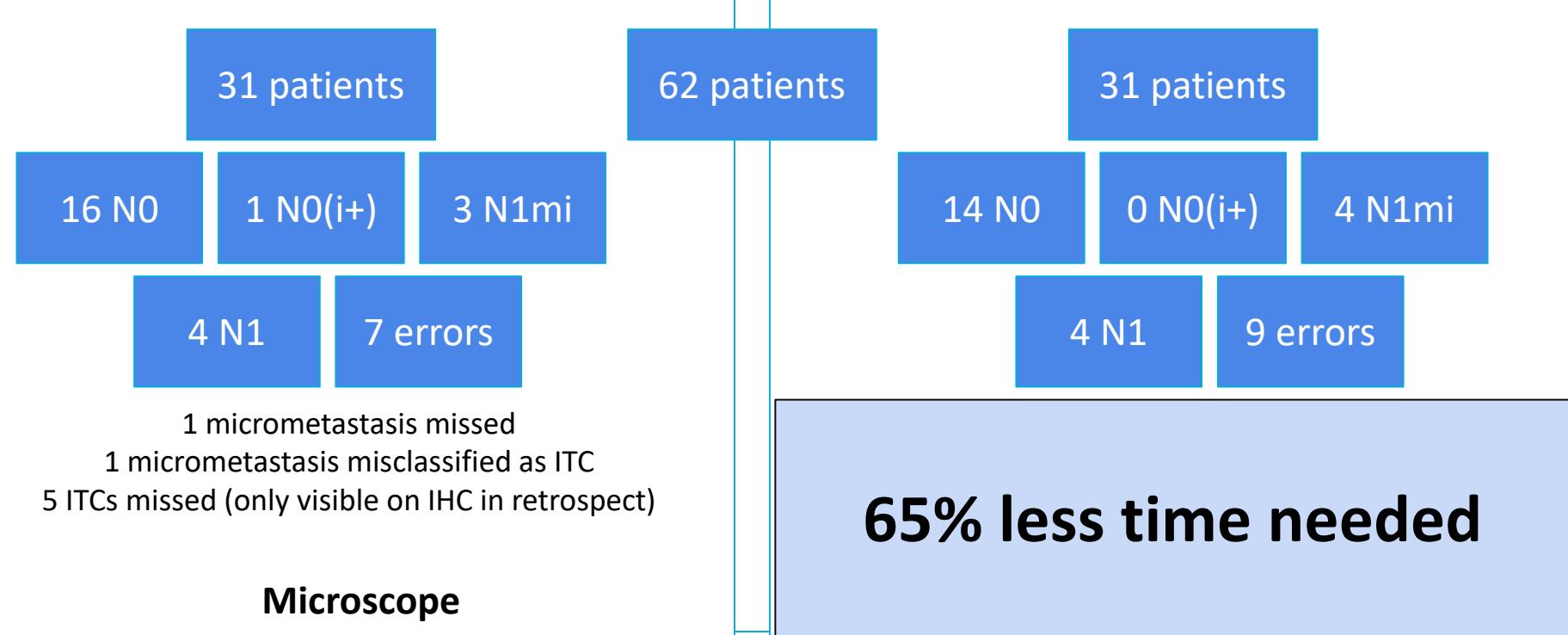
METU

NLP LOGIX





Implemented in clinical practice



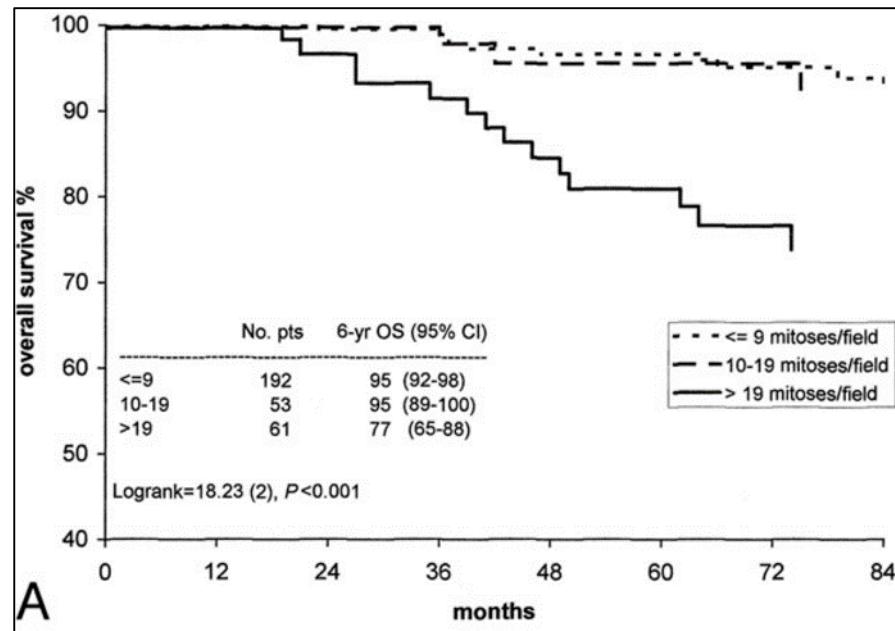
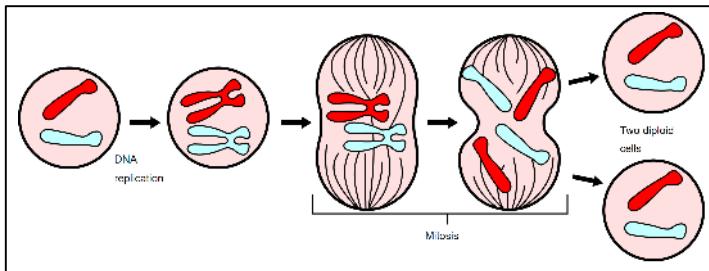
Practical applications of computation pathology

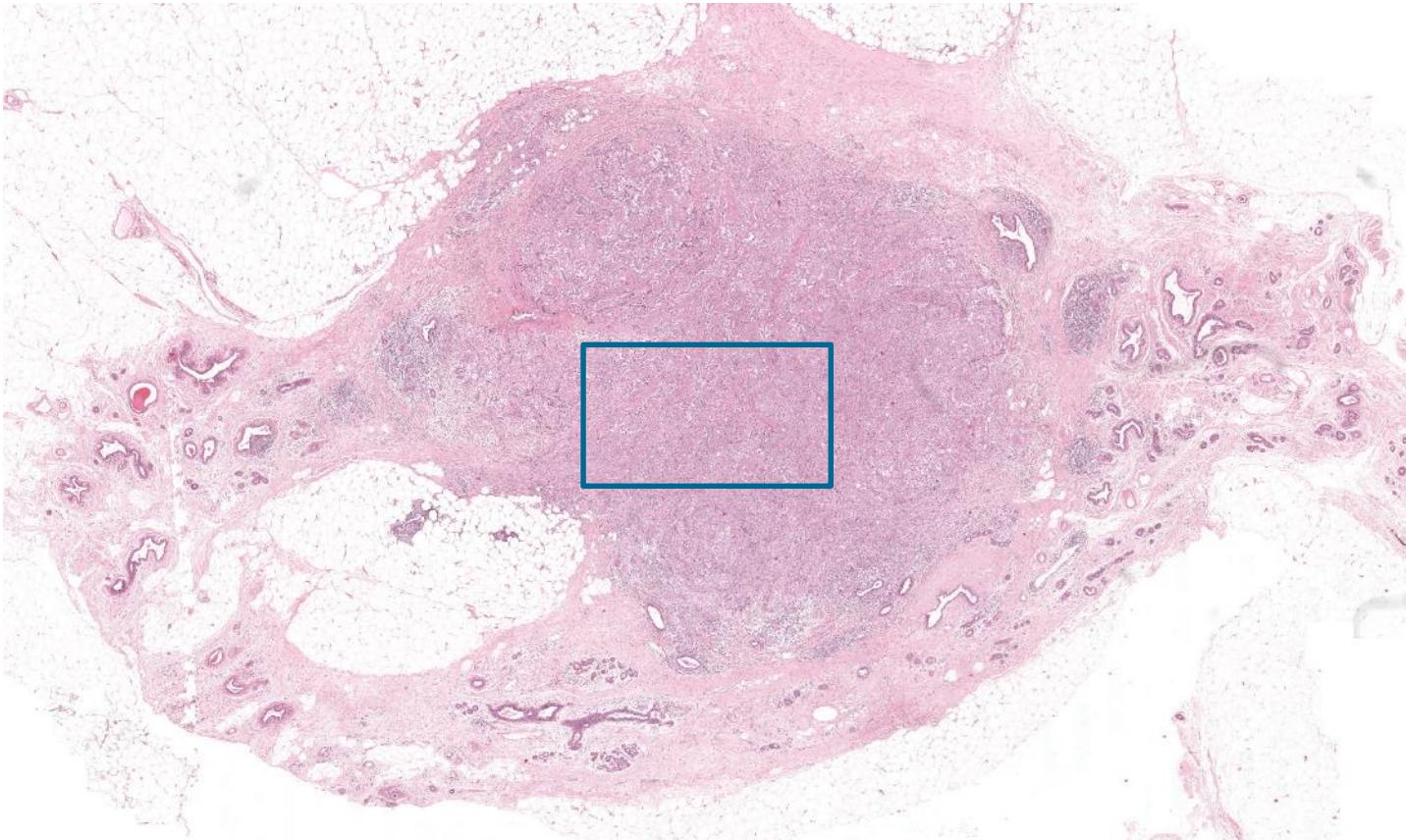
Detection of
metastases in lymph
nodes

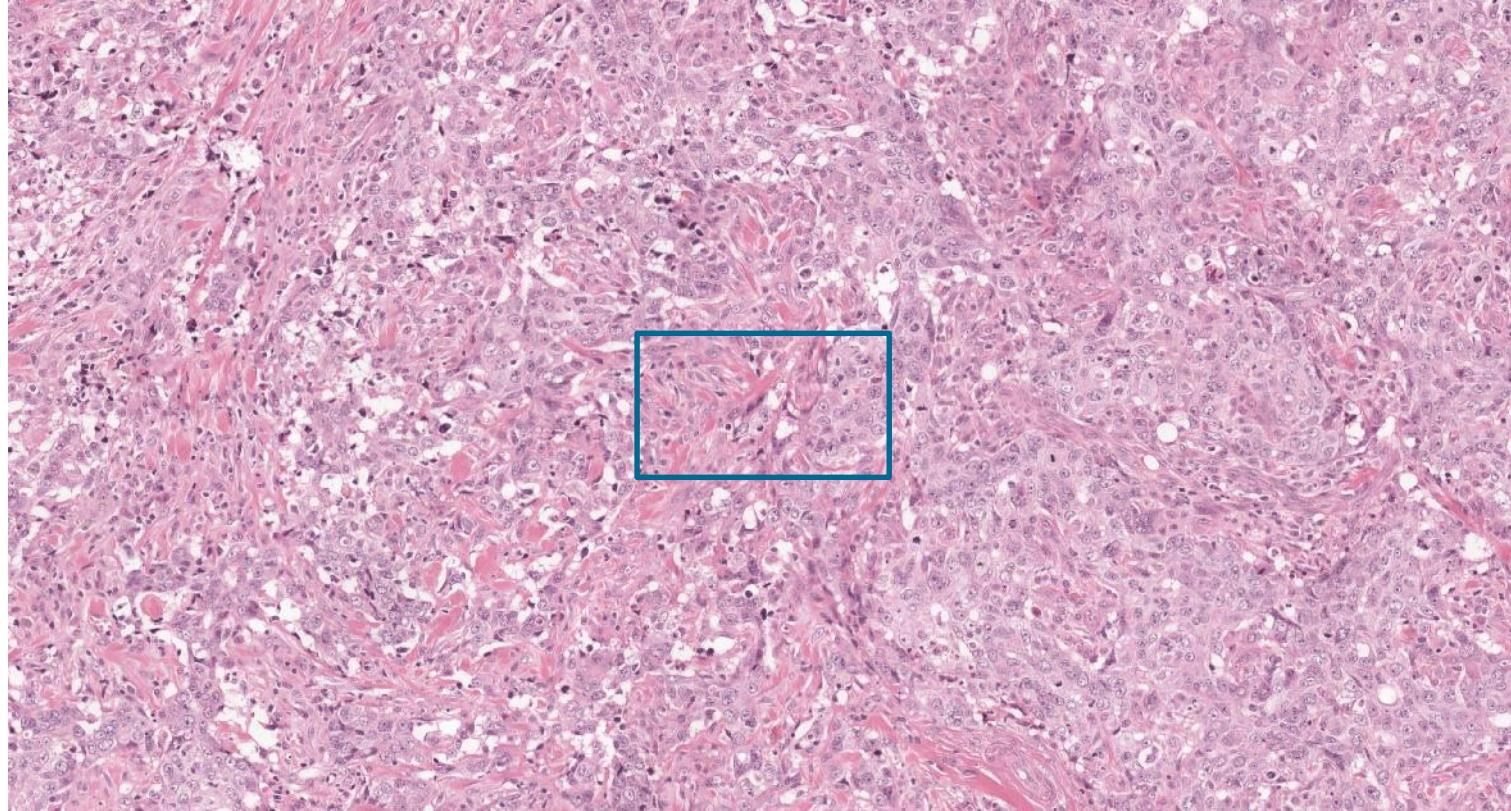
Automatic mitotic
counts

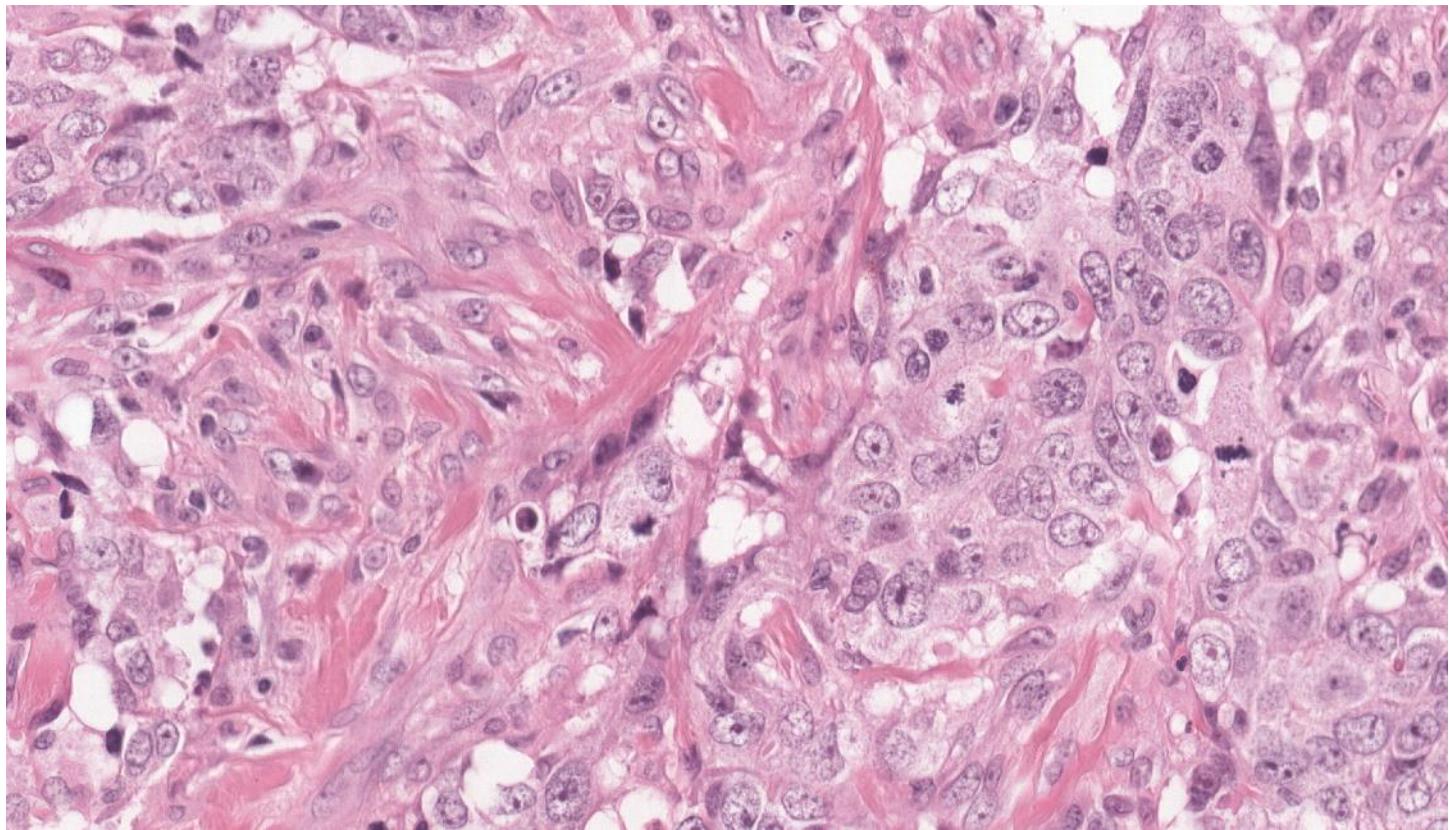
Tumor
qua

Automatic mitotic counts









Automatic mitotic counts



Assessment of Mitosis Detection Algorithms 2013

AMIDAt3 | MICCAI Grand Challenge

D. C. Cireşan, A. Giusti, L. M. Gambardella, and J. Schmidhuber, “Mitosis detection in breast cancer histology images with deep neural networks,” in *International Conference on Medical Image Computing and Computer-assisted Intervention*. Springer, 2013, pp. 411–418.

M. Veta, P. J. van Diest, M. Jiwa, S. Al-Janabi, and J. P. Pluim, “Mitosis counting in breast cancer: Object-level interobserver agreement and comparison to an automatic method,” *PloS one*, vol. 11, no. 8, p. e0161286, 2016.



