

deepG tutorial

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Introduction

The deepG library can be used for applying deep learning on genomic data. The library supports creating neural network architecture, automation of data preprocessing (data generator), network training, inference and visualizing feature importances (integrated gradients).

Create a model

deepG supports three functions to create a keras model.

create_model_lstm_cnn

The architecture of this model is k * LSTM, m * CNN and n * dense layers, where $k, m \geq 0$ and $n \geq 1$.

The user can choose the size of the individual LSTM, CNN and Dense layers and add additional features to each layer; for example the LSTM layer may be bidirectional (runs input in two ways) or stateful (considers dependencies between batches).

The last dense layer has a softmax activation and determines how many targets we want to predict. This output gives a vector of probabilities, i.e. the sum of the vector is 1 and each entry is a probability for one class.

The following implementation creates a model with 3 CNN layer (+ batch normalization), 1 LSTM and 1 dense layer.

```
model <- create_model_lstm_cnn(  
  maxlen = 500, # number of nucleotides processed in one sample  
  layer_lstm = c(32), # number of LSTM cells  
  layer_dense = c(4), # number of neurons in last layer (4 targets: A,C,G,T)  
  vocabulary.size = 4, # input vocabulary has size 4 (A,C,G,T)  
  kernel_size = c(12, 12, 12), # size of individual CNN windows for each layer  
  filters = c(32, 64, 64), # number of CNN filters per layer  
  pool_size = c(3, 3, 3) # size of max pooling per layer  
)
```

```
## Model: "model"
```

```
## -----  
## Layer (type)                Output Shape                Param #  
## -----  
## input_1 (InputLayer)        [(None, 500, 4)]           0  
## -----  
## conv1d (Conv1D)             (None, 500, 32)            1568  
## -----  
## max_pooling1d (MaxPooling1D) (None, 166, 32)            0  
## -----  
## batch_normalization (BatchNormaliza (None, 166, 32)            128  
## -----  
## conv1d_1 (Conv1D)           (None, 166, 64)            24640  
## -----  
## batch_normalization_1 (BatchNormalali (None, 166, 64)            256  
## -----  
## max_pooling1d_1 (MaxPooling1D) (None, 55, 64)            0  
## -----  
## conv1d_2 (Conv1D)           (None, 55, 64)            49216  
## -----  
## batch_normalization_2 (BatchNormalali (None, 55, 64)            256  
## -----  
## max_pooling1d_2 (MaxPooling1D) (None, 18, 64)            0  
## -----  
## lstm (LSTM)                 (None, 32)                 12416  
## -----  
## dense (Dense)               (None, 4)                  132  
## -----  
## Total params: 88,612  
## Trainable params: 88,292  
## Non-trainable params: 320
```

```
## -----
```

The model expects an input with dimensions (NULL (batch size), maxlen, vocabulary size) and a target with dimension (NULL (batch size), number of targets). Maxlen specifies the length of the input sequence.

```
batch_size <- 3
maxlen <- 500
vocabulary.size <- 4
input <- array(rnorm(maxlen * batch_size * vocabulary.size),
              dim = c(batch_size, maxlen, vocabulary.size))
pred <- predict(model, input) # make a prediction with random data
dim(pred)
```

```
## [1] 3 4
```

```
colnames(pred) <- c("A", "C", "G", "T")
pred # prediction for initial random weights
```

```
##           A           C           G           T
## [1,] 0.2418842 0.2160256 0.3328353 0.2092548
## [2,] 0.2363858 0.2228332 0.3235803 0.2172006
## [3,] 0.2369523 0.2181571 0.3296198 0.2152708
```

create_model_lstm_cnn_target_middle

This architecture is closely related to `create_model_lstm_cnn_target` with the main difference that the model has two input layers (provided `label_input = NULL`).

```
model <- create_model_lstm_cnn_target_middle(
  maxlen = 500,
  layer_lstm = c(32),
  layer_dense = c(4),
  vocabulary.size = 4,
  kernel_size = c(12, 12, 12),
  filters = c(32, 64, 64),
  pool_size = c(3, 3, 3)
)
```

```
## Model: "model_1"
## -----
## Layer (type)           Output Shape      Param #   Connected to
## =====
## input_2 (InputLayer)    [(None, 250, 4)]  0
## -----
## input_3 (InputLayer)    [(None, 250, 4)]  0
## -----
## conv1d_3 (Conv1D)        (None, 250, 32)   1568      input_2[0][0]
## -----
## conv1d_6 (Conv1D)        (None, 250, 32)   1568      input_3[0][0]
## -----
## max_pooling1d_3 (MaxPool1d) (None, 83, 32)    0          conv1d_3[0][0]
```

```

## -----
## max_pooling1d_6 (MaxPooli (None, 83, 32)    0      conv1d_6[0][0]
## -----
## batch_normalization_3 (Ba (None, 83, 32)    128     max_pooling1d_3[0][0]
## -----
## batch_normalization_6 (Ba (None, 83, 32)    128     max_pooling1d_6[0][0]
## -----
## conv1d_4 (Conv1D)          (None, 83, 64)    24640    batch_normalization_3[0][0]
## -----
## conv1d_7 (Conv1D)          (None, 83, 64)    24640    batch_normalization_6[0][0]
## -----
## max_pooling1d_4 (MaxPooli (None, 27, 64)    0      conv1d_4[0][0]
## -----
## max_pooling1d_7 (MaxPooli (None, 27, 64)    0      conv1d_7[0][0]
## -----
## batch_normalization_4 (Ba (None, 27, 64)    256     max_pooling1d_4[0][0]
## -----
## batch_normalization_7 (Ba (None, 27, 64)    256     max_pooling1d_7[0][0]
## -----
## conv1d_5 (Conv1D)          (None, 27, 64)    49216    batch_normalization_4[0][0]
## -----
## conv1d_8 (Conv1D)          (None, 27, 64)    49216    batch_normalization_7[0][0]
## -----
## max_pooling1d_5 (MaxPooli (None, 9, 64)     0      conv1d_5[0][0]
## -----
## max_pooling1d_8 (MaxPooli (None, 9, 64)     0      conv1d_8[0][0]
## -----
## batch_normalization_5 (Ba (None, 9, 64)     256     max_pooling1d_5[0][0]
## -----
## batch_normalization_8 (Ba (None, 9, 64)     256     max_pooling1d_8[0][0]
## -----
## lstm_1 (LSTM)              (None, 32)        12416    batch_normalization_5[0][0]
## -----
## lstm_2 (LSTM)              (None, 32)        12416    batch_normalization_8[0][0]
## -----
## concatenate (Concatenate) (None, 64)        0      lstm_1[0][0]
##                                     lstm_2[0][0]
## -----
## dense_1 (Dense)            (None, 4)         260     concatenate[0][0]
## =====
## Total params: 177,220
## Trainable params: 176,580
## Non-trainable params: 640
## -----

