

Fall 2019

Global Conservation Status and Threat Patterns of the World's Most Prominent Forage Fishes (Teleostei, Clupeiformes)

Tiffany L. Birge
Old Dominion University, tbirg001@odu.edu

Follow this and additional works at: https://digitalcommons.odu.edu/biology_etds



Part of the [Biodiversity Commons](#), [Biology Commons](#), [Ecology and Evolutionary Biology Commons](#), and the [Natural Resources and Conservation Commons](#)

Recommended Citation

Birge, Tiffany L.. "Global Conservation Status and Threat Patterns of the World's Most Prominent Forage Fishes (Teleostei, Clupeiformes)" (2019). Master of Science (MS), Thesis, Biological Sciences, Old Dominion University, DOI: 10.25777/8m64-bg07
https://digitalcommons.odu.edu/biology_etds/109

This Thesis is brought to you for free and open access by the Biological Sciences at ODU Digital Commons. It has been accepted for inclusion in Biological Sciences Theses & Dissertations by an authorized administrator of ODU Digital Commons. For more information, please contact digitalcommons@odu.edu.

GLOBAL CONSERVATION STATUS AND THREAT PATTERNS OF THE WORLD'S MOST
PROMINENT FORAGE FISHES (TELEOSTEI, CLUPEIFORMES)

by

Tiffany L. Birge
A.S. May 2014, Tidewater Community College
B.S. May 2016, Old Dominion University

A Thesis Submitted to the Faculty of
Old Dominion University in Partial Fulfillment of the
Requirements for the Degree of

MASTER OF SCIENCE

BIOLOGY

OLD DOMINION UNIVERSITY
December 2019

Approved by:

Kent E. Carpenter (Advisor)

Sara Maxwell (Member)

Thomas Munroe (Member)

ABSTRACT

GLOBAL CONSERVATION STATUS AND THREAT PATTERNS OF THE WORLD'S MOST PROMINENT FORAGE FISHES (TELEOSTEI, CLUPEIFORMES)

Tiffany L. Birge
Old Dominion University, 2019
Advisor: Dr. Kent E. Carpenter

Conserving biodiversity is one of the greatest ethical responsibilities and challenges humans face. Understanding the conservation status of taxonomic groups provides a systematic way to prioritize efforts to combat biodiversity loss. The 405 species within the order Clupeiformes are the herrings, shads, sardines, anchovies, menhadens and relatives that include many of the most important marine forage fishes. These small, schooling fishes are economically, ecologically and culturally significant globally. Despite their contribution to global fisheries and our increasing reliance on these fishes for food and industrial commodities, they are generally poorly known with limited information regarding basic biology and population trends. I applied IUCN Red List methodology, a comprehensive and systematic approach to assessing extinction risk of species, to all clupeiform species. I then used these assessments to synthesize and address their global conservation status and to highlight the potential for improvements to conservation and fisheries management. The best estimate of nearly 11% of species are of elevated conservation concern, although this could be as high as 34% if Data

Deficient species are all threatened. The Caribbean and the Indo-Malay-Philippine Archipelago both have high concentrations of either threatened or Data Deficient species and are areas of particular conservation concern. Major threats include exploitation, pollution and habitat modification for human use although the intensity of a specific threat differs between freshwater, estuarine and marine environments. Life history and ecological traits of threatened and Near Threatened species were characterized between primary habitat systems. Immediate conservation priorities include: 1) the evaluation of current fisheries management strategies, with a strong recommendation toward ecosystem-based management protocols that incorporate group-specific life history traits, and 2) local, intensive habitat restoration to reduce pollution and remove dams. These extinction risk assessments and subsequent analyses should be used to monitor conservation progress and as an informative tool for fisheries and conservation managers.

Copyright, 2019, by Tiffany L. Birge and Kent E. Carpenter, All Rights Reserved

This thesis is dedicated to my father, Bruce P. Birge and to my mother, Jaqueline R. Birge for always inspiring and encouraging my sense of adventure by putting up with my many childhood experiments and for providing continual love and support.

ACKNOWLEDGEMENTS

This research would not have been possible without the continuous support and encouragement from many important people. First, and foremost, I would like to thank my advisor, Dr. Kent Carpenter, for providing me with this incredible opportunity and for showing me that great things can be accomplished without taking life too seriously. I also thank my committee members, Dr. Thomas Munroe and Dr. Sara Maxwell, each of whom provided great insight throughout this process. Tom, thank you for constantly challenging me and for pushing me to think outside of the box. Sara, thank you for always reminding me to take a step back and to enjoy the ride. A special and heartfelt thank you to my mentor and dear friend Dr. Gina Ralph, whom without her support and guidance, I would not be where I am today.

Thanks also to the main funding sources: Toyota Motor Corporation provided the funding for this research and the Philippines' Bureau of Fisheries and Aquatic Resources (BFAR) provided funding and logistical aid for the clupeiform workshop in 2017.

Many thanks are extended to Rob Bullock for the facilitation of species assessments and for his invaluable friendship throughout this process. To my current and previous lab mates, C. Linardich, M. Harvey, C. Gorman, J. Buchannan, E. Stump, A. Ackiss, E. Biesack, B. Stockwell, J. Whalen, J. Baldisimo, M. Kenton, I. Lopez and E. Garcia, thanks for all the advice and laughs you've provided me with. To the members of the BGSO, you have become my biology family; thank you for helping me navigate the roller coaster that is graduate school. My deepest gratitude goes to my family, Dad, Mom, Courtney, Corbin, Brian, Dustin and Aunt Sharon; thank

you for the many sacrifices you have made for me and for your uplifting support during this journey.

Special thanks for the 132 experts who made these assessments possible: M. Santos, D. Gaughan, W. Mohd Arsaad, G. Allen, R. Raghavan, N. Boguskaya, I.G. Priede, D.J. Allen, H. Larson, L.R. Casten, M.N. Alava, N. Richman, A. Beresford, A. Chenery, M. Ram, R. Reis, F. Lima, D. Grubbs, J. Simons, J. Caruso, J. Brenner, L. Tornabene, J. Brown, J. Freyhof, M. Kottelat, L. Grijalba Bendeck, T. Iwamoto, W. Eschmeyer, J. Alvarado, R. Wagner, W. Bussing, B. Collette, P. Chakrabarty, J. Carlson, J.D. McEachran, G.A. Hammerson, C. Vidthayanon, T. Moelants, S. Chaudhry, B.R. Jha, N.C. Datta, R. Britz, S.C. Dey, M. Entsua-Mensah, P. Laléyé, T.A. Adeofe, K. Camara, Y.H. Camara, K. Cissoko, L. de Moraes, R. Djiman, E. Mbye, A. Sidibe, P. Tous, A. Sagna, M. Sylla, J.G. Nielsen, F. Kaymaram, J. Bishop, M. Al-Husaini, S. Hartmann, S. Alam, K. Al-Kalaf, Q. Alghawzi, A. Salarpouri, J. Quilang, J. Torres, D. Milton, A. Cotto, E. Medina, O. Bernal, M. Loeb, C. van der Lingen, P. Borsa, E. Abdulqader, H. Al-Nazry, M. Al-Mukhtar, K. Nedreaas, A.-B. Florin, R. Cook, P. Fernandes, P. Lorange, K.A. Aiken, A. Ali, B. Collen, K.R. Devi, F. Pezold, R. Robertson, K. Cleary, T. Sandell, D. Hay, R. Gustafson, J. Williams, H. Palla, R. Deligero, M. Alcantara, M. Doyola, L. Gatlabayan, M. Villarao, A. Tambihasan, G. Lopez, J. Villanueva, F. Buccat, L. Parido, N. Lanzuela, P. Belga, A. Gapuz, A.K. Acosta, M. Casini, P. Henderson, H.H. Ng, K.E. Carpenter, G. Ralph, J.S., Sparks, A.J. Crivelli, R. Bills, B. Smith-Vaniz, D. Herdson, S. Molur, N. Dahanukar, G. Ntakimazi, L. de Costa, B.D. Olaosebikan, T. Ravelomanana, P.C. Heemstra, J. Tyler, M. Pal and A. Sidibé. Specific recognition and thanks go to Dr. Fabio Di Dario, Dr. Thomas Munroe and Dr. Harutaka Hata, whose collaboration, dedication and extensive review of many species assessments greatly improved this research.

NOMENCLATURE

<i>AOO</i>	Area of Occupancy
<i>CAP</i>	Canonical Analysis of Principal Coordinates
<i>CAT</i>	Red List Category
<i>COO</i>	Countries of Occurrence
<i>CR</i>	Critically Endangered
<i>DD</i>	Data Deficient
<i>E</i>	Euryhaline
<i>EC</i>	Elevated Concern
<i>EN</i>	Endangered
<i>EOO</i>	Extent of Occurrence
<i>EW</i>	Extinct in the Wild
<i>EX</i>	Extinct
<i>F</i>	Freshwater
<i>FAO</i>	Food and Agriculture Organization of the United Nations
<i>GBIF</i>	Global Biodiversity Information Facility
<i>GEBCO</i>	General Bathymetric Chart of the Oceans
<i>GL</i>	Generation Length
<i>IUCN</i>	International Union for Conservation of Nature
<i>IUU</i>	Illegal, Unreported and Unregulated

<i>IWP</i>	Indo-West Pacific
<i>LC</i>	Least Concern
<i>LME</i>	Large Marine Ecosystem
<i>M</i>	Marine
<i>MaxSL</i>	Maximum Standard Length (cms)
<i>MEOW</i>	Marine Ecoregions of the World
<i>MSY</i>	Maximum Sustainable Yield
<i>NT</i>	Near Threatened
<i>OZCAM</i>	Online Zoological Collections of Australian Museums
<i>PERMANOVA</i>	Permutational Multivariate Analysis of Variance
<i>PRIMER-e</i>	Plymouth Routines In Multivariate Ecological Research
<i>RL</i>	Red List
<i>SSB</i>	Spawning Stock Biomass
<i>SAU</i>	Sea Around Us
<i>SYS</i>	Habitat System
<i>VU</i>	Vulnerable
<i>WDFW</i>	Washington Department of Fish and Wildlife

TABLE OF CONTENTS

	Page
LIST OF FIGURES.....	xii
 Chapter	
1. INTRODUCTION.....	1
2. GLOBAL CONSERVATION STATUS OF THE WORLD’S MOST PROMINENT FORAGE FISHES (TELEOSTEI: ORDER CLUPEIFORMES).....	9
INTRODUCTION.....	9
METHODS	13
RED LIST METHODS	13
DISTRIBUTION MAPPING METHODOLOGY	16
SPECIES RICHNESS ANALYSES	17
RESULTS	17
GLOBAL IUCN RED LIST STATUS OF CLUPEIFORMS	17
MAJOR THREATS	20
SPATIAL ANALYSES	21
DISCUSSION	25
3. CHARACTERIZING CLUPEIFORM THREATS, LIFE HISTORY TRAITS AND HABITAT PREFERENCE TO INFORM MANAGEMENT OF DATA DEFICIENT SPECIES.....	31
INTRODUCTION	31
METHODS	34
RESULTS	38
DISCUSSION	45
4. DISCUSSION AND CONCLUSIONS.....	52

REFERENCES.....	57
-----------------	----

APPENDICIES

A. LIST OF ALL CLUPEIFORM IUCN RED LIST CATEGORIES	76
B. IUCN RED LIST METHODS AND DATA USE.....	87
C. LIST OF ALL SPECIES WITH KNOWN THREATS USED IN CAP ANALYSIS	92
D. LIST OF ALL SPECIES OF ELEVATED CONSERVATION CONCERN USED IN CAP ANALYSIS...	99
 VITA.....	 101

LIST OF FIGURES

Figure	Page
1. The nine extinction risk categories from the IUCN Red List of Threatened Species	14
2. Proportion of species (n = 405) listed in each Red List Category	18
3. Proportion of species listed in each Red List Category separated by family.....	19
4. Proportion of species listed in each Red List Category by major habitat (fresh, euryhaline or marine)	22
5. Number of species impacted by major threats	23
6. Number of clupeiforms in each Large Marine Ecoregion (LME) and freshwater hydrobasin for A) All species, B) all species of elevated concern (CR, EN, VU, NT), and C) all Data Deficient species	24
7. Canonical Analysis of Principal Coordinates (CAP) ordination of species with known threats in multivariate threat space by primary habitat system (n = 144)	41
8. Canonical Analysis of Principal Coordinates (CAP) ordination of threatened and Near Threatened species-specific life history and ecological traits in multivariate space by primary habitat system (n = 33).....	43
9. Mean maximum standard length (cm) of clupeiforms (n = 394) as a function of primary habitat system (marine, euryhaline, freshwater) and exploitation status (exploited or not exploited)	44

CHAPTER 1

INTRODUCTION

Global biodiversity is under threat. From grasses and dragonflies to sharks and mangroves, and nearly all taxa in between, there is growing concern regarding the survival status of biodiversity. Population declines have been documented in such diverse taxa as mammals (Davidson et al., 2009; Ripple et al., 2016), birds (Dunn et al., 2002), large predatory fishes (Myers and Worm, 2005), amphibians (Bielby et al., 2008), flying insects (Hallmann et al., 2017) and plants (Willis, 2017), leading to local extirpations and global extinctions (Young et al., 2016). Although up to 100 million species are estimated as extant (May, 1992; Mace et al., 2005), with the best working estimate between 8 – 9 million species (Chapman, 2009; Hilton-Taylor et al., 2009; Mora et al., 2011), only around 2 million species have been described to date (Hilton-Taylor et al., 2009). As a result, there is risk that species will disappear before we are aware they exist. Given that current rates of extinction are over 1000 times that of the background rate of extinction (Pimm et al., 2014), the future of biodiversity is bleak.

Despite the dominant aquatic global surface area (~ 71% of earth's surface) and even larger inhabitable volume (Polidoro et al., 2009; Darwall et al., 2009), our knowledge of and concern for these ecosystems lags far behind that of terrestrial systems. Historically, active conservation of aquatic resources lags behind terrestrial conservation effort. This is partly due to proximity and ease of study of terrestrial systems, but also because the aquatic realm is a

vast environment, whose size alone was thought to be a buffer to impacts (Myers and Worm, 2005). As a result, global species conservation status is heavily biased towards terrestrial vertebrates and plants (Hilton-Taylor et al., 2009; Polidoro et al., 2009). Presently, major anthropogenic threats to marine and freshwater species are recognized (Young et al., 2016), particularly from the fishing sector (e.g., Atlantic cod: Shelton et al., 2006).

Fisheries are important to the financial and nutritional security of billions of people globally (FAO, 2018). Capture fisheries generate substantial local and national revenue (total estimated at USD 130 billion in 2016: FAO, 2018), mostly from landings of marine fisheries (FAO, 2018). These landings play a vital role in global nutritional security by providing a valuable source of protein and micronutrients (FAO, 2018). Fish provide over 3.2 billion people with about 20% of their average per capita animal protein intake (FAO, 2018) and consumption has steadily grown in developing regions and low-income food-deficit countries (FAO, 2018). However, overharvesting of our fish stocks has resulted in population declines up to 90% for pelagic fish species (Sadovy, 2001; Myers and Worm, 2003; Pauly et al., 2005; Sadovy et al., 2013). Taxa- and region-specific studies increasingly express that exploitation is the most prominent threat and is of growing concern as our reliance on fishery resources continues to expand (Sadovy et al., 2013; Lynch et al., 2016; FAO, 2018).

One extremely important, but often underappreciated, component of fisheries are the forage fishes. These highly numerous small- and medium-sized, pelagic species support global economies through direct fisheries exploitation and also by serving as a major food source for higher predators that are important to these economies as well (Pikitch et al., 2014). Forage fishes comprise over 30% of the total global marine fish catch (Alder et al., 2008; Smith et al.,

2011). Species of the order Clupeiformes, including herrings, shads, menhadens, sardines, anchovies, and their relatives, make up a major component of forage fishes and dominate worldwide forage fish landings (Tacon and Metian, 2009a). Three distinct contributions of forage fishes have been recognized, including: 1) ecological support for predators as a vital food resource, 2) economic value to forage fisheries, and 3) support for the catch and value of other commercially targeted predators, such as fishes, mammals and squid (Cury et al., 2011; Smith et al., 2011; Pikitch et al., 2014; Hilborn et al., 2017).

Clupeiform fisheries have a long history of nutritional, cultural and economic importance (Whitehead, 1985; Alder, et al., 2008). Their presence has been associated with persistent human settlement, growth and survival for thousands of years (Finney et al., 2002; Bassett, 2014) and is well documented in the northeastern Pacific (Thornton et al., 2010; Levin et al., 2016), the northwestern (Bassett, 2014) and northeastern Atlantic (Bloch, 1809; Coull, 2003) and the tropical western Pacific (Ruddle and Ishige, 2010). Due to their overall importance and abundance, some have been considered as cultural keystone species and have been given local nicknames like 'silver of the sea' and 'silver darlings' (Coull, 2003; Smyllie, 2004; Murray, 2015; Levin et al., 2016) to reflect the important status of these species.

Today, many millions of people rely on clupeiform catches across the world for food, industrial commodities, and everyday items. The majority of clupeiform resources are 'reduced' or processed and turned into fishmeal and related products (van der Meer et al., 2015), which makes them one of the largest species groups targeted for non-food uses (Tacon and Metian 2009a, 2009b). Currently, the largest consumer of reduced fish product is the aquaculture sector (Tacon and Metian, 2009b), which is rapidly increasing to keep pace with the growing

demand for fish products (FAO, 2018). Aside from aquaculture, agricultural industries use reduced material as fertilizer and as fishmeal or fish oil to support livestock in direct or compound animal feed (Tydemers, 2004; Tacon and Metian, 2009a; Pikitch et al., 2012; FAO, 2016a). Fish oil is also used in a wide array of industrial applications including fuel, glue production, paint manufacture, and as a vitamin supplement (Tydemers, 2004).

To keep up with the high demand for these products, clupeiforms comprise some of the world's largest fisheries and continue to be the principal group of non-domesticated vertebrates harvested by man (Whitehead, 1985; Tacon and Metian, 2009a, 2009b; FAO, 2018). In general, the largest fisheries exploit cold-water clupeoids, such as species of *Sardinops* and *Clupea* (Whitehead, 1985; FAO, 2018). Historically, the largest fishery by volume was the famous Peruvian anchoveta (*Engraulis ringens*), which contributed an annual estimated 16 million tonnes during peak harvest years (Castillo and Mendo, 1987; Tsukayama and Palomares, 1987; Pikitch et al., 2012).

Of growing concern to fisheries and conservation managers is the tendency, particularly of the cold-water species that support large fisheries, to exhibit highly variable, albeit natural, population fluctuations (Whitehead, 1985; McKechnie et al., 2013). The episodic trends in population fluctuations are thought to be heavily influenced by environmental conditions (Pikitch et al., 2014), such as long-term, decadal-scale physical processes (e.g., El Niño: Alheit et al., 2009). Knowledge is limited on how the excessive removal of these species by fisheries may impact aquatic ecosystems (Alder et al., 2008). However, heavy industrial fishing pressure is known to exacerbate the population flux and has recently been shown to increase the

likelihood of population collapse in small pelagic fishes that have been exploited by long-term fisheries (Pinsky et al., 2011).

The overwhelming importance of cold-water clupeoid fisheries often overshadows the contributions of tropical and freshwater clupeiform fisheries (Whitehead, 1985). These warm-water fisheries tend to dominate the landings of artisanal and subsistence sectors, rather than the industrial sector (Whitehead, 1985). When reported in fisheries landings, multiple species are often lumped in landings data (e.g., *Stolephorus* spp., FAO, 2018), making analysis of species-specific trends problematic.

Aside from most cold-water species that tend to represent the landings of the larger fisheries, we know very little about clupeiforms globally despite our overwhelming reliance on them and their known importance in nearly every aquatic ecosystem (Whitehead, 1985). Research has been hindered by confusing taxonomy and challenging identifications (Whitehead, 1985). Overall, it is relatively easy to distinguish clupeiforms from other fish groups because nearly all lack a visible lateral line on the body; however, it is difficult to tell them apart from each other, particularly in regions where clupeiform species richness is high (Whitehead, 1985). Ironically, these areas coincide with the fastest growing human populations and their reliance on fisheries, and often these ecosystems represent regions most in need of conservation (Darwall et al., 2009).

The Clupeomorpha (Greenwood et al., 1966), along with Alepocephali and Ostariophysii make up the Otocephala, one of four extant lineages of Teleostei (Nelson et al., 2016). Representatives of the Order Clupeiformes are characterized within two suborders: the

Denticipitoidei, a monotypic group with only one extant representative, *Denticeps clupeioides* Clausen 1959, and the Clupeioidi which comprises all other extant species in the Order Clupeiformes (Whitehead, 1985; Grande, 1985; Di Dario, 2004; Di Dario and de Pinna, 2006; Lavouè et al., 2014; Bloom and Egan, 2018). Since Fowler's attempt to list all valid clupeoid species (Fowler, 1973), Whitehead (1985) and Whitehead et al., (1988) have been the only comprehensive works to compile species-specific information on valid species in the suborder Clupeioidi, representing nearly the entire Order Clupeiformes.

Historically, clupeiform systematics largely relied on morphometric, meristic, and other morphological characters, which sometimes classified taxa based more on overall similarity or geographical convenience rather than on rigorous scientific support (Bloom and Egan, 2018). Grande's (1985) five proposed subfamilies of Clupeidae (Alosinae, Clupeinae, Pellonulinae, Dorosomatinae and Dussumieriinae), and the description of what is now considered to be a species-complex of the genus *Sardinops* (Whitehead, 1985), are examples of such taxonomic convenience. Further, given that numerous and often similar species are known in many genera, some valid species may have long been obscured within the synonymies of others, and many more proposed names exist than are needed (Whitehead, 1985).

Current advancement of molecular and genetic methods and recent morphological analyses aided in the description of several species and rearrangement of groups (e.g., Loeb et al., 2017; Li and Ortí, 2007; Lavouè et al., 2014; Di Dario, 2009; Hata and Motomura, 2018; Bloom and Egan, 2018). However, some systematic relationships remain unresolved (Malabara and Di Dario, 2016; Bloom and Egan, 2018). Given this taxonomic and systematic uncertainty, for the purposes of this thesis, I followed the family group names outlined in the study by Van

der Laan et al., (2014) that recognizes seven families (Denticipitidae, Clupeidae, Engraulidae, Pristigasteridae, Chirocentridae, Dussumieriidae and Sundasalangidae). Recent and ongoing analyses, including the reassignment of the Sundasalangidae within the Clupeidae (Lavoué et al., 2014), elevation of the subfamily Spratelloidinae to the family Spratelloididae (Bloom and Egan, 2018) and re-examinations of genera (e.g., revision of *Sardinella* and *Stolephorus* by Hata and Motomura, 2017, 2018, and 2019; revision of *Anchoviella* by Loeb et al., 2017) will likely improve our understanding of the taxonomic and systematic relationships within the Order Clupeiformes.