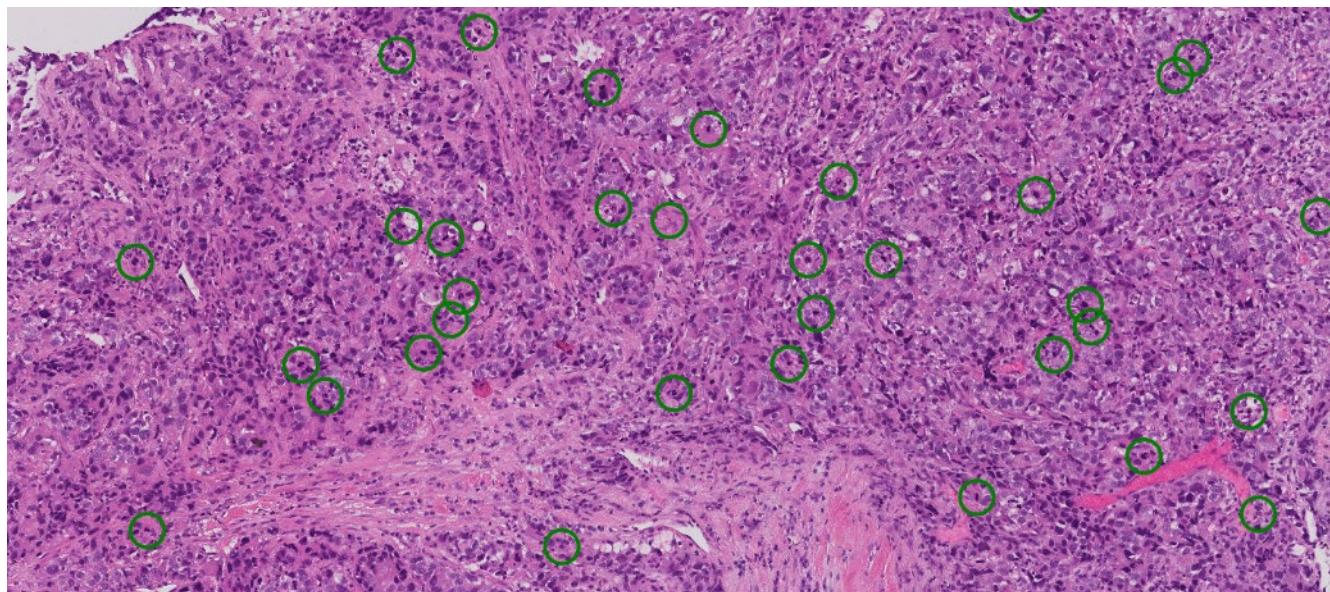
Tumor Proliferation Assessment Challenge 2016

TUPAC16 | MICCAI Grand Challenge

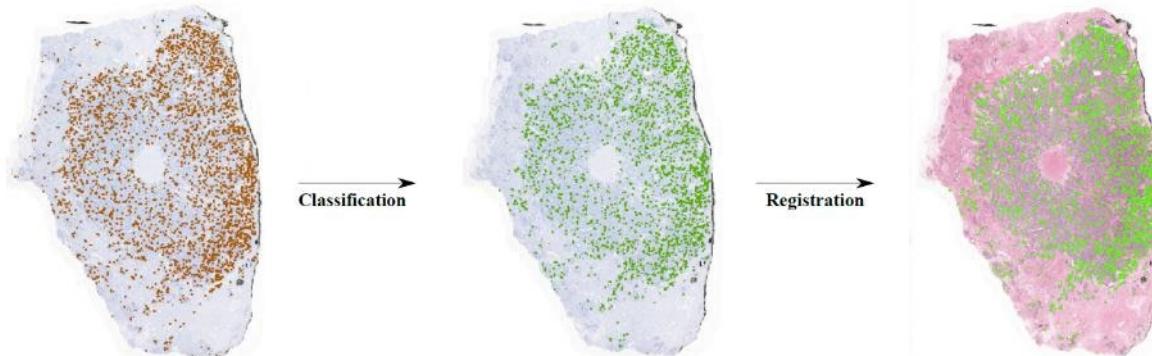
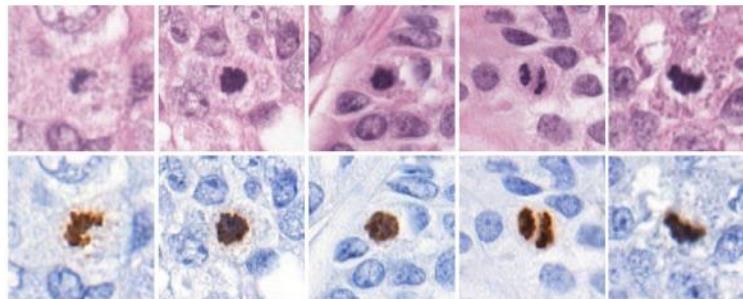
E. Zerhouni, D. Lányi, M. Viana, and M. Gabrani, “Wide residual networks for mitosis detection,” in *Biomedical Imaging (ISBI 2017), 2017 IEEE 14th International Symposium on*. IEEE, 2017, pp. 924–928.

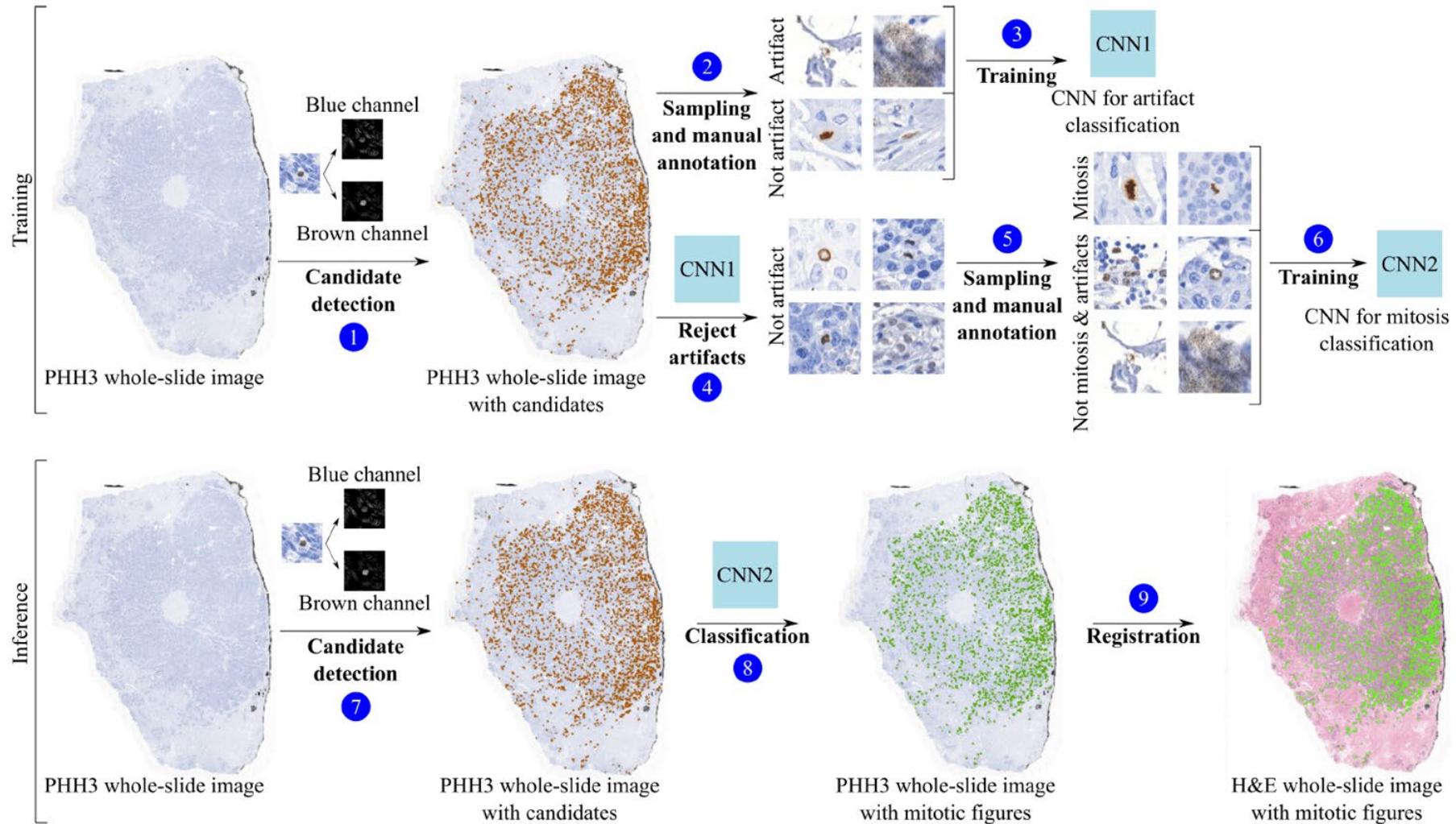
K. Paeng, S. Hwang, S. Park, M. Kim, and S. Kim, “A unified framework for tumor proliferation score prediction in breast histopathology,” *arXiv preprint arXiv:1612.07180*, 2016.