```

This architecture can be used to predict a character in the middle of a sequence. For example sequence: ACCG**T**GGAA

then the first input should correspond to ACCG, the second input to GGAA and T to the target. This can be used to combine the 2 tasks

1. predict T given ACCG
2. predict T given AAGG (note reversed order of input)

in one model.

create_model_wavenet

This model uses causal dilated convolution layers, which is suitable to handle long sequences. The original paper can be found [here](#)

```
model <- create_model_wavenet(filters = 16, kernel_size = 2, residual_blocks = 2^(2:4),
                              maxlen = 500, input_tensor = NULL, initial_kernel_size = 32,
                              initial_filters = 32, output_channels = 4,
                              output_activation = "softmax", solver = "adam",
                              learning.rate = 0.001, compile = TRUE)
```

model

```
## Model
## Model: "model_2"
## -----
## Layer (type)           Output Shape      Param #   Connected to
## -----
## input_4 (InputLayer)   [(None, 500, 4)]  0
## -----
## conv1d_9 (Conv1D)      (None, 500, 32)   4096      input_4[0][0]
## -----
## r_layer (RLayer)       [(None, 500, 32), 0
## -----
## r_layer_1 (RLayer)     [(None, 500, 32), 0
## -----
## r_layer_2 (RLayer)     [(None, 500, 32), 0
## -----
## add (Add)              (None, 500, 32)   0          r_layer[0][1]
##                               r_layer_1[0][1]
##                               r_layer_2[0][1]
## -----
## activation (Activation) (None, 500, 32)   0          add[0][0]
## -----
## conv1d_11 (Conv1D)     (None, 500, 16)   512        activation[0][0]
## -----
## conv1d_10 (Conv1D)     (None, 500, 4)    68         conv1d_11[0][0]
## =====
## Total params: 4,676
## Trainable params: 4,676
## Non-trainable params: 0
## -----
```

The model expects an input and output of dimension (batch size, maxlen, vocabulary.size). The target sequence should be equal to input sequence shifted by one position. For example, given a sequence ACCGGTC and maxlen = 6, the input should correspond to ACCGGT and target to CCGGTC.

Training

Preparing the data

Input data must be files in FASTA or FASTQ format and file names must have .fasta or .fastq ending; otherwise files will be ignored. All training and validation data should each be in one folder. deepG uses a data generator to iterate over files in train/validation folder.

Before we train our model, we have to decide what our training objective is. It can be either a language model or label classification.

```
path <- "/home/rmreches/tutorial"
path_16S_train <- file.path(path, "16s/train")
path_16S_validation <- file.path(path, "16s/validation")
path_bacteria_train <- file.path(path, "bacteria/train")
path_bacteria_validation <- file.path(path, "bacteria/validation")

checkpoint_path <- file.path(path, "checkpoints")
tensorboard.log <- file.path(path, "tensorboard")
dir_path <- file.path(path, "outputs")
if (!dir.exists(checkpoint_path)) dir.create(checkpoint_path)
if (!dir.exists(tensorboard.log)) dir.create(tensorboard.log)
if (!dir.exists(dir_path)) dir.create(dir_path)
```

Language model

With language model, we mean a model that predicts a character in a sequence. The target can be at the end of the sequence, for example

ACGTCAG

or in the middle

ACGTCAG

Language model for 16S (predict next character)

Say we want to predict the next character in a sequence given the last 500 characters and our text consists of the letters A,C,G,T. First we have to create a model. We may use a model with 1 LSTM, 3 CNN and 1 dense layer for predictions.

```
model <- create_model_lstm_cnn(
  maxlen = 500,
  layer_lstm = c(32),
  layer_dense = c(4),
  vocabulary.size = 4,
  kernel_size = c(12, 12, 12),
  filters = c(32, 64, 64),
  pool_size = c(3, 3, 3),
  learning.rate = 0.001
)
```

```
## Model: "model_3"
```

```
## -----
```

## Layer (type)	Output Shape	Param #
## =====		
## input_5 (InputLayer)	[(None, 500, 4)]	0
## -----		
## conv1d_12 (Conv1D)	(None, 500, 32)	1568
## -----		
## max_pooling1d_9 (MaxPooling1D)	(None, 166, 32)	0
## -----		
## batch_normalization_9 (BatchNormali	(None, 166, 32)	128
## -----		
## conv1d_13 (Conv1D)	(None, 166, 64)	24640
## -----		
## batch_normalization_10 (BatchNormal	(None, 166, 64)	256
## -----		
## max_pooling1d_10 (MaxPooling1D)	(None, 55, 64)	0
## -----		
## conv1d_14 (Conv1D)	(None, 55, 64)	49216
## -----		
## batch_normalization_11 (BatchNormal	(None, 55, 64)	256
## -----		
## max_pooling1d_11 (MaxPooling1D)	(None, 18, 64)	0
## -----		
## lstm_3 (LSTM)	(None, 32)	12416
## -----		
## dense_2 (Dense)	(None, 4)	132
## =====		
## Total params: 88,612		
## Trainable params: 88,292		
## Non-trainable params: 320		
## -----		

Next we have to specify the location of our training and validation data and the output format of the data generator

```

trainNetwork(train_type = "lm", # train a language model
             model = model,
             path = path_16S_train, # location of training data
             path_val = path_16S_validation, # location of validation data
             checkpoint_path = checkpoint_path,
             tensorboard_log = tensorboard_log,
             validation_split = 0.2, # use 20% of samples for validation compared to train size
             run_name = "lm_16S_target_right",
             batch_size = 256,
             epochs = 4,
             steps_per_epoch = 10, # 1 epoch = 10 batches
             step = 500, # take a sample every 500 steps
             output = list(none = FALSE,
                           checkpoints = TRUE,
                           tensorboard = TRUE,
                           log = FALSE,
                           serialize_model = FALSE,
                           full_model = FALSE
                           ),
             tb_images = TRUE,