Given the taxonomic challenges presented by the clupeiforms, species-specific threats can go undocumented particularly in face of overexploitation and in their dependence on often degraded coastal ecosystems. Information about which species are at risk and what factors are most threatening is particularly important to successfully and strategically plan and implement conservation management policies (Venter et al., 2006). Therefore, to evaluate the conservation status of clupeiform fishes, I used the most widely accepted standard for assessing the symptoms of extinction risk, the International Union for Conservation of Nature (IUCN) Red List Criteria (Hoffman et al., 2008). This thesis analyzes the conservation status of the clupeiforms, accounting for species-specific characteristics and population trends. In Chapter 2, I evaluated the global extinction risk for all species using the IUCN Red List methodology. I hypothesized that major threats would vary by family group and by the primary habitat type occupied by the species. In Chapter 3, I used data and results from the Red List Assessments to test the influence of habitat type and natural history traits on susceptibility to threats. I hypothesized that species can be characterized into groups based on these ecological

and natural history traits, and that such analyses could be used to inform management measures.

CHAPTER 2

GLOBAL CONSERVATION STATUS OF THE WORLD'S MOST PROMINENT FORAGE FISHES (TELEOSTEI: ORDER CLUPEIFORMES)

INTRODUCTION

Forage fishes directly link primary production to keystone predators in marine environments (Pikitch et al., 2014). These small- to medium-sized, typically very numerous pelagic species also support the global economy by directly and indirectly sustaining many fisheries (Pikitch et al., 2014). Forage fishes make up over 30% of the global marine catch (Alder et al., 2008; Smith et al., 2011). They also play a key role as prey for many commercially targeted predators, such as fishes, mammals and squids (Cury et al., 2011; Smith et al., 2011; Pikitch et al., 2014; Hilborn et al., 2017).

Species of the order Clupeiformes, including herrings, shads, menhadens, sardines, anchovies, and their relatives, are a major component of forage fishes and dominate worldwide forage fish landings (Tacon and Metian, 2009a). Additional to providing ecological and economic support, clupeiforms contribute to food security worldwide given their abundance, access and exceptionally high nutrient content (FAO, 2018); in some communities, clupeiforms make up the major or the sole protein source (Mohan Dey et al., 2005; Alder et al., 2008; Kawarazuka and Béné, 2011; Mohanty et al., 2019). Historically, clupeiform presence has been associated with persistent human settlement, growth and survival for thousands of years

(Bloch, 1809; Coull, 2003; Thornton et al., 2010; Ruddle and Ishige, 2010; Bassett, 2014; Levin et al., 2016). To meet the needs of a projected rising human global population (United Nations, Department of Economic and Social Affairs, Population Division, 2017), demand for fisheries resources is expected to continue to grow (FAO, 2018). Given the overall ecological, cultural, nutritional, and economic importance of clupeiforms worldwide, their conservation status warrants greater attention.

The teleost fish order Clupeiformes includes 405 species that are globally distributed with tropical, temperate and sub-Arctic representatives (Whitehead, 1985; Blaber et al., 1996; Wongratana et al., 1999; Munroe et al., 1999; Lavoué et al., 2013; Pikitch et al., 2014). Members of this Order are ecologically diverse and span all aquatic habitats, including freshwater rivers and lakes, estuaries, coastal marine areas, and the open ocean (Whitehead, 1985; Lavoué et al., 2013; Bloom and Egan, 2018). Clupeiform species can be restricted to fresh, estuarine, or marine waters, or they can exhibit diadromy (Whitehead, 1985). This ability to navigate between marine and freshwater habitats is shared with other groups such as stingrays, needlefishes, silversides, drums and pufferfishes (Lovejoy et al., 2006; Bloom and Lovejoy, 2017; Bloom and Egan, 2018). Strictly marine clupeiforms (32% of all species) are distributed in every ocean, except for the Southern Ocean (Whitehead, 1985), strictly freshwater species (18% of all species) are found on every continent except for Antarctica (Bloom and Lovejoy, 2012, 2014; Bloom and Egan, 2018).

In general, life history traits such as high fecundity, widespread distributions, adaptability to diverse habitats, and high dispersal ability are features that are thought to

increase survivability in face of anthropogenic stresses (Stearns, 1992; Hutchins, 2000; Sadovy, 2001; Denney et al., 2002; Reynolds et al., 2005; Alder et al., 2008; Comeros-Raynal et al., 2016). In contrast, slow growth, large body size, and high longevity are life history features thought to increase a species' vulnerability to extinction (Roberts and Hawkins, 1999; Reynolds et al., 2005; Harnik et al., 2012; Juan-Jorda et al., 2015; Comeros-Raynal et al., 2016). These innate traits have also been used to determine a species' ability to cope with, and recover from, human-induced and environmental disturbances (Cardillo et al., 2005, 2008; Reynolds et al., 2005). However, high fecundity, early age at maturation and similar demographic traits do not reliably predict a species' vulnerability to, or ability to recover from, overexploitation (Jennings et al., 1998; Kindsvater et al., 2016; Sadovy, 2001; Juan-Jorda et al., 2012, Comeros-Raynal et al., 2016).

Despite the global importance of clupeiforms, basic biological information, fisheries data, and management efforts are severely deficient compared to those of other commercially important fishes such as tunas and billfishes. This disparity may be due in part to perception of extinction resistant traits and taxonomic complexity of clupeiforms (Whitehead, 1985; Alder et al., 2008). Clupeiform value per pound is also far less than that for other commercial fishes, which may further disincentivize the contribution of resources to research and conservation for the clupeiform fishes. For example, the average commercial landed value of all tunas in the U.S. for 2017 was about USD \$ 2.8/pound, while the average value for clupeiforms was roughly USD \$0.09/pound (NOAA Fisheries, 2019). The paradox between worldwide clupeiform importance and lack of available study resources and reliable data reinforces the need to invest effort into understanding the current conservation status of species within the Order. The International

Union for Conservation of Nature Red List of Threatened Species provides an ideal starting point for highlighting and addressing conservation needs for fish species, including the clupeiforms (Mace et al., 2008).

The IUCN Red List is a comprehensive repository of open-access, species-specific assessments that conveys a species' symptoms of extinction (Rodrigues et al., 2006; Vié et al., 2009). Red List assessments are the most widely accepted standard for species-level risk evaluations (Hoffman et al., 2008). By illuminating knowledge gaps regarding the conservation status of species (e.g., Carpenter et al., 2008; Schipper et al., 2008; Polidoro et al., 2010; Short et al., 2011), the assessments can be used to inform and influence decisions regarding biodiversity conservation (Rodrigues et al., 2006; Mace et al., 2008; Vié et al., 2009).

Limited species-specific information on the conservation status of clupeiforms hampers our ability to proactively manage and conserve these essential components of aquatic food webs. Therefore, the extinction risk of all 405 species within the Order was evaluated following the IUCN Red List methodology to provide a baseline from which to monitor changes. The resulting information was then used to evaluate: 1) variability in the proportion of species at an elevated risk of extinction as a function of family (Denticipitidae, Pristigasteridae, Engraulidae, Chirocentridae, Clupeidae, Dunsumieriidae, Sundasalangidae), and as a function of habitat (freshwater, marine and euryhaline); 2) major threats to all species; and 3) spatial trends in clupeiform species richness.

METHODS

Red List Methods

A comprehensive species list was compiled based on the online version of the Catalog of Fishes (Eschmeyer et al., 2017) and in consultation with taxonomic experts. Individual clupeiform species assessments were collated from information on the geographic distribution, population status, life history, utilization and quality of habitat, potential threats and the conservation measures of each species. The assessment process required input and involvement from 132 international experts from more than 20 countries who systematically evaluated extinction risk indicators for all 405 species. Three nominal species recently described after December 2018 as new or elevated as distinct from synonymy with another species are not included within these analyses, but Red List assessments have been completed for these species (Appendix A).

The IUCN Red List includes eight levels of extinction risk (Fig. 1): Extinct (EX), Extinct in the Wild (EW), Critically Endangered (CR), Endangered (EN), Vulnerable (VU), Near Threatened (NT), Least Concern (LC), and Data Deficient (DD: IUCN, 2012). A species can qualify for a threatened category (CR, EN, VU) by meeting at least one of the five quantitative thresholds that fall under IUCN Criteria (A – E: Mace et al., 2008). The criteria evaluate population decline (A), restricted geographic distribution (B), small population size and decline (C), very small or restricted population size (D), and the high probability of potential extinction (E: Akçakaya et al., 2000; Mace et al., 2008).

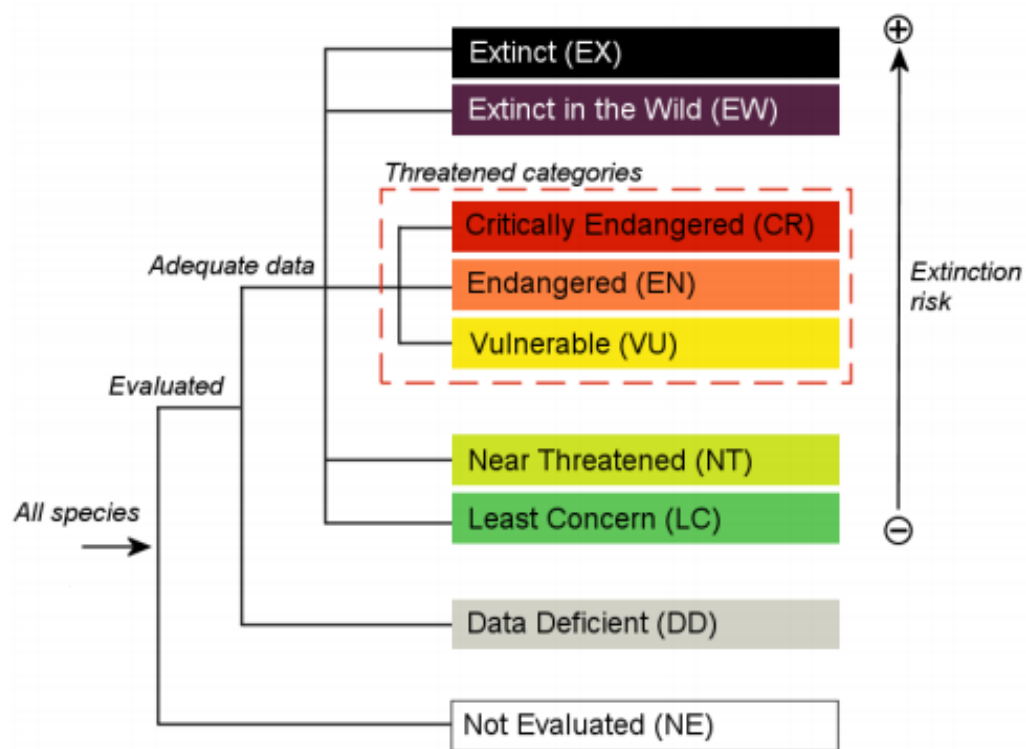


Fig. 1. The nine extinction risk categories from the IUCN Red List of Threatened Species.

A category of NT can be applied if the quantified estimates of population decline or range size nearly meet the thresholds for assigning a threatened category under at least one of the criteria. The DD category is applied if a species is only known from few specimens, lacks available information to assess under any of the criteria, or if there is uncertainty regarding its taxonomic status. This category can also be applied if declines are likely due to a known threat (e.g., fishing pressure), but the threat could not be quantified, such that a more appropriate category could be applied.

All five Red List Criteria were considered during the assessment process; however, almost all species were assessed under criteria A (population decline) or B (restricted range). Data required to assess a species under the remaining criteria (C, D or E) were often unavailable given the difficulty of quantifying the number of mature individuals present in fish populations. As of July, 2019, all species are published on the Red List website (www.iucnredlist.org), where species data, maps and extinction risk categories are freely available.

Quantifying Threats

As part of the Red List process, threats were identified for each species based on the published literature and in consultation with experts. Threats were quantified within the Red List assessments using a hierarchical process by coding an individual threatening event to the finest resolution level possible (IUCN, 2016). Major threats were then summarized and the proportion of threatened and near threatened species was explored for all species, as well as by clupeiform family and major habitat system. The proportion of threatened and NT species is expressed using both a midpoint and a range to address the uncertainty surrounding the true status of a DD species. The midpoint was calculated by removing the species listed as DD, whereas the lower and upper bound were calculated by either excluding or including the DD species with the threatened species, respectively. The lower bound assumes that none of the DD species are threatened, while the upper bound assumes that all DD species are threatened.

A species was assigned a major habitat category using the information in the Red List assessments. Given the known or suspected tolerance for salinity fluctuations exhibited by many clupeiforms, I modified the IUCN Red List classification scheme from two aquatic

categories (freshwater, including inland estuarine waters, and marine, including coastal estuarine waters) to three categories. Therefore, the freshwater system includes those known to occupy only freshwater environments and the marine system includes species restricted to marine waters. I added a third, euryhaline category that includes estuarine species, diadromous species, and species known or suspected to tolerate changes in salinity.

Distribution Mapping Methodology

Maps were created for each species using ArcMAP 10.3 by compiling data from published and grey literature, expert knowledge, and online databases (e.g., FishNet2; OZCAM; GBIF) on known occurrence along with habitat and depth limits. As marine clupeiforms are primarily coastal, the distribution polygons for strictly marine species were standardized using a base map that represents either the 200 m bathyline or 100 km from the shore, whichever is further from the coast. Bathymetric layers were extracted from two global level sources, the National Geophysical Data Center's ETPO1 (Amante and Eakins, 2008) and the General Bathymetric Chart of the Oceans (GEBCO: IOC et al., 2003). Maps for freshwater species were created using hydrobasins because these areas are considered as minimum management units for freshwater conservation (Lévêque et al., 2008; Carrizo et al., 2013). For species that utilize both marine and freshwater habitats (e.g., diadromous species), maps separately followed the marine and freshwater protocols, and were then combined to encompass the entirety of the species' range.

Species Richness Analyses

Global maps of overall species richness, Data Deficient richness, and richness of elevated concern species were created using ArcMap 10.3 based on two biogeographic systems. Species with a freshwater extent ($n = 74$) were summarized within the Global HydroBASINS (Leher and Grill, 2013), using Level 3, the largest river basins of each continent. Species with a marine extent ($n = 130$) were summarized within the Marine Ecosystems of the World at the province level (Spalding et al., 2007). This shapefile was modified to include a region for the Caspian Sea, as it is excluded from the Global HydroBASINS and Marine Ecosystems of the World. Freshwater and marine layers were merged to summarize species with both a freshwater and marine extent ($n = 201$).

RESULTS

Global IUCN Red List status of clupeiforms

The best estimate of the proportion of elevated concern for clupeiforms species is 11% ($n = 33$), which includes those assessed as threatened or Near Threatened. Given the uncertainty of an appropriate Red List Category for all Data Deficient (DD) species, the true proportion of elevated concern species could lie between 8 – 34%. Of all species ($n = 405$), three (0.7%) are listed as Critically Endangered (CR), 11 (2.7%) as Endangered (EN), 13 (3.2%) as Vulnerable (VU) and six species (1.5%) as Near Threatened (NT) (Fig. 2). Species are primarily listed as threatened or Near Threatened due to a restricted range size with an ongoing threat (criterion B; $n = 18$) or due to population decline (criterion A; $n = 10$); two species (*Sardinella tawilis* and *Alosa vistonica*) are listed as threatened under both criteria A and B (Appendix A).

Three species are listed as Vulnerable (VU) given that they have a very restricted range and a serious plausible future threat (criterion D). Of the remaining 372 species, 266 (65.7%) are categorized as Least Concern (LC) and another 106 (26.2%) are considered as Data Deficient (DD).

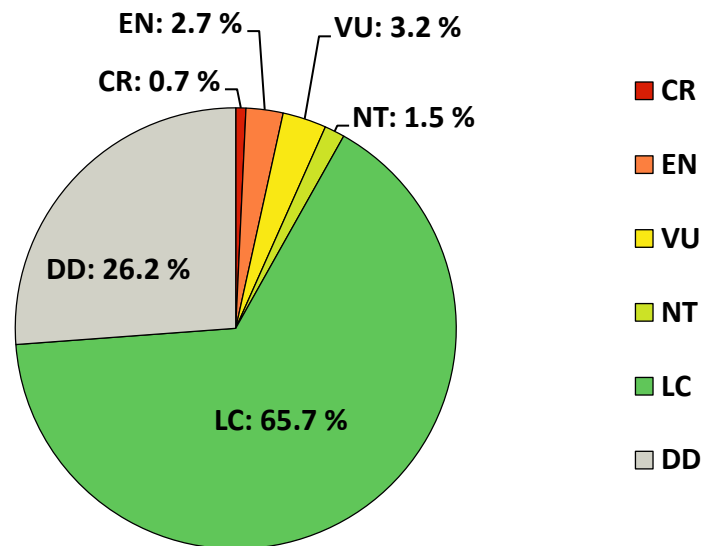


Fig. 2. Proportion of species (n = 405) listed in each Red List Category. Abbreviations of Red List Categories are as follows: CR = Critically Endangered, EN = Endangered, VU = Vulnerable, NT = Near Threatened, LC = Least Concern and DD = Data Deficient.

Among the families of clupeiform fishes, the family Denticipitidae consists of only one species (*Denticeps clupeoides*), which is listed as VU; as such, it is the family with the highest proportion of elevated concern overall (Fig. 3). However, excluding *D. clupeoides*, the Clupeidae

(26 of 195 species; midpoint = 16.6%) has the highest proportion of elevated concern species, followed by the Engraulidae (5 of 154 species; midpoint = 4.9%) and the Pristigasteridae (1 of 36 species; midpoint = 3.8%). None of the Chirocentridae ($n = 2$), Dussumieriidae ($n = 10$) or Sundasalangidae ($n = 7$) are considered threatened. However, the high proportion of DD species, especially within the Sundasalangidae, may be obscuring trends in threat patterns and compromising the accuracy of the overall conservation status estimated for these species.

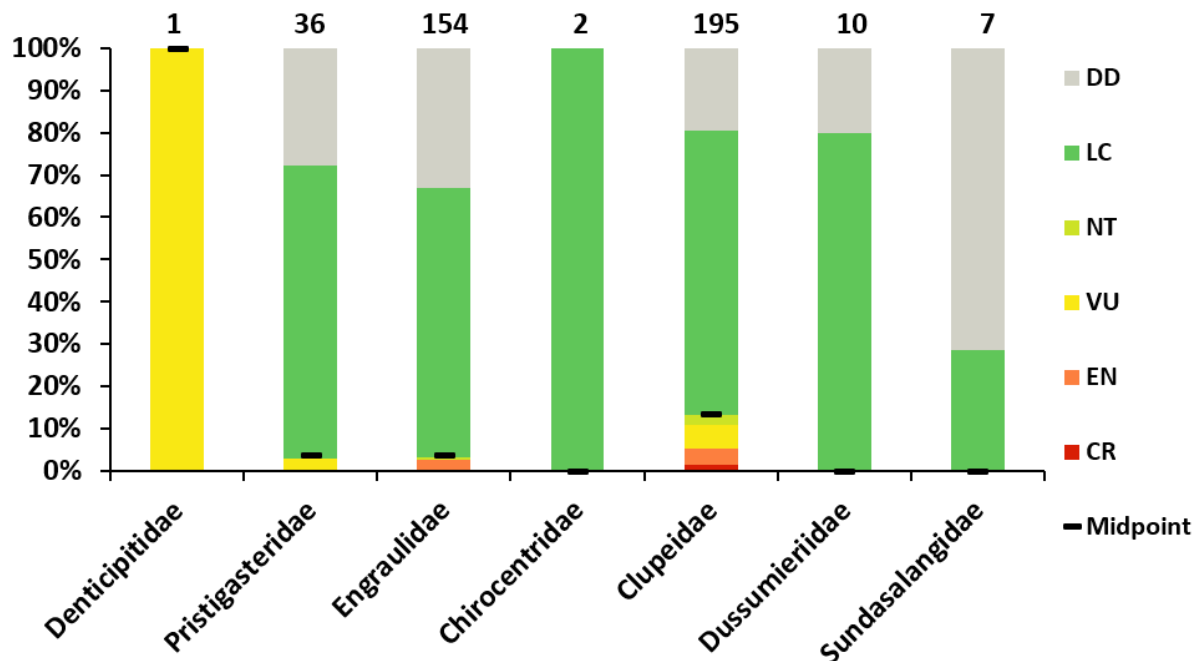


Fig. 3. Proportion of species listed in each Red List Category separated by family. The total number of species in each family is represented by the number at the top of each bar. Abbreviations of Red List Categories are as follows: CR = Critically Endangered, EN = Endangered, VU = Vulnerable, NT = Near Threatened, LC = Least Concern and DD = Data Deficient. The midpoint is represented by the black bar and was calculated by the following equation: $(CR + EN + VU) / (Total - DD)$.

Species classified as euryhaline (i.e., diadromous or estuarine) constituted nearly half of the species within the order (n = 201; 49.6%), followed by marine (n = 130; 32.1%) and freshwater species (n = 74; 18.3%) (Fig. 4). Euryhaline species have the largest proportion of Least Concern species (n = 147; 73.1%) followed by marine (n = 80; 61.5%), and then by freshwater species (n = 39; 52.7%). Overall, despite having the lowest number of representatives, the freshwater inhabitants have the highest proportion of elevated concern species (n = 16; 21.6%), more than double that of the species inhabiting marine and euryhaline habitats combined (5.4% and 5.0%, respectively). Additionally, all species assessed as CR, the highest threat level, are found in freshwater habitats.

Major threats

Of the 405 species, 144 have at least one coded threat; the remaining 261 species either have no major threats causing significant impacts or threats are unknown for these species. Overall, the most prominent threat by a significant margin impacting all clupeiforms, is exploitation (Fig. 5). Pollution and natural system changes (e.g., dams) impact nearly the same number of species (47 and 42, respectively). Despite having the highest proportion of LC species, euryhaline species are disproportionately impacted by pollution and natural system modifications relative to freshwater and marine species. For example, the number of euryhaline species impacted by one of these threats is more than 1.5 times the number of fresh and marine species combined. Likewise, euryhaline species impacted by both threats (pollution and natural system modifications) is double that of the combined number of marine and freshwater species impacted by these factors. Climate change and invasive species make up the fourth and

fifth most common threat to all species, respectively. However, invasive species impact more threatened and NT species than climate change.

Spatial Analyses

Global species richness follows two general distribution patterns; a longitudinal gradient, where the highest tropical richness is within the Indo-West Pacific, and a latitudinal gradient where richness decreases with an increasing latitude from the tropics. The highest global species richness of all 405 clupeiforms is located along the coast of India and throughout the Indo-West Pacific from the eastern Andaman Sea, east to the Philippines, Indonesia and northeastern Papua New Guinea (Fig. 6A). High richness also occurs in the central eastern Pacific from Mexico to northern Peru, and the central western Atlantic from the greater Caribbean to northern Brazil. Areas of lowest species richness are within the southern and northernmost limits of the global range for species of this order (e.g., the Arctic and sub-Arctic region, and north of the Southern Ocean), further inland (e.g., the rivers of China, Australia, and parts of Africa), and off Polynesian Islands in the central and south Pacific (e.g., Hawaii, New Zealand, Society Islands, etc.).