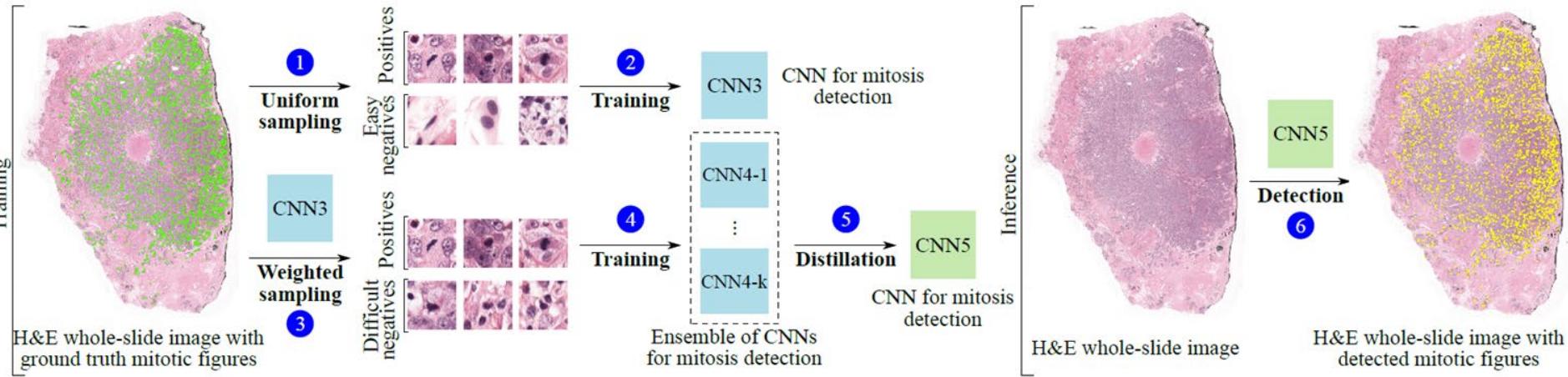
Challenge 1: Reference standard



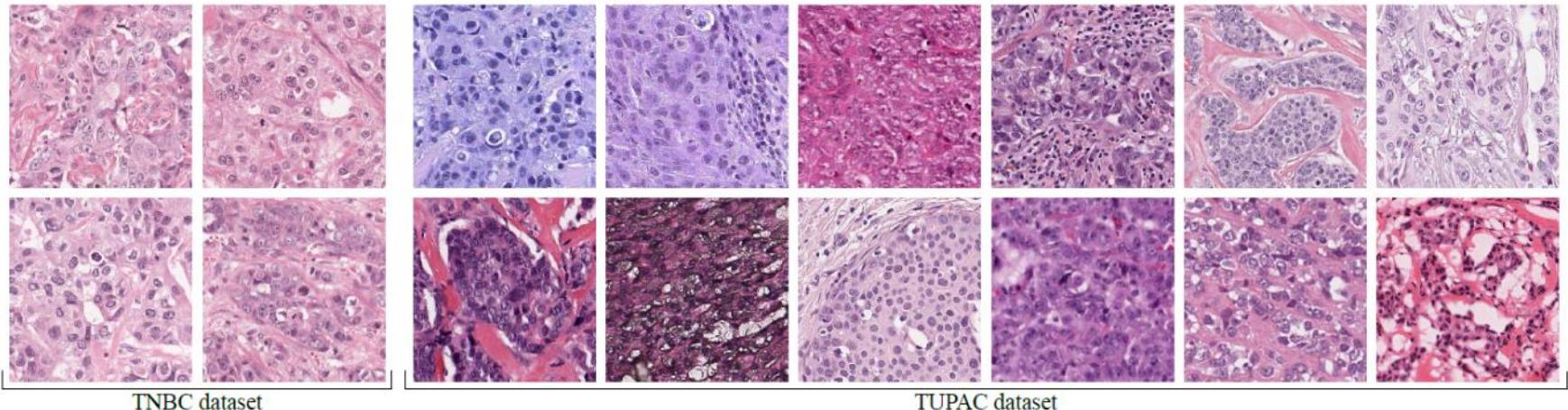
IHC offers a solution



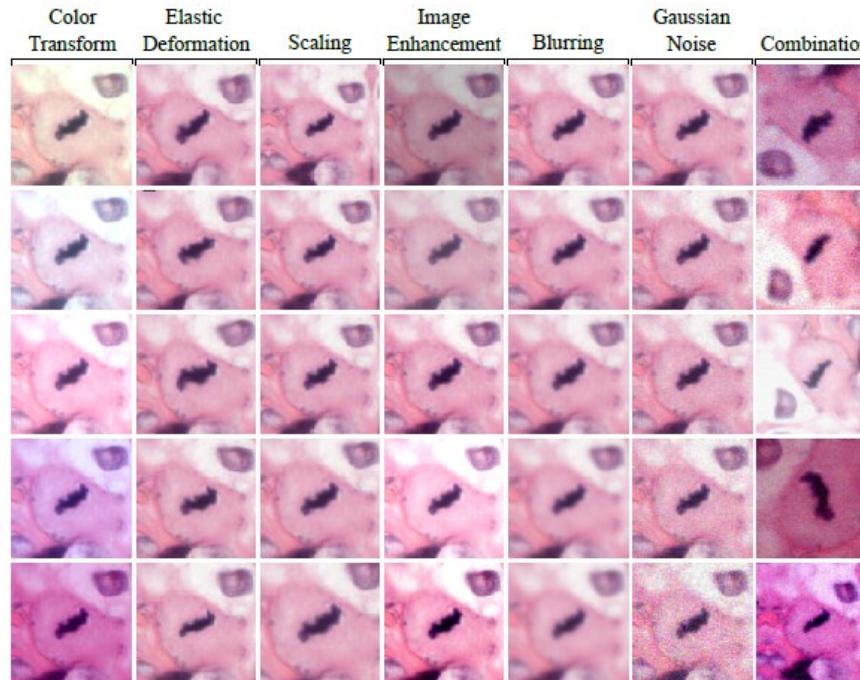


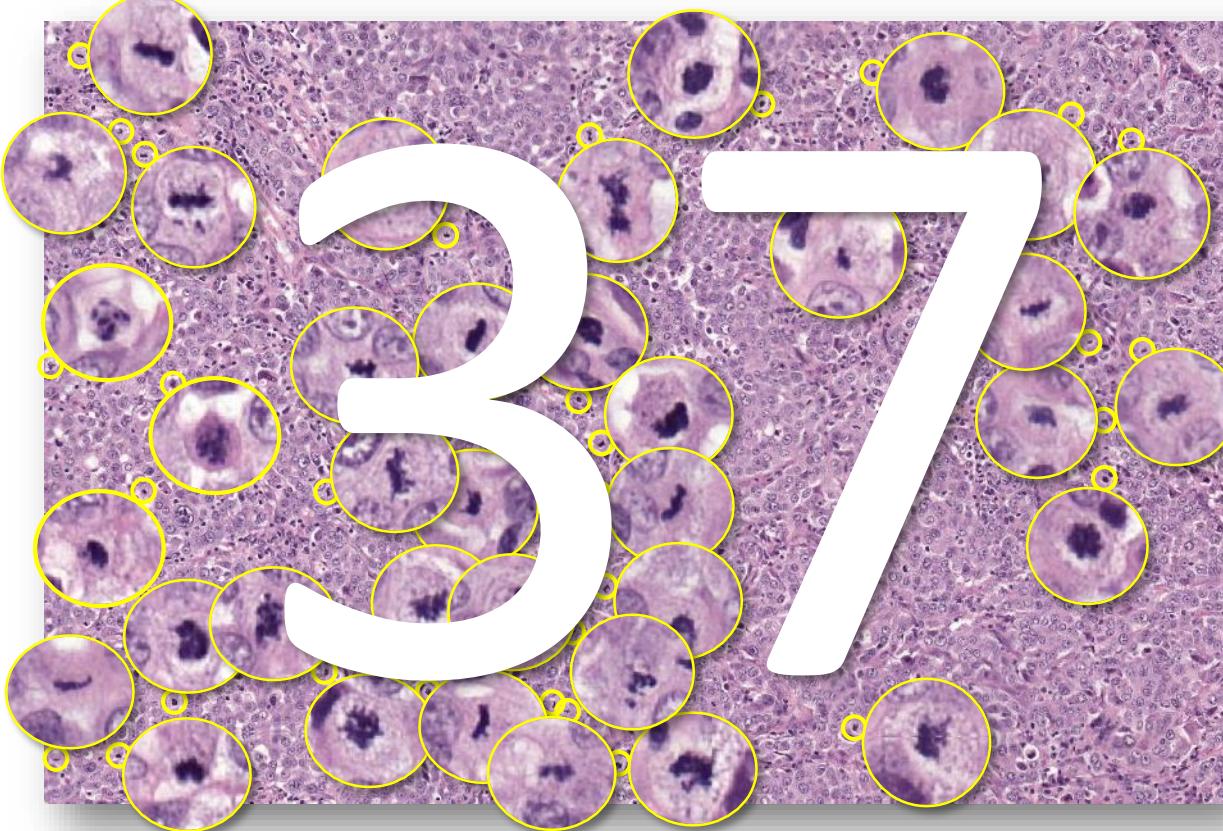


Challenge 2: staining differences

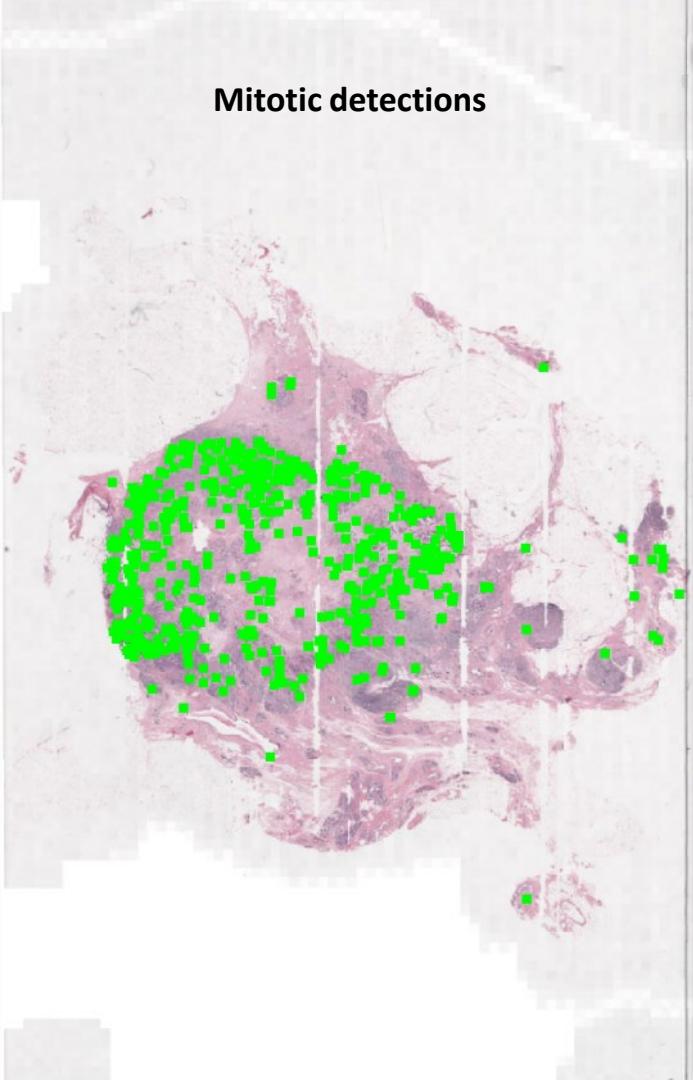


Solution 2: Data augmentation

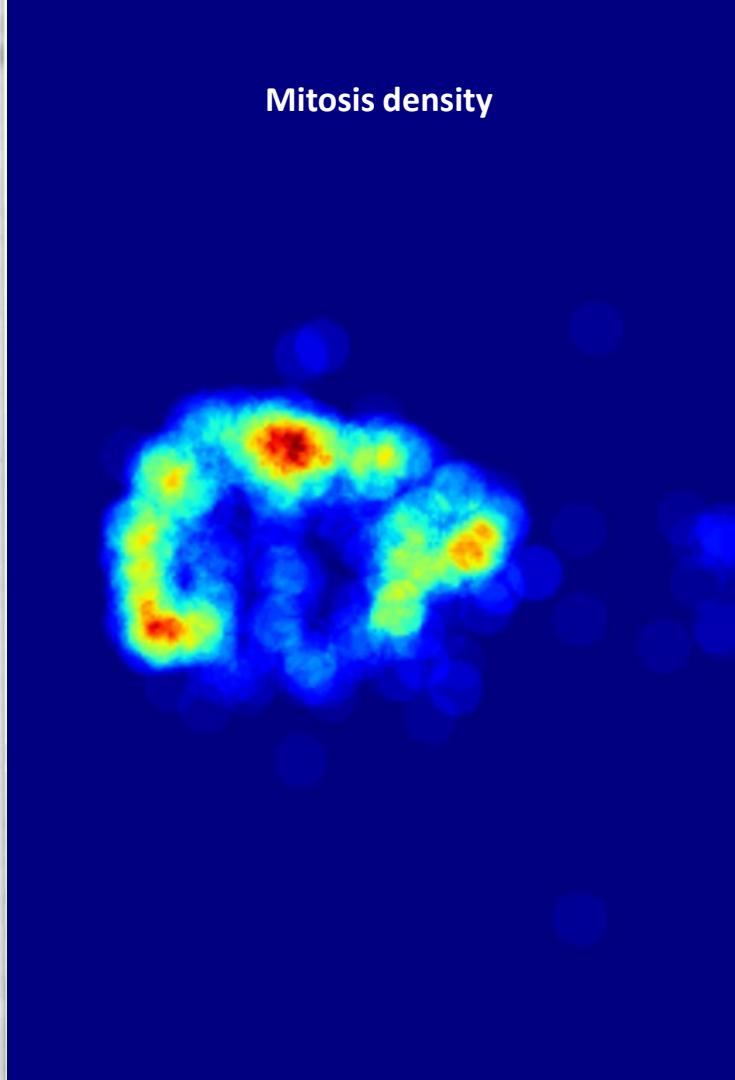




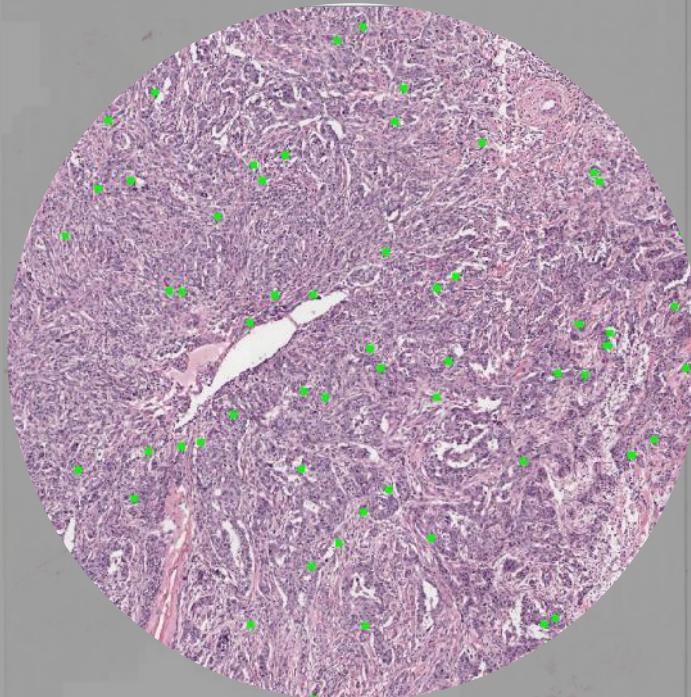
Mitotic detections



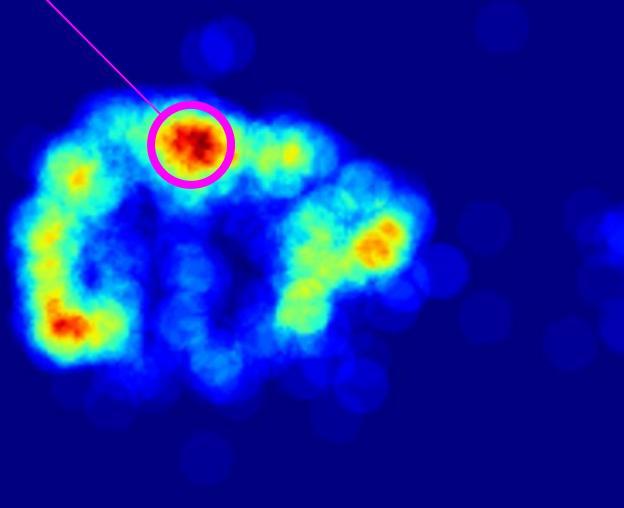
Mitosis density



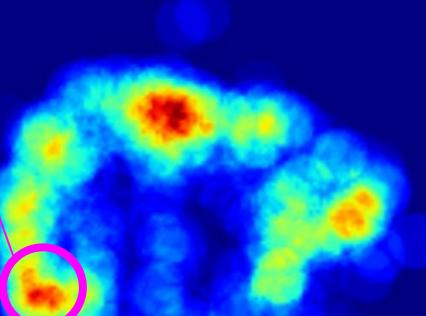
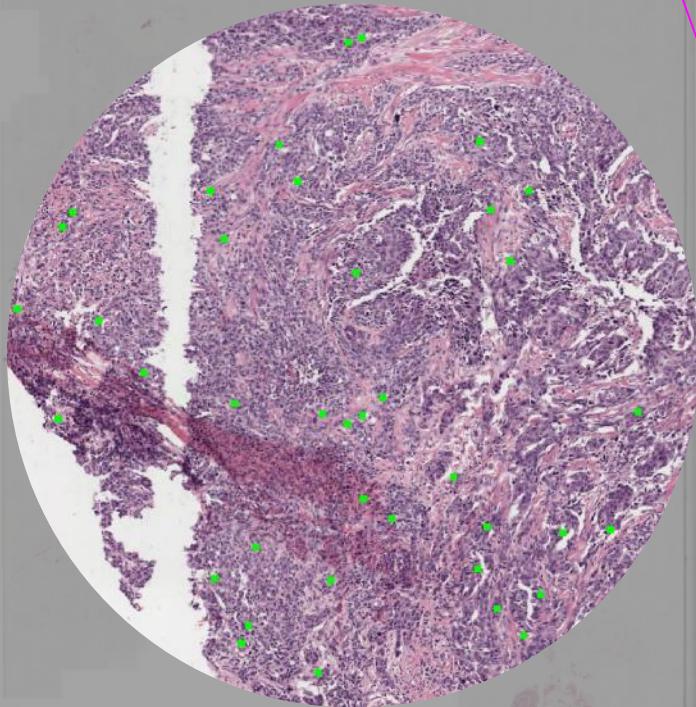
56 mitoses



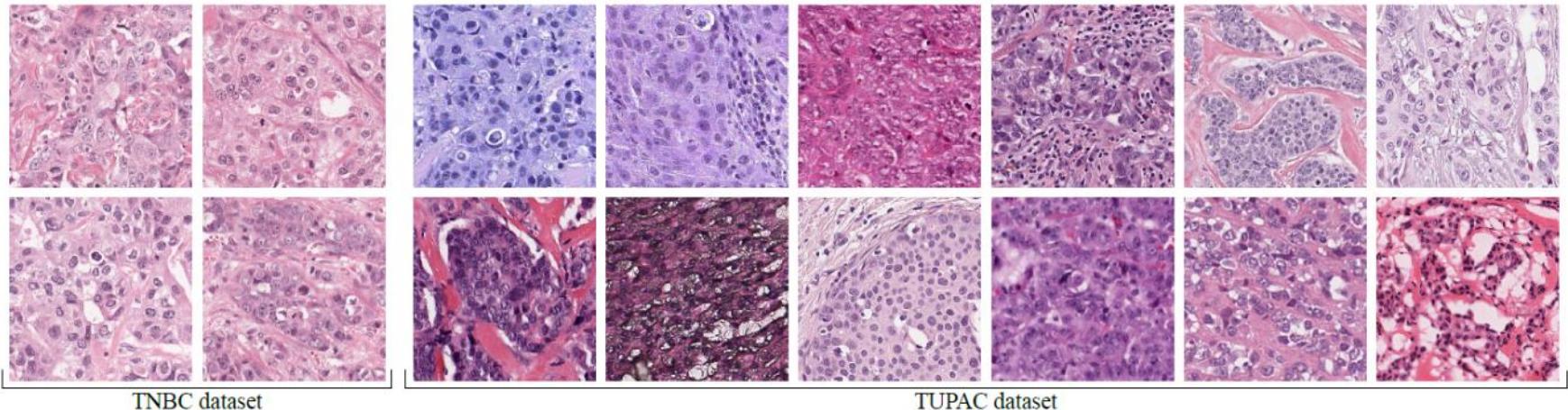
Direct visibility of hotspots



38 mitoses

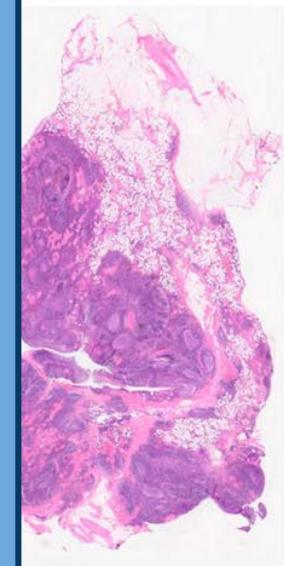
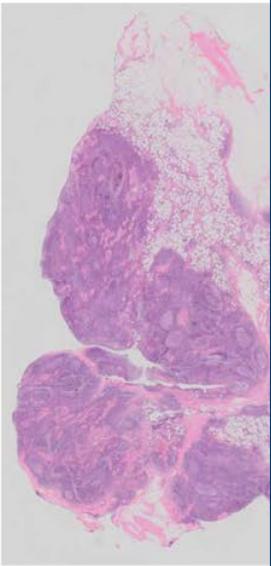


Challenge 2: staining differences

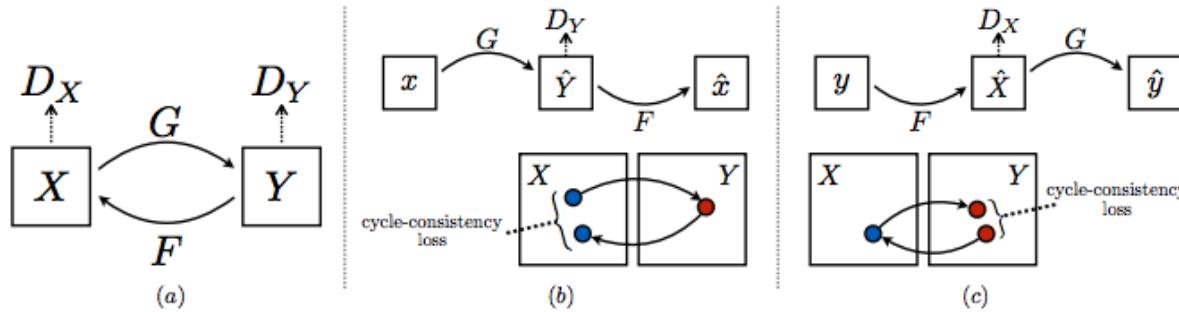


'Traditional' stain normalization

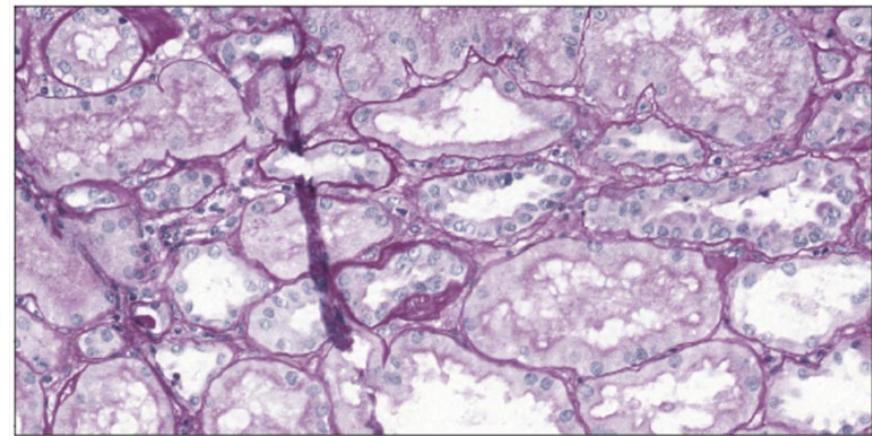
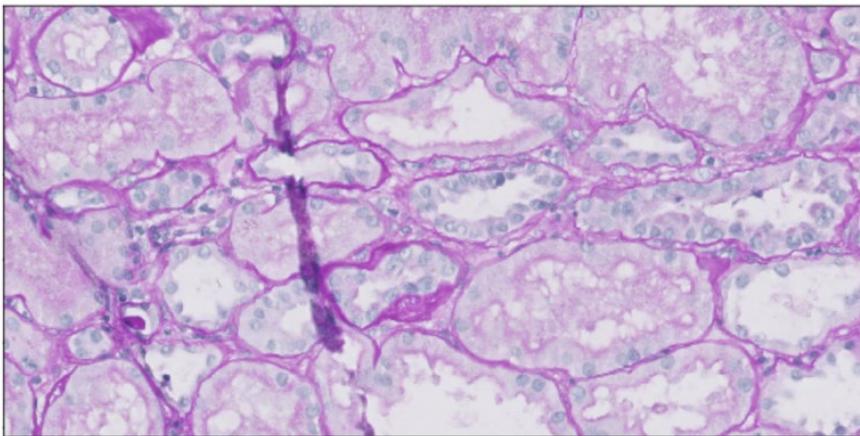
- Only modifies color information
- Very time-consuming algorithm
- Dependent on presence of nuclei
- Parameter tweaking for new datasets



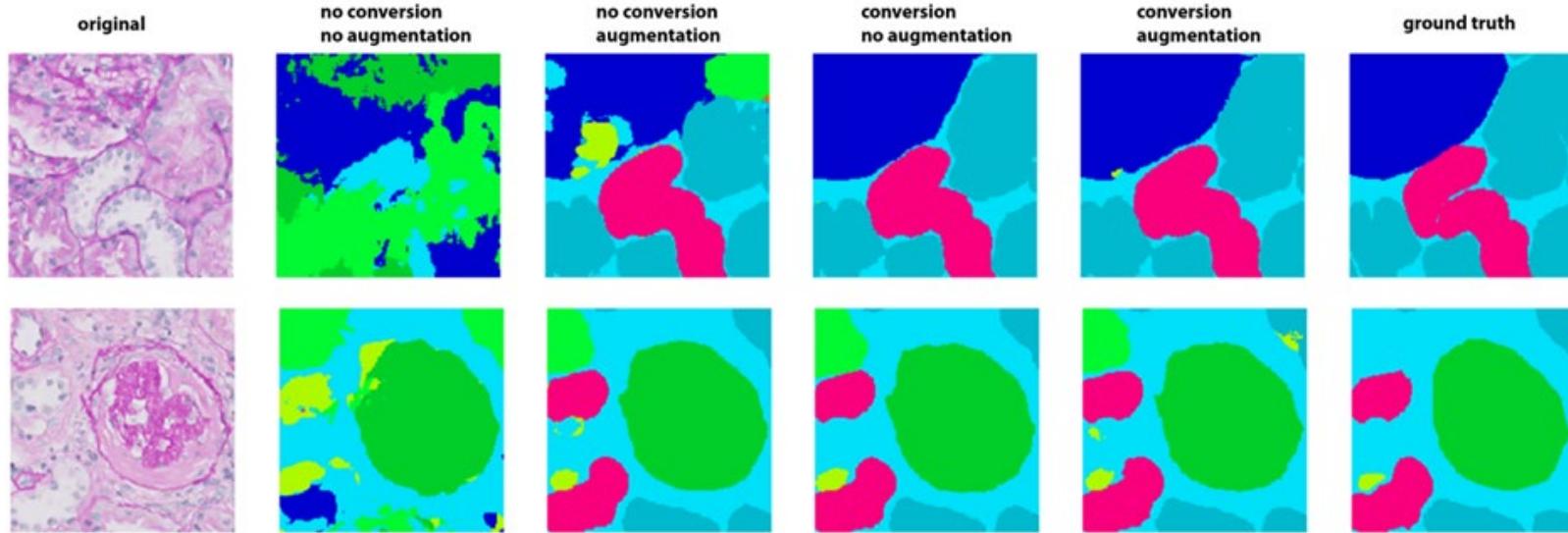
Stain normalization using cycleGANs



Stain normalization using cycleGANs



Stain normalization using cycleGANs



Experiment		Dice coefficient AMC			
Augmentations	Stain transformed	Mean	Std	Min	Max
x	x	0.36	0.21	0.09	0.65
x	✓	0.85	0.06	0.69	0.91
✓	x	0.78	0.08	0.65	0.87
✓	✓	0.85	0.05	0.72	0.91

Practical applications of computation pathology

Detection of metastases in lymph nodes

Automatic mitotic counts

Tumor/stroma ratio quantification

Identification of associations

Tumor/stroma ratio quantification

Annals of Oncology

original articles

Annals of Oncology 24: 179–185, 2013

doi:10.1093/annonc/mds246

Published online 2 August 2012

The proportion of tumor-stroma as a strong prognosticator for stage II and III colon cancer patients: validation in the VICTOR trial