```

```
output_format = "target_right" # predict target at end of sequence
)
```

```
## Trained on 10 samples (batch_size=NULL, epochs=4)
## Final epoch (plot to see history):
##      loss: 0.1063
##      acc: 0.9934
##      f1: Inf
## val_loss: 0.5382
## val_acc: 0.8145
## val_f1: Inf
##      lr: 0.001
```

```
tensorflow::tensorboard(tensorboard.log)
```

```
## Started TensorBoard at http://127.0.0.1:4527
```

Predict character in middle of sequence

If we want to predict a character in the middle of a sequence and use LSTM layers, we should split our input into two layers. One layer handles the sequence before and one the input after the target. If, for example

sequence: ACCG**T**GGAA

then first input corresponds to ACCG and second to AAGG. We may create a model with two input layers using the `create_model_cnn_lstm_target_middle`

```
model <- create_model_lstm_cnn_target_middle(
  maxlen = 500,
  layer_lstm = c(32),
  layer_dense = c(4),
  vocabulary.size = 4,
  kernel_size = c(12, 12, 12),
  filters = c(32, 64, 64),
  pool_size = c(3, 3, 3),
  learning.rate = 0.001
)
```

```
## Model: "model_4"
```

```
## -----
## Layer (type)           Output Shape      Param #   Connected to
## =====
## input_6 (InputLayer)   [(None, 250, 4)]  0
## -----
## input_7 (InputLayer)   [(None, 250, 4)]  0
## -----
## conv1d_15 (Conv1D)      (None, 250, 32)   1568      input_6[0][0]
## -----
## conv1d_18 (Conv1D)      (None, 250, 32)   1568      input_7[0][0]
## -----
## max_pooling1d_12 (MaxPool (None, 83, 32)    0          conv1d_15[0][0]
## -----
```



```

## max_pooling1d_15 (MaxPool (None, 83, 32)    0      conv1d_18[0] [0]
## -----
## batch_normalization_12 (B (None, 83, 32)    128     max_pooling1d_12[0] [0]
## -----
## batch_normalization_15 (B (None, 83, 32)    128     max_pooling1d_15[0] [0]
## -----
## conv1d_16 (Conv1D)          (None, 83, 64)    24640    batch_normalization_12[0] [0]
## -----
## conv1d_19 (Conv1D)          (None, 83, 64)    24640    batch_normalization_15[0] [0]
## -----
## max_pooling1d_13 (MaxPool (None, 27, 64)    0      conv1d_16[0] [0]
## -----
## max_pooling1d_16 (MaxPool (None, 27, 64)    0      conv1d_19[0] [0]
## -----
## batch_normalization_13 (B (None, 27, 64)    256     max_pooling1d_13[0] [0]
## -----
## batch_normalization_16 (B (None, 27, 64)    256     max_pooling1d_16[0] [0]
## -----
## conv1d_17 (Conv1D)          (None, 27, 64)    49216    batch_normalization_13[0] [0]
## -----
## conv1d_20 (Conv1D)          (None, 27, 64)    49216    batch_normalization_16[0] [0]
## -----
## max_pooling1d_14 (MaxPool (None, 9, 64)     0      conv1d_17[0] [0]
## -----
## max_pooling1d_17 (MaxPool (None, 9, 64)     0      conv1d_20[0] [0]
## -----
## batch_normalization_14 (B (None, 9, 64)     256     max_pooling1d_14[0] [0]
## -----
## batch_normalization_17 (B (None, 9, 64)     256     max_pooling1d_17[0] [0]
## -----
## lstm_4 (LSTM)               (None, 32)        12416    batch_normalization_14[0] [0]
## -----
## lstm_5 (LSTM)               (None, 32)        12416    batch_normalization_17[0] [0]
## -----
## concatenate_1 (Concatenat (None, 64)        0      lstm_4[0] [0]
##                                lstm_5[0] [0]
## -----
## dense_3 (Dense)             (None, 4)         260     concatenate_1[0] [0]
## =====
## Total params: 177,220
## Trainable params: 176,580
## Non-trainable params: 640
## -----

```

The `trainNetwork` call is identical to the previous model, except we have to change the output format of the generator by setting `output_format = "target_middle_lstm"`. This reverses the order of the sequence after the target.

```

trainNetwork(train_type = "lm", # train a language model
             model = model,
             path = path_16S_train, # location of training data
             path.val = path_16S_validation, # location of validation data
             checkpoint_path = checkpoint_path,
             tensorboard.log = tensorboard.log,

```

```

validation.split = 0.2, # use 20% of samples for validation compared to train size
run.name = "lm_16S_target_middle_lstm",
batch.size = 256,
epochs = 4,
steps.per.epoch = 10, # 1 epoch = 10 batches
step = 500, # take a sample every 500 steps
output = list(none = FALSE,
              checkpoints = TRUE,
              tensorboard = TRUE,
              log = FALSE,
              serialize_model = FALSE,
              full_model = FALSE
            ),
tb_images = TRUE,
output_format = "target_middle_lstm" # predict character in middle of sequence
)

```

```

## Trained on 10 samples (batch_size=NULL, epochs=4)
## Final epoch (plot to see history):
##      loss: 0.04586
##      acc: 0.9988
##      f1: Inf
## val_loss: 0.3642
## val_acc: 0.873
## val_f1: Inf
##      lr: 0.001

```

Label classification

With label classification, we describe the task of mapping a label to a sequence. For example: given the sequence ACGACCG, does the sequence belong to a viral or bacterial genome?

deepG offers two options to map a label to a sequence

1. the label gets read from the fasta header
2. files from every class are in separate folders

Label by folder

We put all data from one class into separate folders. In the following example, we want to classify if a sequence belongs to 16s or bacterial genome. We have to put all 16s/bacteria files into their own folder. In this case the `path` and `path.val` arguments should be vectors, where each entry is the path to one class.

```

model <- create_model_lstm_cnn(
  maxlen = 500,
  layer_lstm = c(32),
  layer_dense = c(2), # predict two classes
  vocabulary.size = 4,
  kernel_size = c(12, 12, 12),
  filters = c(32, 64, 64),
  pool_size = c(3, 3, 3),