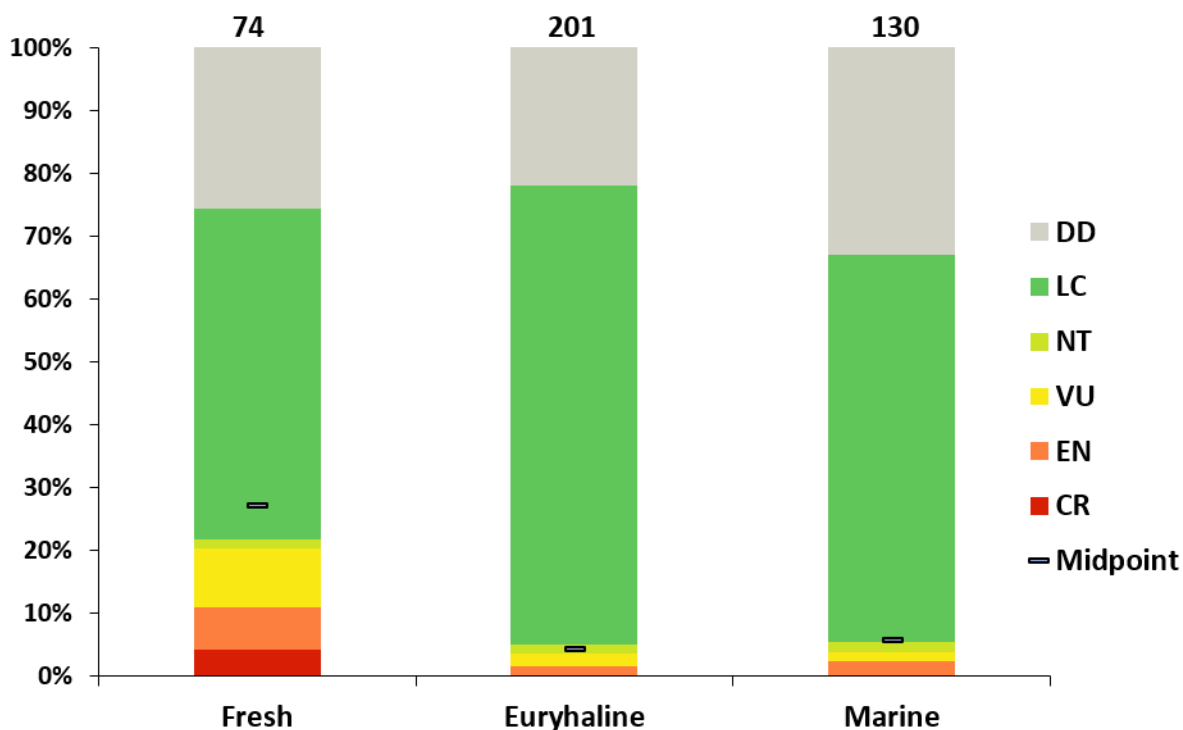


Fig. 4. Proportion of species listed in each Red List Category by major habitat (fresh, euryhaline or marine). The total number of species is represented by the number at the top of each bar. Abbreviations of Red List Categories are as follows: CR = Critically Endangered, EN = Endangered, VU = Vulnerable, NT = Near Threatened, LC = Least Concern and DD = Data Deficient. The midpoint is represented by the black bar and was calculated by the following equation: $(CR + EN + VU) / (Total - DD)$.

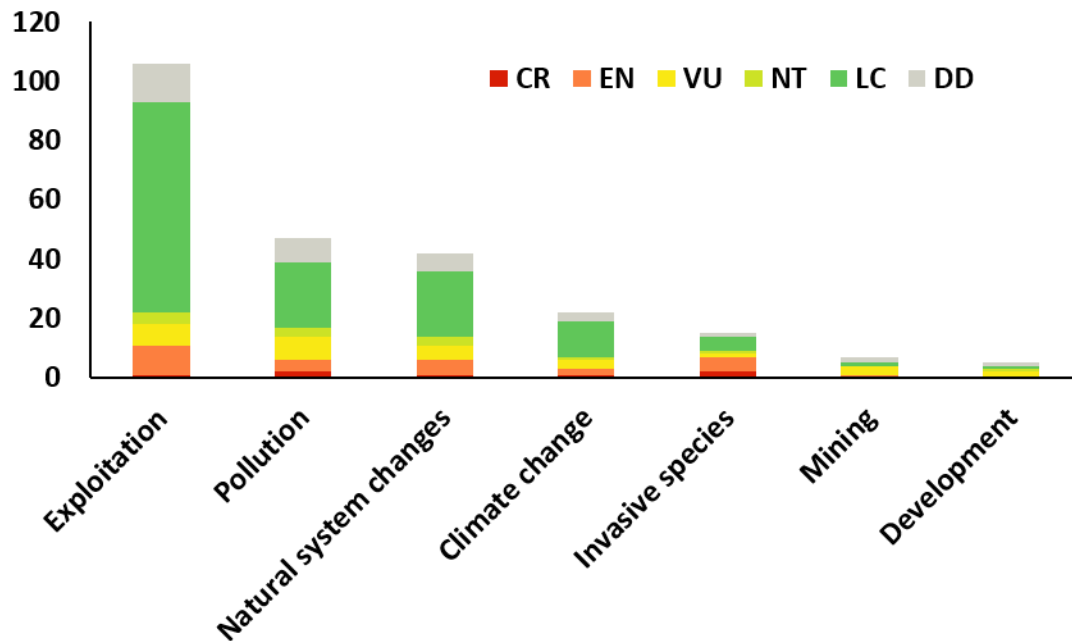


Fig. 5. Number of species impacted by major threats. Each threat is represented by the number of species listed in each Red List category. Threats that impact less than five species (Human intrusion and Transportation) are excluded. Abbreviations of Red List Categories are as follows: CR = Critically Endangered, EN = Endangered, VU = Vulnerable, NT = Near Threatened, LC = Least Concern and DD = Data Deficient.

In general, richness of DD species closely follows that of the total species richness (Fig. 6C). However, the richness of DD species is higher in northern Australian rivers relative to the total species richness. In contrast, the high species richness in Europe, eastern United States and South American rivers is not mirrored by high DD species richness.

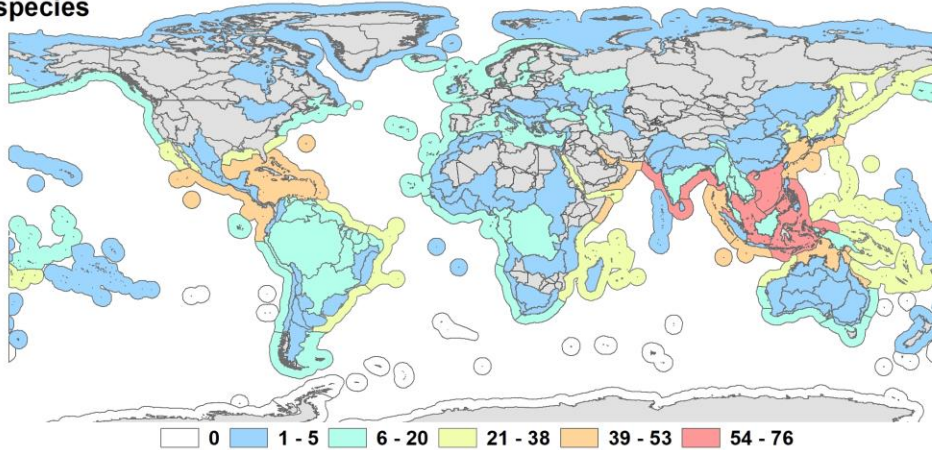
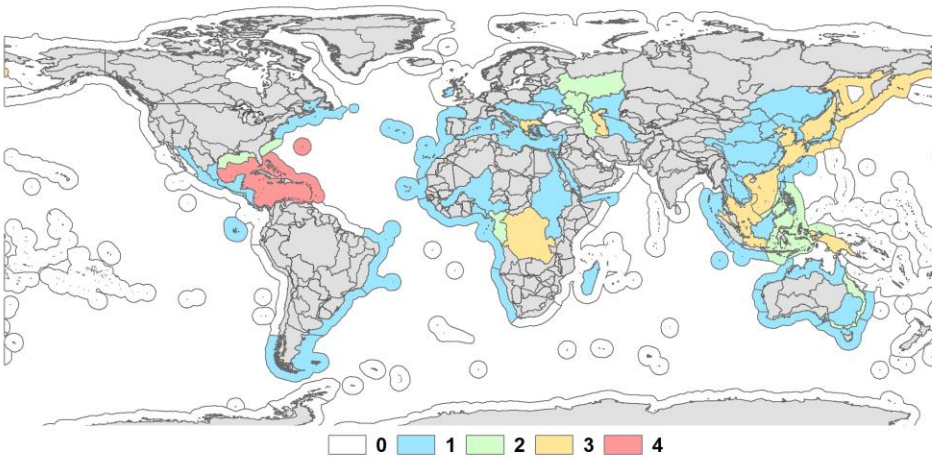
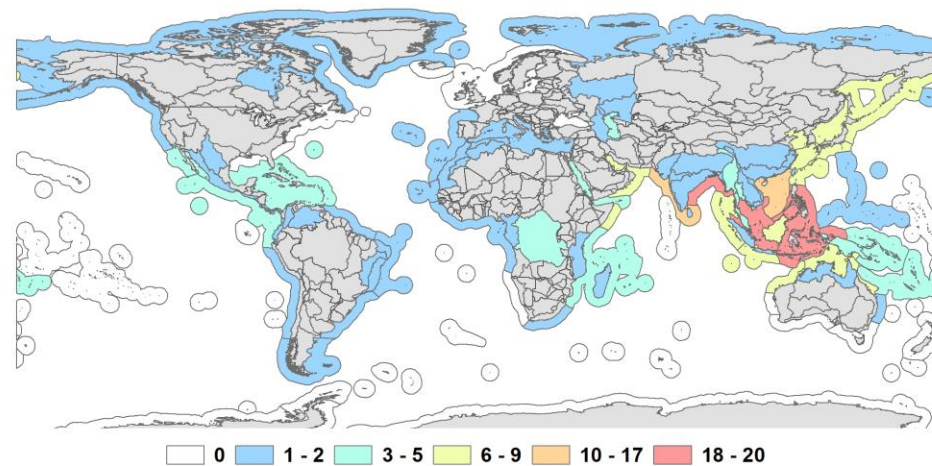
A - All species**B - Threatened and NT species****C- DD species**

Fig. 6. Number of clupeiforms in each Large Marine Ecoregion (LME) and freshwater hydrobasin for A) All species, B) all species of elevated concern (CR, EN, VU, NT), and C) all Data Deficient species. Colors correspond to the number of species listed at the bottom of each map. The Marine Ecosystems of the World (MEOW) at the province level was used for marine species, Hydrobasins of the world at level three was used for freshwater species. The freshwater and marine extents were created separately and merged to represent the total global extent for euryhaline species.

Conversely, the highest richness of species of elevated concern (threatened and NT, $n = 33$) occurs within the greater Caribbean (Fig. 6B). Other areas of high richness of species of elevated concern are along the western Pacific continental coast (Russia south to Indonesia), and inland areas including the Caspian Sea, rivers of Croatia to Greece and Bulgaria, the Congo River in Central Africa, and the rivers of Borneo. A low richness of species of elevated concern is found along the northeastern United States, within Central America, along the eastern and southern coasts of South America, the western coast of Africa, parts of Europe including the Mediterranean and Black Seas, southern Australia, and within some freshwater areas in China and Russia.

DISCUSSION

When compared to other economically and ecologically important fish groups globally assessed using the IUCN Red List methodology (e.g., Collette et al., 2011; Sadovy de Mitcheson et al., 2013; Comeros-Raynal et al., 2016), clupeiforms have the lowest percentage of threatened and Near Threatened (NT) species overall. Just over 8% are currently known to be at high risk of potential future extinction as compared to roughly 18% of tunas and billfishes (Collette et al., 2011), 26% of groupers (Sadovy de Mitcheson et al., 2013) and 17% of sparids (Comeros-Raynal et al., 2016). However, major threats to clupeiforms are nearly identical to those found in previous analyses of the conservation status of other fishes (e.g., Roberts and Hawkins, 1999; Reynolds et al., 2005; Dulvy et al., 2009; Harnik et al., 2012), with exploitation as the leading threat for all clupeiforms in all habitats. While exploitation may be the most prolific threat by impacting the highest number of clupeiforms, pollution may be the most

detrimental, as it affects a greater number of species assessed as Critically Endangered (CR) (Fig. 5).

The lower proportion of threatened species in clupeiforms compared to other bony fishes may be a function of uncertainty of conservation status and is likely an underestimate of the true threatened status. The high percentage of data deficiency in clupeiforms (26.2%) surpasses that of the tunas and billfishes (Collette et al., 2011) and sparids (Comeros-Raynal et al., 2016), which have less than 20% of species that are DD. A DD listing is most often related to taxonomic uncertainty, low number of known specimens, unknown geographical range, or inability to quantify a threat or decline in population (IUCN, 2012), all of which occur within the clupeiforms.

For individual species, the paucity of data on distribution, status, ecology and threats may be a consequence of taxonomic uncertainty (IUCN, 2017). For example, *Alosa curensis* is DD because it was previously recognized as a synonym of *Alosa brashnikowi* and is known only from a few specimens. Thus, information associated with what was previously thought to be the single global population of the nominal *A. brashnikowi*, may not also be applicable to *A. curensis*. Challenges associated with taxonomic uncertainty or recent revision, such as estimating decline or geographic range, may allow species-specific threats to go undocumented.

The high proportion of DD clupeiform species coincides with geographic areas of both dense clupeiform biodiversity and areas of depressed economic status. In general, global biodiversity is unevenly distributed; the most biodiverse places are often areas of high human

populations of relatively low per capita income (Baille et al., 2004; Brooks et al., 2006) and tend to have the highest number of threatened species (Hoffmann et al., 2010). This pattern is reflected in clupeiforms. Countries with high human populations and high biodiversity are less likely to have financial resources available for research and conservation purposes (Baille et al., 2004). In contrast, countries such as those in the advanced economies of Europe invest substantially in conservation research and management and have few globally threatened species (Baille et al., 2004), including clupeiforms.

In many parts of the world, particularly in highly biodiverse areas, stock assessments and fishery effort data are lacking or unreported for many clupeiforms. Where data are available, it is often in the form of raw fishery landings (FAO, 2016) or reconstructed catches (Pauly and Zeller, 2016a). These landings often include many species lumped together because many clupeiforms that co-occur look very similar, are difficult to identify taxonomically, and are known to school together in some cases (e.g. sardines and anchovies: Bakun and Cury, 1999). Teasing apart landings from multi-species fisheries is a difficult task and when identifications contain many errors can lead to a false estimation of species-specific catch data (Gaichas et al., 2012). Exploitation is a major threat to over 25% of clupeiform species and this may be an underestimate given uncertainties in catch data and the population status of DD species (26.2%). Clupeiforms also contribute to many unreported artisanal fisheries (Whitehead, 1985; Whitehead et al., 1988), represent a significant portion of bycatch in other industrial trawl fisheries (e.g., Stobutzki et al., 2001) and are taken in illegal, unreported and unregulated fisheries (IUU: Agnew et al., 2009). Accidental and IUU fishing, along with lumped landings adversely affect our ability to quantify global fishing pressure on these species. It can further

impact conclusions drawn regarding population trends by underestimating true catches (Pauly and Zeller, 2016b) and ultimately impacting the efficacy of conservation or management decisions.

The highest concentration of threatened species in this analysis is centered in the Caribbean region, but this estimate does not take into consideration uncertainty concerning species listed as DD. The highest species richness and number of species listed as DD is concentrated in the central Indo-West Pacific region. Given that the Caribbean and the Indo-West Pacific are both areas of high species richness, but that about one-tenth of the Caribbean species are assessed as DD compared to roughly one-third of Indo-West Pacific species, we therefore know more about the species in the Caribbean in general. Currently, clupeiforms in the Caribbean would benefit most from threat mitigation as this region has the highest number of threatened and NT species present. It has been noted that the most diverse areas often have the highest number of threatened species (Baille et al., 2004). As data become available to adequately assess species currently listed as DD, it is possible that we may find a higher proportion of threatened and NT species within the Indo-West Pacific rather than within the Caribbean. However, currently clupeiforms in the Indo-West Pacific region may benefit most from emergent research to fill in our knowledge gaps presented by the high number of DD species.

In addition to the high proportion of DD species, traditional perceptions of intrinsic life history traits have been an impediment to the conservation concerns of clupeiforms overall. Their typical high fecundity, multiple spawning, and early age at maturation are regarded as resilience factors even though these traits often do not reflect vulnerability to extinction

(Jennings et al., 1998; Kindsvater et al., 2016; Sadovy, 2001; Juan-Jorda et al., 2012; Comeros-Raynal et al., 2016). For example, the Pacific herring (*Clupea pallasii*) is a widely distributed species that is exploited to a varying degree throughout a large portion of its range. In some regions where this species has experienced drastic declines, subpopulations have not recovered even decades after fishing pressure has ceased (see Hay et al., 2001 for description of Yellow Sea and Hokkaido – Sakhalin herring). Overall, the intrinsic life history characteristics of many clupeiforms may be providing them with a buffer against extinction relative to other taxa such as sharks, rays, tunas, billfishes and groupers, but this buffer does not hold for all clupeiform subpopulations.

Synergistic influences of threats can be detrimental to the survival of a population (Brook et al., 2008). Often, a freshwater or euryhaline species is threatened by both pollution and natural system modifications, indicating a potential cumulative effect between threats. This interaction was not explored in this study. Genera with many anadromous representatives such as *Alosa* and *Tenualosa* appear to be most negatively impacted by one or both threats (e.g., Freyhof and Kottelat, 2008b; NatureServe, 2013; Di Dario, 2018b; Mohd Arshaad et al., 2018). In line with previous studies of other freshwater fishes (e.g., Collen et al., 2014), freshwater clupeiforms have over double the proportion of threatened and NT species compared with marine and euryhaline clupeiform species combined (% threatened and NT = 21.6% of FW, 5.4% for marine, and 5.0% for euryhaline). Given that all species listed as Critically Endangered (CR) are freshwater clupeids, the responses of these fishes to stresses should be examined more closely.

Given the overall importance of clupeiform fishes and their ubiquity as an important fishery resource, there should be concern regarding these species despite the lower percentage of threatened species compared to other fish groups of higher economic value. Many species threatened with exploitation have monitoring in place, which may not be sufficient; therefore, it is urgent that the efficiency of current management measures is evaluated. An increase in species-specific landings and catch statistics would also further improve our abilities to assess exploitation as a threat for a larger number of species. Additionally, a few large-scale industrial fisheries, such as those for the Peruvian anchoveta (*Engraulis ringens*) and for the Pacific herring (*Clupea pallasii*), may benefit from increased multi-national cooperative regulations. Species listed as an elevated conservation concern should be monitored more closely and anthropogenic pressure strictly managed although prioritizing research and conservation initiatives in areas of high biodiversity can be difficult given limited resources. Fishery managers in areas with a large proportion of exploited DD species should prioritize research initiatives to fill gaps in our understanding of these species. At a local level, species with limited ranges, such as *Alosa killarnensis* (Freyhof and Kottelat, 2008) and *Sardinella tawilis* (Santos et al., 2018), should be a priority for stringent protection, especially regarding habitat quality, which impacts mainly freshwater and estuarine species.

CHAPTER 3

CHARACTERIZING CLUPEIFORM THREATS, LIFE HISTORY TRAITS, AND HABITAT PREFERENCE TO INFORM MANAGEMENT OF DATA DEFICIENT SPECIES

INTRODUCTION

Aquatic biodiversity supports ecosystem health and the ecosystem services we rely upon (Brooks et al., 2006). Covering more than 71% of the Earth's surface and even more inhabitable space by volume, freshwater, estuarine and marine environments supply more than 40 million jobs with an estimated contribution of several trillion dollars annually to the global economy (Darwall et al., 2011; FAO, 2018). The services provided by our aquatic ecosystems include food provisioning, climate and atmospheric regulation, carbon sequestration, flood control, storm protection, nutrient cycling and waste removal (Aladin et al., 2005; Worm et al., 2006; Palumbi et al., 2009). Despite our reliance on aquatic resources, conservation initiatives have lagged far behind those of the terrestrial realm (Hilton-Taylor et al., 2009; Polidoro et al., 2009; Darwall et al., 2011). Resource limitation and exigent needs have resulted in prioritized conservation within marine and freshwater environments to support species groups such as mammals (Freeman, 2008), sharks (Dulvy et al., 2008) and turtles (Seminof and Shanker, 2008), or regions of most concern such as the Mediterranean Sea (Smith and Darwall, 2006; Abdul Malak et al., 2011). However, the conservation and management dilemmas of priority species cannot be solved without also incorporating the complexities and trade-offs of the ecosystem,

including the effects of predator-prey interactions. Thus, recent interest has shifted toward ecosystem-based management to account for ecological, economic and societal challenges associated with fisheries management (Link, 2002; Pikitch et al., 2004, 2012, 2014; Palumbi et al., 2009).

Clupeiforms, including herrings, shads, sardines, anchovies and their relatives make up the bulk of what we consider to be forage fishes (Whitehead, 1985) as they are a major food source for many aquatic predators (Pikitch et al., 2012). In addition to providing support for many other, often commercially important species, clupeiforms make up lucrative fisheries on every continent where they are distributed (Whitehead, 1985). They have supported the world's largest fishery in history (the Peruvian anchoveta, *Engraulis ringens*: Whitehead, 1985) and continue to support substantial fisheries worldwide (FAO, 2018). Clupeiform fisheries make major contributions to international industrial commodities and provide nutritional security for billions of people globally each year (Alder, 2008; Tacon and Metian, 2009).

Management and conservation regulations for clupeiforms are often lacking in the places where needed most, such as in tropical areas of highest biodiversity with the lowest capacity to fund such initiatives (Worm and Branch, 2012). Current management efforts are often species-specific for the well-known or heavily exploited species of clupeiforms. Management objectives using Maximum Sustainable Yield, biomass cutoff limits, or gear restrictions have worked well for managing and rebuilding some stocks including, for example, the Pacific herring, *Clupea pallasii* (WDFW, 2018) which is in a low biodiversity temperate region. However, regions of high biodiversity of clupeiforms generally lack species-specific management capabilities.

In tropical regions with high biodiversity, multispecies catches of clupeiforms are often very difficult to identify to the species level (Whitehead, 1985; FAO, 2018) confounding fishery management efforts. Instead, species are categorized with variable resolution into taxonomic groups by genus or family, or into functional groups such as ‘forage fishes’ or ‘small-to medium-sized pelagics’, with similar management strategies applied to all species in the group (Beverton, 1990; Patterson, 1992). However, clupeiforms express an extensive spectrum of diversity of life history features among species (Whitehead, 1985; Bloom and Egan, 2018). For example, maximum known lengths vary from about 2 cm in species of *Sundasanx* (Roberts, 1981) to 100 cm in *Chirocentrus nudus* and *C. dorab* (Munroe et al., 1999) with known longevity spanning from less than one year in *Spratelloides gracilis* (Milton et al., 1991; Meekan et al., 2006) to up to 25 years in *Sardinops sagax* (Whitehead, 1985). Total geographic distributions extend from a single small lake as in the case of *Alosa killarensis* (Freyhof and Kottelat, 2008) to the entire Indo-West Pacific in *Sardinella gibbosa* (Whitehead, 1985). Clupeiforms span maximum depths from less than 10 meters in *Anchoa analis* (Whitehead et al., 1988) to more than 400 meters in *Clupeonella grimmi* (Aliasghari et al., 2017). Habitat preference and tolerance of ecological conditions also vary widely in species throughout the order with representatives from freshwater, estuarine, marine and diadromous groups (Whitehead, 1985).