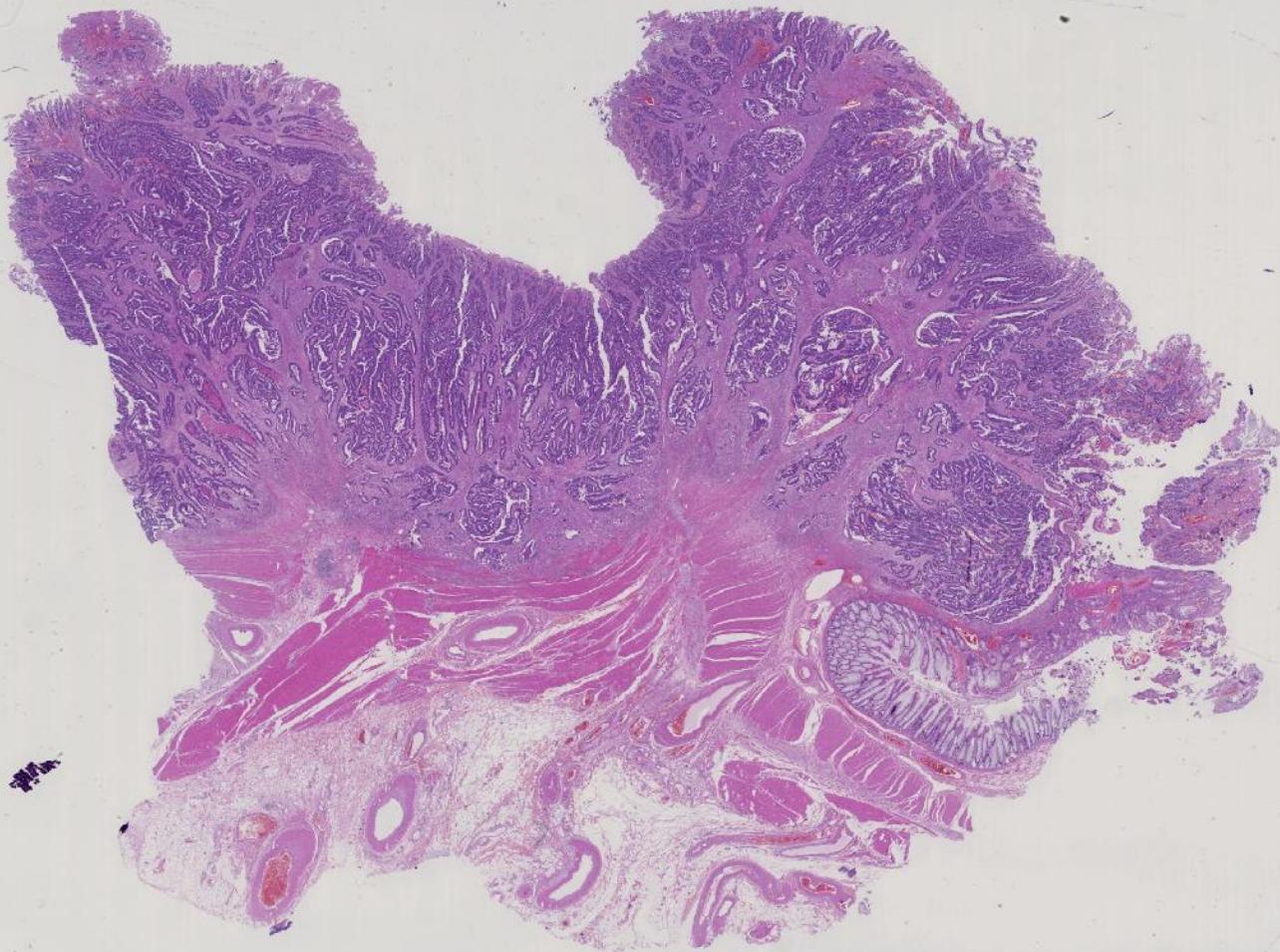
A. Huijbers¹, R. A. E. M. Tollenaar¹, G. W. v Pelt¹, E. C. M. Zeestraten¹, S. Dutton³,
C. C. McConkey⁶, E. Domingo⁷, V. T. H. B. M. Smit², R. Midgley⁴, B. F. Warren⁸, E. C. Johnstone⁴,
D. J. Kerr⁵ & W. E. Mesker^{1*}

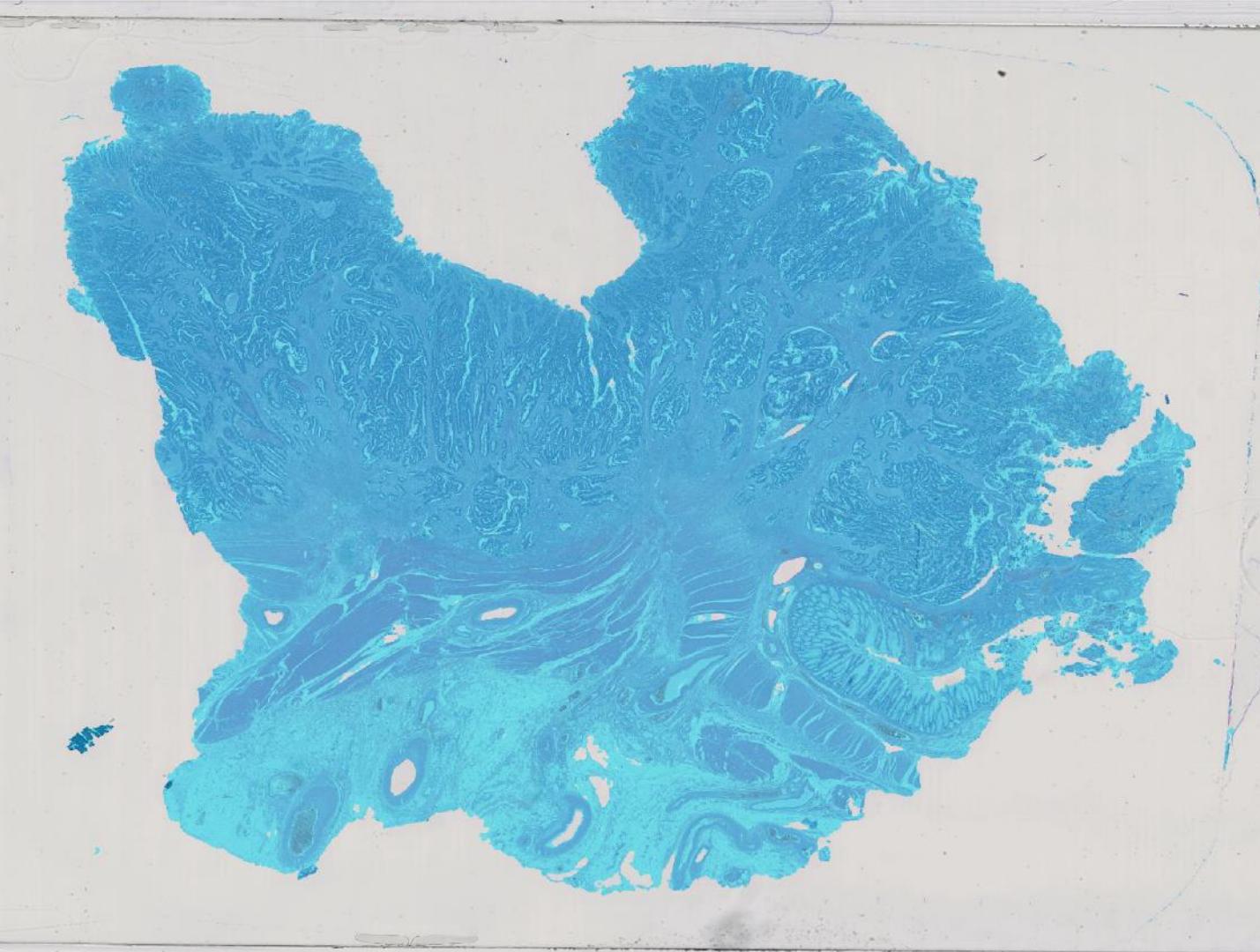
Departments of ¹Surgery; ²Pathology, Leiden University Medical Center (LUMC), Leiden, The Netherlands; ³Centre for Statistics in Medicine, University of Oxford, Oxford;

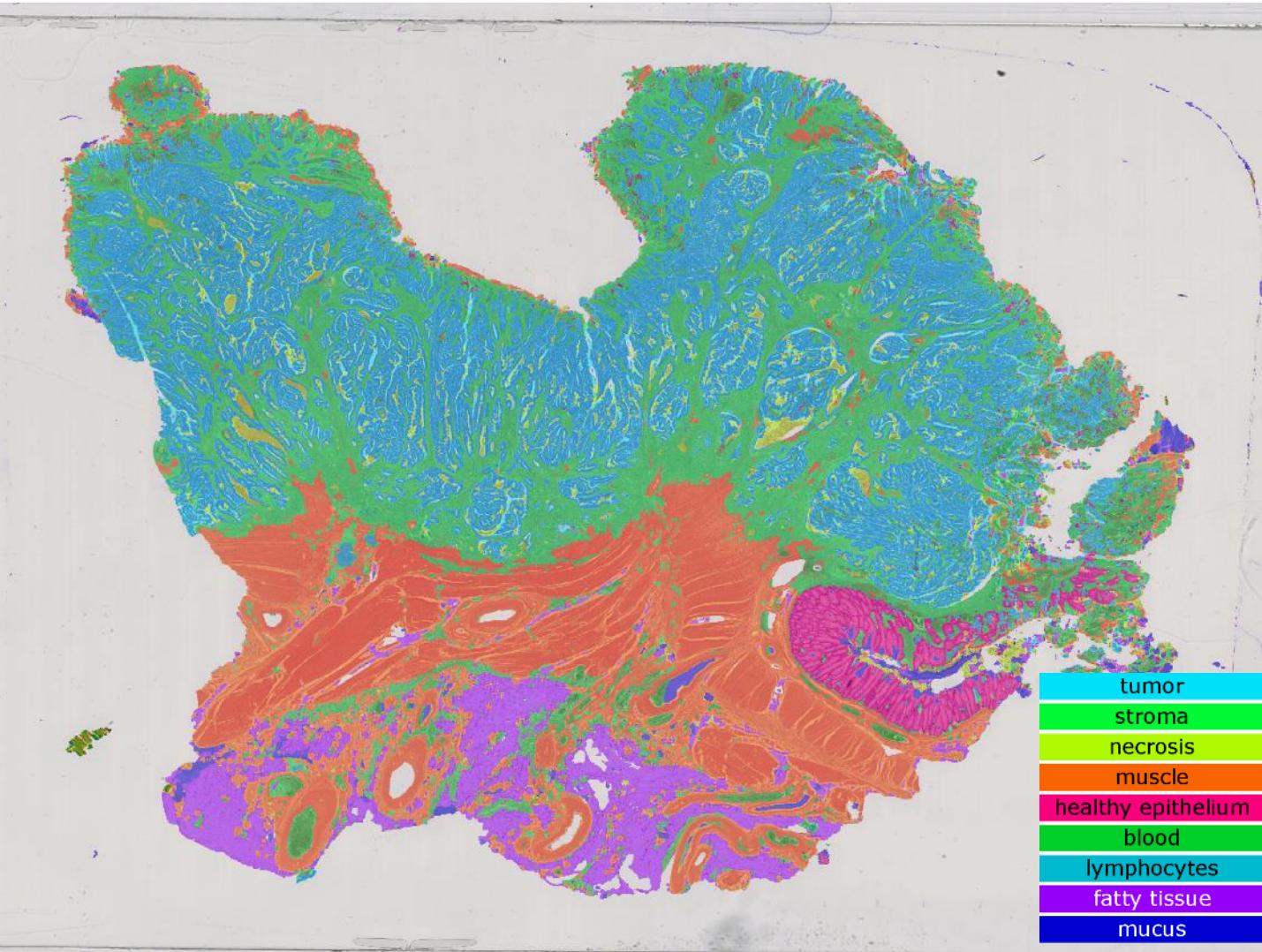
Departments of ⁴Oncology; ⁵Clinical Pharmacology, University of Oxford, Oxford; ⁶Clinical Trials Unit, University of Warwick, Coventry; ⁷Molecular and Population

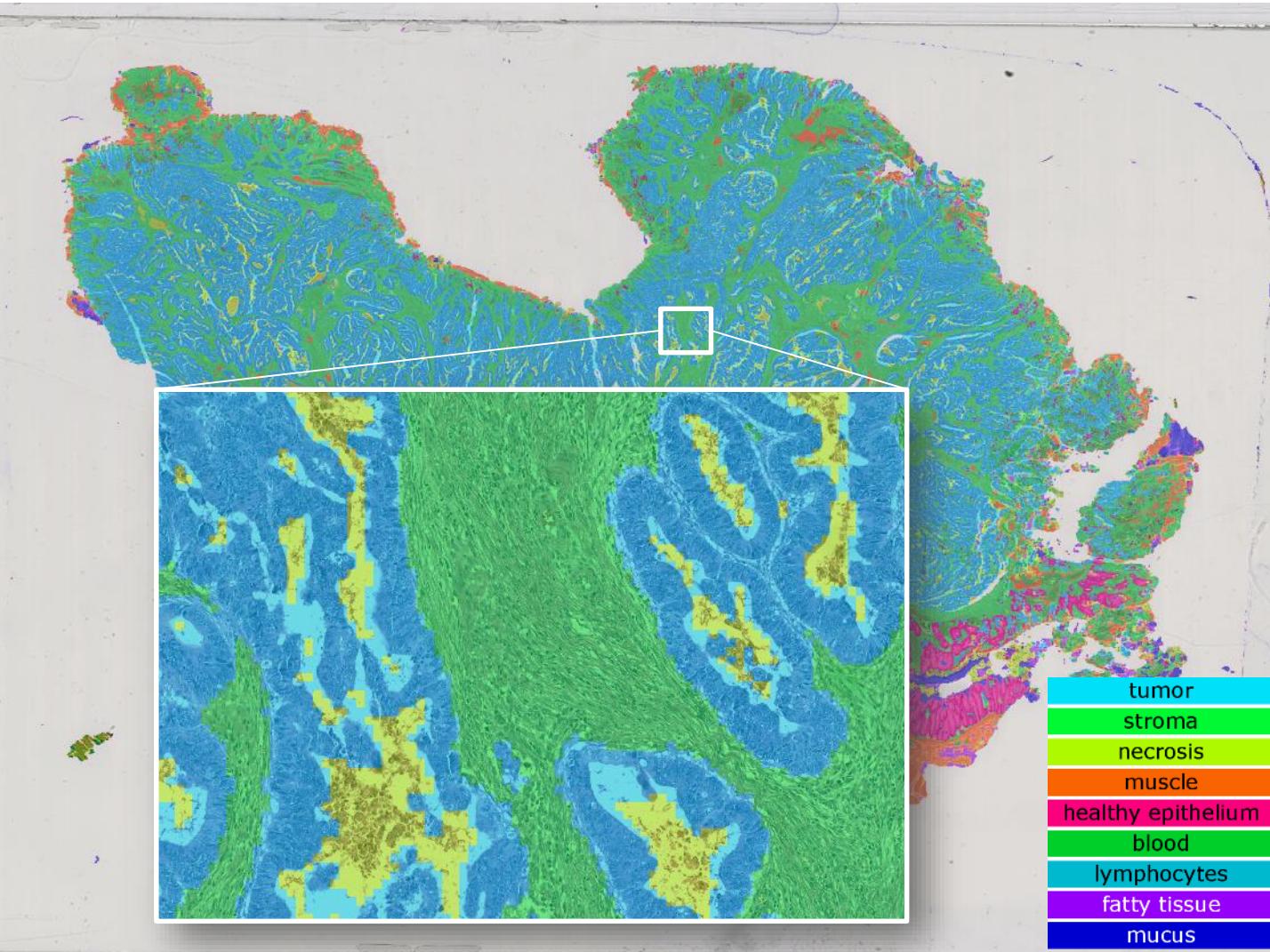
Genetics, Wellcome Trust Center for Human Genetics, Oxford; ⁸Department of Pathology, John Radcliffe Hospital, Headington, Oxford, UK

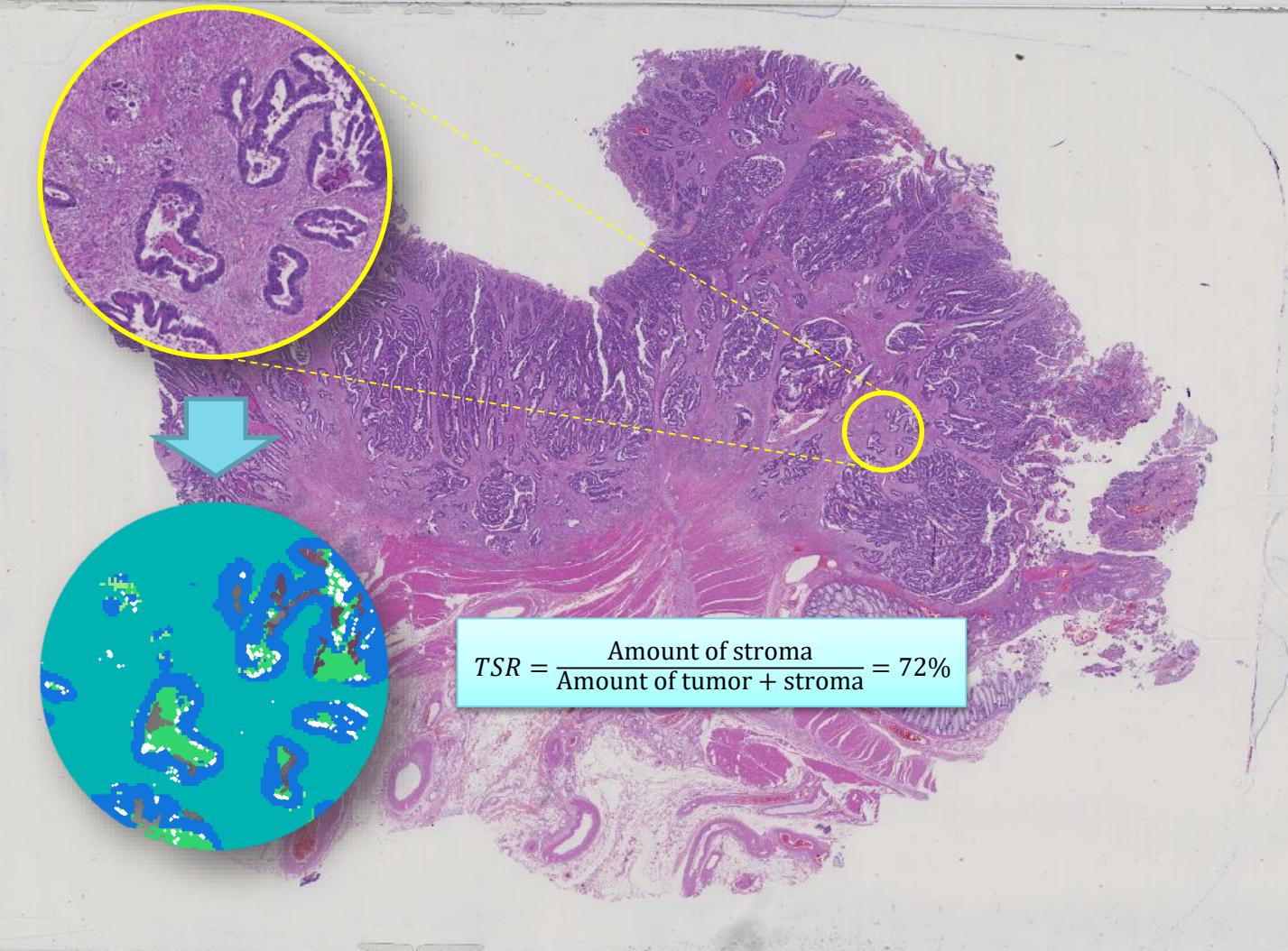
Received 28 February 2012; revised 15 June 2012; accepted 18 June 2012