```

```
learning.rate = 0.001
)
```

```
## Model: "model_5"
##
## -----
## Layer (type)                Output Shape                Param #
## -----
## input_8 (InputLayer)        [(None, 500, 4)]           0
## -----
## conv1d_21 (Conv1D)           (None, 500, 32)            1568
## -----
## max_pooling1d_18 (MaxPooling1D) (None, 166, 32)           0
## -----
## batch_normalization_18 (BatchNormal (None, 166, 32)           128
## -----
## conv1d_22 (Conv1D)           (None, 166, 64)            24640
## -----
## batch_normalization_19 (BatchNormal (None, 166, 64)           256
## -----
## max_pooling1d_19 (MaxPooling1D) (None, 55, 64)            0
## -----
## conv1d_23 (Conv1D)           (None, 55, 64)            49216
## -----
## batch_normalization_20 (BatchNormal (None, 55, 64)           256
## -----
## max_pooling1d_20 (MaxPooling1D) (None, 18, 64)            0
## -----
## lstm_6 (LSTM)                (None, 32)                 12416
## -----
## dense_4 (Dense)              (None, 2)                  66
## =====
## Total params: 88,546
## Trainable params: 88,226
## Non-trainable params: 320
## -----
```

```
trainNetwork(train_type = "label_folder", # reading label from folder
             model = model,
             path = c(path_16S_train, # note that path has two entries
                     path_bacteria_train),
             path.val = c(path_16S_validation,
                         path_bacteria_validation),
             checkpoint_path = checkpoint_path,
             tensorboard.log = tensorboard.log,
             validation.split = 0.2,
             run.name = "16S_vs_bacteria",
             batch.size = 256, # half of batch is 16s and other half bacteria data
             epochs = 6,
             steps.per.epoch = 25,
             step = 500,
             labelVocabulary = c("16s", "bacteria"), # label names
             output = list(none = FALSE,
                          checkpoints = TRUE,
```

```

        tensorboard = TRUE,
        log = FALSE,
        serialize_model = FALSE,
        full_model = FALSE
    ),
    tb_images = TRUE,
    proportion_per_file = c(1, 0.05) # randomly select 5% of bacteria file
)

```

```

## Trained on 25 samples (batch_size=NULL, epochs=6)
## Final epoch (plot to see history):
##      loss: 0.004071
##      acc: 0.9991
##      f1: 0.9991
## val_loss: 0.05052
## val_acc: 0.9852
## val_f1: 0.9849
##      lr: 0.001

```

Inference

Once we have trained a model, we may use the model to get the activations of a certain layer and write the states to an h5 file. In the following example we use the binary model trained to classify 16S/bacteria data.

```
print(model)
```

```

## Model
## Model: "model_5"
## -----
## Layer (type)                Output Shape                Param #
## =====
## input_8 (InputLayer)        [(None, 500, 4)]            0
## -----
## conv1d_21 (Conv1D)          (None, 500, 32)             1568
## -----
## max_pooling1d_18 (MaxPooling1D) (None, 166, 32)            0
## -----
## batch_normalization_18 (BatchNormal (None, 166, 32)            128
## -----
## conv1d_22 (Conv1D)          (None, 166, 64)             24640
## -----
## batch_normalization_19 (BatchNormal (None, 166, 64)            256
## -----
## max_pooling1d_19 (MaxPooling1D) (None, 55, 64)              0
## -----
## conv1d_23 (Conv1D)          (None, 55, 64)             49216
## -----
## batch_normalization_20 (BatchNormal (None, 55, 64)            256
## -----
## max_pooling1d_20 (MaxPooling1D) (None, 18, 64)              0
## -----

```

```
## lstm_6 (LSTM)                                (None, 32)                12416
## -----
## dense_4 (Dense)                              (None, 2)                  66
## =====
## Total params: 88,546
## Trainable params: 88,226
## Non-trainable params: 320
## -----
```

```
num_layers <- length(model$get_config()$layers)
layer_name <- model$get_config()$layers[[num_layers]]$name
cat("get output at layer", layer_name)
```

```
## get output at layer dense_4
```

```
fasta.path <- list.files(path_16S_validation, full.names = TRUE)[1] # make predictions for 16S file
fasta.file <- microseq::readFasta(fasta.path)
head(fasta.file)
```

```
## # A tibble: 1 x 2
##   Header                      Sequence
##   <chr>                      <chr>
## 1 16S_rRNA::CP015410.2:15033~ TATGAGAGTTTGATCCTGGCTCAGGACGAACGCTGGCGGCGTGCCTAAT~
```

```
sequence <- fasta.file$Sequence[1]
filename <- file.path(dir_path, "states.h5")
```

```
if (!file.exists(filename)) {
  writeStates(
    model = model,
    layer_name = layer_name,
    sequence = sequence,
    round_digits = 4,
    filename = filename,
    batch.size = 10,
    mode = "lm")
}
```

```
## Computing output for model at layer dense_4
```

```
## Model
## Model: "model_6"
## -----
## Layer (type)                Output Shape                Param #
## =====
## input_8 (InputLayer)        [(None, 500, 4)]            0
## -----
## conv1d_21 (Conv1D)          (None, 500, 32)             1568
## -----
## max_pooling1d_18 (MaxPooling1D) (None, 166, 32)            0
## -----
## batch_normalization_18 (BatchNormal (None, 166, 32)            128
## -----
```

```
## conv1d_22 (Conv1D)                (None, 166, 64)                24640
## -----
## batch_normalization_19 (BatchNormal (None, 166, 64)                256
## -----
## max_pooling1d_19 (MaxPooling1D)    (None, 55, 64)                0
## -----
## conv1d_23 (Conv1D)                (None, 55, 64)                49216
## -----
## batch_normalization_20 (BatchNormal (None, 55, 64)                256
## -----
## max_pooling1d_20 (MaxPooling1D)    (None, 18, 64)                0
## -----
## lstm_6 (LSTM)                     (None, 32)                    12416
## -----
## dense_4 (Dense)                   (None, 2)                     66
## =====
## Total params: 88,546
## Trainable params: 88,226
## Non-trainable params: 320
## -----
```

We can access the h5 file as follows

```
states <- readRowsFromH5(h5_path = filename, complete = TRUE)
```

```
## states matrix has 1058 rows and 2 columns
```

```
colnames(states) <- c("16S", "bacteria")
head(states)
```

```
##          16S bacteria
## [1,] 0.9995    5e-04
## [2,] 0.9996    4e-04
## [3,] 0.9997    3e-04
## [4,] 0.9993    7e-04
## [5,] 0.9995    5e-04
## [6,] 0.9995    5e-04
```

The matrix shows the models confidence in its predictions. Every row corresponds to one sample. If the value in the 16s column is > 0.500 , the model will classify the sample as 16s.