The highly variable life history traits of clupeiforms suggests that diverse management approaches that account for this variation may help solve management hurdles in data poor fisheries (Siple et al., 2018). Identifying differences and similarities among species to group them based on shared life history, ecological characteristics, and response to threats may

provide tractable management strategies. Particularly in areas of high clupeiform diversity where ecological and biomass data are relatively limited, traditional management strategies are challenged by data limitation (Smith et al., 2009; Carruthers et al., 2014). Methods dealing with data-poor fisheries are often in the form of a ‘Robin-hood’ approach, where borrowed information from a similar, well-known species is used to make decisions for the lesser known species (Smith et al., 2009). However, given the diversity of clupeiforms, (Whitehead, 1985; Bloom and Egan, 2018) information from many of the well-known species may not be applicable to those that are data limited. Therefore, an alternative ‘basket’ approach, where similar, data-poor species are binned and managed together (Smith et al., 2009), may prove to be the more useful management approach.

METHODS

The IUCN Red List is a globally recognized standard for assessing species-level extinction risk and acts as a baseline from which to monitor change (Vié et al., 2009). IUCN Red List assessments were conducted for the 405 valid clupeiform species following Eschmeyer et al., (2017) and taxonomic expertise (Appendix A). Four nominal species were described as new or elevated from synonymy since December 2018; species Red List assessments for these taxa have been completed, but the information from these assessments is not included in this analysis. Each assessment includes expert-vetted information on geographic distribution, population trends, ecology, potential threats and existing conservation measures (for detailed Red List methodology, see Ch. 2 and Appendix B).

Multivariate analyses are widely used in ecology to address increasingly complex questions and are used here to explore patterns within available clupeiform data. These sophisticated ordination techniques are required to reduce dimensionality and visualize patterns in multivariate data (Anderson and Willis, 2003). Analysis options include unconstrained methods such as principal component analysis, principal coordinate analysis and nonmetric multidimensional scaling, whereas constrained methods include such analyses as canonical discriminant analysis, canonical correlation analysis and canonical analysis of principal coordinates (Anderson and Willis, 2003). Unconstrained methods are typically used to discover unknown or suspected patterns in data (Anderson and Willis, 2003). In general, constrained ordinations use *a priori* hypotheses from which to produce a plot so that a matrix of response variables such as community or species data can be related to some predictor variable or variables, such as measured ecological data. Canonical analysis of principal coordinates (CAP) is a flexible constrained method that allows the use of any distance or dissimilarity measure and accounts for underlying correlation structuring among response variables (Anderson and Willis, 2003).

To explore which known characters from well-studied species can be used to help bin together and possibly improve conservation measures for poorly-known species, two CAP analyses were conducted. Species-specific data were exported from the International Union for Conservation of Nature (IUCN) Red List of Threatened Species open-sourced database (available at: www.iucnredlist.org) and organized into matrices. These matrices of species data were then imported into PRIMER-e with PERMANOVA+, a multivariate statistical software for ecological sciences (Anderson et al., 2008; Clarke and Gorley, 2015). A Bray-Curtis similarity test was then

run on the matrix data to quantify the similarities between species relative to the input variables. I tested for significance between groups in *a priori* hypotheses using a permutational multivariate analysis of variance (PERMANOVA). Finally, CAP was used to visually compare species assemblages and to ascertain which axes in a multivariate space effectively discriminate between *a priori* groups. A unit circle and vectors of the response variables were overlaid on the CAP figure to determine which variables most influence the observed patterns.

Additionally, I explored the effects of habitat on the maximum size of 394 exploited and non-exploited species (all species for which maximum size data were available). I used a two-way analysis of variance (ANOVA), followed by a Tukey HSD post-hoc test for pairwise comparisons. Analyses were completed using the R Project version 3.4.3 (R Core Team, 2017) and RStudio.

To determine how major threats vary as a function of the primary occupied habitat system (freshwater, marine or euryhaline), the species with major threats identified as part of the IUCN Red List assessments (n=144) were included in a CAP analysis. The remaining 261 species have either no major threats identified, or threats are suspected but unconfirmed. The threats identified for each species were organized based on the IUCN threat classification hierarchy (IUCN, 2012) and include exploitation, climate change, mining, human disturbance, invasive species, natural system modifications, which primarily refers to dam placement or water abstraction, pollution, residential and commercial development, and transportation corridors.

I included two explanatory variables: the IUCN Red List categories and the primary occupied habitat system (freshwater, marine, euryhaline). The IUCN Red List categories include Data Deficient (DD), Least Concern (LC), Near Threatened (NT) and the threatened categories: Vulnerable (VU), Endangered (EN), Critically Endangered (CR) Extinct in the Wild (EW) and Extinct (EX) (IUCN, 2012; Chapter 2, Fig. 1). Given the high degree of plasticity in salinity tolerance that is known or suspected for many clupeiforms, I modified the habitat system classification used in the IUCN Red List methodology. Instead of including just two aquatic categories (freshwater, including inland, brackish and upper estuarine waters, and marine, including coastal or lower estuarine waters: IUCN, 2013), I added a third, euryhaline category that separates the estuarine component from the Red List classifications. Therefore, the freshwater system refers to species currently known to occupy freshwater habitats with no documented tolerance of an increased salinity; marine species are known to tolerate only marine waters. The euryhaline category comprises a variety of species including strictly estuarine species, diadromous species and those known or presumed to tolerate wide salinity fluctuations. Separating estuarine and diadromous groups was problematic as several species could not be easily classified into one of these two groups. For example, a species may be known to tolerate a wide range in salinity with records from both marine and freshwaters, but diadromy is unconfirmed. Therefore, a single, euryhaline category was implemented to account for all estuarine species and those known to withstand salinity fluctuations to a varying degree.

To address which life history characteristics are most important in determining clustering of species of elevated conservation concern, life history and ecological traits of all threatened and NT species (n=33) were included in a second CAP analysis. I included the

following as response variables: maximum standard length, number of coded threats, habitat system preference (freshwater, marine or euryhaline), habitat specificity (generalist or specialist) and proxies for distribution and relative clupeiform richness in an area where a species is found. Habitat system preference and habitat specificity (generalist or specialist) are numerical variables coded within the input matrix. Habitat system data were carried over from the species-threat matrix. Within Red List assessments, the number and type of habitats occupied by a species are coded based on information from available literature; this information was pulled from assessments and used to assign a species as either a habitat generalist or specialist. The attribution of a species to one of the two categories follows Stump et al., (2018); if a species occupies only one coded habitat type (i.e., freshwater lakes/rivers) it was considered a specialist and if it occupies more than one habitat type (i.e., freshwater rivers and coastal marine waters), it was considered a generalist. A numerical estimate of total geographic distribution area was measured by determining the number of countries where a species is known, inferred or suspected to occur. Using the distribution shapefiles of each species, estimated clupeiform diversity in an area where a species is found was measured in ArcMAP 10 by adding the number of other clupeiforms that have an overlapping distribution with an individual species.

RESULTS

For clupeiforms with at least one identified threat ($n = 144$), the CAP analysis revealed a separation of species assemblages as a function of habitat system, specifically between threats

impacting freshwater and marine species (Fig. 7). The euryhaline species assemblage overlaps with both the freshwater and marine species clusters. This partitioning is supported by a significant difference between habitat systems (PERMANOVA, $p = 0.001$, 999 permutations). A pair-wise comparison indicates that threats experienced by marine species are significantly different from those of both freshwater ($p = 0.001$) and euryhaline species ($p = 0.001$), but euryhaline and freshwater threats are not significantly different from each other ($p = 0.432$).

Three of the nine major threats identified were highlighted as the most pervasive by examining the vector length of explanatory variables: exploitation, pollution and natural system modifications. In general, the primary threat to marine species is exploitation, impacting 34 out of 38 species (89.5%) impacted by a threat. Comparatively, freshwater species tend to be more collectively impacted by pollution and natural system modifications than marine species (Fig. 7), both of which individually affect 37.5% of the freshwater species that are impacted by at least one major threat. However, exploitation is the most prevalent threat to freshwater clupeiforms, impacting 62.5% (20 of 32 species) of those with a recorded threat. Euryhaline species are impacted by all three major threats. Exploitation impacts the largest proportion of euryhaline species (70.3%), while the proportion affected by pollution and natural system modifications (41.9% and 36.5%, respectively) rivals that of the freshwater species. Of the marine species with a recorded threat, only two, *Harengula jaguana* and *Sardinella maderensis*, are impacted by pollution (Tous et al., 2015; Munroe et al., 2019). Both species occur at or near estuary mouths and are impacted by various sources of pollution, such as agricultural and industrial effluents. Likewise, an additional two marine species, *Anchoa helleri* and *Nemalosa*

japonica, are impacted by natural system modifications that include heavy coastal land reclamation and water diversions (Iwamoto et al., 2010; Di Dario, 2018a).

Seven distinct groups have been identified among the clupeiforms with major known threats based on the string of threats impacting each species (Fig. 7). Except for group 7, at least one species from each primary habitat (freshwater, euryhaline or marine) is represented in every group. All species within a group share similar threats. For example, groups 1, 3, and 6 represent 14, 64, and 12 species, respectively that are all impacted by one of the most prominent threats (natural habitat system changes, exploitation, or pollution). Species within group 1 are primarily only impacted by changes to the natural habitat such as dams and water abstraction. Group 3 represents species that are all primarily threatened by exploitation and group 6 represents those mostly impacted by pollution. Groups 2 and 7 are characterized by species likely impacted by two of the most prominent threats, pulling them in between the two threat vectors in space. For example, group 2 represents 11 species threatened by both exploitation and natural system modifications, whereas species in group 7 are impacted by natural system changes and pollution. Groups 4 and 5 include species threatened by two of the most prominent threats and by at least one of the less influential threats, such as climate change or mining.

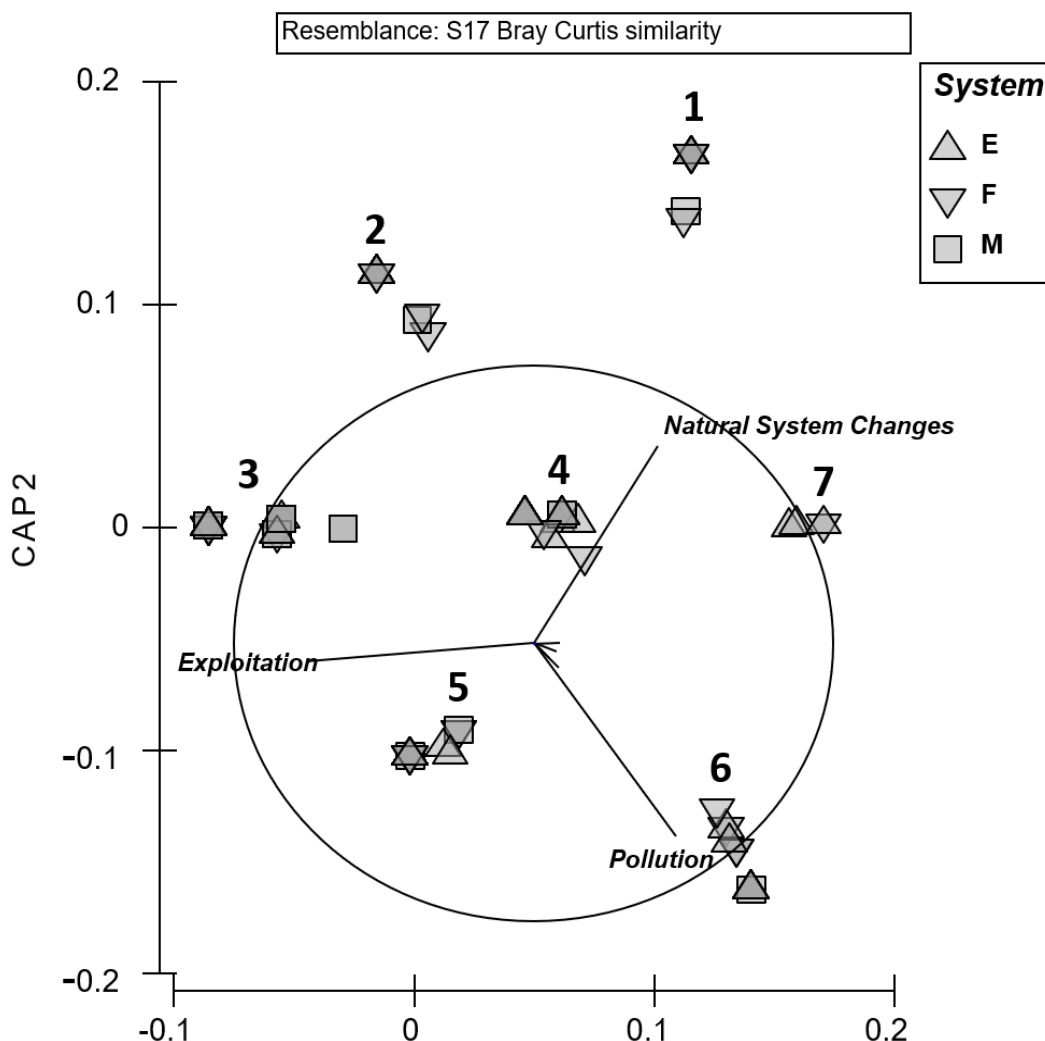


Fig. 7. Canonical Analysis of Principal Coordinates (CAP) ordination of species with known threats in multivariate threat space by primary habitat system ($n = 144$). Habitat system abbreviations are E = Euryhaline, F = Freshwater and M = Marine. The three most prominent threats – Exploitation, Pollution and Natural system modifications are labelled; major threats including climate change, mining, human disturbance, invasive species, residential/commercial development and transportation corridors impact a smaller proportion of species and are not labelled. Each species is represented by a single symbol; however, as there is substantial overlap, symbol transparency was set at 50% to indicate where overlaps occur. Thus, symbols that appear darker in color represent more species than lighter symbols. Each group of species, indicated by the numerical value above the group, represents a different number of species: group 1 – 14 species (E = 7, F = 6, M = 1); group 2 – 11 species (E = 6, F = 4, M = 1); group 3 – 64 species (E = 25, F = 9, M = 30); group 4 – 21 species (E = 16, F = 2, M = 3); group 5 – 18 species (E = 10, F = 6, M = 2); group 6 – 12 species (E = 7, F = 4, M = 1); group 7 – 4 species (E = 3, F = 1, M = 0). The direction and length of the vectors represent the relationship between the ordination axes and threat type.

Results of the CAP analysis on known life history characteristics of elevated concern species ($n = 33$) show slight partitioning between the three habitat systems (freshwater, marine, euryhaline), but is most notable between marine and freshwater fishes (Fig. 8). A significant difference occurs between characteristics of species within the three habitat systems (PERMANOVA, $p = 0.001$, 999 permutations). Based on a pair-wise comparison, characters exhibited by freshwater species are significantly different from those of both euryhaline ($p = 0.001$) and marine ($p = 0.002$) species, whereas marine and euryhaline species are not significantly different from each other ($p = 0.079$).

Two explanatory variables, maximum standard length and relative clupeiform diversity, have the most influence on the species assemblage pattern. In general, freshwater species of concern tend to have smaller maximum standard lengths compared to euryhaline and marine species. *Clupeonella grimmi*, *Anchoa choerostoma* and *Opisthonema berlangai* are the only three marine species that cluster with the freshwater species, likely because they are some of the smallest marine species included in the study and are also those with the lowest number of other clupeiforms present in their ranges.

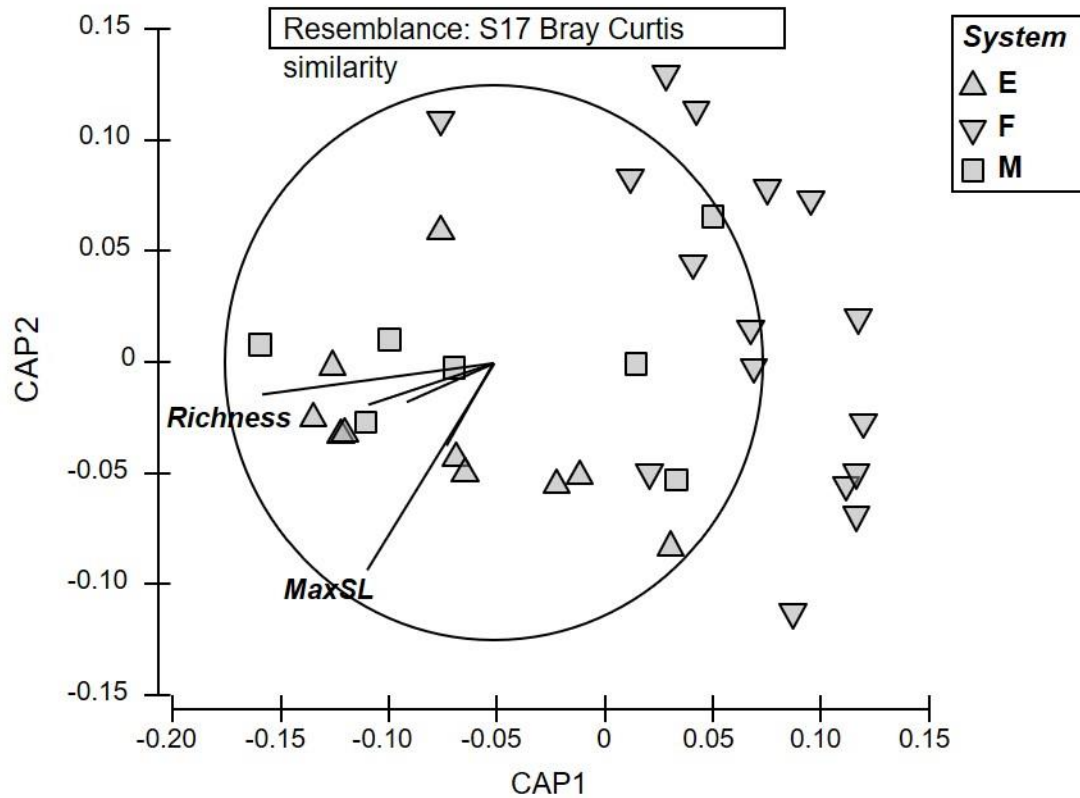


Fig. 8. Canonical Analysis of Principal Coordinates (CAP) ordination of threatened and Near Threatened species-specific life history and ecological traits in multivariate space by primary habitat system ($n = 33$). Habitat system abbreviations are E = Euryhaline, F = Freshwater and M = Marine. The two most prominent traits – maximum size in cm (MaxSL) and number of other clupeiforms within a species distribution (Richness) are labeled; additional traits represented by the unlabeled vectors include habitat requirements (e.g., generalist or specialist), the number of country waters a species is distributed within and total number of threats known to impact a species. Individual symbols represent a single species; symbol transparency was set a 50% to indicate where species are overlapping. The direction and length of the vectors represent the relationship between the ordination axes and the life history and ecological characters.

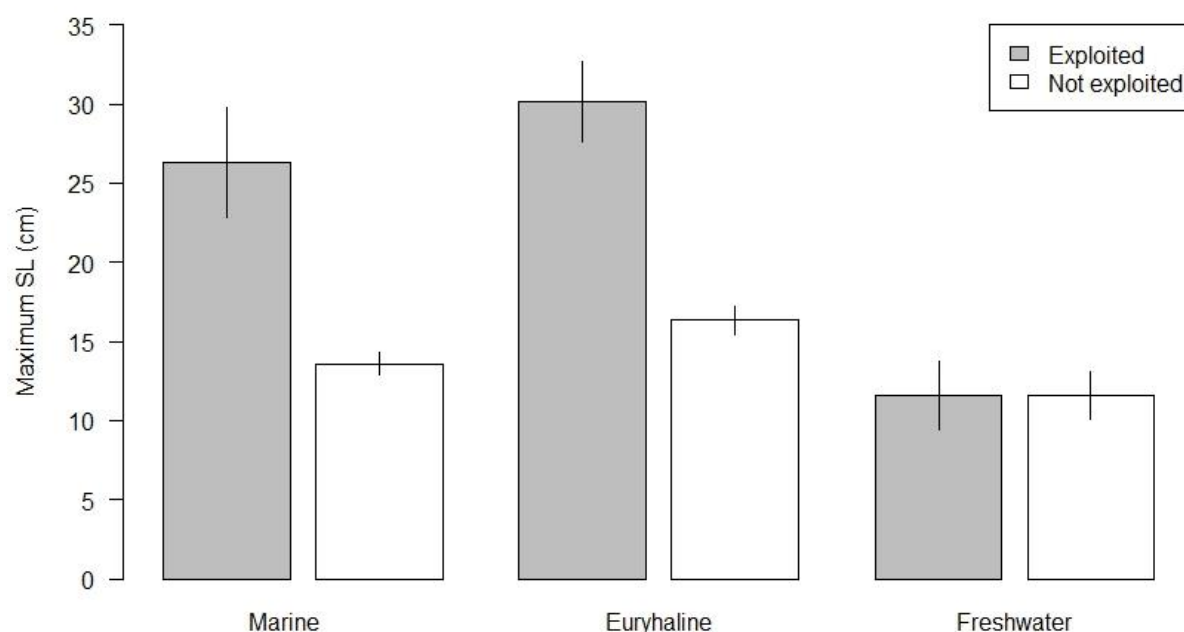


Fig. 9. Mean maximum standard length (cm) of clupeiforms ($n = 394$) as a function of primary habitat system (marine, euryhaline, freshwater) and exploitation status (exploited or not exploited). Grey bars indicate mean size of exploited species; white bars represent mean size of unexploited species. Standard error is indicated by the black vertical lines.

Table 1

Tukey HSD pair-wise comparisons of exploited vs. non-exploited species within each primary habitat system. Difference is the difference in means of standard length, Lower bound and Upper bound refer to the lower and upper confidence intervals, and p-adjusted indicates the adjusted p-values for the possible pairs. The marine and euryhaline systems show a significant difference between the size of exploited vs. non-exploited species; the difference in size between exploited and non-exploited freshwater species is not significant.

Habitat	Difference	Lower bound	Upper bound	P-adjusted
Marine	12.73	5.82	19.64	<<0.05
Euryhaline	13.81	8.04	19.58	<<0.05
Freshwater	-0.01	-9.22	9.20	1.00

DISCUSSION

Marine, freshwater and euryhaline clupeiforms are influenced by threats differently. Additionally, life history characteristics influencing susceptibility to threats also differ according to these major habitat types. In general, marine and freshwater species exhibited different responses to threats, and were influenced by different life history characteristics. Euryhaline species had similar responses to threats as freshwater species but were more similar to marine species in susceptible life history characteristics. Some shared threats such as exploitation affect species differently among these habitat systems.

Within each habitat system, size can influence how threats impact species. While exploitation affects species of all sizes within each habitat system, larger-bodied species are more impacted in marine and euryhaline systems (Fig. 8 and 9). For example, large-scale commercial fisheries often target the larger-bodied marine and euryhaline species (e.g., Atlantic herring, *Clupea harengus*) or multi-species groups and genera (e.g., *Stolephorus* spp.). Some of the largest-bodied exploited clupeiforms tend to be diadromous species in euryhaline waters where the passage through narrow estuaries makes them easily harvestable and increases their vulnerability as many are purposely targeted throughout various stages of ontogeny (McDowall, 1999). For example, the anadromous *Tenualosa macrura* (NT) and *T. toli* (VU) are a delicacy in Malaysia and Indonesia where small males are fished in marine waters and large ripe females are targeted for roe during spawning runs (Di Dario, 2018b; Mohd Arshaad et al., 2018) limiting their ability to repopulate. In contrast, freshwater species tend to be exploited based on geographic availability instead of size; the smallest known exploited clupeiform, *Nannothrissa stewarti*, is a freshwater species with a maximum known standard

length of 2.3 cm. Freshwater and inland fisheries typically support small-scale commercial, artisanal and subsistence fisheries of clupeiforms for food rather than reduction-type fisheries.