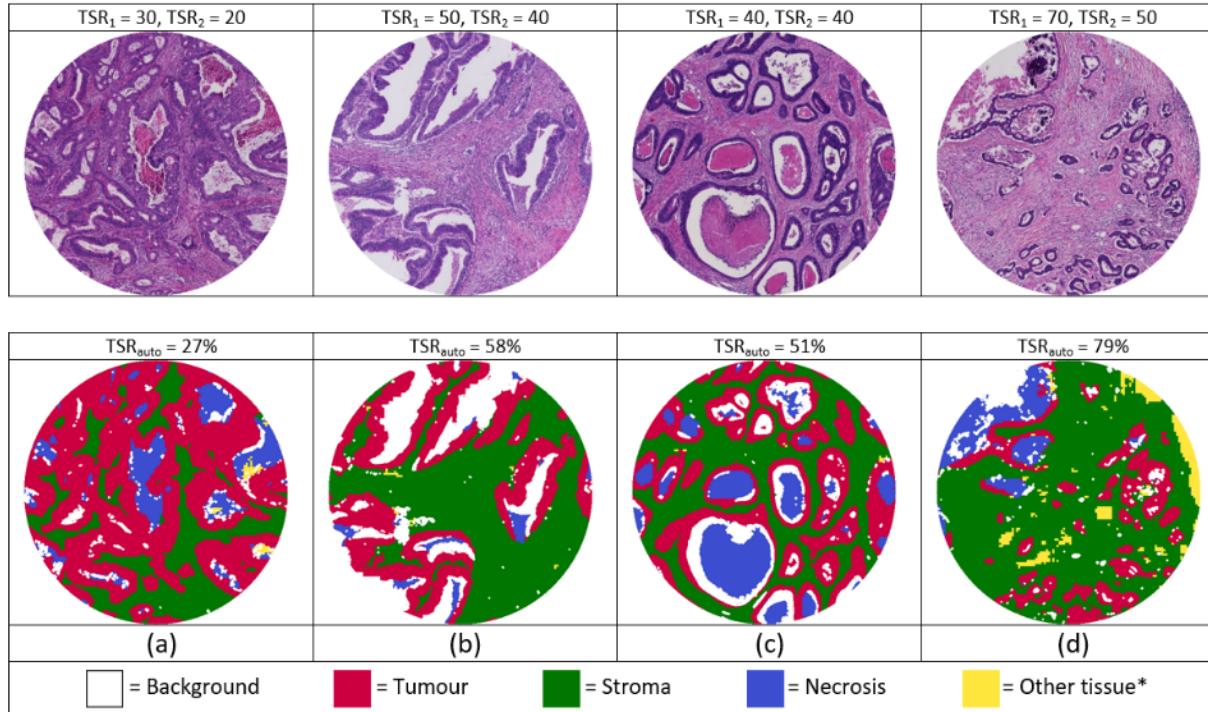
$$TSR = \frac{\text{Amount of stroma}}{\text{Amount of tumor} + \text{stroma}} = 72\%$$

Tumor/stroma ratio quantification

125 patients with rectal carcinoma

- Stage I-III
- At least five year follow-up
- No neo-adjuvant therapy

Tumor/stroma ratio quantification

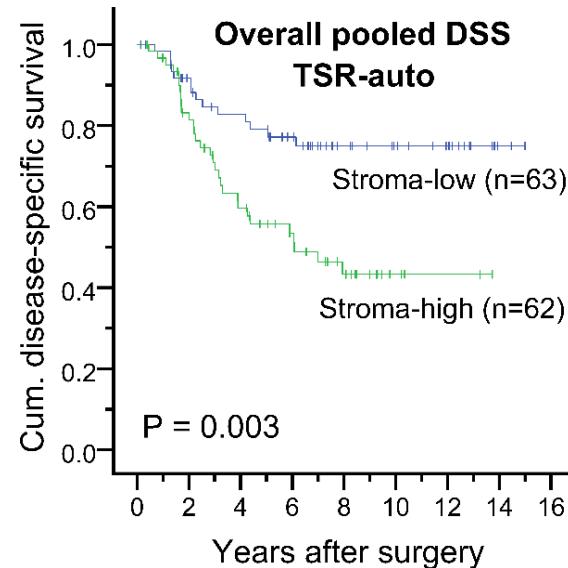
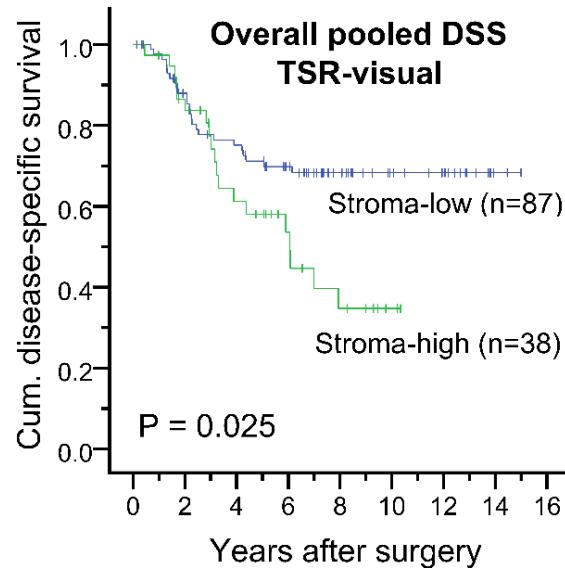


Tumor/stroma ratio quantification

Crosstab: Observer 1 versus Observer 2					
		Observer 2			
		Stroma-low	Stroma-high	Total	
Observer 1	Stroma-low	75	8	83	
	Stroma-high	16	26	42	
	Total	91	34	125	

Crosstab: TSR-Visual (consensus) versus TSR-auto					
		TSR-auto			
		Stroma-low	Stroma-high	Total	
TSR-visual (consensus)	Stroma-low	60	27	87	
	Stroma-high	3	35	38	
	Total	63	62	125	

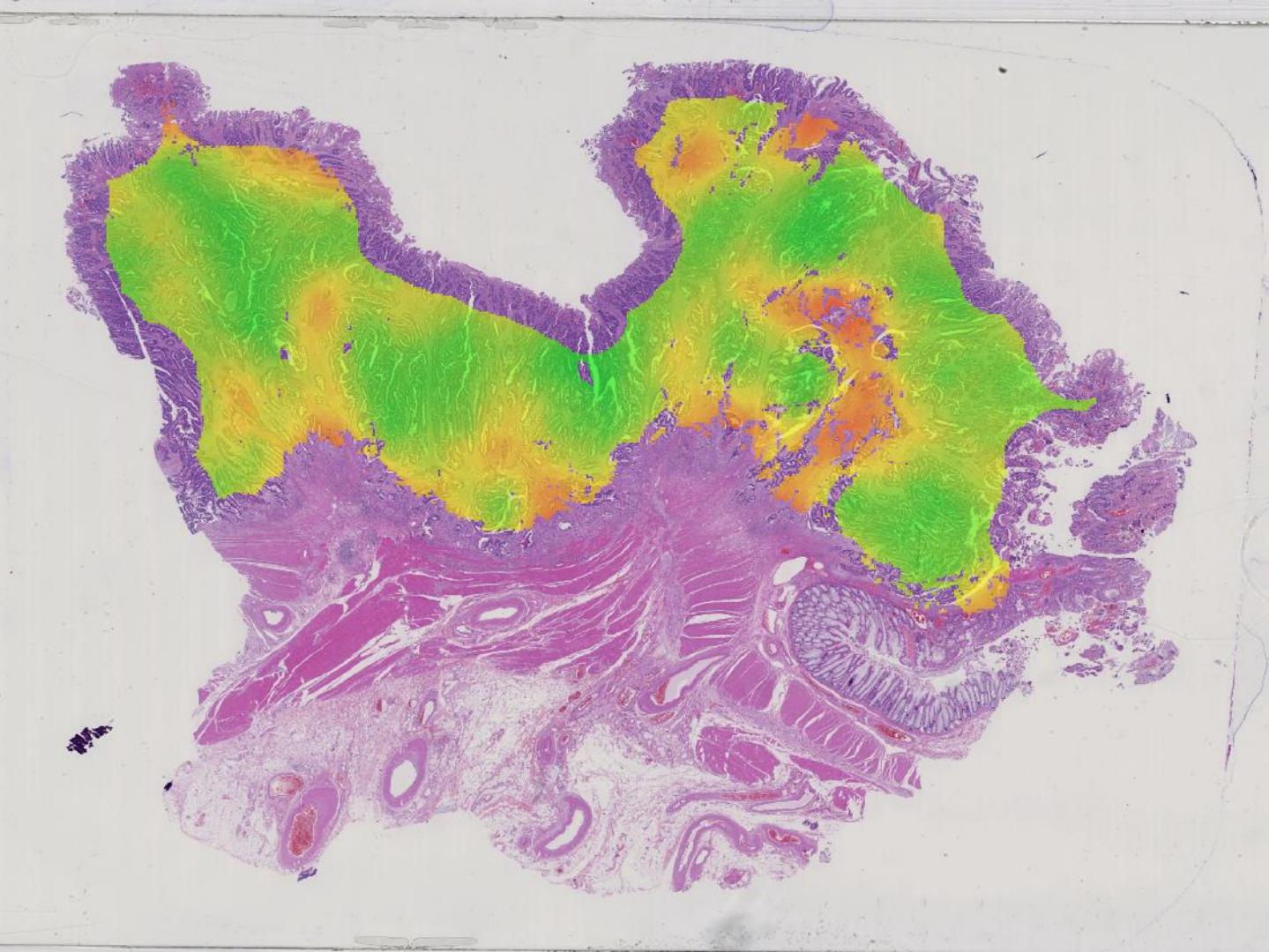
Tumor/stroma ratio quantification



Tumor/stroma ratio quantification

Table 5. Uni- and multivariate Cox regression analysis for disease-specific survival.

	Univariate analysis		Multivariate analysis			
	HR (95% CI)	P-val.	Visual	P-val.	Auto	P-val.
Age	1.01 (0.98-1.04)	0.376				
Gender	0.85 (0.45-1.60)	0.604				
T-stage	2.42 (1.47-3.99)	0.001	1.97 (1.16-3.34)	0.012	2.05 (1.24-3.38)	0.005
N-stage	2.16 (1.49-3.14)	0.0001	2.06 (1.13-3.75)	0.018	2.12 (1.17-3.84)	0.014
Surgical procedure	1.48 (0.94-2.31)	0.090				
Tumour grade	2.96 (1.42-6.17)	0.004	2.40 (1.05-5.48)	0.038	2.23 (0.99-5.00)	0.052
Adj. <u>chemoth.</u>	1.17 (0.28-4.82)	0.831				
Adj. <u>radioth.</u>	2.56 (1.41-4.63)	0.002	0.72 (0.27-1.88)	0.496	0.68 (0.27-1.72)	0.417
TSR-visual	1.96 (1.08-3.58)	0.027	2.07 (1.09-3.93)	0.026		
TSR-auto	2.57 (1.36-4.86)	0.004			2.75 (1.44-5.27)	0.002



Practical applications of computation pathology

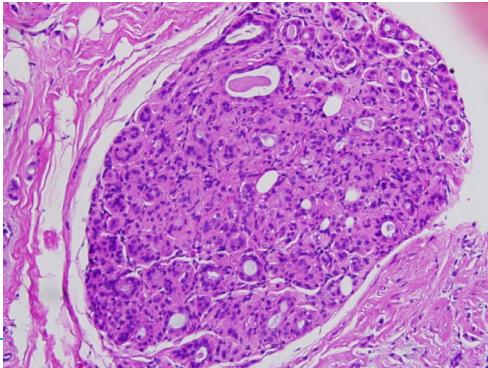
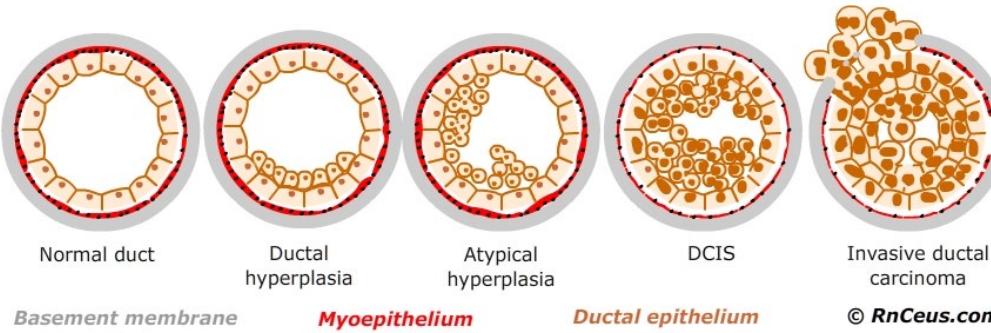
Automatic mitotic counts

Tumor/stroma ratio quantification

Identification of tumor associated stroma

Prec...
ex...

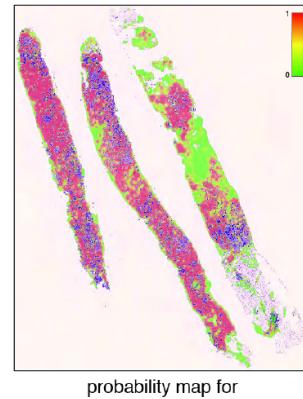
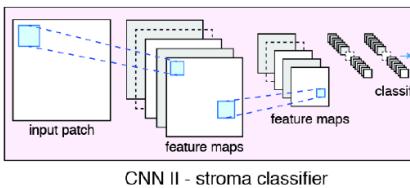
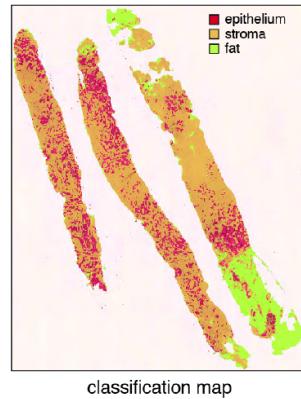
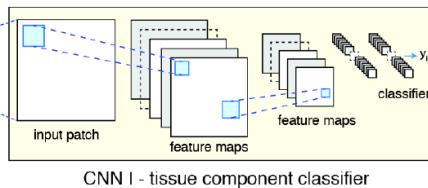
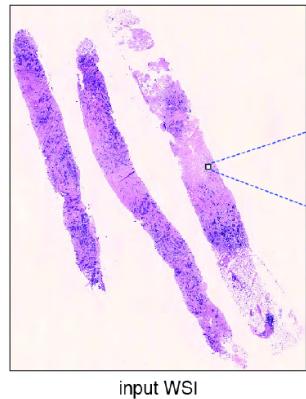
Prognosis of in-situ lesions

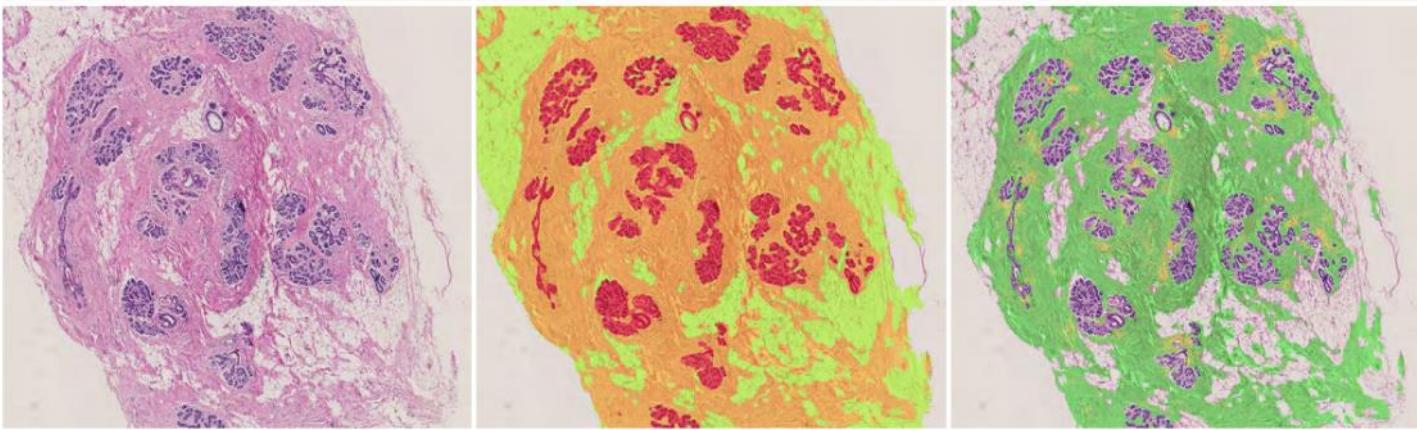
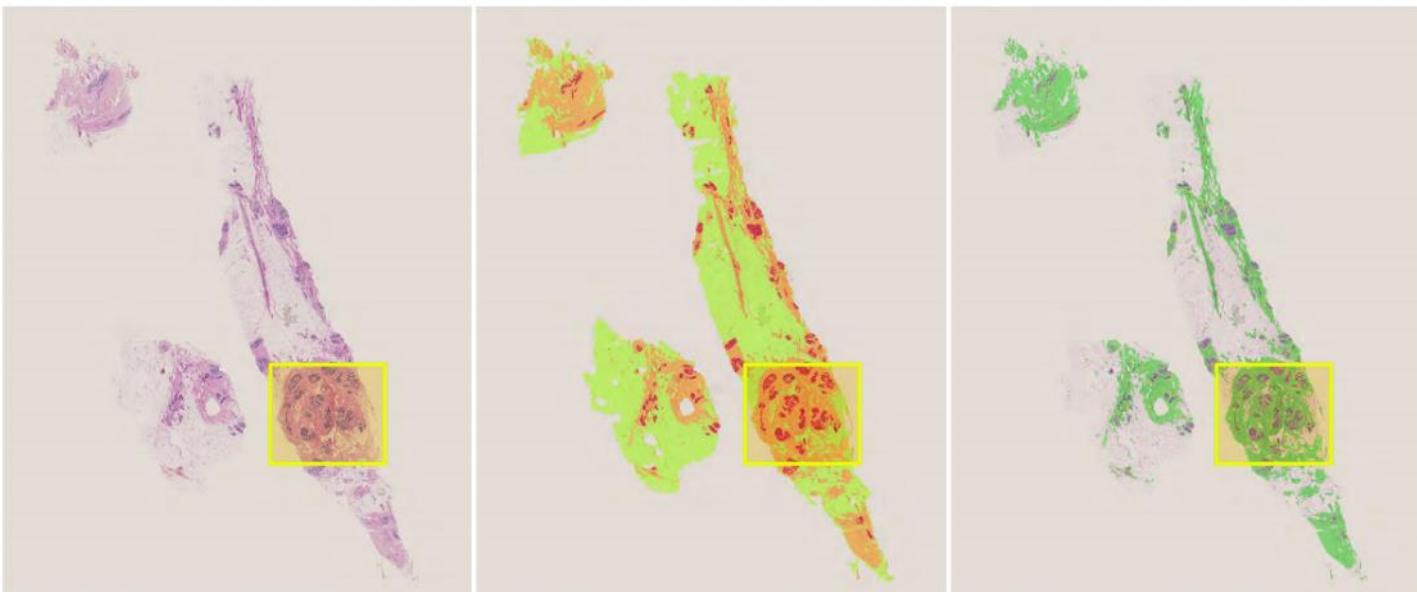