Inference II

We can use our trained model to detect 16S sequences in a bacterial genome. First, we search for the true rRNA region in the corresponding gff file.

```
fasta.path<- file.path(path, "E_faecalis.fasta")
gff.file <- file.path(path, "E_faecalis.gff")
gff.data <- rtracklayer::readGFF(gff.file, version = 0,
                                columns = NULL, tags = NULL, filter = NULL, nrows = -1,
                                raw_data = FALSE)
```

```
rRNA_index <- stringr::str_detect(gff.data$product, "^16S ribosomal") & (gff.data$strand == "+")
start <- gff.data[rRNA_index, "start"]
end <- gff.data[rRNA_index, "end"]
start; end
```

```
## [1] 2189670 2933745
```

```
## [1] 2191227 2935302
```

We iterate over the bacteria file and make a predictions every 100 steps

```
fasta.file <- microseq::readFasta(fasta.path)
sequence <- fasta.file$Sequence[1]
filename <- file.path(dir_path, "bacteria_states.h5")

if (!file.exists(filename)) {
  writeStates(
    model = model,
    layer_name = layer_name,
    sequence = sequence,
    round_digits = 4,
    filename = filename,
    batch.size = 500,
    step = 100)
}
```

```
## Computing output for model at layer dense_4
```

```
## Model
```

```
## Model: "model_7"
```

```
##
```

## Layer (type)	Output Shape	Param #
## input_8 (InputLayer)	[(None, 500, 4)]	0
## conv1d_21 (Conv1D)	(None, 500, 32)	1568
## max_pooling1d_18 (MaxPooling1D)	(None, 166, 32)	0
## batch_normalization_18 (BatchNormal	(None, 166, 32)	128
## conv1d_22 (Conv1D)	(None, 166, 64)	24640
## batch_normalization_19 (BatchNormal	(None, 166, 64)	256
## max_pooling1d_19 (MaxPooling1D)	(None, 55, 64)	0
## conv1d_23 (Conv1D)	(None, 55, 64)	49216
## batch_normalization_20 (BatchNormal	(None, 55, 64)	256
## max_pooling1d_20 (MaxPooling1D)	(None, 18, 64)	0

```
## lstm_6 (LSTM)                                (None, 32)                                12416
## -----
## dense_4 (Dense)                              (None, 2)                                66
## =====
## Total params: 88,546
## Trainable params: 88,226
## Non-trainable params: 320
## -----

states <- readRowsFromH5(h5_path = filename, complete = TRUE, getTargetPositions = TRUE)
```

```
## states matrix has 30252 rows and 2 columns
```

```
pred <- states[[1]]
position <- states[[2]] - 1
df <- cbind(pred, position) %>% as.data.frame()
colnames(df) <- c("conf_16S", "conf_bacteria", "seq_end")
head(df)
```

```
##   conf_16S conf_bacteria seq_end
## 1  0.0027      0.9973     500
## 2  0.0005      0.9995     600
## 3  0.0006      0.9994     700
## 4  0.0010      0.9990     800
## 5  0.0005      0.9995     900
## 6  0.0007      0.9993    1000
```

```
index_16S_pred <- df[, 1] > 0.5
df_16S <- df[index_16S_pred, ]
df_16S
```

```
##   conf_16S conf_bacteria seq_end
## 49  0.5550      0.4450     5300
## 448  0.9753      0.0247    45200
## 1601  0.7632      0.2368   160500
## 2189  0.8609      0.1391   219300
## 2342  0.5041      0.4959   234600
## 2441  0.5160      0.4840   244500
## 3234  0.9874      0.0126   323800
## 3723  0.9799      0.0201   372700
## 3895  0.7928      0.2072   389900
## 3943  0.5679      0.4321   394700
## 11328 0.6072      0.3928  1133200
## 11576 0.9819      0.0181  1158000
## 11577 0.6458      0.3542  1158100
## 11578 0.5642      0.4358  1158200
## 12083 0.6474      0.3526  1208700
## 12741 0.7449      0.2551  1274500
## 12795 0.9168      0.0832  1279900
## 12810 0.6043      0.3957  1281400
## 12817 0.5382      0.4618  1282100
## 15366 0.8659      0.1341  1537000
```


## 18514	0.6147	0.3853	1851800
## 18915	0.8063	0.1937	1891900
## 18942	0.8041	0.1959	1894600
## 18944	0.9505	0.0495	1894800
## 19111	0.7423	0.2577	1911500
## 19220	0.5358	0.4642	1922400
## 19340	0.5661	0.4339	1934400
## 19373	0.6013	0.3987	1937700
## 19460	0.6366	0.3634	1946400
## 19461	0.6180	0.3820	1946500
## 19462	0.9567	0.0433	1946600
## 19504	0.7333	0.2667	1950800
## 19505	0.6498	0.3502	1950900
## 19593	0.6163	0.3837	1959700
## 20050	0.6790	0.3210	2005400
## 20052	0.8426	0.1574	2005600
## 20055	0.9363	0.0637	2005900
## 20056	0.9832	0.0168	2006000
## 20059	0.8733	0.1267	2006300
## 20061	0.9587	0.0413	2006500
## 20062	0.9788	0.0212	2006600
## 20063	0.9648	0.0352	2006700
## 20065	0.9047	0.0953	2006900
## 20067	0.9768	0.0232	2007100
## 20068	0.7019	0.2981	2007200
## 20070	0.9415	0.0585	2007400
## 20071	0.9259	0.0741	2007500
## 20074	0.9231	0.0769	2007800
## 20076	0.7860	0.2140	2008000
## 20077	0.8755	0.1245	2008100
## 20078	0.9354	0.0646	2008200
## 20079	0.5822	0.4178	2008300
## 20080	0.9389	0.0611	2008400
## 20082	0.9058	0.0942	2008600
## 20083	0.6154	0.3846	2008700
## 20084	0.7146	0.2854	2008800
## 20085	0.8570	0.1430	2008900
## 20086	0.9619	0.0381	2009000
## 20088	0.6570	0.3430	2009200
## 20089	0.7229	0.2771	2009300
## 20090	0.7644	0.2356	2009400
## 20091	0.8078	0.1922	2009500
## 20092	0.8127	0.1873	2009600
## 20093	0.8853	0.1147	2009700
## 20095	0.9585	0.0415	2009900
## 20097	0.7148	0.2852	2010100
## 20098	0.5631	0.4369	2010200
## 20099	0.6140	0.3860	2010300
## 20100	0.9221	0.0779	2010400
## 20101	0.9921	0.0079	2010500
## 20103	0.5066	0.4934	2010700
## 20104	0.6941	0.3059	2010800
## 20105	0.7754	0.2246	2010900
## 20107	0.8940	0.1060	2011100