For clupeiforms, exploitation is the most important threat in all three habitats both in terms of numbers of species listing fisheries as the most prominent threat (Chapter 1) and in numbers of species corresponding to the exploitation axis in the CAP analysis (Fig. 7). However, 30 out of 38 (79%) marine species with a known threat are only or mostly impacted by exploitation (group 3 in Fig. 7) making this group proportionately the most heavily impacted by fisheries among the three habitats, compared to less than 40% of euryhaline species and less than 30 % of freshwater species in group 3. Fishery collapse of low trophic-level species has been linked to high fishing mortality and a long history of a developed fishery (Pinsky et al., 2011), as is the case for many clupeiforms. For example, the Pacific herring, *Clupea pallasii*, a temperate marine species widely distributed in the northern Pacific Ocean, has a complex population structure with multiple spawning stocks that have been fished for millennia and have supported industrial fisheries since the early 1900s (Hay et al., 2001). Despite long-term management and monitoring throughout most of its range, some spawning stocks are increasing in abundance (e.g., Quilicene Bay stock: WDFW, 2018), while a neighboring stock may be in a critically low state even after long periods of closed fisheries (e.g., Cherry Point stock: WDFW, 2018). In contrast, the world's only freshwater sardine (Bombon sardine, *Sardinella tawilis*) is a tropical species endemic to a single lake in the Philippines (Whitehead, 1985; Papa et al., 2008) and is one of the most commercially important fish in the country (Mutia, 2015) with limited monitoring and regulation until relatively recently (Villanueva et al., 1996; Willette et al., 2011; Mutia, 2015). Illegal and over-fishing practices have resulted in

declining catches of *S. tawilis* since the late 1990s (Marmaril, 2001; Mutia et al., 2004, 2015). An example of estuarine species that also appear to be heavily exploited with inadequate management are the anadromous species of *Tenuolosa* in Malaysia and Indonesia (Di Dario, 2018b; Mohd Arshaad et al., 2018). These three ecologically different species (*C. pallasii*, *S. tawilis* and *Tenuolosa spp.*) have responded similarly to exploitation with apparently unsustainable population declines in some cases, both with and without long-term complex fisheries management. It is suspected that the response of other, lesser-known clupeiforms to high fishing pressure across habitat systems is comparable.

Exploitation is a much more prevalent threat for marine clupeiforms than freshwater and estuarine clupeiforms even though exploitation is the most ubiquitous threat in all three major habitat types. In IUCN Red List assessments, exploitation threats are essentially nullified if the population is managed sustainably. Typical fisheries management practices such as those based on Maximum Sustainable Yield, fishing effort, or gear restrictions may not be appropriate for some exploited clupeiforms, particularly data-limited species that are targeted in un- or poorly regulated fisheries. Instead, simple strategies based on what is known about species such as primary habitat type and other easily recognizable traits such as body size may be a useful approach to manage clupeiforms. This may allow management measures to be tailored to species groups with limited data if the response to threats within the group is similar to what we observe for other data-rich clupeiforms within the same habitat.

Grouping species for management purposes using readily available traits as I show here in an exploratory CAP approach is consistent with existing fisheries management schemes. Recent proposed management methods are shifting from single-species to multi-species and

ecosystem-based management which would better support the trophic interactions of forage fishes and their predators (Coll et al., 2008; Pikitch et al., 2012, 2014; Essington et al., 2015; Siple et al., 2018). However, the complex data needed to implement these strategies is only available for data-rich forage fishes. Recent work by Siple et al., (2018) suggests that the best management strategies for forage fishes incorporate species-specific life history traits. Given that life history data are limited for the majority of clupeiforms, this exploratory CAP analysis shows that groups can be separated as a function of habitat system and available natural history traits. By binning the data-limited clupeiforms into simple, discrete groups based on what we currently know, our ability to efficiently manage and conserve these species may improve. Similar approaches may also be useful for other ‘small pelagic’ taxa characterized by high ecological diversity and data-limitations.

Aside from the threat of exploitation, pollution and natural system modifications that degrade habitats and their ecosystem services are most detrimental to freshwater, euryhaline, and some nearshore marine species. Neither of these threats show size-specificity; however, larger euryhaline species likely experience threats differently than smaller freshwater species. For example, dams indiscriminately impact freshwater and euryhaline species of all sizes by fragmenting suitable habitat and preventing migration away from a threat (van Puijenbroek et al., 2019). However, typically large diadromous species would be heavily impacted by dams during spawning runs (McDowall, 1999; Marmulla et al., 2001). For example, severe reductions in population abundance and local extirpations were observed in the American shad, *Alosa sapidissima* because of damming (Haro and Castro-Santos, 2012). Similar trends have been reported for the Pontic (*Alosa pontica*) and Allis (*Alosa alosa*) shads in Europe (van Puijenbroek

et al., 2019) as well as for the tropical Hilsa shad (*Tenualosa ilisha*) in the Indian Ocean (Hossain et al., 2019). Dam modifications (e.g., fish passages) to accommodate shad spawning migrations have existed for more than 250 years (Haro and Castro-Santos, 2012); however, these mitigations do not help spawning populations already driven away from natal streams (Sprankle, 2005; Monk et al., 1989).

Maximizing differences among groups using CAP (Anderson and Willis, 2003) helps examine conservation questions of clupeiforms; however, it is not without its limitations. This method has been used previously in conservation studies of a regional assemblage of all threatened and NT shallow water bony fishes (Linardich et al., 2019) and a variety of imperiled Canadian species (McCune et al., 2013). Here it was applied to a single Order of predominantly forage fishes with a more limited variability of natural history characteristics than what is observed in a diverse regional group of species. In addition, data is limited by information compiled in the IUCN Red List species accounts. Common life history traits used to assess population trends in the Red List assessments (e.g., age or size at maturity, fecundity, longevity, etc.) are unknown for many clupeiforms. Therefore, only widely available natural history traits were used, such as maximum length, number of threats, habitat system preference, habitat specialization, and proxies for distribution and relative clupeiform diversity.

Assigning habitat system categories was also challenging, particularly when distinguishing between true diadromous and salinity-tolerant species. Many species were lumped into a single euryhaline category because the existence of diadromy is possible, but unknown for many clupeiforms. Additional research on habitat requirements and basic biology would greatly benefit our ecological understanding of this group and improve future analyses of

threats and conservation needs. A high degree of local knowledge is suspected to be sequestered in unpublished and gray literature. Efforts to make local and indigenous knowledge accessible to the public would also work to expand our understanding of the conservation status of this group.

Threats to clupeiform fishes will continue to worsen without comprehensive mitigation and improved fisheries management should be highest priority given its prevalence while remaining cognizant of other threats. While more information is needed regarding the negative impacts of processes like climate change, this analysis suggests that short-term conservation efforts should also focus on minimizing localized threats in all habitats. Specifically, national and local measures should be taken to reduce the impact of habitat degradation on freshwater, euryhaline, and nearshore marine fishes. By mandating local pollution mitigation and dam removals, suitable habitat can be restored, which can substantially contribute to the local economy by increasing recreational use and ecotourism. Compounding strategies that limit pollution and remove multiple dams have shown to restore natural fish populations in the Cuyahoga River, Ohio after many years of severe degradation (State of Ohio Environmental Protection Agency, 2008) and may also prove successful for clupeiform fishes.

Given that our reliance on clupeiform fishery resources is expected to increase, future work should build upon the CAP results by refining the characters used as an approach to develop new or improve existing management of similar data-limited fisheries. A broad management scheme that provides at least somewhat effective regulation to many similar species is preferred over a complete lack of management or monitoring, as is currently the case

for many of these clupeiform fisheries. Following this approach, we may also inch closer to the goal of ecosystem-based fisheries management.

CHAPTER 4

DISCUSSION AND CONCLUSIONS

This thesis represents the first evaluation of the global conservation status of all members of the Order Clupeiformes and the first attempt to characterize known threats and life history traits by preferred habitat system. Despite many recent morphological, phylogenetic and group-specific works (e.g., Di Dario, 2004; Lavoué et al., 2013, 2014; Hata and Motomura 2017; Loeb et al., 2018; Bloom and Egan, 2018), the compiled species-specific IUCN Red List assessments represent the first review of all species since Whitehead (1985) and Whitehead et al., (1988) assembled taxonomic and biological information on all valid clupeoid species.

This also represents the first initiative to synthesize conservation information from IUCN Red List assessments for a single aquatic taxonomic group with representatives of all habitat types and a particularly high number of diadromous species. Many Red List assessments exist for diadromous species, including representatives of sturgeons, salmon, lampreys, anguillid eels and now, clupeiforms. However, except for the global conservation status of the mostly catadromous, anguillid eels ($n = 13$: Jacoby et al., 2015), diadromous species are often included within regional freshwater initiatives (e.g., Freyhof and Brooks, 2011; Kottelat et al., 2008), even if they are anadromous and spend most of their life cycle in marine waters. While this method may work to address major regional and system-wide conservation issues, it can

undermine the true conservation status of specific taxonomic groups by excluding part of their range from analyses. For example, because all anadromous lampreys were assessed with the European freshwater fishes (Freyhof and Brooks, 2011), the respective European marine fishes initiative did not include them (Nieto et al., 2015), despite that their marine ranges were excluded from freshwater analyses (Freyhof and Brooks, 2011—Appendix 4). The addition of clupeiforms to the IUCN Red List increases the representation of diadromous species within global analyses of overall species conservation.

Highlighting large-scale species patterns from a conservation perspective is a beneficial tool that can answer broad questions with more certainty. By looking at these patterns across an entire taxonomic group, underlying relationships have been uncovered (e.g., widespread major threats, geographic areas of most concern). Ultimately, this synthesized information may be used to influence management and implement better informed conservation measures with a higher probability of success. For example, we now know which threats are the most pervasive to all clupeiforms globally – exploitation, pollution and natural system modifications, and that the impacts of these threats are heavily influenced by primary habitat system and the size of the species. This highlights strategies to address specific threats within each habitat system.

The geographic areas of most concern for clupeiforms (e.g., the Caribbean and the Indo-Malay-Philippine Archipelago) were also identified in this study and support the findings of other Red List syntheses of taxonomic groups, including the groupers (Sadovy de Mitcheson et al., 2013), coastal sharks and rays (Dulvy et al., 2013), freshwater fishes (Collen et al., 2014),

and bonefishes (Adams et al., 2014). Across all assessed taxa, it is becoming increasingly apparent that these large regions with the highest biodiversity warrant management prioritization, especially given that these areas are also where cumulative human impacts are increasing (Halpern et al., 2015). Resources are needed both in research, because of the high number of species and subsequent large proportion of Data Deficient species, and for conservation planning, due to the high number of threatened and Near Threatened species. These emphasized patterns and subsequent increases in our knowledge base will allow us to direct attention to overexploited stocks, heavily degraded waterways and regions most in need of conservation.

Piecing together patterns at a global scale can be extremely useful in assessing broad consistencies but is not without its challenges. Limited by the underlying data, results are subject to shift in response to an increase in available information. Aside from missing data, a plethora of relevant and potentially useful information remains sequestered in unpublished or gray literature. An increase in open-access knowledge regarding geographic distribution, habitat utilization (specifically for spawning and migratory behavior), and total catches would elevate our understanding of clupeiform conservation status. Also, increased impact assessments of local threats on biodiversity would provide opportunity to better quantitatively assess threats to the clupeiforms present in those areas. Additional to insufficient data for many species, the methodology and level of detail, length and consistency of monitoring available are sources of variation both within and between countries, which can make an accurate synthesis of conservation status difficult.

As with many taxonomic groups, clupeiforms have long been plagued with unresolved taxonomy (Whitehead, 1985), hindering biological assessments. Despite recent attempts to untangle taxonomic relationships, some species-level distinctions are still questioned, such as those within the genus *Sardinops* (Whitehead, 1985; Parrish, 1989) and new species continue to be described such as within the genera *Sardinella* and *Stolephrous* (Hata and Motomura, 2018, 2019a, 2019b). While taxonomic changes advance our understanding of a species group, they can have implications within extinction risk assessments. For example, the two most important criteria of distribution and population size often change with information provided in taxonomic revisions and decreases in either of these criteria may result in an increase of risk of extinction.

In this study, the challenges stemming from data limitation and taxonomic uncertainty resulted in many clupeiform species assessed as Data Deficient. The resulting uncertainty in the overall threat status of clupeiforms presents faults in our understanding of conservation status for these species. Uncertainty may result in an underestimate of the true risk of extinction, leading to missed opportunities to apply appropriate mitigation (Davidson et al., 2012; Bland et al., 2014; Dulvy et al., 2014). Therefore, threatened and DD species impacted by multiple threats, particularly those that migrate between habitat systems, should take priority for future research and re-assessments similar to what was determined for the porgies (Comeros-Raynal et al., 2016).

This study represents the current picture of conservation status of clupeiforms based on the best available data. It is a starting point and will provide a more comprehensive

representation as more data are funneled into the species-specific re-assessments. By monitoring changes in conservation status of many taxa and continuing to add whole taxonomic groups onto the IUCN Red List, we can refine and enrich our understanding of biodiversity conservation and redress the declining state of biodiversity across the globe by providing better information for making more informed decisions. Outside of the scientific community, the IUCN Red List assessments and subsequent analyses may inform all stakeholders and end-users, including, fishers, processors, and consumers. Embracing biodiversity conservation will allow us to see maximum benefits for all parties, including the long-term sustainable use of our aquatic resources as well as helping to maintain or replenish the balance of the ecosystem.

Addition of this group to the Red List acts as further evidence in support of overarching conservation dilemmas and as a catalyst for change such as decreasing local human impact on aquatic ecosystems and resources. For example, a common finding among these species-specific initiatives is that the major threats most prevalent to clupeiforms are also those that negatively impact many other taxa. While the individual effects and the intensity of the threat may vary by species and locality, the overall outcomes tend to be similar, further supporting the need for local as well as multinational threat mitigation. It will not only benefit clupeiforms to revisit management protocols as well as water use and waste removal protocols, but it will also benefit many other important freshwater, estuarine and marine taxa and fishery resources.

REFERENCES

- Adams, A.J., Horodysky, A. Z., McBride, R.S., Guindon, K., Shenker, J., MacDonald, T.C., Harwell, H.D., Ward, R., and Carpenter, K., 2013. Global conservation status and research needs for tarpons (Megalopidae), ladyfishes (Elopidae) and bonefishes (Albulidae). *Fish Fish* 15, 280 – 311. doi: 10.1111/faf.12017.
- Agnew, D.J., Pearce, J., Pramod, G., Peatman, T., Watson, R., Beddington, J.R. and Pitcher, T.J., 2009. Estimating the worldwide extent of illegal fishing. *PLoS ONE* 4. doi: 10.1371/journal.pone.0004570.
- Akçakaya, H.R., Ferson, S., Murgman, M.A., Keith, D.A., Mace, G.M., and Todd, C.R., 2000. Making consistent IUCN classifications under uncertainty. *Conserv Biol* 14, 1001-1013. doi: 10.1046/j.1523-1739.2000.99125.x.
- Aladin, N., Barker, D.R., Beltram, G., Brouwer, J., Davidson, N., Duker, L., Junk, W., Kaplowitz, M.D., Ketelaars, H., Kreuzberg-Mukhina, E., Espino, G.L., Lévêque, C., Lopex, A., Milton, R.G., Mirabzadeh, P., Pritchard, D., Revenga, C., Rivera, M., Hussainy, A.S., Silvius, M. and Steinkamp, M., 2005. Inland Water Systems. Ch. 20. In: Hassan, R., Scholes, R. and Ash, N. (Eds.) *Ecosystems and Human Well-being: Current State and Trends*, Volume 1. Island Press, Washington, D.C.
- Alder, J., Campbell, B., Karpouzi, V., Kaschner, K., and Pauly, D., 2008. Forage Fish: From Ecosystems to Markets. *Annu Rev Env Resour* 33, 153-166. doi: 10.1146/annurev.enviro.33.020807.
- Alheit, J. Roy, C. and Kifani, S., 2009. Decadal-scale variability in populations. In: Checkley, D.M. Jr., Oozeki, Y., and Roy, C. *Climate change and small pelagic fish* (Eds.), Cambridge University Press, Cambridge, 285 – 299.
- Aliasghari, M., AnvariFar, H., and Mir, J.I., 2017. Effect of fishing and natural factors on the population of the fish *Clupeonella grimmi* (Clupeiformes: Clupeidae) in southern waters of the Caspian Sea. *Cuad UNED* 9, 185 - 191. ISSN: 1659-4266.
- Amante, C., Eakins, B.W., 2009. ETOPO1 1 Arc-Minute Global Relief Model: Procedures, Data Sources and Analysis NOAA Technical Memorandum NESDIS NGDC-24. doi:10.7289/V7285C8276M: National Geophysical Data Center, NOAA.
- Anderson, M.J. and Willis, T.J., 2003. Canonical analysis of principal coordinates: a useful method of constrained ordination for ecology. *Ecology* 84, 511 – 525.

Anderson, M.J., Gorley, R.N. and Clarke, K.R., 2008. PERMANOVA+ for PRIMER: Guide to software and statistical methods. PRIMER-E Limited: Plymouth, UK.

Baille, J. Hilton-Taylor, C. and Stuart, S.N., 2004. 2004 IUCN Red List of Threatened Species: A Global Species Assessment. IUCN, Gland, Switzerland and Cambridge, UK.

Baker, J.L., 2013. Status report of rare and endemic species and other marine fauna of conservation concern in the Northern Rivers CMA region, New South Wales. Northern Rivers Catchment Management Authority, New South Wales.

Bakun, A., & Cury, P., 1999. The 'school trap': A mechanism promoting large-amplitude out-of-phase population oscillations of small pelagic fish species. *Ecol* 2, 349-351. doi: 10.1046/j.1461-0248.1999.00099.x.

Bartley, D.M., De Graaf, G.J., Valbo-Jørgensen, J., and Marmulla, G., 2015. Inland capture fisheries: status and data issues. *Fisheries Manage Ecol* 22, 71–77. doi: 10.1111/fme.12104.

Bassett, E., 2014. Cultural importance of river herring to the Passamaquoddy people. Sipayik Environmental Department, Pleasant Point Reservation, Passamaquoddy Tribe.

Bielby, J., Cooper, N., Cunningham, A.A., Garner, T.W.J. and Purvis, A., 2008. Predicting susceptibility to future declines in the world's frogs. *Conserv Lett* 1, 82 – 90. doi: 10.1111/j.1755-263X.2008.00015.x.

Blaber, S.J.M., Milton, D.A., Pang, J., Wong, P., Boon-Teck, O., Nyigo, L., and Lubim, D., 1996. The life history of the tropical shad *Tenulosa toli* from Sarawak: first evidence of protandry in the Clupeiformes? *Environ Biol Fishes* 46, 225-242.

Bloch, M., 1809. Natural history of the herring. *The Belfast Monthly Magazine* 2, 241-245.

Bloom, D.D. and Egan, J.P., 2018. Systematics of Clupeiformes and testing for ecological limits on species richness in a trans-marine/freshwater clade. *Neotrop Ichthyol* 16, e180095. doi: 10.1590/1982-0224-20180095.

Bloom, D.D. and Lovejoy, N.R., 2012. Molecular phylogenetics reveals a pattern of biome conservatism in New World anchovies (family Engraulidae). *J Evol Biol* 25, 701 - 775. doi: 10.1111/j.1420-9101.2012.02464.x.

Bloom, D.D. and Lovejoy, N.R., 2014. The evolutionary origins of diadromy inferred from a time-calibrated phylogeny for Clupeiformes (herring and allies). *Proc R Soc B* 281, 20132081. doi: 10.1098/rspb.2013.2081.

Bloom, D.D. and Lovejoy, N.R., 2017. On the origins of marine-derived freshwater fishes in South America. *J Biogeogr* 44, 1927 – 1938. doi: 10.1111/jbi.12954.

Beverton, R.J.H., 1990. Small marine pelagic fish and the threat of fishing; are they endangered? *J Fish Biol* 37, 5 – 16.

Brooks, T.M., Mittermeier, R.A., da Fonseca, G.A.B., Gerlach, J., Hoffmann, M., Lamoreux, J.F., Mittermeier, C.G., Pilgrim, J.D., Rodrigues, A.S.L., 2006. Global biodiversity conservation priorities. *Science* 313, 58-61. doi: 10.1126/science.1127609.

Cardillo, M., Mace, G.M., Jones, K.E., Bielby, J., Bininda-Emonds, O.R.P., Sechrest, W., Orme, A.D.J. and Purvis, A., 2005. Multiple causes of high extinction risk in large mammal species. *Science* 309, 1239-1241. doi: 10.1126/science.116030.

Cardillo, M., Mace, G.M., Gittleman, J.L., Jones, K.E., Bielby, J. and Purvis, A., 2008. The predictability of extinction: biological and external correlates of decline in mammals. *Proc R Soc B* 275, 1441 – 1448. doi: 10.1098/rspb.2008.0179.

Carpenter, K.E., Abrar, M., Aeby, G., Aronson, R.B., Banks, S., Bruckner, A., . . . Wood, E., 2008. One-third of reef-building corals face elevated extinction risk from climate change and local impacts. *Science* 321, 560-563. doi: 10.1126/science.1159196.

Castillo, S. and Mendo, J., 1987. Estimation of unregistered Peruvian anchoveta (*Engraulis ringens*) in official catch statistics, 1951 to 1982. 109 – 116. In D. Pauly and I. Tsukayama (Eds.), *The Peruvian anchoveta and its upwelling ecosystem: three decades of changes*. ICLARM Stud Rev 15, 351. Instituto del Mar del Peru (IMARPE), Callao, Peru; Deutsche Gesellschaft für Technische Zusammenarbeit (GTZ), GmbH, Eschborn, Federal Republic of Germany; and International Center for Living Aquatic Resources Management (ICLARM), Manila, Philippines.

Carrizo, S.F., Smith, K.G., and Darwall, W.R.T., 2013. Progress towards a global assessment of the status of freshwater fishes (Pisces) for the IUCN Red List: application to conservation programmes in zoos and aquariums. *International Zoo Yearbook* 47, 46 – 64. doi: 10.1111/izy.12019.

Carruthers, T.R., Punt, A.E., Walters, C.J., MacCall, A., McAllister, M.K., Dick, E.J. and Cope, J., 2014. Evaluating method for setting catch limits in data-limited fisheries. *Fish Res* 153, 48 – 68. doi: 10.1015/j.fisres.2013.12.014.

Chapman, A.D., 2009. Numbers of living species in Australia and the world. 2nd Edition. Report for the Australian Biological Resources Study. Canberra, Australia.

Clarke, K.R. and Gorley, R.N., 2015. PRIMER v7: User Manual/Tutorial. PRIMER-E Limited: Plymouth, UK.

Coll, M., Libralato, S., Tudela, S., Palomera, I. and Pranovi, F., 2008. Ecosystem overfishing in the ocean. *PLoS ONE* 3, e3881. doi:10.1371/journal.pone.0003881.