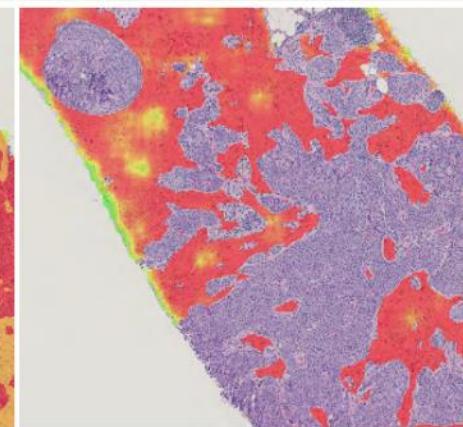
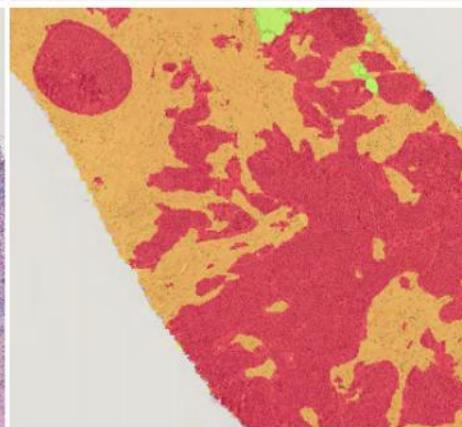
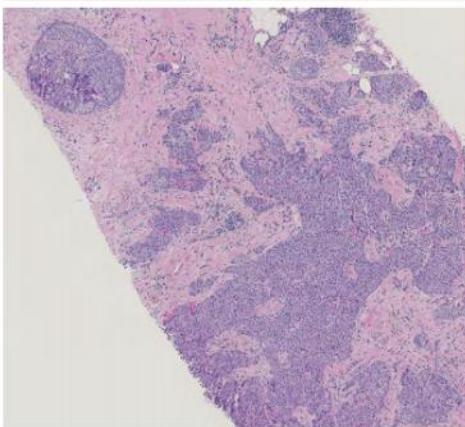
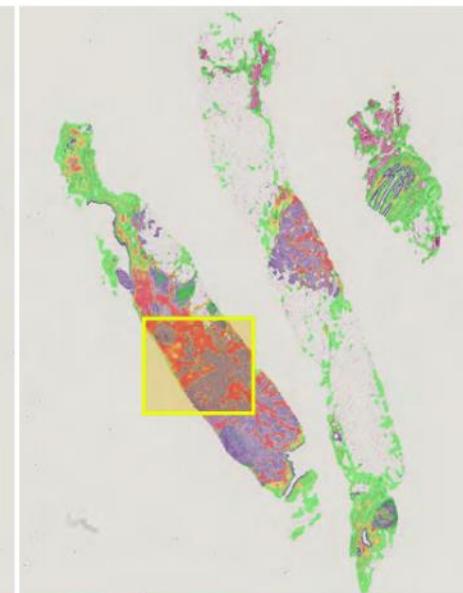
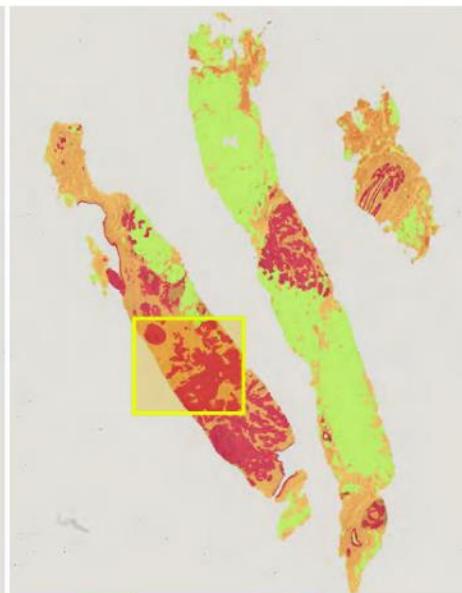
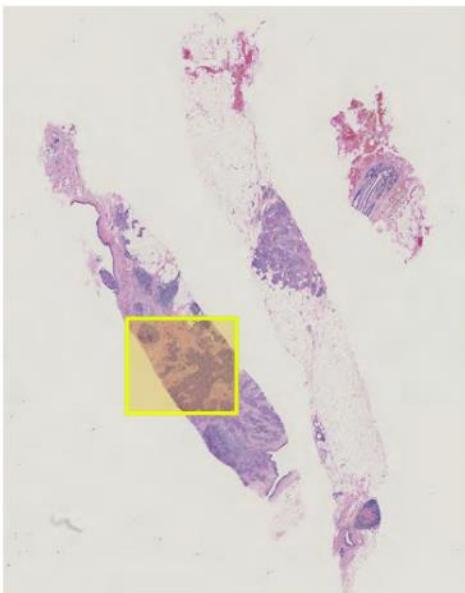
Diagnosis	Entire dataset			Training dataset			Testing dataset		
	# Patient	# WSI	%	# Patient	# WSI	%	# Patient	# WSI	%
Benign	321	675	36.4	209	437	37.9	112	238	33.9
Proliferative	312	937	35.4	209	608	37.9	103	329	32.3
Proliferative with atypia	57	212	6.5	42	171	7.6	15	41	4.5
Ductal carcinoma in-situ	58	222	6.6	—	—	—	58	222	17.6
Lobular carcinoma in-situ	10	29	1.1	7	21	1.2	3	8	0.9
Invasive breast cancer	124	312	14.0	85	222	15.4	39	90	11.8
Total	882	2387	100	552	1459	100	330	928	100

Tumor-associated stroma

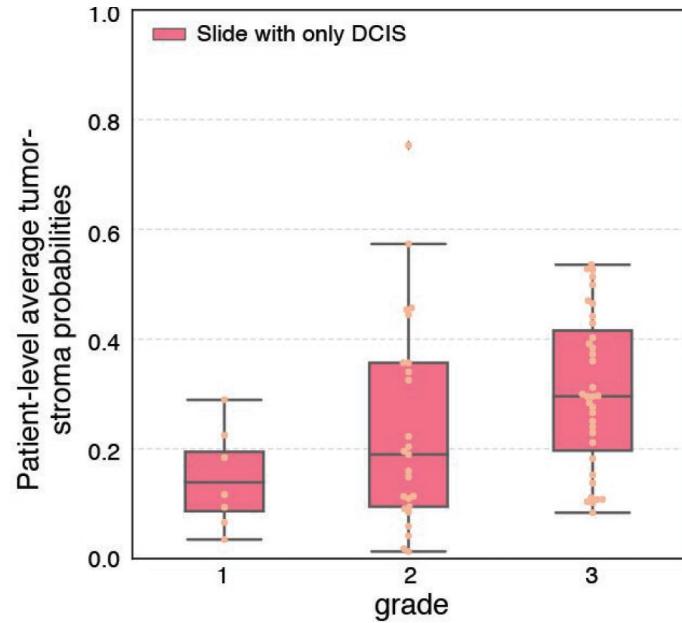
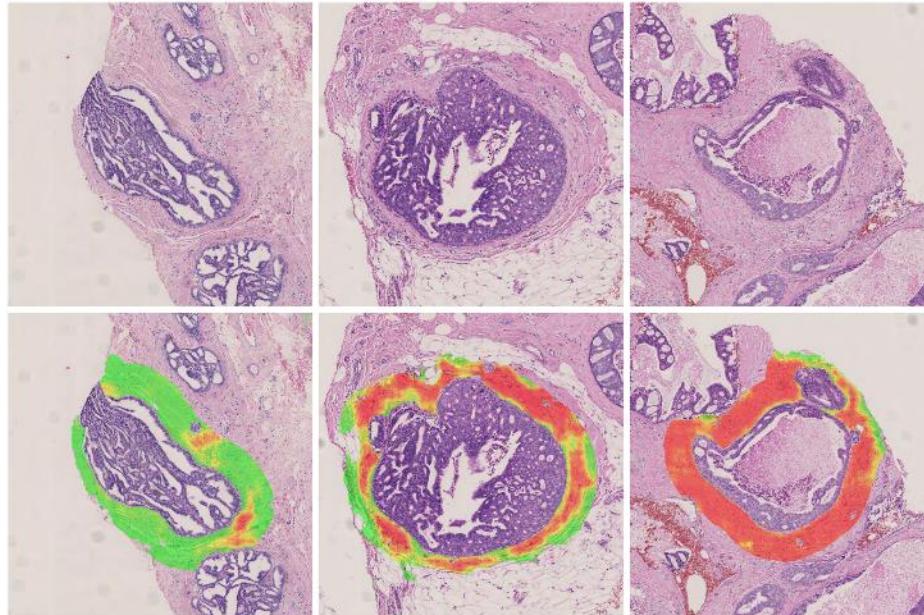
Tumor stroma identification pipeline







Tumor-associated stroma



Practical applications of computation pathology

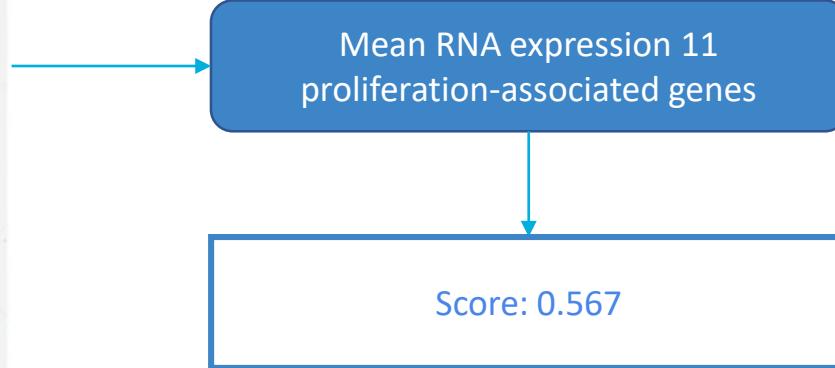
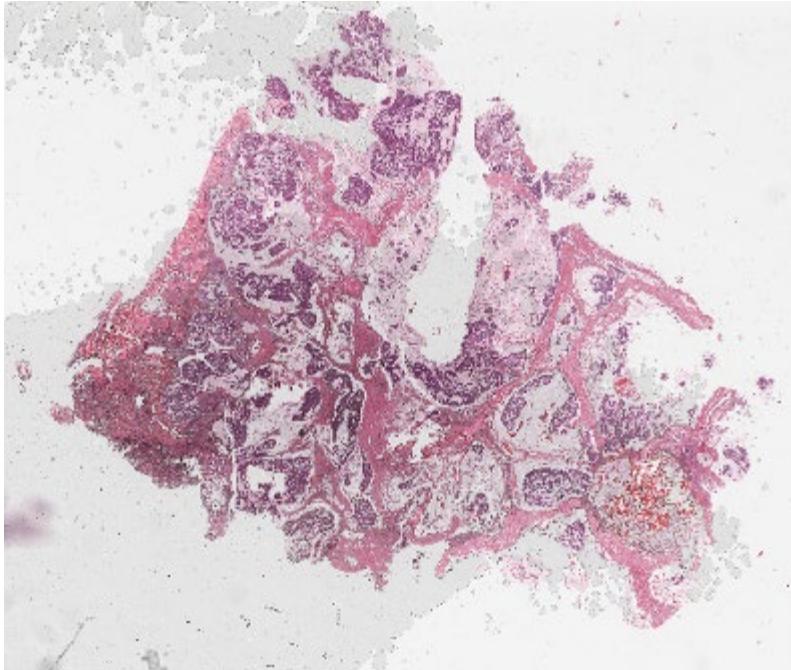
Tumor/stroma ratio
quantification

Identification of tumor
associated stroma

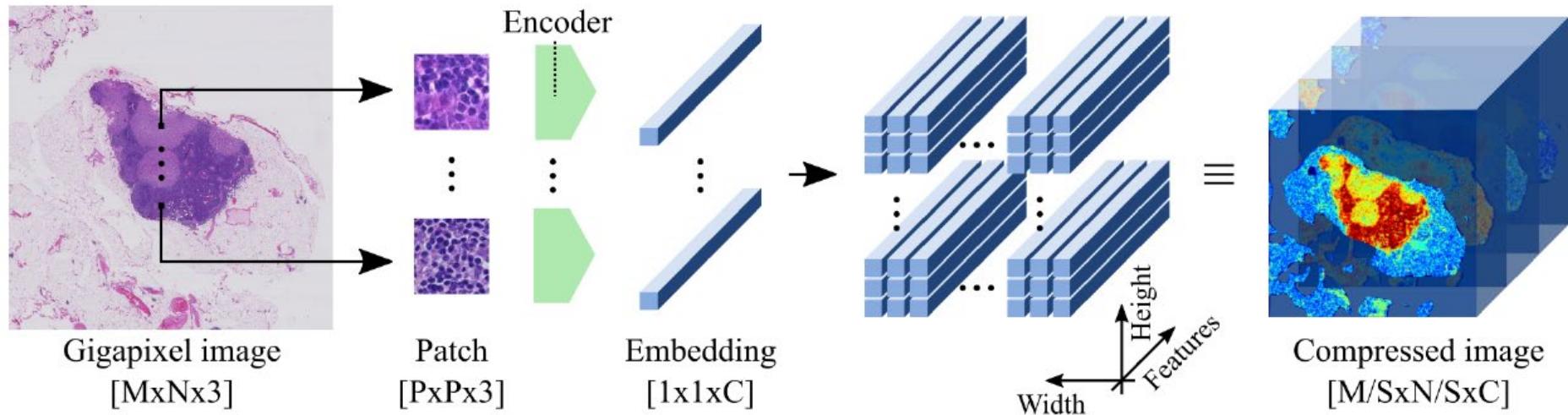
Prediction of gene
expression

Gleas...

Prediction of gene expression



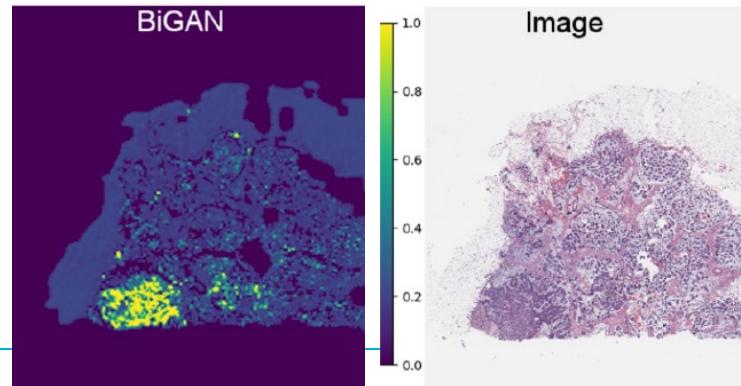
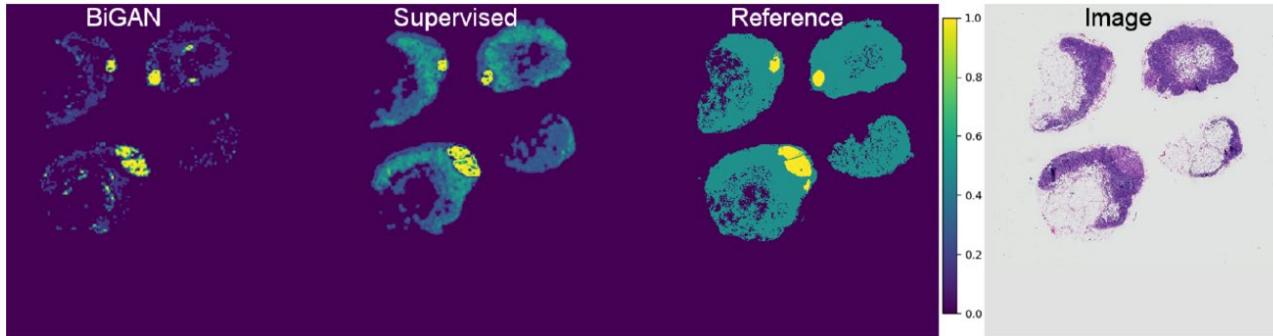
Neural compression



Prediction of gene expression

Team	Spearman's ρ
Lunit (mitosis counting)	0.617
Radboud (neural compression)	0.557
Radboud (regular CNN)	0.516
ContextVision	0.503

Explainability of ML systems



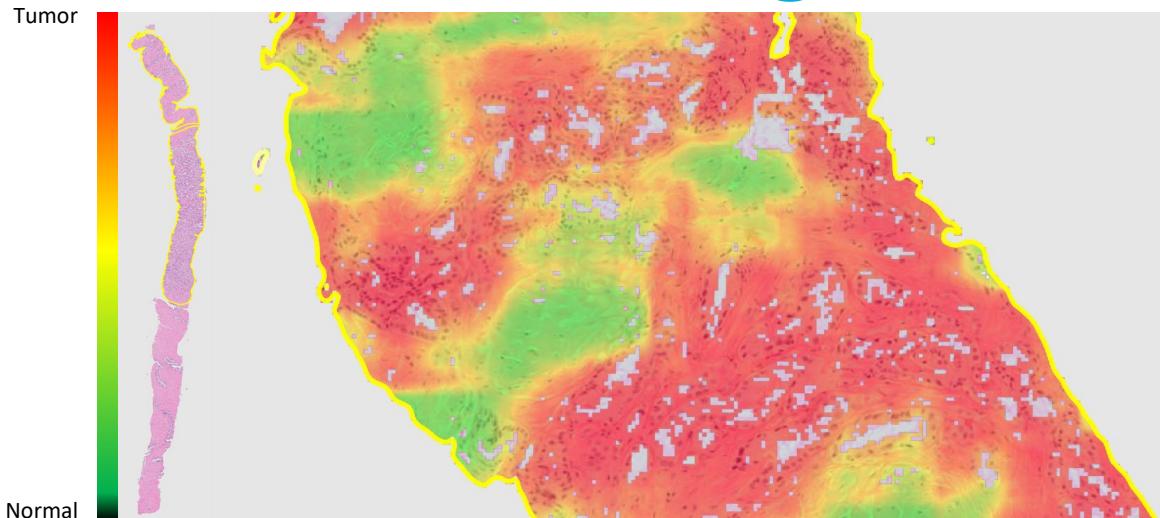
Practical applications of computation pathology

Identification of tumor
associated stroma

Prediction of gene
expression

Gleason grading of
prostate cancer

Prostate cancer segmentation



Training set

150 slides

- 83 normal
- 67 cancer

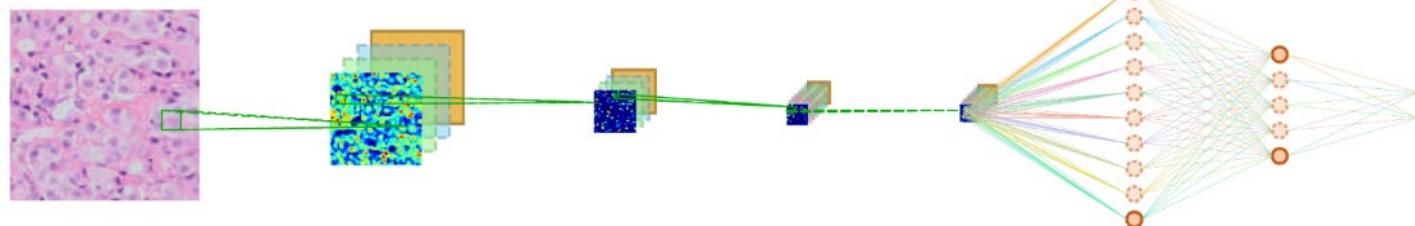
Test set

75 slides

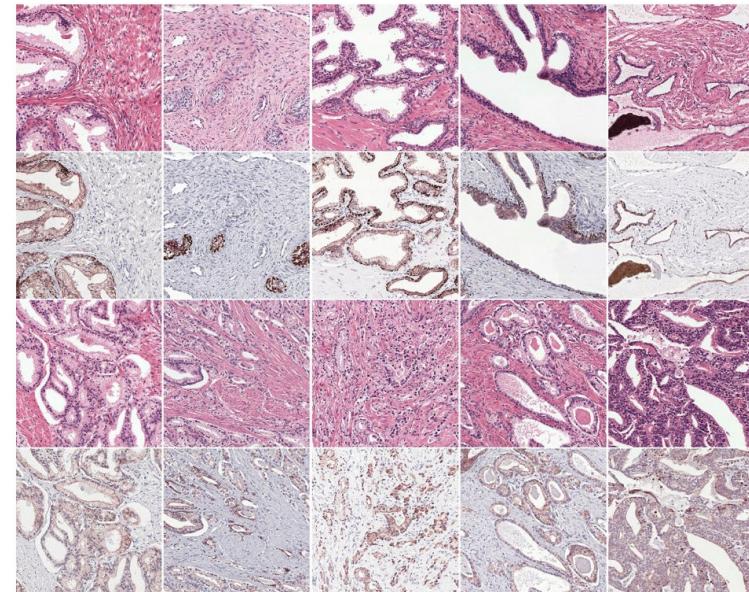
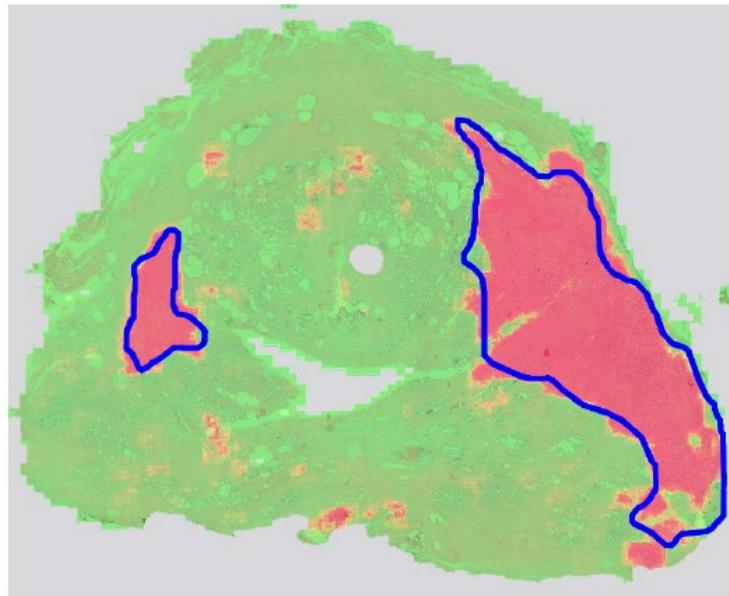
- 30 normal
- 45 cancer

Results

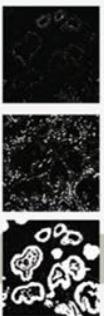
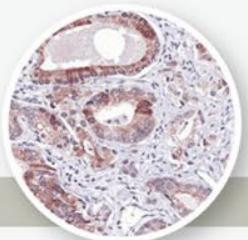
AUC of 0.99...