##	20108	0.8964	0.1036	2011200
##	20110	0.9441	0.0559	2011400
##	20115	0.8897	0.1103	2011900
##	20116	0.9839	0.0161	2012000
##	20118	0.7172	0.2828	2012200
##	20119	0.8098	0.1902	2012300
##	20120	0.6868	0.3132	2012400
##	20122	0.9634	0.0366	2012600
##	20123	0.8128	0.1872	2012700
##	20125	0.8023	0.1977	2012900
##	20128	0.6296	0.3704	2013200
##	20239	0.9303	0.0697	2024300
##	20240	0.5410	0.4590	2024400
##	20241	0.9728	0.0272	2024500
##	20242	0.8442	0.1558	2024600
##	20243	0.8978	0.1022	2024700
##	20397	0.9945	0.0055	2040100
##	20398	0.9903	0.0097	2040200
##	20399	0.8585	0.1415	2040300
##	20435	0.9347	0.0653	2043900
##	21126	0.9948	0.0052	2113000
##	21376	0.7458	0.2542	2138000
##	21404	0.7766	0.2234	2140800
##	21417	0.5835	0.4165	2142100
##	21418	0.9573	0.0427	2142200
##	21419	0.9791	0.0209	2142300
##	21420	0.9201	0.0799	2142400
##	21436	0.9436	0.0564	2144000
##	21439	0.5111	0.4889	2144300
##	21442	0.6153	0.3847	2144600
##	21465	0.8563	0.1437	2146900
##	21478	0.7410	0.2590	2148200
##	21480	0.6853	0.3147	2148400
##	21517	0.6290	0.3710	2152100
##	21532	0.9173	0.0827	2153600
##	21681	0.5189	0.4811	2168500
##	21882	0.7297	0.2703	2188600
##	21896	0.9561	0.0439	2190000
##	21897	0.9985	0.0015	2190100
##	21898	0.9994	0.0006	2190200
##	21899	0.9952	0.0048	2190300
##	21900	0.9973	0.0027	2190400
##	21901	0.9991	0.0009	2190500
##	21902	0.9962	0.0038	2190600
##	21903	0.9997	0.0003	2190700
##	21904	0.9971	0.0029	2190800
##	21905	0.9946	0.0054	2190900
##	21906	0.9538	0.0462	2191000
##	21907	0.9883	0.0117	2191100
##	21908	0.9997	0.0003	2191200
##	21909	0.9891	0.0109	2191300
##	21914	0.9787	0.0213	2191800
##	21915	0.9914	0.0086	2191900
##	21916	0.9925	0.0075	2192000

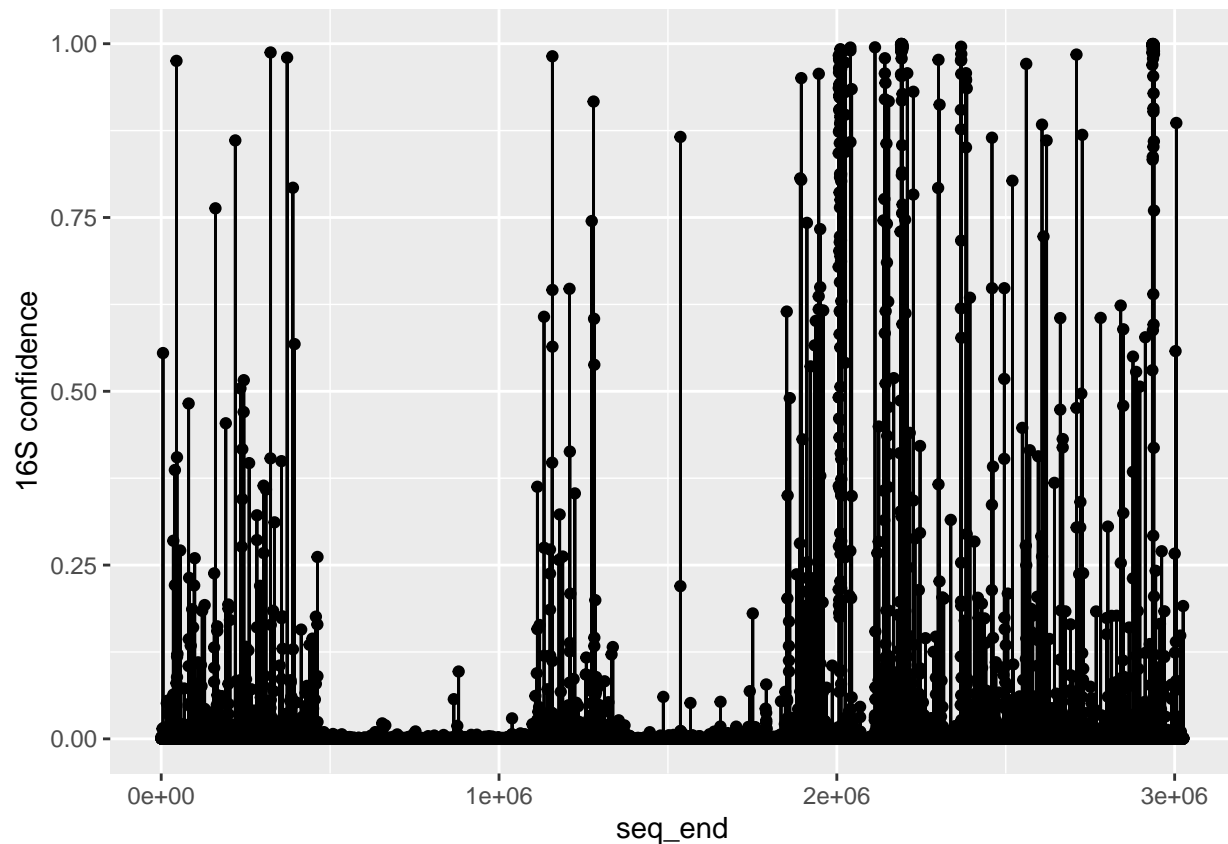
##	21917	0.9929	0.0071	2192100
##	21918	0.9970	0.0030	2192200
##	21919	0.9994	0.0006	2192300
##	21920	0.9964	0.0036	2192400
##	21922	0.8111	0.1889	2192600
##	21923	0.9183	0.0817	2192700
##	21924	0.9982	0.0018	2192800
##	21925	0.9935	0.0065	2192900
##	21926	0.9538	0.0462	2193000
##	21927	0.8153	0.1847	2193100
##	21928	0.9978	0.0022	2193200
##	21929	0.7561	0.2439	2193300
##	21930	0.8540	0.1460	2193400
##	21931	0.9986	0.0014	2193500
##	21932	0.9974	0.0026	2193600
##	21933	0.9994	0.0006	2193700
##	21934	0.9914	0.0086	2193800
##	21935	0.9940	0.0060	2193900
##	21936	0.5965	0.4035	2194000
##	21937	0.9959	0.0041	2194100
##	21938	0.7306	0.2694	2194200
##	21939	0.9948	0.0052	2194300
##	21940	0.9957	0.0043	2194400
##	21941	0.9278	0.0722	2194500
##	21942	0.7687	0.2313	2194600
##	22017	0.7469	0.2531	2202100
##	22027	0.6121	0.3879	2203100
##	22081	0.9575	0.0425	2208500
##	22260	0.7830	0.2170	2226400
##	22261	0.9309	0.0691	2226500
##	22995	0.7925	0.2075	2299900
##	23005	0.9768	0.0232	2300900
##	23037	0.9123	0.0877	2304100
##	23669	0.9049	0.0951	2367300
##	23670	0.6192	0.3808	2367400
##	23671	0.9759	0.0241	2367500
##	23672	0.8768	0.1232	2367600
##	23674	0.9957	0.0043	2367800
##	23676	0.7168	0.2832	2368000
##	23677	0.9766	0.0234	2368100
##	23679	0.9565	0.0435	2368300
##	23680	0.9852	0.0148	2368400
##	23682	0.5770	0.4230	2368600
##	23818	0.9488	0.0512	2382200
##	23819	0.9576	0.0424	2382300
##	23820	0.8508	0.1492	2382400
##	23821	0.9476	0.0524	2382500
##	23842	0.9358	0.0642	2384600
##	23937	0.6346	0.3654	2394100
##	24589	0.8648	0.1352	2459300
##	24591	0.6483	0.3517	2459500
##	24953	0.5178	0.4822	2495700
##	24954	0.6483	0.3517	2495800
##	25194	0.8029	0.1971	2519800