Collen, B., Whitton, F., Dyer, E.E., Baillie, J.E.M., Cumberlidge, N., Darwall, W.R.T., Pollock, C., Richman, N.I., Soulsby, A.-M. and Böhm, M., 2014. Global patterns of freshwater species diversity, threat and endemism. *Global Ecol Biogeogr* 23, 40 – 51. doi: 10.1111/geb.12096.

Collette, B.B., Carpenter, K.E., Polidoro, B.A., Juan-Jordá, M.J., Boustany, A., Die, D.J., ... Yáñez, E., 2011. High value and long life-double jeopardy for tunas and billfishes. *Science* 333, 291-292.

Coull, J.R. 2003. Silver Darlings: The History of Herring Fishing on the East Coast of Scotland. Scotland. https://sites.scran.ac.uk/secf_final/silver/index.php. (accessed 16 January 2018).

Comeros-Raynal, M.T., Polidoro, B.A., Broatch, J., Mann, B.Q., Gorman, C., Buxton, C.D., Goodpaster, A.M., Iwatsuki, Y., MacDonald, T.C., Pollard, D., Russell, B. and Carpenter, K.E., 2016. Key predictors of extinction risk in sea breams and porgies (Family: Sparidae). *Biol Conserv* 202, 88 – 98. doi: 10.1016/j.biocon.2016.08.027.

Cury, P.M., Boyd, I.L., Bonhommeau, S., Anker-Nilssen, T., Crawford, R.J.M., Furness, R.W., Mills, J.A., Murphy, E.J., Osterblom, H., Paleczny, M., Piatt, J.F., Roux, J.-P., Shannon, L., and Sydeman, W.J., 2011. Global seabird response to forage fish depletion -- one-third for the birds. *Science* 334, 1703-1706. doi: 10.1126/science.1212928.

Darwall, W.R.T., Smith, K.G., Allen, D., Seddon, M.B., Reid, G.M., Clausnitzer, V. and Kalkman, V.J., 2009. Freshwater biodiversity: a hidden resource under threat. In: Vié, J.-C., Hilton-Taylor, C. and Stuart, S.N. (Eds.), 2009. *Wildlife in a Changing World – An Analysis of the 2008 IUCN Red List of Threatened Species*. IUCN, Gland, Switzerland.

Darwall, W.R.T., Holland, R.A., Smith, K.G., Allen, D., Brooks, E.G.E., Katarya, V., Pollock, C.M., Shi, Y., Clausnitzer, V., Cumberlidge, N., Cuttelod, A., Dijkstra, K.-D.B., Diop, N.D., García, N., Seddon, M.B., Skelton, P.H., Snoeks, J., Tweddle, D. and Vié, J.-C., 2011. Implications of bias in conservation research and investment for freshwater species. *Conserv Lett* 4, 474 – 482. doi: 10.1111/j.1755-263X.00202.x.

Davidson, A.D., Hamilton, M.J., Boyer, A.G., Brown, J.H. and Ceballos, G., 2009. Multiple ecological pathways to extinction in mammals. *PNAS* 106, 10702 – 10705. doi: 10.1073/pnas.0901956106.

Davidson, A.D., Boyer, A.G., Kim, H., Pompa-Mansilla, S., Hamilton, M.J., Costa, D.P., Ceballos, G. and Brown, J.H., 2012. Drivers and hotspots of extinction risk in marine mammals. *PNAS* 109, 3395 – 3400. doi: 10.1073/pnas.1121469109.

Denney, N.H., Jennings, S. and Reynolds, J.D., 2002. Life-history correlated of maximum population growth rates in marine fishes. *Proc R Soc Lond B* 269, 2229 – 2237. doi: 10.1098/rspb.2002.2138.

Di Dario, F., 2004. Homology between the recessus lateralis and cephalic sensory canals, with the proposition of additional synapomorphies for the Clupeiformes and the Clupeoidei. *Zool J Linn Soc* 141, 257 – 270.

Di Dario, F. and de Pinna, M.C.C., 2006. The supratemporal system and the pattern of ramification of cephalic sensory canals in *Denticeps clupeoides* (Denticipitoidei, Teleostei): additional evidence for monophyly of Clupeiformes and Clupeoidei. *Papéis Avulsos de Zoologica. Museu de Zoologia da Universidade de São Paulo* 46, 107 – 123. doi: 10.1590/S0031-10492006001000001.

Di Dario, F., 2018a. *Nematalosa japonica* (errata version published in 2019). The IUCN Red List of Threatened Species 2018: e.T75153777A143835471. doi: 10.2305/IUCN.UK.2018-2.RLTS.T75153777A143835471.en. (accessed 6 June 2019).

Di Dario, F., 2018b. *Tenuulosa toli* (errata version published in 2019). The IUCN Red List of Threatened Species 2018: e.T187944A143832449. doi: 10.2305/IUCN.UK.2018-2.RLTS.T187944A143832449.en. (accessed 6 June 2019).

Dulvy, N. K., Baum, J. K., Clarke, S., Compagno, L. J., Cortés, E., Domingo, A., ... & Martínez, J., 2008. You can swim but you can't hide: the global status and conservation of oceanic pelagic sharks and rays. *Aquat Conserv* 18, 459-482. doi: 10.1002/aqc.975.

Dulvy, N.K., Pinnegar, J.K. and Reynolds, J.D., 2009. Holocene extinctions in the sea. Chapter 6. In: Turvey, S.T. (Ed.) *Holocene Extinctions*. Oxford Biology. Oxford University Press, Oxford, 129 – 150.

Dulvy, N.K., Fowler, S.L., Musick, J.A., Cavanagh, R.D., Kyne, P.M., Harrison, L.R., . . . White, W. T., 2014. Extinction risk and conservation of the world's sharks and rays. *eLife* 3, 1-34.

Dunn, E. H., 2002. Using decline in bird populations to identify needs for conservation action. *Conserv Biol* 16, 1632 – 1637.

Eschmeyer, W.N., Fricke, R., and van der Laan, R. (Eds.), 2017. *Catalog of Fishes: genera, species, references*.

<http://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp>.
(accessed 14 December 2017).

Eschmeyer, W.N., Fricke, R. and Van der Laan, R. (Eds.), 2018. Catalog of Fishes: genera, species, references. Updated 31 May 2018. <http://researcharchive.calacademy.org/research/ichthyology/catalog/fishcatmain.asp>.
(accessed 19 January 2019).

Essington, T.E., Moriarty, P.E., Froehlich, H.E., Hodgson, E.E., Koehn, L.E., Oken, K.L., Siple, M.C. and Stawitz, C.C., 2015. Fishing amplifies forage fish population collapses. PNAS 112, 6648 – 6652. doi: 10.1073/pnas.1422020112.

FAO, 2016. Fishery and Aquaculture Statistics. Global capture production 1950-2014 (FishStatJ). FAO Fisheries and Aquaculture Department [online or CD-ROM], Rome. Updated 2016. <http://www.fao.org/fishery/>. (Accessed 6 March 2018).

FAO, 2018. The State of World Fisheries and Aquaculture 2018 – Meeting the sustainable development goals. Rome. License: CC BY-NC-SA 3.0 IGO.

Finney, B.P., Gregory-Eaves, I., Douglas, M.S.V., and Smol, J.P., 2002. Fisheries productivity in the northeastern Pacific Ocean over the past 2,200 years. Nature 416, 729 – 733.

FishNet2 Portal, 2017. FishNet2. www.fishnet2.net. (accessed 17 May 2017).

Fowler, H.W., 1973. A catalog of world fishes (XVIII). Taiwan Sheng Bo Wu Guan Ji Khan. 26.

Freeman, M. R., 2008. Challenges of assessing cetacean population recovery and conservation status. Endanger Species Res 6: 173–184. doi: 10.33354/esr001052.

Freyhof, J. & Kottelat, M., 2008a. *Alosa killarnensis*. The IUCN Red List of Threatened Species 2008. e.T135582A4152432. doi: 10.2305/IUCN.UK.2008.RLTS.T135582A4152432.en. (accessed 6 June 2019).

Freyhof, J. & Kottelat, M., 2008b. *Alosa immaculata*. The IUCN Red List of Threatened Species 2008. e.T907A13093654. doi: 10.2305/IUCN.UK.2008.RLTS.T907A13093654.en. (accessed 6 June 2019).

Freyhof, J. and Brooks, E., 2011. European Red List of Freshwater Fishes. Publications Office of the European Union, Luxembourg.

Gaichas, S., Gamble, R., Fogarty, M., Benoit, H., Essington, T., Fu, C., Koen-Alonso, M. and Link, J., 2012. Assembly rules for aggregate-species production models: simulations in support of management strategy evaluation. *Mar Ecol* 459, 275 – 292. doi: 10.3354/meps09650.

GBIF, 2017. Global Biodiversity Information Facility. <http://gbif.org>. (accessed 15 September 2017).

Grant, W.S. and Leslie, R.W, 1996. Late Pleistocene dispersal of Indian - Pacific sardine populations in an ancient lineage of the genus *Sardinops*. *Mar Biol* 126, 133 - 142.

Grande, L., 1985. Recent and fossil clupeomorph fishes with materials for revision of the subgroup of clupeoids. *Bull Am Mus Nat Hist* 181, 235 – 365.

Greenwood PH, Rosen DE, Weitzman SH, Myers GS., 1966. Phyletic studies of teleostean fishes, with a provisional classification of living forms. *Bull Am Mus Nat Hist* 131, 339–456.

Hallmann, C.A., Sorg, M., Jongejans, E., Siepel, H., Hofland, N., Schwan, H., Stenmans, E., Müller, A., Sumser, H., Hörrén, T., Goulson, D., and de Kroon, H., 2017. More than 75 percent decline over 27 years in total flying insect biomass in protected areas. *PLoS ONE* 12, e0185809. doi: 10.1371/journal.pone.0185809.

Halpern, B.S., Frazier, M., Potapenko, J., Casey, K.S., Koenig, K., Longo, C., Lowndes, J.S., Rockwood, R.C., Selig, E.R., Selkoe, K.A. and Walbridge, S., 2015. Spatial and temporal changes in cumulative human impacts on the world's ocean. *Nat Commun* 6, 7615. doi: 10.1038/ncomms8615.

Hata, H. and Motomura, H., 2017. Validity of *Encrasicholina pseudoheteroloba* (Hardenburg 1933) and redescription of *Encrasicholina heteroloba* (Ruppell 1837), a senior synonym of *Encrasicholina devisi* (Whitley 1940) (Clupeiformes: Engraulidae). *Ichthyol Res* 64, 18 – 28. doi: 10.1007/s10228-016-0529-4.

Hata, H. and Motomura, H., 2018. *Stolephorus continentalis*, a new anchovy from the northwestern South China Sea, and redescription of *Stolephorus chinensis* (Gunther 1880) (Clupeiformes: Engraulidae). *Ichthyol Res* 65, 374 – 382. doi: 10.1007/s10228-018-0621-z.

Hata, H. and Motomura, H., 2019a. *Stolephorus insignis*, a new anchovy from the western Pacific, and redescription of *Stolephorus apiensis* (Jordan and Seale 1906) (Clupeiformes: Engraulidae). *Ichthyol Res* 66: 280 – 288. doi: 10.1007/s10228-018-00675-5.

Hata, H. and Motomura, H., 2019b. A new species of sardine, *Sardinella pacifica* from the Philippines (Teleostei, Clupeiformes, Clupeidae). ZooKeys 829: 75 – 83. doi: 10.3897/zookeys.829.30688.

Hay, D.E., Torenson, R., Stephenson, R., Thompson, M., Claytor, R., Funk, F., Ivshina, E., Jakobsson, J., Kobayashi, T., McQuinn, I., Melvin, G., Molloy, J., Naumenko, N., Oda, K.T., Parmanne, R., Power, M., Radchenko, V., Schweigert, J., Simmonds, J., Sjöstrand, B., Stevenson, D.K., Tanasichuk, R., Tang, Q., Watters, D.L. and Wheeler, J., 2001. Taking Stock: An Inventory and Review of World Herring Stocks in 2001. Herring: Expectations for a New Millennium. Alaska Sea Grant College Program. AK-SG-01-04.

Harnik, P.G., Lotze, H.K., Anderson, S.C., Finkel, Z.V., Finnegan, S., Lindberg, D.R., Liow, L.H., Lockwood, R., McClain, C.R., McGuire, J.L., O’Dea, A., Pandolfi, J.M., Simpson, C., Tittensor, D. P., 2012. Extinctions in ancient and modern seas. Trends Ecol Evol 27, 608-617. doi: 10.1016/j.tree.2012.07.010

Haro, A. and Castro-Santos, T., 2012. Passage of American Shad: Paradigms and Realities. Mar Coast Fish 4, 252 – 261. doi: 10.1080/19425120.2012.675975.

Hilborn, R., Amoroso, R.O., Bogazzi, E., Jensen, O.P., Parma, A.M., Szuwalski, C., and Walters, C.J., 2017. When does fishing forage species affect their predators? Fish Res 191, 211-221. doi: 10.1016/j.fishres.2017.01.008.

Hilton-Taylor, C., Pollock, C.M., Chanson, J.S., Butchart, S.H.M., Oldfield, T.E.E. and Katariya, V., 2009. Status of the world’s species. In: Vié, J.-C., Hilton-Taylor, C. and Stuart, S.N. (Eds.). Wildlife in a Changing World – An Analysis of the 2008 IUCN Red List of Threatened Species. IUCN, Gland, Switzerland, 15 – 42.

Hoffmann, M., Brooks, T.M., da Fonseca, G.A., Gascon, C., Hawkins, A.F.A., James, R.E., Langhammer, P., Mittermeier, R.A., Pilgrim, J.D., Rodrigues, A.S.L., Silva, J.M.C., 2008. Conservation planning and the IUCN Red List. Endanger Species Res 6, 113-125.

Hoffmann, M., Hilton-Taylor, C., Angulo, A., Böhm, M., Brooks, T.M., Butchart, S.H., ... & Darwall, W.R., 2010. The impact of conservation on the status of the world’s vertebrates. Science 330, 1503-1509.

Hossain, M.S., Sharifuzzaman, S.M., Rouf, M.A., Pomeroy, R.S., Hossain, M.D., Chowdhury, S.R. and AftabUddin, S., 2018. Tropical hilsa shad (*Tenualosa ilisha*): Biology, fishery and management. Fish Fish 20, 44 – 65. doi: 10.1111/faf.12323.

Hutchins, J.A., 2000. Collapse and recovery of marine fishes. Nature 406, 882 – 885.

IOC, IHO and BODC, 2003. Centenary Edition of the GEBCO Digital Atlas, published on CD-ROM on behalf of the Intergovernmental Oceanographic Commission and the International Hydrographic Organization as part of the General Bathymetric Chart of the Oceans. British Oceanographic Data Centre, Liverpool.

IUCN., 2012. IUCN Red List Categories and Criteria: Version 3.1. Second edition. IUCN, Gland, Switzerland and Cambridge, UK, pp. 32.

IUCN., 2013. Documentation standards and consistency checks for IUCN Red List assessments and species accounts (Version 2).

IUCN, 2016. Rules of Procedure for IUCN Red List Assessments 2017 – 2020. Version 3.0. Annex 1: Required and Recommended Supporting Information for IUCN Red List Assessments. Approved by the IUCN SSC Steering Committee in September 2016.

IUCN, 2019. Summary Statistics. Table 1a: Number of species evaluated in relation to the overall number of described species, and numbers of threatened species by major groups. <https://www.iucnredlist.org/resources/summary-statistics#Summary%20Tables>. (accessed 4 September 2019).

Iwamoto, T., Eschmeyer, W., Alvarado, J., Bussing, W., 2010. *Anchoa helleri*. The IUCN Red List of Threatened Species 2010: e.T183382A8103626. doi: 10.2305/IUCN.UK.2010-3.RLTS.T183382A8103626.en. (accessed 6 June 2019).

Jacoby, D.M.P., Casselman, J.M., Crook, V., Delucia, M.-B., Ahn, H., Kaifu, K., Kurwie, T., Sasai, P., Silfvergrip, M.C., Smith, K.G., Uchinda, K., Walker, A.M. and Gollock, M.J., 2015. Synergistic patterns of threat and the challenges facing global anguillid eel conservation. *Glob Ecol Conserv* 4, 321 – 333. doi: 10.1016/j.gecco.2015.07.009.

Jennings, S., Reynolds, J.D., Mills, S.C., 1998. Life history correlates of responses to fisheries exploitation. *Proc R Soc B* 265, 333–339.

Juan-Jorda, M.J., Mosquiera, I., Freire, J. and Dulvy, N.K., 2012. Life history correlates of marine fisheries vulnerability: a review and a test with tunas and mackerel species. In: Briand, F. (Ed.), *Marine extinctions – patterns and processes*. CIESM Workshop Monograph n°45, 113 – 128. CIESM Publisher, Monaco.

Juan-Jorda, M.J., Mosquiera, I., Freire, J. and Dulvy, N.K., 2015. Population declines of tuna and relatives depend on their speed of life. *Proc R Soc B* 282, 20150322. doi: 10.1098/rspb.2015.0322.

- Kawarazuka, N. and Béné, C., 2011. The potential role of small fish species in improving micronutrient deficiencies in developing countries: building evidence. *Public Health Nutr* 14, 1927 – 1938. doi: 10.1017/S1368980011000814.
- Kent, G., 1986. The industrialization of fisheries. *Peasant Stud* 13, 133 – 143.
- Kindsvater, H.K., Mangel, M., Reynolds, J.D., Dulvy, N.K., 2016. Ten principles from evolutionary ecology essential for effective marine conservation. *Ecol Evol* 6, 2125–2138. doi: 10.1002/ece3.2012.
- Kottelat, M., Baird, I.G., Kullander, S.O., Ng, H.H., Parenti, L.R., Rainboth, W.J., Vidthayanon, C., 2008. The status and distribution of freshwater fishes of Indo-Burma. In: Allen, D.J., Smith, K.G., Darwall, W.R.T., 2008. The status and distribution of freshwater biodiversity in Indo-Burma. IUCN, Cambridge, UK and Gland, Switzerland, 39 - 64.
- Lavoué, S., Miya, M., Musikasinthorn, P., Chen, W.-J., and Nishida, M., 2013. Mitogenomic Evidence for an Indo-West Pacific Origin of the Clupeoidei (Teleostei: Clupeiformes). *PLoS ONE* 8, e56485. doi: 10.1371/journal.pone.0056485.
- Lavoué, S., Konstantinidis, P. and Chen, W.-J., 2014. Progress in clupeiform systematics. Chapter 1. In: Ganas, K. Biology and ecology of sardines and anchovies, 3 – 42. CRC Press. Taylor & Francis Group. Boca Raton, Florida.
- Lehner, B. and Grill, G., 2013. Global river hydrography and network routing: baseline data and new approaches to study the world's large river systems. *Hydrological Processes* 27, 2171 – 2186. doi: 10.1002/hyp.9740. Data is available at www.hydrosheds.org. (accessed 5 June 2018).
- Lévêque, C., Oberdorff, T., Paugy, D., Stiassny, M.L.J., and Tedesco, P.A., 2008. Global diversity of fish (Pisces) in freshwater. *Hydrobiologia* 595, 545 – 567. doi: 10.1007/s10750-007-9034-0.
- Levin, P.S., Francis, T.B. and Taylor, N.G., 2016. Thirty-two essential questions for understanding the social-ecological system of forage fish: the case of Pacific Herring. *EHS* 2, e01213. doi:10.1002/ehs2.1213.
- Li, C. and Ortí, G., 2007. Molecular phylogeny of Clupeiformes (Actinopterygii) inferred from nuclear and mitochondrial DNA sequences. *Mol Phylogenet Evol* 44, 386 – 398. doi: 10.1016/j.ympev.2006.10.030.
- Linardich, C., Ralph, G.M., Robertson, D.R., Harwell, H., Polidoro, B.A., Lindeman, K.C. and Carpenter, K.E., 2019. Extinction risk and conservation of marine bony shorefishes of the Greater Caribbean and Gulf of Mexico. *Aquat Conserv* 29, 85 – 101. 10.1002/aqc.2959.
- Link, J., 2002. What does ecosystem-based fisheries management mean? *Fisheries* 27, 18 – 21.

Loeb, M.V., Varella, H.R. and Menezes, N.A., 2018. A new species of *Anchoviella* (Clupeiformes: Engraulidae) from the western Amazon River in Peru, with comments on congeners in the Peruvian Amazon River. *J Fish Biol* 92, 1720 – 1730. doi: 10.1111/jfb.13601.

Lovejoy, N.R., Albert, J.S. and Crampton, W.J.S., 2006. Miocene marine incursions and marine/freshwater transitions: Evidence from Neotropical fishes. *J S Am Earth Sci* 21, 5-13.

Lynch, A.J., Cooke, S.J., Deines A.M., Bower, S.D., Bunnell, D.B., Cowx, I.G., Nguyen, V.M., Nohner, J., Phouthavong, K., Riley, B., Rogers, M.W., Taylor, W.W., Woelmer, W., Youn, S.-J. and Beard, Jr., T.D., 2016. The social, economic, and environmental importance of inland fish and fisheries. *ER* 24, 115 – 121. doi: 10.1139/er-2015-0064.

Mace, G.M., Maundire, H., Baillie, J.E.M., Ricketts, T.H., Brooks, T.M., Hoffmann, M., Stuart, S.N., et al., 2005. Ecosystems and human well-being: current state and trends. Millennium Ecosystem Assessment. Washington, Island Press.

Mace, G.M., Collar, N.J., Gaston, K.J., Hilton-Taylor, C., Akcakaya, H.R., Leader-Williams, N., Milner-Gulland, E.J., and Stuart, S.N., 2008. Quantification of Extinction Risk: IUCN's system for classifying threatened species. *Conserv Biol* 22, 1424-1442. doi: 10.1111/j.1523-1739.2008.01044.x

Malabara, M.C. and Di Dario, F., 2016. A new predatory herring-like fish (Teleostei: Clupeiformes) from the early Cretaceous of Brazil, and implications for relationships in the Clupeoidei. *Zool J Linn Soc* 180, 175 – 194.

Malak, D., Livingstone, S.R., Pollard, D., Polidoro, B.A., Cuttelod, A., Bariche, M., Bilecenoglu, M., Carpenter, K.E., Collette, B.B., Francour, P., Goren, M., Kara, M.H., Massuti, E., Papaconstantinou, C., Tunesi, L., 2011. Overview of the Conservation Status of the Marine Fishes of the Mediterranean Sea. IUCN, Gland, Switzerland and Malaga, Spain.

Mamaril, A. C., 2001. Translocation of the clupeid *Sardinella tawilis* to another lake in the Philippines: A proposal and ecological considerations. In: Santigao, C. B., Cuvin-Aralar, M. L., and Basiao, Z. U. (Eds.), *Conservation and ecological management of Philippine lakes in relation to fisheries and aquaculture*, 133 – 147. Southeast Asian Fisheries Development Center, Aquaculture Department, Iloilo, Philippines; Los Baños, Laguna, Philippines: Philippine Council for Aquatic and Marine Research and Development (PCAMRD), Department of Science and Technology; Quezon City, Philippines: Bureau of Fisheries and Aquatic Resources (BFAR), Department of Agriculture, Quezon City, Philippines.