Prostate cancer: epithelium segmentation



1) Training of IHC network



Input data: 25 IHC WSIs
(20 training, 5 validation)

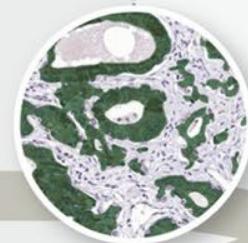
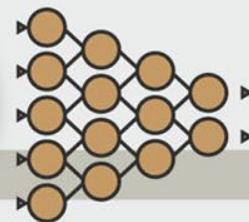
Specimens are stained with CK8/18 and P63 to mark epithelial tissue and basal cell layer.

Color deconvolution is applied to each slide. Only the channel representing the epithelial tissue is used, the rest is discarded.

Artifacts are introduced due to imperfections in the staining and color deconvolution method (Example: top left corner).

Artifacts are removed manually in selected regions. Training data is sampled from these regions.

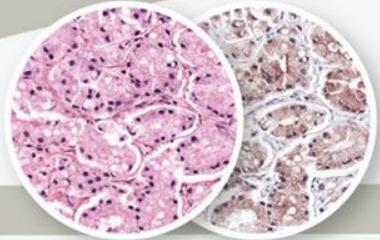
Network training



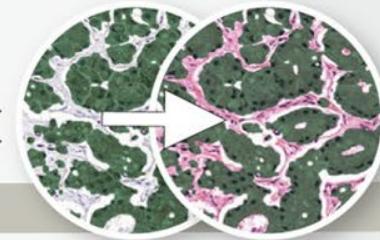
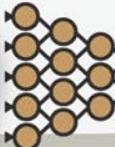
A 5-layer deep U-Net is trained on the corrected IHC regions. Areas with artifacts are sampled more.

The IHC network produces precise segmentation masks given an IHC slide, independent of the color deconvolution.

2) Training of H&E network

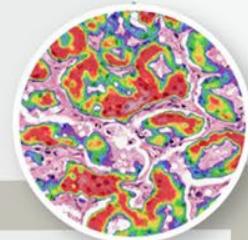
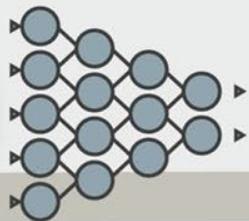


Input data: 62 restained and registered IHC/H&E pairs (50 training, 12 validation)



The trained IHC network is applied to each IHC slide. The network output is used as the training mask for the H&E network. No additional post processing or manual annotations are used.

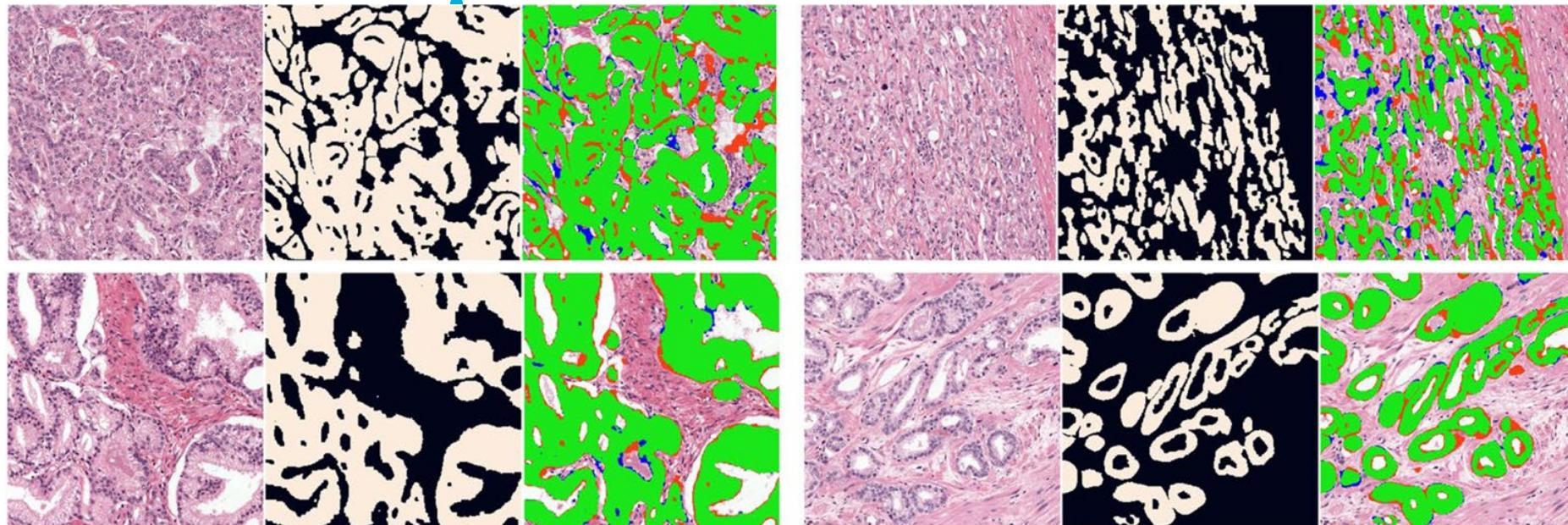
Network training



A 6-layer deep U-Net is trained on H&E and the masks generated by the IHC network.

The trained H&E network segments epithelial tissue on H&E.

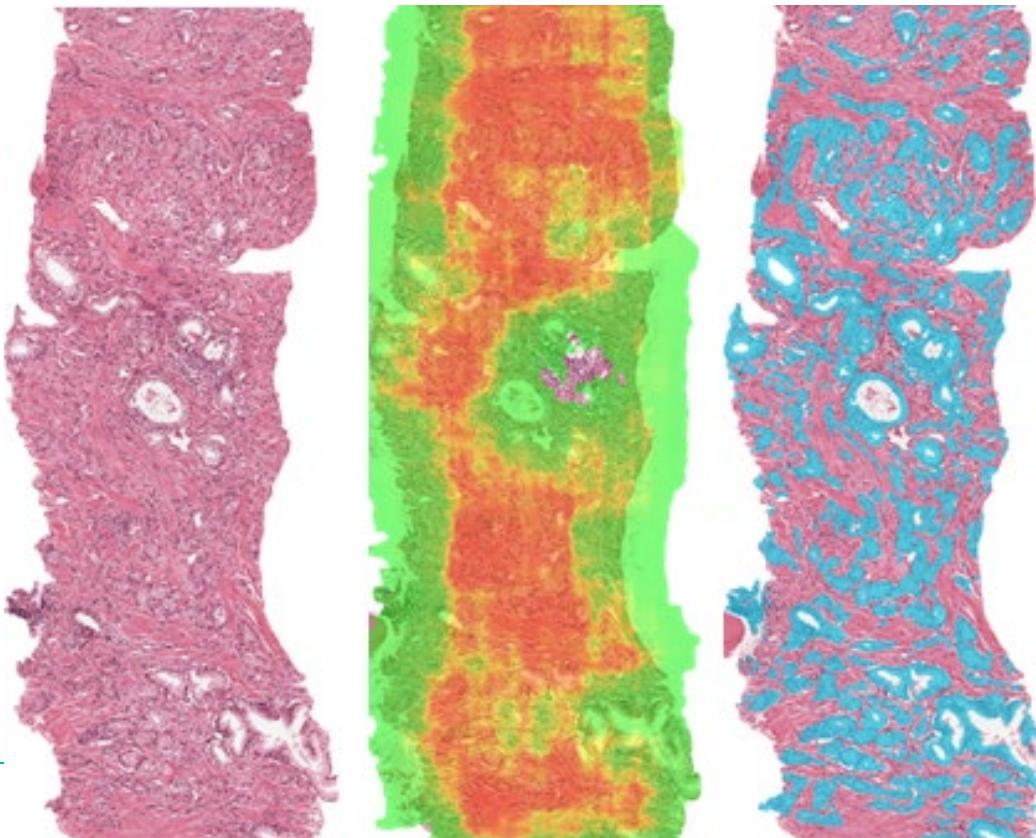
Prostate cancer: epithelium



Regions	N	F1 score mean (min, max)	Accuracy	Jaccard
All regions	160	0.893 ± 0.05 (0.661, 0.959)	0.940	0.811

Network	Evaluation	Accuracy	F1	Jaccard
Gertych <i>et al.</i> ⁸	Cross-validation	—	—	0.595 ± 0.15
Li <i>et al.</i> ¹²	Cross-validation	—	—	0.737^*
Our method	Hold-out validation	0.866 ± 0.07	0.835 ± 0.13	0.735 ± 0.16

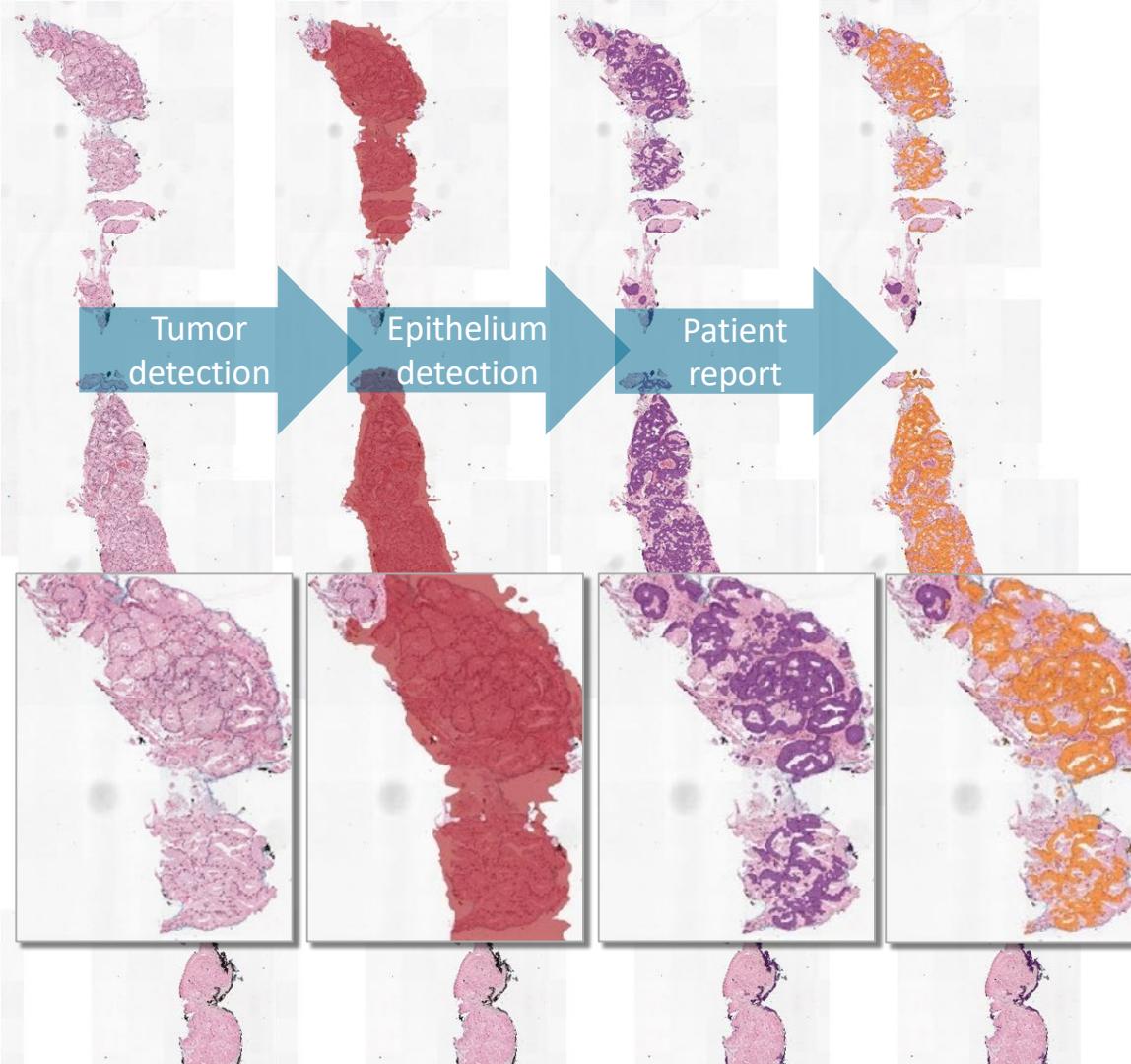
Prostate cancer: Gleason Grading



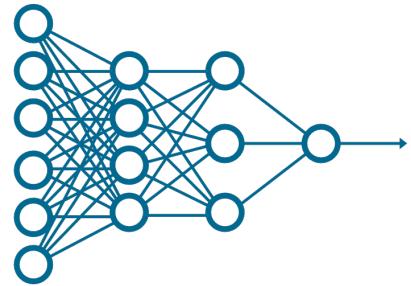
Prostate cancer: Gleason Grading

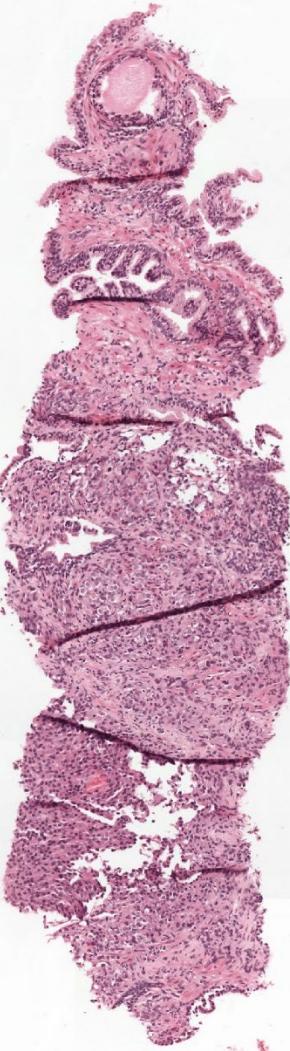
Collected prostate biopsies from 1271 patients

Grade	Training Set	Validation Set	Test Set
No cancer	777	200	271
3	1508	139	120
4	2102	138	134
5	329	42	100
Totals	4716	519	625



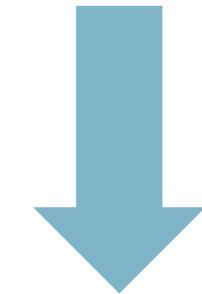
Input to system





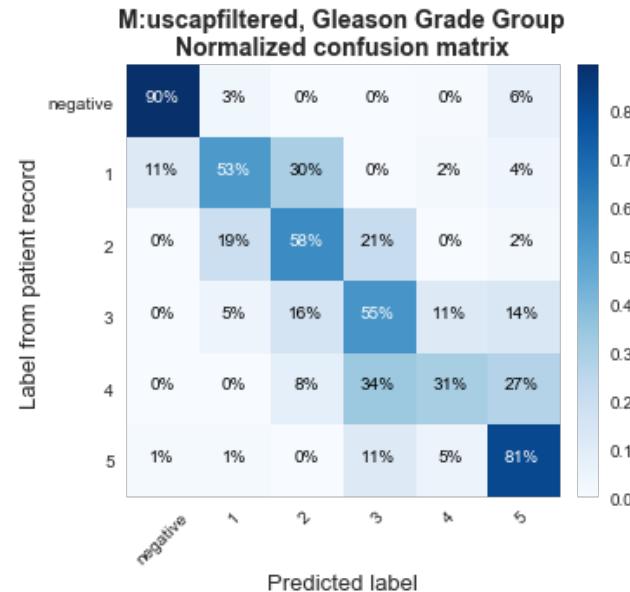
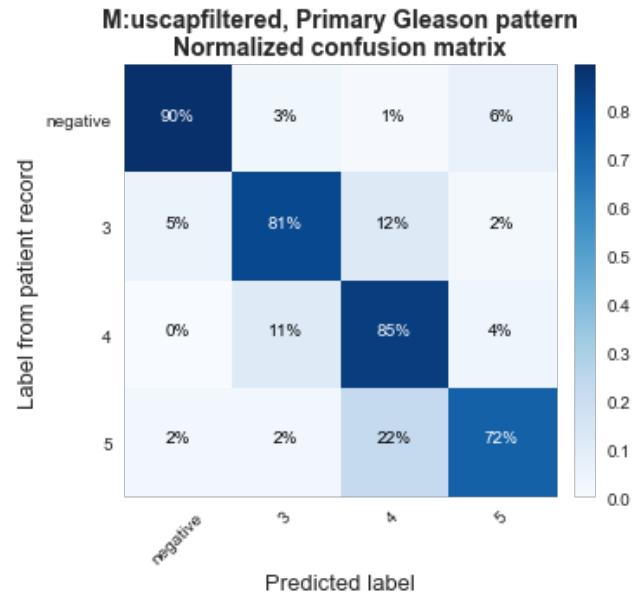
Count

20% benign
15% Gleason 4
65% Gleason 5



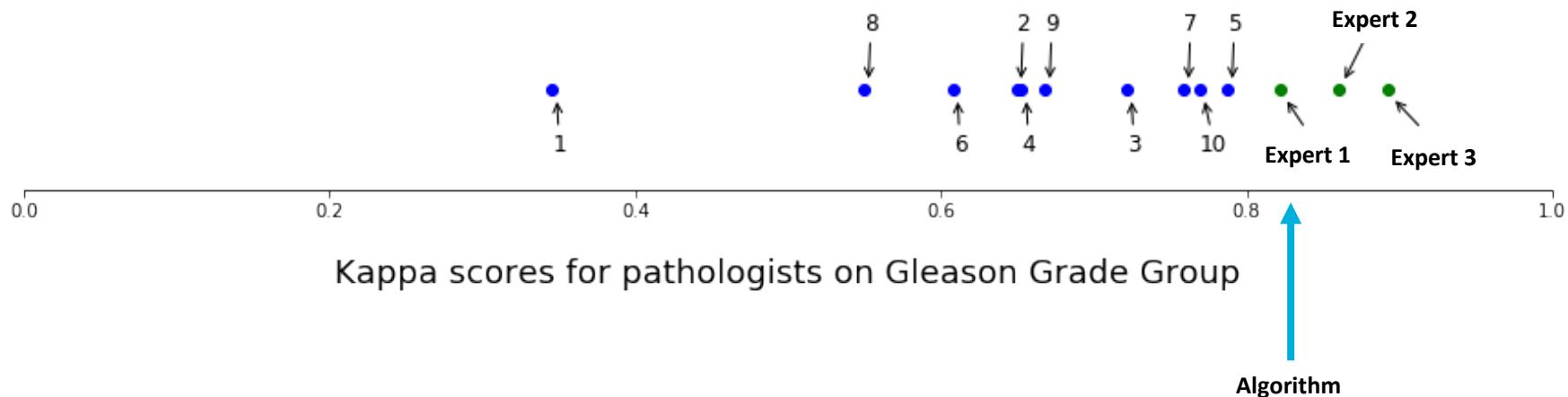
Gleason 5+4

Gleason Grading



Performance of model on GGG: acc 0.84, k 0.83

Observer experiment



The people who do all the work...