##	25603	0.9710	0.0290	2560700
##	26070	0.8836	0.1164	2607400
##	26114	0.7227	0.2773	2611800
##	26206	0.8608	0.1392	2621000
##	26611	0.6053	0.3947	2661500
##	27090	0.9844	0.0156	2709400
##	27260	0.8689	0.1311	2726400
##	27806	0.6056	0.3944	2781000
##	28400	0.6233	0.3767	2840400
##	28475	0.5892	0.4108	2847900
##	28763	0.5499	0.4501	2876700
##	28855	0.5279	0.4721	2885900
##	28966	0.5067	0.4933	2897000
##	29127	0.5777	0.4223	2913100
##	29336	0.5303	0.4697	2934000
##	29337	0.9695	0.0305	2934100
##	29338	0.9992	0.0008	2934200
##	29339	0.9994	0.0006	2934300
##	29340	0.9871	0.0129	2934400
##	29341	0.9990	0.0010	2934500
##	29342	0.9986	0.0014	2934600
##	29343	0.9964	0.0036	2934700
##	29344	0.9975	0.0025	2934800
##	29345	0.9876	0.0124	2934900
##	29346	0.9987	0.0013	2935000
##	29347	0.8332	0.1668	2935100
##	29348	0.9925	0.0075	2935200
##	29349	0.9997	0.0003	2935300
##	29350	0.8373	0.1627	2935400
##	29353	0.5879	0.4121	2935700
##	29356	0.9786	0.0214	2936000
##	29357	0.9901	0.0099	2936100
##	29358	0.9958	0.0042	2936200
##	29359	0.9968	0.0032	2936300
##	29360	0.9969	0.0031	2936400
##	29361	0.9995	0.0005	2936500
##	29362	0.9532	0.0468	2936600
##	29364	0.6397	0.3603	2936800
##	29365	0.9947	0.0053	2936900
##	29366	0.9985	0.0015	2937000
##	29367	0.9957	0.0043	2937100
##	29368	0.8518	0.1482	2937200
##	29369	0.9069	0.0931	2937300
##	29370	0.9975	0.0025	2937400
##	29371	0.9022	0.0978	2937500
##	29372	0.5960	0.4040	2937600
##	29373	0.9286	0.0714	2937700
##	29374	0.9967	0.0033	2937800
##	29375	0.9986	0.0014	2937900
##	29376	0.9917	0.0083	2938000
##	29377	0.9892	0.0108	2938100
##	29379	0.9937	0.0063	2938300
##	29380	0.9958	0.0042	2938400
##	29381	0.9982	0.0018	2938500

```
## 29382    0.8596        0.1404 2938600
## 29383    0.9893        0.0107 2938700
## 29384    0.9850        0.0150 2938800
## 29385    0.7600        0.2400 2938900
## 30023    0.5578        0.4422 3002700
## 30046    0.8860        0.1140 3005000
```