Marmulla, G. (ed.), 2001. Dams, fish and fisheries: Opportunities, challenges and conflict resolution. FAO Fisheries Technical Paper. No. 419. Rome, FAO.

May, R. M., 1992. How many species inhabit the earth? *Sci Am* 267, 42 – 49.

McCune, J.L., Harrower, W. L., Avery-Gomm, S., Brogan, J.M., Csergo, A.-M., Davidson, L.N.K., Garani, A., Halpin, L.R., Lipsen, L.P.J., Lee, C., Nelson, J.C., Prugh, L.R., Stinson, C.M., Whitney, C.K. Whitton, J., 2013. Threats to Canadian species at risk: An analysis of finalized recovery strategies. *Biol Conserv* 166, 254 – 266. doi: 10.1016/j.biocon.2013.07.006.

McDowall, R.M., 1999. Different kinds of diadromy: Different kinds of conservation problems. *ICES J Mar Sci* 56, 410 – 413.

McKechnie, I., Lepofsky, D., Moss, M.K., Butler, V.L., Orchard, T.J., Coupland, G., Foster, F., Caldwell, M. and Lertzman, K., 2013. Archaeological data provide alternative hypotheses on Pacific herring (*Clupea pallasii*) distribution, abundance, and variability. *PNAS* 111, E807 – E816. doi: 10.1073/pnas.1316072111.

Meekan, M.G., Vigliola, L., Hansen, A., Doherty, P.J., Halford, A. and Carleton, J.H., 2006. Bigger is better: size-selective mortality throughout the life history of a fast-growing clupeid, *Spratelloides gracilis*. *Mar Ecol* 317, 237-244.

Milton, D.A., Blaber, S.J.M. and Rawlinson N.J.F., 1991. Age and growth of three species of tuna baitfish (genus: *Spratelloides*) in tropical Indo-Pacific. *J Fish Biol* 39, 849-866.

Mohan Dey, M., Rab, M.A., Paraguas, F.J., Piumsombun, S., Bhatta, R., Alam, M.F. and Ahmed, M., 2005. Fish consumption and food security: A disaggregated analysis by types of fish and classes of consumers in selected Asian countries. *Aquacult Econ Manage* 9, 89 – 111. doi: 10.1080/13657300590961537.

Mohanty, B.P., Ganguly, S., Mahanty, A., Mitra, T., Patra, S., Karunakaran, D., Mathew, S., Chakraborty, K., Paul, B.N., Sarma, D., Dayal, S., Singh, S., and Ayyappan, S., 2019. Fish in human health and nutrition. *Adv Fish Res* 7, 189 – 218.

Mohd Arshaad, W., Gaughan, D. & Munroe, T.A., 2018. *Tenuulosa macrura* (errata version published in 2019). The IUCN Red List of Threatened Species 2018: e.T98842673A143840186. doi: 10.2305/IUCN.UK.2018-2.RLTS.T98842673A143840186.en. (accessed 6 June 2019).

Monk, B. Weaver, D., Thompson, C., and Ossiander, F., 1989. Effects of flow and weir design on the passage behavior of American shad and salmonids in an experimental fish ladder. *N Am J Fish Manag* 9, 60-67.

Mora, C., Tittensor, D.P., Adl, S., Simpson, A.G.B. and Worm, B., 2011. How many species are there on Earth and in the ocean? *PLoS Biol* 9, e1001127. doi: 10.1371/journal.pbio.1001127.

Munroe, T.A., Wongratana, T., and Nizinski, M.S., 1999a. Pristigasteridae: Ilishas, pellonas. In: Carpenter, K.E. and Niem, V.H. (Eds.), FAO species identification guide for fishery purposes. The living marine resources of the Western Central Pacific. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (Elopidae to Linophrynidae), 1754 – 1770. FAO, Rome.

Munroe, T.A., Nizinski, M.S. and Wongratana, T., 1999b. Chirocentridae: Wolf-herrings. In: Carpenter, K.E. and Niem, V.H. (Eds.), FAO species identification guide for fishery purposes. The living marine resources of the Western Central Pacific. Vol. 3. Batoid fishes, chimaeras and bony fishes part 1 (Elopidae to Linophrynidae), 1754 – 1770. FAO, Rome.

Munroe, T.A., Aiken, K.A., Brown, J., Grijalba Bendeck, L. & Vega-Cendejas, M., 2019. *Harengula jaguana*. The IUCN Red List of Threatened Species 2019: e.T190478A86377366. doi: 10.2305/IUCN.UK.2019-2.RLTS.T190478A86377366.en. (accessed 6 June 2019).

Murray, D.M., 2015. Herring Tales: How the Silver Darlings Shaped Human Taste and History. Bloomsbury. London, Oxford, New York, New Delhi, Sydney.

Mutia, T.M., Magistrado, M.L. and Muyot, M.C., 2004. Status of *Sardinella tawilis* in Taal Lake, Philippines. 8th ZONAL Center II Research and Development Review. De La Salle University, Taft, Manila.

Mutia, M.A.T.M., 2015. Environmental influences on the reproductive biology and population dynamics of *Sardinella tawilis* in lake Taal, Philippines. Department of Environmental Sciences, University of the Philippines Los Banos.

Myers, R.A. and Worm, B., 2003. Rapid worldwide depletion of predatory fish communities. Lett Nat 423, 280 – 283.

Myers, R.A. and Worm, B., 2005. Extinction, survival or recovery of large predatory fishes. Philos Trans R Soc B 360, 13 – 20. doi: 10.1098/rstb.2004.1573.

NatureServe, 2013. *Alosa aestivalis*. The IUCN Red List of Threatened Species 2013: e.T201946A2730890. doi: 10.2305/IUCN.UK.2013-1.RLTS.T201946A2730890.en. (accessed 6 June 2019).

Nieto, A., Ralph, G.M., Comeros-Raynal, M.T., Kemp, J., García Criado, M., Allen, D.J., Dulvy, N. K., . . . Scott, J., Serena, F., Smith-Vaniz, W.F., Soldo, A., Stump, E. and Williams, J.T., 2015. European Red List of marine fishes. Publications Office of the European Union Luxembourg, IUCN.

NOAA Fisheries, 2019. National Oceanic and Atmospheric Administration: Commercial landings. <https://foss.nmfs.noaa.gov/apexfoss/f?p=215:200:11056708372560:::> (accessed 12 December 2018).

OZCAM database, 2007. Atlas of Living Australia Tools. Available at: <http://www.ozcam.org>. (accessed 15 July 2017).

Palumbi, S.R., Sandifer, P.A., Allan, J.D., Beck, M.W., Fautin, D.G., Fogarty, M.J., Halber, S.B., Incze, L.S., Leong, J.-A., Norse, E., Stachowicz, J.J. and Wall, D.H., 2009. Managing for ocean biodiversity to sustain marine ecosystem services. *Front Ecol Environ* 7, 204 – 211. doi: 10.1890/070135.

Papa, R.D.S., Pagulayan, R.C. and Pagulayan, A.E.J., 2008. Zooplanktivory in the endemic freshwater sardine, *Sardinella tawilis* (Herre 1927) of Taal Lake, the Philippines. *Zool Stud* 47, 535 – 543.

Parrish, J.D., Serra, R. and Grant, W.S., 1989. The monotypic sardines, *Sardina* and *Sardinops*: their taxonomy, distribution, stock structure, and zoogeography. *Can J Fish Aquat Sci* 46, 2019-2036.

Patterson, K., 1992. Fisheries for small pelagic species: an empirical approach to management targets. *Rev Fish Biol Fisher* 2, 321 – 338.

Pauly, D., Watson, R. and Alder, J., 2005. Global trends in world fisheries: impacts on marine ecosystems and food security. *Philos Trans R Soc B* 360, 5 – 12. doi: 10.1098/rstb.2004.1574.

Pauly, D. and Zeller, D., 2016a. Sea Around Us Concepts, Design and Data. Available at: seaaroundus.org. (accessed 12 December 2018).

Pauly, D. and Zeller, D., 2016b. Catch reconstructions reveal that global marine fisheries catches are higher than reported and declining. *Nat Commun* 7, 1 – 9. doi: 10.1038/ncomms10244.

Pikitch, E.K., Santora, C., Babcock, E.A., Bakun, A., Bonfil, R., Conover, D.O., Dayton, O., Doukakis, P., Fluharty, D., Heneman, B., Houde, E.D., Link, J., Livingston, P.A., Mangel, M., McAllister, M.K., Pope, J. and Sainsbury, K.J., 2004. Ecosystem-based fishery management. *Science* 305, 346 – 347. doi: 10.1126/science.1098222.

Pikitch, E., Boersma, P.D., Boyd, I.L., Conover, D.O., Cury, P., Essington, T., Heppell, S.S., Houde, E.D., Mangel, M., Pauly, D., Plagányi, É., Sainsbury, K., and Steneck, R.S., 2012. Little Fish, Big Impact: Managing a Crucial link in Ocean Food Webs. Lenfest Ocean Program, Washington, DC.

Pikitch, E.K., Rountos, K.J., Essington, T.E., Santora, C., Pauly, D., Watson, R., Sumalia, U.R., Boersma, P.D., Boyd, I.L., Conover, D.O., Cury, P., Heppell, S.S., Houde, E.D., Mangel, M., Plagányi, E., Sainsbury, K., Steneck, R.S., Geers, T.M., Gownaris, N., and Munch, S.B., 2014. The

global contribution of forage fish to marine fisheries and ecosystems. *Fish Fish* 15: 43-64. doi: 10.1111/faf.12004.

Pimm, S.L., Jenkins, C.N., Abell, R., Brooks, T.M., Gittleman, J.L., Joppa, L.N., Raven, P.H., Roberts, C.M., Sexton, J.O., 2014. The biodiversity of species and their rates of extinction, distribution, and protection. *Science* 344. doi: 10.1126/science.1246752

Pinsky, M.L., Jensen, O.P., Ricard, D. and Palumbi, S.R., 2011. Unexpected patterns of fisheries collapse in the world's oceans. *PNAS* 108, 8317 – 8322. doi: 10.1073/pnas.1015313108.

Polidoro, B.A., Linvingstone, S.R., Carpenter, K.E., Hutchinson, B., Mast, R.B., Pilcher, N.J., Sadovy de Mitcheson, Y. and Valenti, S.V., 2009. Status of the world's marine species. In: Vié, J.-C., Hilton-Taylor, C. and Stuart, S.N. (Eds.). *Wildlife in a Changing World – An Analysis of the 2008 IUCN Red List of Threatened Species*. IUCN, Gland, Switzerland.

Polidoro, B.A., Carpenter, K.E., Collins, L., Duke, N.C., Ellison, A.M., Ellison, J.C., Farnsworth, E.J., Fernando, E.S., Kathiresan, K., Koedam, N.E., Livingstone, S.R., Miyago, T., Moore, G.E., Ngoc Nam, V., Eong Ong, J., Primavera, J.H., Salmo, S.G., Sanciangco, J.C., Sukardjo, S., Wang, Y. and Hong Yong, J.W., 2010. The loss of species: mangrove extinction risk and geographic areas of global concern. *PLoS One* 5, e10095. doi: 10.131/journal.pone.0010095.

R Core Team, 2017. R version 1.0.153. A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

Reynolds, J.D., Dulvy, N.K., Goodwin, N.B., Hutchings, J.A., 2005. Biology of extinction risk in marine fishes. *Proc R Soc B* 272, 2337-2344.

Ripple, W.J., Abernethy, K., Betts, M.G., Chapron, G., Dirzo, R., Galetti, M., Levi, T., Lindsey, P.A., Macdonald, D.W., Machovina, B., Newsome, T.M., Peres, C.A., Wallach, A.D., Wolf, C. and Young, H., 2016. Bushmeat hunting and extinction risk to the world's mammals. *R Soc Open Sci* 3, 160498. doi: 10.1098/rsoc.160498.

Roberts, T. R., 1981. *Sundasalangidae*, a new family of minute freshwater salmoniform fishes from Southeast Asia. *Proc Calif Acad Sci* 59.

Roberts, C.M., Hawkins, J.P., 1999. Extinction risk in the sea. *Trends Ecol Evol* 14, 241-246. doi: 10.1016/S0169-5347(98)01584-5.

Rodrigues, A.S.L., Pilgrim, J.D., Lamoreux, J.F., Hoffmann, M., Brooks, T.M., 2006. The value of the IUCN Red List for conservation. *Trends Ecol Evol* 21, 71-76. doi: 10.1016/j.tree.2005.10.010.

Ruddle, K. and Ishige, N., 2010. On the origins, diffusion and cultural context of fermented fish products in Southeast Asia. In: Ferrer, J. (ed.), *Globalization, food and social identities in the Asia Pacific region*. Sophia University Institute of Comparative Culture, Tokyo, 1 – 17.

Sadovy, Y., 2001. The threat of fishing to highly fecund fishes. *J Fish Biol* 59, 90 – 108. doi: 10.1006/jfbi.2001.1760.

Sadovy de Mitcheson, Y., Craig, M.T., Bertoncini, A.A., Carpenter, K.E., Cheung, W.W.L., Choat, J.H., . . . Sanciangco, J., 2013. Fishing groupers towards extinction: a global assessment of threats and extinction risks in a billion dollar fishery. *Fish Fish* 14, 119-136. doi: 10.1111/j.1467-2979.2011.00455.x.

Schipper, J., Chanson, J.S., Chiozza, F., Cox, N.A., Hoffmann, M., Katariya, V., . . . Young, B.E., 2008. The status of the world's land and marine mammals: Diversity, threat, and knowledge. *Science* 322, 225-230. doi: 10.1126/science.1165115.

Santos, M., Munroe, T.A., Di Dario, F., Hata, H., Torres, F. and Quilang, J.P., 2018. *Sardinella tawilis* (errata version published in 2019). The IUCN Red List of Threatened Species 2018: e.T98836352A143839946. doi: 10.2305/IUCN.UK.20182.RLTS.T98836352A143839946.en. (accessed 6 June 2019).

Seminoff J, Shanker K., 2008. Marine turtles and IUCN Red Listing: A review of the process, the pitfalls, and novel assessment approaches. *J Exp Mar Biol Ecol* 356: 52–68. doi: 10.1016/j.jembe.2007.12.007.

Shelton, P.A., Sinclair, A.F., Chouninard, G.A., Mohn, R. and Duplisea, D.E., 2006. Fishing under low productivity conditions is further delaying recovery of Northwester Atlantic cod (*Gadus morhua*). *Can J Fish Aquat Sci* 63: 235 – 238. doi: 10.1139/F05-253.

Short, F.T., Polidoro, B., Linvingstone, S.R., Carpenter, K.E., Bandeira, S., Bujang, J.S., Calumpong, H.P., Carruthers, T.J.B., Coles, R.G., Dennison, W.C., Erftemeijer, P.L.A., Fortes, M.D., Freeman, A.S., Jagtap, T.G., Kamal, A.H.M., Kendrick, G.A., Kensworthy, W.J., La Nafie, Y.A., Nasution, I.M., Orth, R.J., Prathep, A., Sanciangco, J.C., van Tussenbroek, B., Vergara, S.G., Waycott, M., and Zieman, J.C., 2011. Extinction risk assessment of the world's seagrass species. *Biol Conserv* 144, 1961 – 1971.

Siple, M.C., Essington, T.E. and Plagányi, É. E., 2018. Forage fish fisheries management requires a tailored approach to balance trade-offs. *Fish Fish* 20, 110 – 124. doi: 10.1111/faf.12326.

Smith, D., Punt, A., Dowling, N., Smith, A., Tuck, G. and Knuckey, I., 2009. Reconciling approaches to the assessment and management of data-poor species and fisheries with Australia's harvest strategy policy. *Mar Coast Fish* 1, 244 – 254. doi: 10.1577/C08-041.1.

Smith, K.G. and Darwall, W.R.T., 2006. The status and distribution of freshwater fish endemic to the Mediterranean basin. IUCN, Gland, Switzerland and Cambridge, UK.

Smith, A.D.M., Brown, C.J., Bulman, C.M., Fulton, E.A., Johnson, P., Kaplan, I.C., Lozano-Montes, H., Mackinson, S., Marzloff, M., Shannon, L.J., Sjin, Y.-J., and Tam, J., 2011. impacts of fishing low-trophic level species on marine ecosystems. *Science* 333, 1147 – 1150. doi: 10.1126/science.1209395.

Smyllie, M., 2004. Herring: A History of the Silver Darlings. Tempus Publishing Limited, Stroud, Gloucestershire.

Spalding, M.D., Fox, H.E., Allen, G.R., Davidson, N., Ferdaña, Z.A., Finlayson, M., . . . Robertson, J., 2007. Marine ecoregions of the world: A bioregionalization of coastal and shelf areas. *BioScience* 57, 573-583. doi: 10.1641/b570707

Sprankle, K., 2005. Inter-dam movements and passage attraction of American shad in the lower Merrimack River main stem. *N Am J Fish Manag* 25, 1456-1466. doi: 10.1577/M04-049.1.

State of Ohio Environmental Protection Agency, 2008. Cuyahoga River aquatic life use attainment following the Kent and Munroe Falls dam modifications. Portage and Summit Counties. Ohio EPA Biological and Water Quality Report NEDO/2008-08-01. Available at: <https://epa.ohio.gov/dsw/tmdl/CuyahogaRiver#116519125-tmdl-report>. (accessed 4 September 2019).

Stearns. S.C., 1992. The evolution of life histories. Oxford University Press, Oxford.

Stobutzki, I., Miller, M. and Brewer, D., 2001. Sustainability of fishery bycatch: a process for assessing highly diverse and numerous bycatch. *Environ Conserv* 28, 167 – 181. doi: 10.1017/S0376892901000170.

Stump, E., Ralph, G.M., Comeros-Raynal, M.T., Matsuura, K. and Carpenter, K.E., 2018. Global conservation status of marine pufferfishes (Tetraodontiformes: Tetraodontidae). *Global Ecol Conserv* 14, e00388. doi: 10.1016/j.gecco.2018.e00388.

Tacon, A.G.J. and Metian, M., 2009a. Fishing for feed of fishing for food: increasing global competition for small pelagic forage fish. *AMBIO* 38, 294-302.

Tacon, A.G.J. and Metian, M., 2009b. Fishing for aquaculture: non-food use of small pelagic forage fish – a global perspective. *Rev Fish Sci* 17, 305 – 317. doi: 10.1080/10641260802677074.

Thornton, T.F., Moss, M.L., Butler, V.L., Hebert, J., and Funk, F., 2010. Local and traditional knowledge and the historical ecology of Pacific herring in Alaska. *J Ecol Anthropol* 14, 81 – 88.

Tous, P., Sidibé, A., Mbye, E., de Morais, L., Camara, K., Munroe, T., Adeofe, T.A., Camara, Y.H., Djiman, R., Sagna, A. & Sylla, M., 2015. *Sardinella maderensis*. The IUCN Red List of Threatened Species 2015: e.T198582A15543624. doi: 10.2305/IUCN.UK.2015-4.RLTS.T198582A15543624.en. (accessed 6 June 2019).

Tsukayama, I. and Palomares, M. L. D., 1987. Monthly catch and catch composition of Peruvian anchoveta (*Engraulis ringens*) (Northern-Central Stock, 4-14 S), 1953 to 1982. In: D. Pauly and L. Tsukayama (Eds.), The Peruvian anchoveta and its upwelling ecosystem: three decades of change. ICLARM Stud Rev 15, 89–108.

Tyedmers, P., 2004. Fisheries and energy use. Encyclopedia of Energy 2, 683 – 693.

United Nations, Department of Economic and Social Affairs, Population Division, 2017. World Population Prospects: The 2017 Revision, World Population 2017 Wallchart. AT/ESA/SER.A/398.

Van der Laan, R., W. N. Eschmeyer & R. Fricke, 2014. Family-group names of recent fishes. Zootaxa 3882, 1 – 230. doi: 10.11646/zootaxa.3882.1.1.

van der Meer, L., Arancibia, H., Zylich, K., and Zeller, D., 2015. Reconstruction of total marine fisheries catches for mainland Chile (1950- 2010). Working Paper Series. Working Paper # 2015-91. Fisheries Centre, University of British Columbia, Vancouver, BC, V6T 1Z4, Canada.

Van Puijenbroek, P.J.T.M., Buijse, A.D., Kraak, M.H.S. and Verdonchot, P.F.M., 2019. Species and river specific effects of river fragmentation on European anadromous fish species. River Res Appl 35, 66 – 77. doi: 10.1002/rra.3386.

Venter, O., Brodeur, N.N., Nemiroff, L., Belland, B., Dolinsek, I.J., and Grant, J.W.A., 2006. Threats to endangered species in Canada. BioScience 56, 904 – 910.

Vié, J.-C., Hilton-Taylor, C. and Stuart, S.N., 2009. Wildlife in a Changing World - An Analysis of the 2008 IUCN Red List of Threatened Species. IUCN, Gland, Switzerland.

Villanueva, L.S., Luistro, A.P. and Calabig, C.S., 1996. Assessment of Lake Taal capture fisheries with emphasis on the exploitation of *Harengula tawilis*. Tanauan, Batangas, the Philippines. STIARC-Experimental Farm for Fisheries.

WDFW, 2018. Washington state herring stock status report. Marine Fisheries Division. Washington Department of Fish and Wildlife.

Welcomme, R.L., Cowx, I.G., Coates, D., Béné, C., Funge-Smith, S., Halls, A., and Lorenzen, K., 2010. Inland capture fisheries. Philos Trans R Soc B 365, 2881 – 2896. doi:10.1098/rstb.2010.0168.

Whitehead, P.J.P., 1985. FAO species catalogue. Vol. 7. Clupeoid fishes of the world (suborder Clupeoidei). An annotated and illustrated catalogue of the herrings, sardines, pilchards, sprats, shads, anchovies and wolf-herrings. Part 1 - Chirocentridae, Clupeidae and Pristigasteridae. Food and Agricultural Organization (FAO) Fisheries Synopsis, Rome, Italy.

Whitehead, P.J.P., Nelson, G.J. and Wongratana, T., 1988. FAO species catalogue. Vol. 7. Clupeoid fishes of the world (Suborder Clupeoidei). An annotated and illustrated catalogue of the herrings, sardines, pilchards, sprats, shads, anchovies and wolf-herrings. Part 2 - Engraulididae. Food and Agricultural Organization (FAO) Fisheries Synopsis, Rome, Italy.

Willette, D.A., Bognot, E.D.C., Mutia, Ma.T.M. and Santos, M. D., 2011. Biology and ecology of sardines in the Philippines: A review. Bureau of Fisheries and Aquatic Resources – National Fisheries Research and Development Institute 13. Philippines.

Willis, K.J., 2017. State of the World's Plants 2017. Report. Royal Botanic Gardens, Kew.

Wongratana, T., Munroe, T.A. and Nizinski, M., 1999. Engraulidae. In: Carpenter, K.E. and Niem, V.H. (Eds.), The living marine resources of the western central Pacific. Volume 3 batoid fishes, chimaeras and bony fishes part 1 (Elopidae to Linophrynidae), 1698 – 1753. FAO, Rome.

Worm, B., Hilborn, R., Baum, J. K., Branch, T. A., Collie, J. S., Costello, C., ... Zeller, D., 2009. Rebuilding global fisheries. *Science* 325, 578 – 585. doi: [10.1126/science.1173146](https://doi.org/10.1126/science.1173146).