Scientific staff



Caner Mercan

Postdoctoral researcher



David Tellez

PhD student



Elke Loskamp-Huntink

Study manager



Hans Pinckaers

PhD student



Jasper Linmans

PhD student



John-Melle Bokhorst

PhD student

Technical staff



Karel Gerbrands

Research Software Engineer



Maud Wekking

Research technician



Merijn van Erp

Scientific programmer

Visiting researchers



Mart van Rijthoven

PhD student



Maschenka Balkenhol

Pathology resident and PhD student



Meyke Hermsen

PhD student



Oscar Geessink

PhD student



Péter Bándi

PhD student



Thomas de Bel

PhD student

Emiel Stoelinga

Master student

Koen Dercksen

Master student

Leander van Eekelen

Master student

Michel Kok

Master student

Patrick Sonsma

Master student

Faculty



Jeroen van der Laak

Associate professor/Group leader



Geert Litjens

Assistant professor



Francesco Ciompi

Assistant Professor



Institute for Health Sciences
Radboudumc

Radboudumc

Computational Pathology Group

The Computational Pathology Group develops, validates and deploys novel medical image analysis methods based on deep learning technology and focusing on computer-aided diagnosis. Application areas include diagnostics and prognostics of breast, prostate and colon cancer. We have rapidly expanded over the last few years, counting over 15 people today. Our group is among the international front runners in the field, witnessed for instance by our highly successful CAMELYON challenges. We have a strong translational focus, facilitated by our close collaboration with clinicians and industry.



Automated tumor detection

computationalpathology.eu

Automating kidney diagnostics

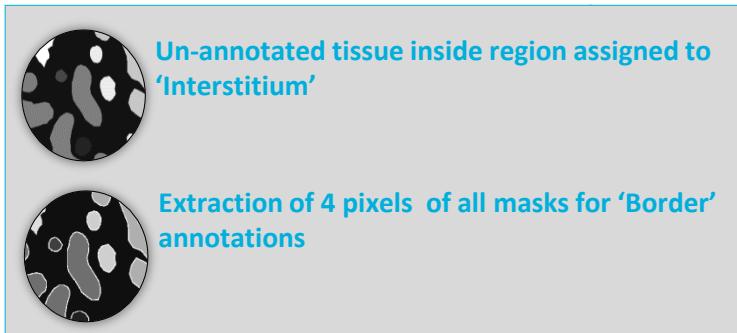
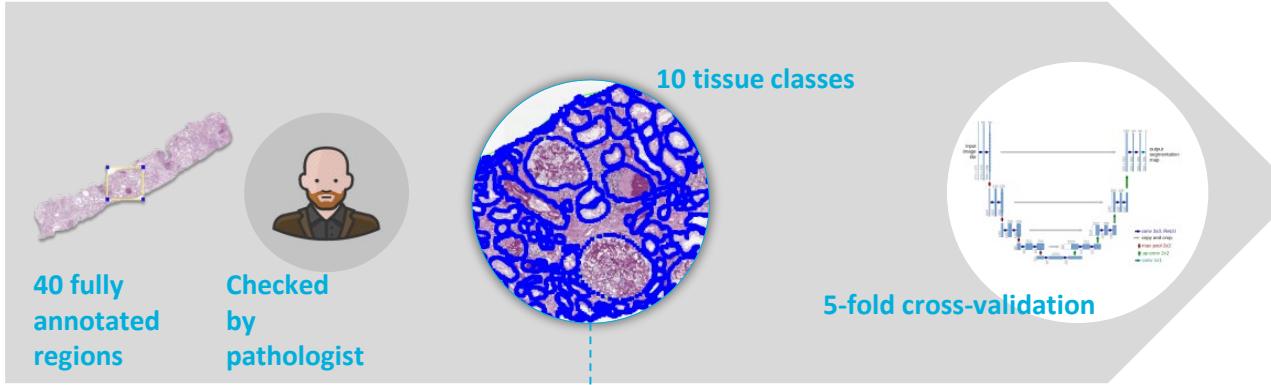
- Glomerular counting
- ct score vs % Atrophic tubuli

Quantitative criteria for tubular atrophy: ct score

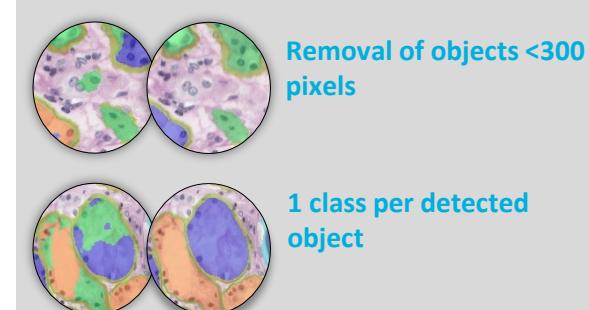
ct0	No tubular atrophy
ct1	Tubular atrophy involving up to 25% of the area of cortical tubules (mild tubular atrophy)
ct2	Tubular atrophy involving up to 26-50% of the area of cortical tubules (moderate tubular atrophy)
ct3	Tubular atrophy involving in >50% of the area of cortical tubules (severe tubular atrophy)



Automating kidney diagnostics

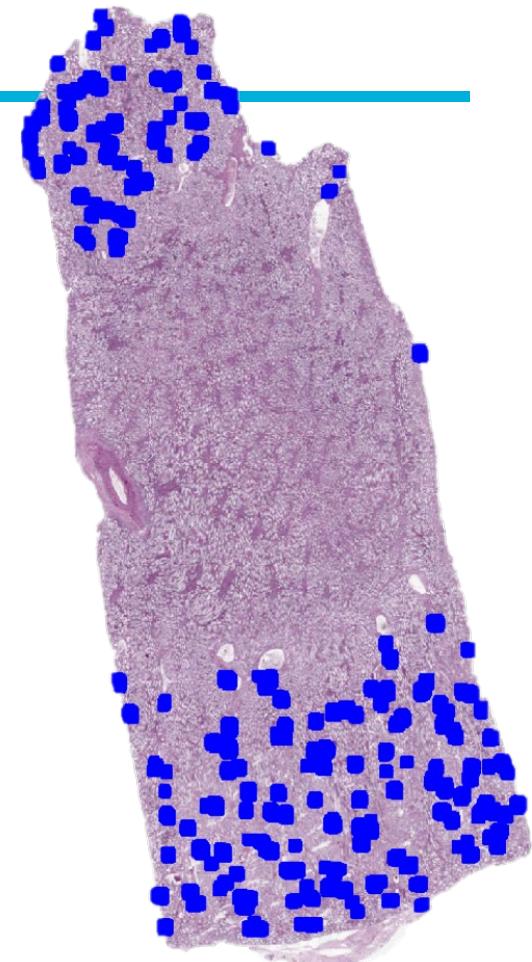


Post-processing

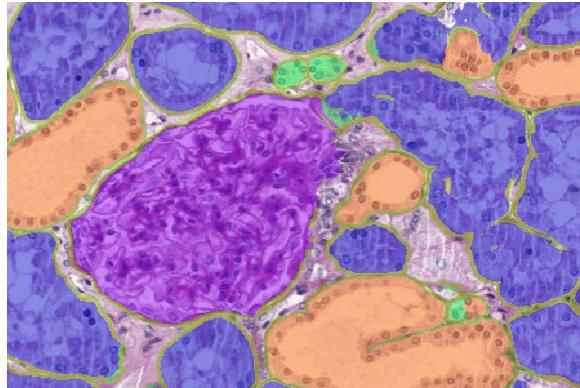
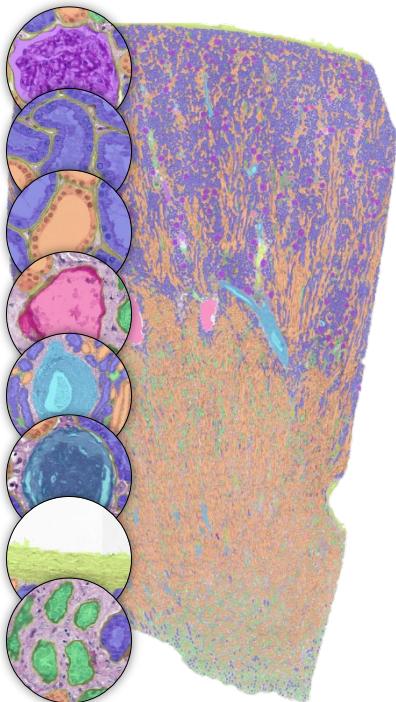


Automating kidney diagnostics

- Applied to 15 WSIs of large tumor nephrectomies
- All glomeruli annotated
 - *1747 Glomeruli and 72 Sclerotic glomeruli*

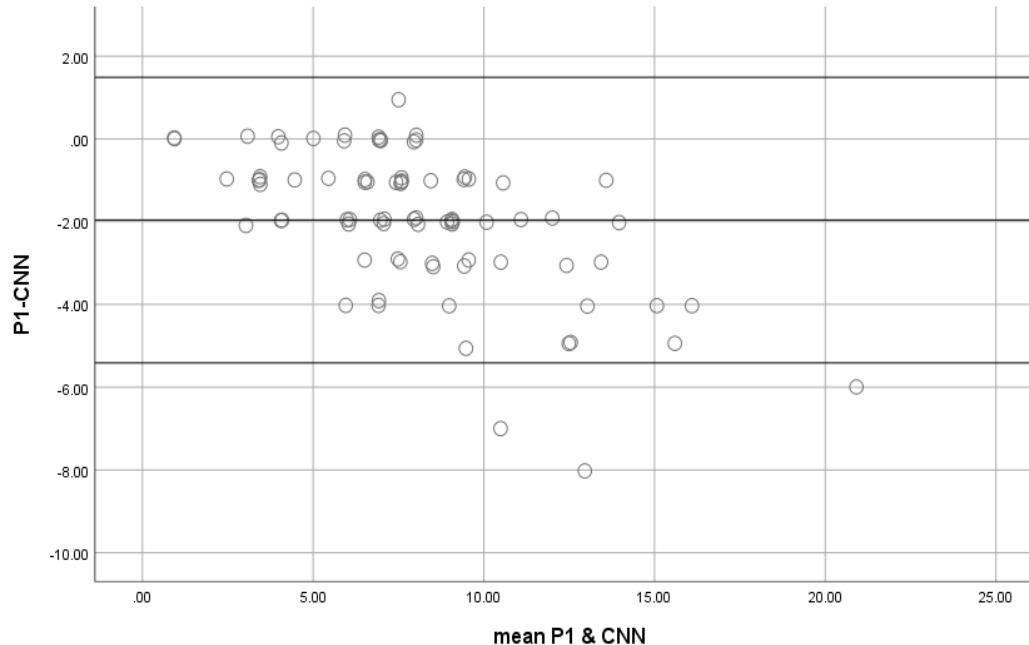


Automating kidney diagnostics



	TP	FP	FN
Glomeruli (n=1747)	93.4% (1632)	8.4 % (149)	6.6 % (115)
Sclerotic glomeruli (n=76)	76.4 % (55)	45.5 % (46)	23.6 % (17)
Total (n=1819)	92.7 % (1687)	10.4 % (192)	7.3 % (132)

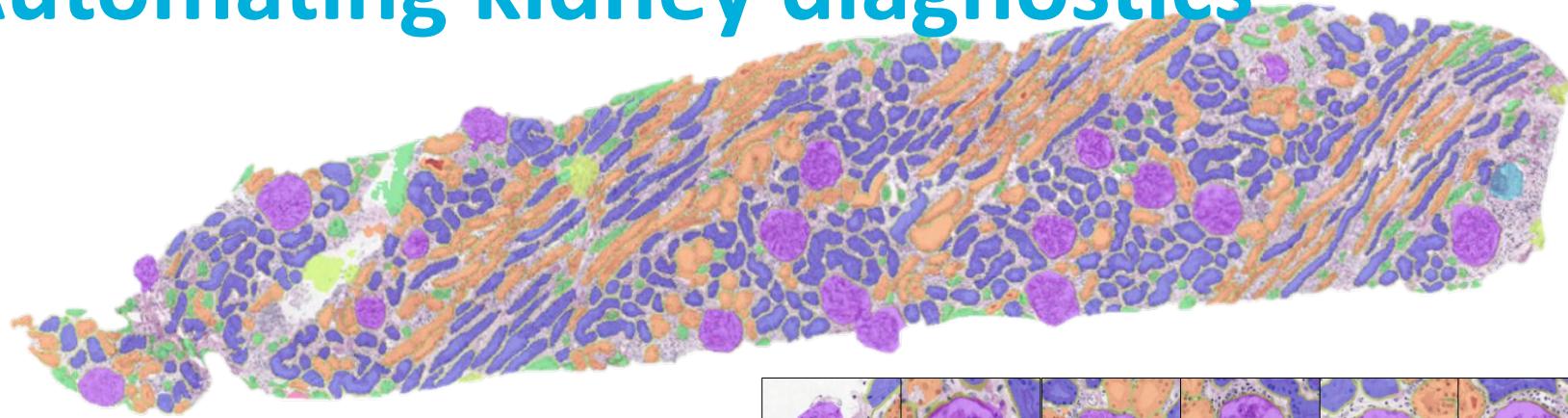
Automating kidney diagnostics



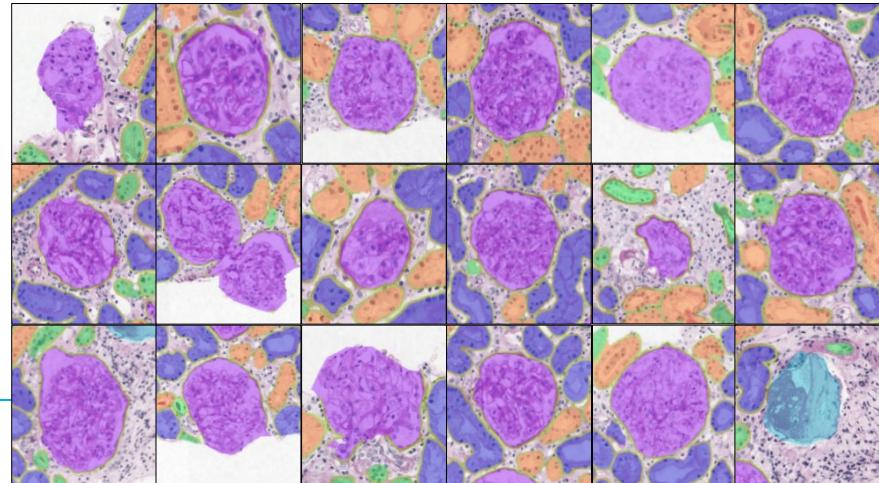
Inter-class correlation coefficient

	P1	P2	P3	CNN
P1		0.94	0.95	0.78
P2			0.95	0.85
P3				0.85
CNN				

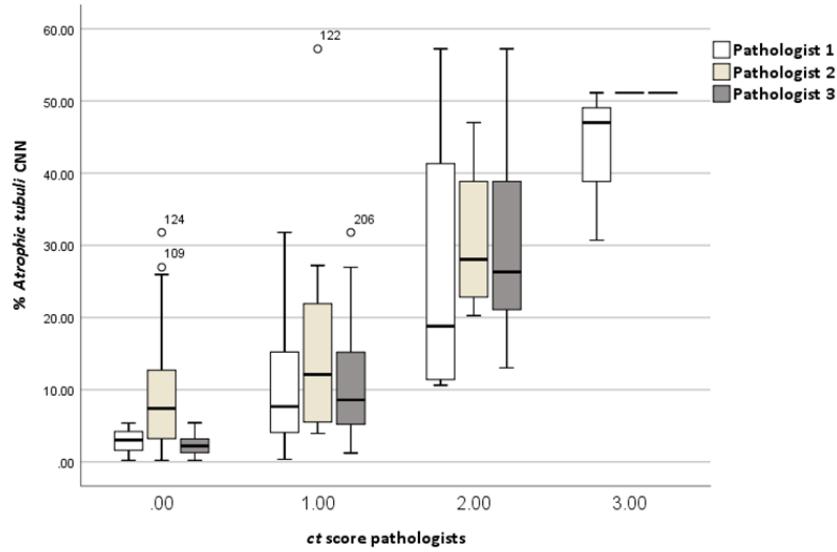
Automating kidney diagnostics



	No.
Pathologist 1	13
Pathologist 2	13
Pathologist 3	14
CNN Glomeruli	17
CNN Sclerotic glomeruli	1



Automating kidney diagnostics



Quantitative criteria for tubular atrophy: ct score

ct0	No tubular atrophy
ct1	Tubular atrophy involving up to 25% of the area of cortical tubules (mild tubular atrophy)
ct2	Tubular atrophy involving up to 26-50% of the area of cortical tubules (moderate tubular atrophy)
ct3	Tubular atrophy involving >50% of the area of cortical tubules (severe tubular atrophy)

Bonferroni analysis

	0	1	2	3
0	0.24	<0.001	<0.001	
1		<0.001	<0.001	
2			<0.01	
3				

Weighted kappa

	P1	P2	P3
P1		0.13	0.34
P2			0.20
P3			