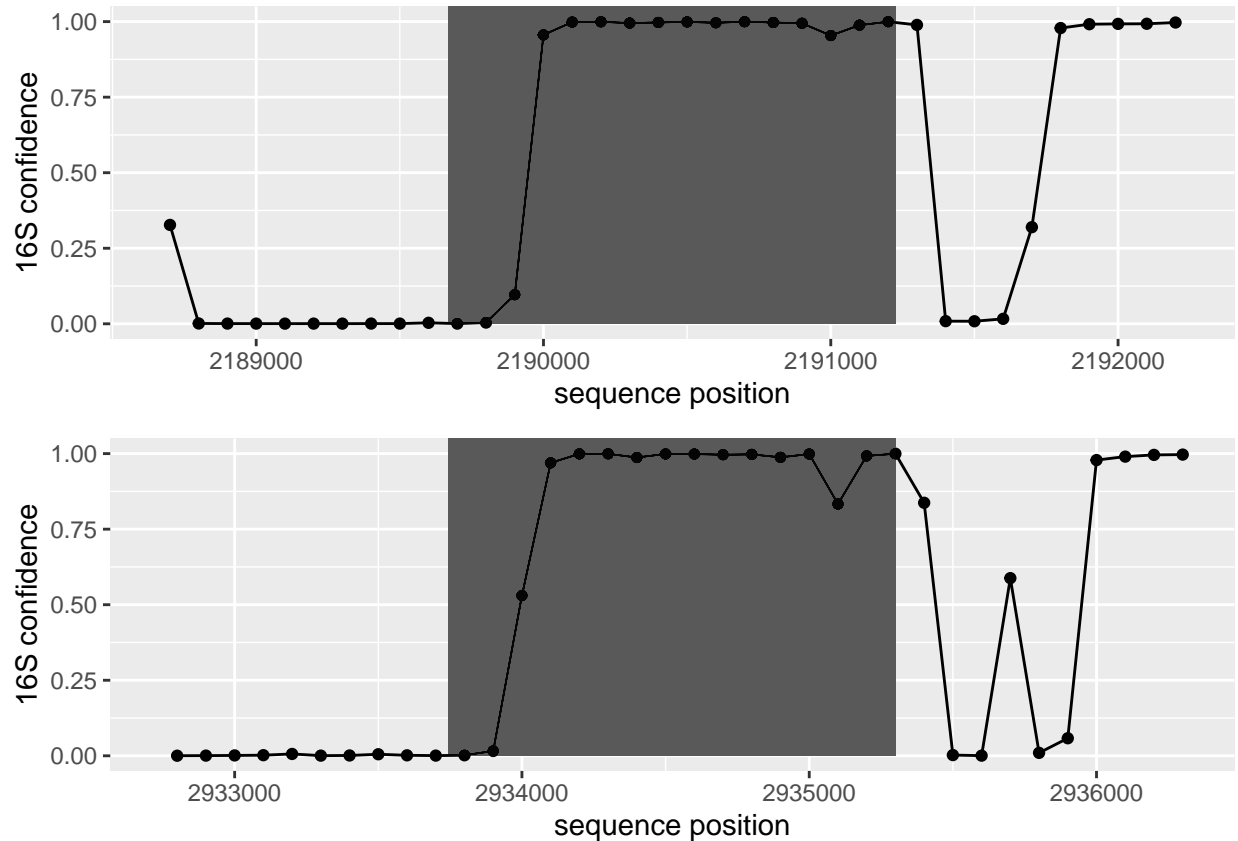
Let's visualize our models predictions and compare them to the true areas. First we look at the confidence in 16S over the whole genome.

```
ggplot(df, aes(x = seq_end, y = conf_16S)) + geom_point() + geom_line() + ylab("16S confidence")
```



Next we may zoom into areas with high 16S confidence.

```
p1 <- ggplot(df, aes(x = seq_end, y = conf_16S)) + geom_point() + geom_line() +
  geom_rect(aes(xmin=start[1], xmax=end[1], ymin=0, ymax=Inf), alpha = 0.01) +
  xlim(c(start[1] - 1000, end[1] + 1000)) +
  ylab("16S confidence") + xlab("sequence position")
p2 <- ggplot(df, aes(x = seq_end, y = conf_16S)) + geom_point() + geom_line() +
  geom_rect(aes(xmin=start[2], xmax=end[2], ymin=0, ymax=Inf), alpha = 0.01) +
  xlim(c(start[2] - 1000, end[2] + 1000)) +
  ylab("16S confidence") + xlab("sequence position")
ggpubr::ggarrange(p1, p2, ncol = 1, nrow = 2)
```



Tensorboard

We can use tensorboard to monitor our training runs. To track the runs, we have to specify a path for tensorboard files and give the run a unique name.

```
# tensorboard_path <- tensorboard.log
# if (!dir.exists(tensorboard_path)) dir.create(tensorboard_path)
# model <- create_model_lstm_cnn()
# run.name <- "run_1"
# trainNetwork(train_type = "lm",
#               model = model,
#               path = train_path_1,
#               path.val = validation_path_1,
#               steps.per.epoch = 5,
#               batch.size = 8,
#               epochs = 10,
#               run.name = run.name,
#               tensorboard.log = tensorboard_path,
#               output = list(none = FALSE,
#                             checkpoints = FALSE,
#                             tensorboard = TRUE, # enable tensorboard
#                             log = FALSE,
#                             serialize_model = FALSE,
#                             full_model = FALSE),
#               output_format = "target_right")
```

```
# )

## open tensorboard in browser
# tensorflow::tensorboard(tensorboard_path)
```

The “SCALARS” tab displays accuracy, loss, learning rate and percentage of files seen for each epoch.

The “TEXT” tab shows the `trainNetwork` call as text.

The “HPARAM” tab tracks the hyperparameters of the different runs (maxlen, batch size etc.).

Further tensorboard documentation can be found [here](#).

Checkpoints

We can save the architecture and weights of a model after every epoch using checkpoints. The checkpoints get stored in h5 format. The file names contain the corresponding epoch, loss and accuracy

```
# checkpoint_path <- file.path(dir_path, "checkpoints")
# if (!dir.exists(checkpoint_path)) dir.create(checkpoint_path)
# model <- create_model_lstm_cnn()
# run.name <- "run_2"
# trainNetwork(train_type = "lm",
#             model = model,
#             path = train_path_1,
#             path.val = validation_path_1,
#             steps.per.epoch = 5,
#             batch.size = 8,
#             epochs = 10,
#             run.name = run.name,
#             checkpoint_path = checkpoint_path,
#             save_best_only = TRUE, # only save model if loss improves
#             save_weights_only = FALSE, # save architecture and weights
#             output = list(none = FALSE,
#                           checkpoints = TRUE, # enable checkpoints
#                           tensorboard = FALSE,
#                           log = FALSE,
#                           serialize_model = FALSE,
#                           full_model = FALSE),
#             output_format = "target_right"
# )
```

After training, we can now load a trained model and continue training or use the model for predictions/inference.

```
# cp_run_path <- file.path(checkpoint_path, paste0(run.name, "_checkpoints"))
# checkpoints <- list.files(cp_run_path)
# checkpoints
# last_checkpoint <- checkpoints[length(checkpoints)]
#
# # load trained model and compile
# model <- keras::load_model_hdf5(file.path(cp_run_path, last_checkpoint), compile = FALSE)
# model <- keras::load_weights_model_hdf5(model, file.path(cp_run_path, last_checkpoint))
```

```

# optimizer <- keras::optimizer_adam(lr = 0.01)
# model %>% keras::compile(loss = "categorical_crossentropy", optimizer = optimizer, metrics = c("acc"))
#
# # continue training
# trainNetwork(train_type = "lm",
#               model = model,
#               path = train_path_1,
#               path.val = validation_path_1,
#               steps.per.epoch = 5,
#               batch.size = 8,
#               epochs = 2,
#               run.name = "continue_from_checkpoint",
#               output_format = "target_right"
# )

```