Worm, B., Barbier, E.B., Beaumont, N., Duffy, J.E., Folke, C., Halpern, B.S., Jackson, J.B.C., Lotze, H.K., Micheli, F., Palumbi, S.R., Sala, E., Selkoe, K.A., Stachowicz, J.J., Watson, R., 2006. Impacts of biodiversity loss on ocean ecosystem services. *Science* 314, 787-790. doi: [10.1126/science.1132294](https://doi.org/10.1126/science.1132294)

Worm, B., Branch, T.A., 2012. The future of fish. *Trends Ecol Evol* 27, 594-599. doi: [10.1016/j.tree.2012.07.005](https://doi.org/10.1016/j.tree.2012.07.005).

Young, H.S., McCauley, D.J., Galetti, M. and Dirzo, R., 2016. Patterns, causes, and consequences of anthropocene defaunation. *Annu Rev Ecol Evol S* 47, 333 – 358.

APPENDIX A

LIST OF ALL CLUPEIFORM IUCN RED LIST CATEGORIES

Table A1: List of all 405 clupeiforms alphabetical by family and then by species name. The global IUCN Red List categories and criteria are listed: CR = Critically Endangered, EN = Endangered, VU = Vulnerable, NT = Near Threatened, LC = Least Concern, DD = Data Deficient, NE = Not Evaluated. Criterion A = population decline in the past, present or future, B = restricted range, C = small population size and decline, D = very small or restricted population, E = quantitative analysis of extinction probability. For further information available on categories and criteria, visit the IUCN Red List website (www.iucnredlist.org). The preferred habitat system is also listed; F = Freshwater, M = Marine, E = Euryhaline which includes estuarine species and diadromous species.

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Chirocentridae	<i>Chirocentrus dorab</i>	LC	M
Chirocentridae	<i>Chirocentrus nudus</i>	LC	M
Clupeidae	<i>Alosa aestivalis</i>	VU A2b	E
Clupeidae	<i>Alosa agone</i>	LC	F
Clupeidae	<i>Alosa alabamae</i>	NT A2ac	E
Clupeidae	<i>Alosa algeriensis</i>	DD	E
Clupeidae	<i>Alosa alosa</i>	LC	E
Clupeidae	<i>Alosa braschnikowi</i>	DD	E
Clupeidae	<i>Alosa caspia</i>	LC	E
Clupeidae	<i>Alosa chrysochloris</i>	LC	E
Clupeidae	<i>Alosa curensis</i>	DD	E
Clupeidae	<i>Alosa fallax</i>	LC	E
Clupeidae	<i>Alosa immaculata</i>	VU B2ab(v)	E
Clupeidae	<i>Alosa kessleri</i>	LC	E
Clupeidae	<i>Alosa killarnensis</i>	CR B1ab(iii)	F
Clupeidae	<i>Alosa macedonica</i>	VU D2	F
Clupeidae	<i>Alosa maeotica</i>	LC	E
Clupeidae	<i>Alosa mediocris</i>	LC	E
Clupeidae	<i>Alosa pontica</i>	LC	E

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Clupeidae	<i>Alosa pseudoharengus</i>	LC	E
Clupeidae	<i>Alosa sapidissima</i>	LC	E
Clupeidae	<i>Alosa saposchnikowii</i>	DD	E
Clupeidae	<i>Alosa sphaerocephala</i>	LC	E
Clupeidae	<i>Alosa suworowi</i>	DD	E
Clupeidae	<i>Alosa tanaica</i>	LC	E
Clupeidae	<i>Alosa vistonica</i>	CR A2ace; B1ab(iii,v)	F
Clupeidae	<i>Alosa volgensis</i>	EN B2ab(iii,v)	E
Clupeidae	<i>Amblygaster clupeoides</i>	LC	M
Clupeidae	<i>Amblygaster indiana</i>	DD	M
Clupeidae	<i>Amblygaster leiogaster</i>	LC	M
Clupeidae	<i>Amblygaster sirm</i>	LC	M
Clupeidae	<i>Anodontostoma chacunda</i>	LC	E
Clupeidae	<i>Anodontostoma selangkat</i>	LC	E
Clupeidae	<i>Anodontostoma thailandiae</i>	LC	E
Clupeidae	<i>Brevoortia aurea</i>	LC	E
Clupeidae	<i>Brevoortia gunteri</i>	LC	M
Clupeidae	<i>Brevoortia patronus</i>	LC	E
Clupeidae	<i>Brevoortia pectinata</i>	LC	E
Clupeidae	<i>Brevoortia smithi</i>	LC	E
Clupeidae	<i>Brevoortia tyrannus</i>	LC	E
Clupeidae	<i>Clupanodon thrissa</i>	LC	E
Clupeidae	<i>Clupea harengus</i>	LC	M
Clupeidae	<i>Clupea pallasii</i>	DD	M
Clupeidae	<i>Clupeichthys aesarnensis</i>	LC	F
Clupeidae	<i>Clupeichthys bleekeri</i>	VU B1ab(iii)	F
Clupeidae	<i>Clupeichthys goniognathus</i>	LC	E
Clupeidae	<i>Clupeichthys perakensis</i>	LC	E
Clupeidae	<i>Clupeoides borneensis</i>	LC	E
Clupeidae	<i>Clupeoides hypselosoma</i>	DD	F
Clupeidae	<i>Clupeoides papuensis</i>	DD	F
Clupeidae	<i>Clupeoides venulosus</i>	VU B2ab(iii,v)	F
Clupeidae	<i>Clupeonella abrau</i>	CR B1ab(ii,iii,v)+2ab(ii,iii,v)	F
Clupeidae	<i>Clupeonella caspia</i>	LC	E
Clupeidae	<i>Clupeonella cultriventris</i>	LC	E
Clupeidae	<i>Clupeonella engrauliformis</i>	EN A2bde	M
Clupeidae	<i>Clupeonella grimmi</i>	EN A2bde	M
Clupeidae	<i>Clupeonella muhlisi</i>	EN B1ab(iii)+2ab(iii)	F
Clupeidae	<i>Clupeonella tscharchalensis</i>	LC	E
Clupeidae	<i>Congothrissa gosse</i>	DD	F

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Clupeidae	<i>Corica laciniata</i>	DD	F
Clupeidae	<i>Corica soborna</i>	LC	E
Clupeidae	<i>Dayella malabarica</i>	LC	E
Clupeidae	<i>Dorosoma anale</i>	LC	F
Clupeidae	<i>Dorosoma cepedianum</i>	LC	E
Clupeidae	<i>Dorosoma chavesi</i>	NT B1ab(iii)	F
Clupeidae	<i>Dorosoma petenense</i>	LC	E
Clupeidae	<i>Dorosoma smithi</i>	DD	F
Clupeidae	<i>Dussumieria acuta</i>	LC	M
Clupeidae	<i>Dussumieria elopsoidea</i>	LC	M
Clupeidae	<i>Ehirava fluviatilis</i>	DD	E
Clupeidae	<i>Escualosa elongata</i>	DD	M
Clupeidae	<i>Escualosa thoracata</i>	LC	E
Clupeidae	<i>Ethmalosa fimbriata</i>	LC	E
Clupeidae	<i>Ethmidium maculatum</i>	DD	M
Clupeidae	<i>Etrumeus acuminatus</i>	LC	M
Clupeidae	<i>Etrumeus golanii</i>	DD	M
Clupeidae	<i>Etrumeus jacksoniensis</i>	LC	M
Clupeidae	<i>Etrumeus makiawa</i>	LC	M
Clupeidae	<i>Etrumeus micropus</i>	LC	M
Clupeidae	<i>Etrumeus sadina</i>	LC	M
Clupeidae	<i>Etrumeus whiteheadi</i>	LC	M
Clupeidae	<i>Etrumeus wongratanai</i>	DD	M
Clupeidae	<i>Gilchristella aestuaria</i>	LC	E
Clupeidae	<i>Gonialosa manmina</i>	LC	E
Clupeidae	<i>Gonialosa modesta</i>	DD	E
Clupeidae	<i>Gonialosa whiteheadi</i>	DD	E
Clupeidae	<i>Gudusia chapra</i>	LC	F
Clupeidae	<i>Gudusia variegata</i>	LC	F
Clupeidae	<i>Harengula clupeola</i>	LC	E
Clupeidae	<i>Harengula humeralis</i>	LC	E
Clupeidae	<i>Harengula jaguana</i>	LC	M
Clupeidae	<i>Harengula thrissina</i>	LC	E
Clupeidae	<i>Herklotsichthys blackburni</i>	DD	E
Clupeidae	<i>Herklotsichthys castelnaui</i>	LC	E
Clupeidae	<i>Herklotsichthys collettei</i>	LC	M
Clupeidae	<i>Herklotsichthys dispilonotus</i>	LC	M
Clupeidae	<i>Herklotsichthys gotoi</i>	LC	E
Clupeidae	<i>Herklotsichthys koningsbergeri</i>	LC	E
Clupeidae	<i>Herklotsichthys lippa</i>	LC	M

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Clupeidae	<i>Herklotsichthys lossei</i>	LC	M
Clupeidae	<i>Herklotsichthys ovalis</i>	DD	M
Clupeidae	<i>Herklotsichthys punctatus</i>	LC	M
Clupeidae	<i>Herklotsichthys quadrimaculatus</i>	LC	M
Clupeidae	<i>Herklotsichthys spilurus</i>	LC	M
Clupeidae	<i>Hilsa kelee</i>	LC	E
Clupeidae	<i>Hyperlophus translucidus</i>	LC	E
Clupeidae	<i>Hyperlophus vittatus</i>	LC	E
Clupeidae	<i>Jenkinsia lamprotaenia</i>	LC	M
Clupeidae	<i>Jenkinsia majua</i>	LC	M
Clupeidae	<i>Jenkinsia parvula</i>	DD	M
Clupeidae	<i>Jenkinsia stolifera</i>	LC	M
Clupeidae	<i>Konosirus punctatus</i>	LC	E
Clupeidae	<i>Laeviscutella dekimpei</i>	LC	E
Clupeidae	<i>Lile gracilis</i>	LC	E
Clupeidae	<i>Lile nigrofasciata</i>	LC	E
Clupeidae	<i>Lile piquitinga</i>	LC	E
Clupeidae	<i>Lile stolifera</i>	LC	E
Clupeidae	<i>Limnothrissa miodon</i>	LC	E
Clupeidae	<i>Limnothrissa stappersii</i>	DD	F
Clupeidae	<i>Microthrissa minuta</i>	VU D2	F
Clupeidae	<i>Microthrissa royauxi</i>	LC	F
Clupeidae	<i>Microthrissa whiteheadi</i>	LC	F
Clupeidae	<i>Minyclupeoides dentibranchialis</i>	LC	E
Clupeidae	<i>Nannothrissa parva</i>	LC	F
Clupeidae	<i>Nannothrissa stewarti</i>	EN B1ab(v)	F
Clupeidae	<i>Nematalosa arabica</i>	DD	M
Clupeidae	<i>Nematalosa come</i>	LC	M
Clupeidae	<i>Nematalosa erebi</i>	LC	F
Clupeidae	<i>Nematalosa flyensis</i>	DD	F
Clupeidae	<i>Nematalosa galathea</i>	LC	E
Clupeidae	<i>Nematalosa japonica</i>	DD	M
Clupeidae	<i>Nematalosa nasus</i>	LC	E
Clupeidae	<i>Nematalosa papuensis</i>	DD	F
Clupeidae	<i>Nematalosa persara</i>	DD	M
Clupeidae	<i>Nematalosa resticularia</i>	DD	M
Clupeidae	<i>Nematalosa vlaminghi</i>	LC	E
Clupeidae	<i>Odaxothrissa ansorgii</i>	LC	F
Clupeidae	<i>Odaxothrissa losera</i>	DD	F
Clupeidae	<i>Odaxothrissa mento</i>	LC	F

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Clupeidae	<i>Odaxothrissa vittata</i>	LC	F
Clupeidae	<i>Opisthonema berlangai</i>	VU D2	M
Clupeidae	<i>Opisthonema bulleri</i>	LC	M
Clupeidae	<i>Opisthonema libertate</i>	LC	M
Clupeidae	<i>Opisthonema medirastre</i>	LC	M
Clupeidae	<i>Opisthonema oglinum</i>	LC	E
Clupeidae	<i>Pellonula leonensis</i>	LC	E
Clupeidae	<i>Pellonula vorax</i>	LC	E
Clupeidae	<i>Platanichthys platana</i>	LC	E
Clupeidae	<i>Pliosteostoma lutipinnis</i>	LC	E
Clupeidae	<i>Poecilothrissa centralis</i>	LC	F
Clupeidae	<i>Poecilothrissa congica</i>	LC	F
Clupeidae	<i>Poecilothrissa moeruensis</i>	VU B1ab(v)	F
Clupeidae	<i>Potamalosa richmondia</i>	NT B2ab(I,ii,iii,iv,v)	E
Clupeidae	<i>Potamothrissa acutirostris</i>	LC	F
Clupeidae	<i>Potamothrissa obtusirostris</i>	LC	F
Clupeidae	<i>Potamothrissa whiteheadi</i>	DD	F
Clupeidae	<i>Ramnogaster arcuata</i>	LC	M
Clupeidae	<i>Ramnogaster melanostoma</i>	LC	F
Clupeidae	<i>Rhinosardinia amazonica</i>	LC	E
Clupeidae	<i>Rhinosardinia bahiensis</i>	LC	E
Clupeidae	<i>Sardina pilchardus</i>	LC	M
Clupeidae	<i>Sardinella albella</i>	LC	M
Clupeidae	<i>Sardinella atricauda</i>	LC	M
Clupeidae	<i>Sardinella aurita</i>	LC	M
Clupeidae	<i>Sardinella brachysoma</i>	LC	M
Clupeidae	<i>Sardinella brasiliensis</i>	DD	E
Clupeidae	<i>Sardinella dayi</i>	DD	M
Clupeidae	<i>Sardinella electra</i>	NE	M
Clupeidae	<i>Sardinella fijiense</i>	LC	M
Clupeidae	<i>Sardinella fimbriata</i>	LC	E
Clupeidae	<i>Sardinella gibbosa</i>	LC	M
Clupeidae	<i>Sardinella goni</i>	DD	M
Clupeidae	<i>Sardinella hualiensis</i>	LC	M
Clupeidae	<i>Sardinella jussieu</i>	DD	M
Clupeidae	<i>Sardinella lemuru</i>	NT A2bd	M
Clupeidae	<i>Sardinella longiceps</i>	LC	M
Clupeidae	<i>Sardinella maderensis</i>	VU A2d	M
Clupeidae	<i>Sardinella marquesensis</i>	LC	M
Clupeidae	<i>Sardinella melanura</i>	LC	E

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Clupeidae	<i>Sardinella neglecta</i>	LC	M
Clupeidae	<i>Sardinella pacifica</i>	NE	M
Clupeidae	<i>Sardinella richardsoni</i>	DD	M
Clupeidae	<i>Sardinella rouxi</i>	DD	M
Clupeidae	<i>Sardinella sindensis</i>	LC	E
Clupeidae	<i>Sardinella tawilis</i>	EN A2bd; B1ab(iii,v)+2ab(iii,v)	F
Clupeidae	<i>Sardinella zunasi</i>	LC	M
Clupeidae	<i>Sardinops sagax</i>	LC	M
Clupeidae	<i>Sauvagella madagascariensis</i>	LC	E
Clupeidae	<i>Sauvagella robusta</i>	EN B2ab(iii)	F
Clupeidae	<i>Sierrathrissa leonensis</i>	LC	F
Clupeidae	<i>Spratelloides atrofasciatus</i>	LC	M
Clupeidae	<i>Spratelloides delicatulus</i>	LC	M
Clupeidae	<i>Spratelloides gracilis</i>	LC	M
Clupeidae	<i>Spratelloides lewisi</i>	LC	M
Clupeidae	<i>Spratelloides robustus</i>	LC	E
Clupeidae	<i>Spratellomorpha bianalis</i>	DD	E
Clupeidae	<i>Sprattus antipodum</i>	LC	M
Clupeidae	<i>Sprattus fuegensis</i>	LC	M
Clupeidae	<i>Sprattus muelleri</i>	LC	M
Clupeidae	<i>Sprattus novaehollandiae</i>	LC	E
Clupeidae	<i>Sprattus sprattus</i>	LC	E
Clupeidae	<i>Stolothrissa tanganicae</i>	LC	F
Clupeidae	<i>Strangomera bentincki</i>	LC	M
Clupeidae	<i>Tenualosa ilisha</i>	LC	E
Clupeidae	<i>Tenualosa macrura</i>	NT B2ab(iii)	E
Clupeidae	<i>Tenualosa reevesii</i>	DD	E
Clupeidae	<i>Tenualosa thibaudeaui</i>	VU A2bcd	F
Clupeidae	<i>Tenualosa toli</i>	VU B2ab(iii,v)	E
Clupeidae	<i>Thrattidion noctivagus</i>	DD	F
Denticipitidae	<i>Denticeps clupeoides</i>	VU B2ab(iii)	F
Engraulidae	<i>Amazonsprattus scintilla</i>	LC	F
Engraulidae	<i>Anchoa analis</i>	DD	E
Engraulidae	<i>Anchoa argentivittata</i>	LC	M
Engraulidae	<i>Anchoa belizensis</i>	LC	F
Engraulidae	<i>Anchoa cayorum</i>	LC	M
Engraulidae	<i>Anchoa chamensis</i>	DD	M
Engraulidae	<i>Anchoa choerostoma</i>	EN B1ab(v)+2ab(v)	M
Engraulidae	<i>Anchoa colonensis</i>	LC	M

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Engraulidae	<i>Anchoa compressa</i>	LC	E
Engraulidae	<i>Anchoa cubana</i>	LC	E
Engraulidae	<i>Anchoa curta</i>	LC	E
Engraulidae	<i>Anchoa delicatissima</i>	LC	E
Engraulidae	<i>Anchoa eigenmannia</i>	LC	M
Engraulidae	<i>Anchoa exigua</i>	LC	M
Engraulidae	<i>Anchoa filifera</i>	LC	E
Engraulidae	<i>Anchoa helleri</i>	LC	M
Engraulidae	<i>Anchoa hepsetus</i>	LC	E
Engraulidae	<i>Anchoa ischana</i>	LC	M
Engraulidae	<i>Anchoa januaria</i>	LC	E
Engraulidae	<i>Anchoa lamprotaenia</i>	LC	M
Engraulidae	<i>Anchoa lucida</i>	LC	E
Engraulidae	<i>Anchoa lyolepis</i>	LC	M
Engraulidae	<i>Anchoa marinii</i>	LC	E
Engraulidae	<i>Anchoa mitchilli</i>	LC	E
Engraulidae	<i>Anchoa mundeola</i>	LC	E
Engraulidae	<i>Anchoa mundeoloides</i>	LC	E
Engraulidae	<i>Anchoa nasus</i>	LC	M
Engraulidae	<i>Anchoa panamensis</i>	LC	E
Engraulidae	<i>Anchoa parva</i>	LC	E
Engraulidae	<i>Anchoa pectoralis</i>	LC	E
Engraulidae	<i>Anchoa scofieldi</i>	LC	E
Engraulidae	<i>Anchoa spinifer</i>	LC	E
Engraulidae	<i>Anchoa starksi</i>	LC	E
Engraulidae	<i>Anchoa tricolor</i>	LC	E
Engraulidae	<i>Anchoa trinitatis</i>	DD	M
Engraulidae	<i>Anchoa walkeri</i>	LC	E
Engraulidae	<i>Anchovia clupeoides</i>	LC	E
Engraulidae	<i>Anchovia landivarensis</i>	DD	E
Engraulidae	<i>Anchovia macrolepidota</i>	LC	E
Engraulidae	<i>Anchovia surinamensis</i>	LC	E
Engraulidae	<i>Anchoviella alleni</i>	LC	F
Engraulidae	<i>Anchoviella balboae</i>	DD	M
Engraulidae	<i>Anchoviella blackburni</i>	DD	E
Engraulidae	<i>Anchoviella brevirostris</i>	LC	E
Engraulidae	<i>Anchoviella carrikeri</i>	LC	F
Engraulidae	<i>Anchoviella cayennensis</i>	LC	E
Engraulidae	<i>Anchoviella elongata</i>	LC	E
Engraulidae	<i>Anchoviella guianensis</i>	LC	F

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Engraulidae	<i>Anchoviella hernanni</i>	LC	F
Engraulidae	<i>Anchoviella jamesi</i>	LC	F
Engraulidae	<i>Anchoviella juruasanga</i>	LC	F
Engraulidae	<i>Anchoviella lepidentostole</i>	LC	E
Engraulidae	<i>Anchoviella manamensis</i>	LC	F
Engraulidae	<i>Anchoviella miarcha</i>	DD	E
Engraulidae	<i>Anchoviella perezii</i>	DD	F
Engraulidae	<i>Anchoviella perfasciata</i>	LC	M
Engraulidae	<i>Anchoviella sanfranciscana</i>	DD	E
Engraulidae	<i>Anchoviella vaillanti</i>	LC	F
Engraulidae	<i>Cetengraulis edentulus</i>	LC	E
Engraulidae	<i>Cetengraulis mysticetus</i>	LC	M
Engraulidae	<i>Coilia borneensis</i>	DD	E
Engraulidae	<i>Coilia coomansi</i>	DD	E
Engraulidae	<i>Coilia dussumieri</i>	LC	E
Engraulidae	<i>Coilia grayii</i>	LC	E
Engraulidae	<i>Coilia lindmani</i>	LC	E
Engraulidae	<i>Coilia macrognathos</i>	DD	E
Engraulidae	<i>Coilia mystus</i>	EN A2bd	E
Engraulidae	<i>Coilia nasus</i>	EN A2bd	E
Engraulidae	<i>Coilia neglecta</i>	LC	E
Engraulidae	<i>Coilia ramcarati</i>	DD	E
Engraulidae	<i>Coilia rebentischii</i>	DD	E
Engraulidae	<i>Coilia reynaldi</i>	LC	E
Engraulidae	<i>Encrasicholina auster</i>	DD	M
Engraulidae	<i>Encrasicholina gloria</i>	DD	M
Engraulidae	<i>Encrasicholina heteroloba</i>	LC	M
Engraulidae	<i>Encrasicholina intermedia</i>	DD	M
Engraulidae	<i>Encrasicholina macrocephala</i>	DD	M
Engraulidae	<i>Encrasicholina oligobranchus</i>	DD	M
Engraulidae	<i>Encrasicholina pseudoheteroloba</i>	LC	M
Engraulidae	<i>Encrasicholina punctifer</i>	LC	M
Engraulidae	<i>Encrasicholina purpurea</i>	LC	E
Engraulidae	<i>Engraulis albidus</i>	DD	E
Engraulidae	<i>Engraulis anchoita</i>	NT A2bd	M
Engraulidae	<i>Engraulis australis</i>	LC	E
Engraulidae	<i>Engraulis capensis</i>	LC	M
Engraulidae	<i>Engraulis encrasicolus</i>	LC	E
Engraulidae	<i>Engraulis eurystole</i>	LC	M
Engraulidae	<i>Engraulis japonicus</i>	LC	M

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Engraulidae	<i>Engraulis mordax</i>	LC	M
Engraulidae	<i>Engraulis ringens</i>	LC	M
Engraulidae	<i>Jurengraulis juruensis</i>	LC	F
Engraulidae	<i>Lycengraulis batesii</i>	LC	E
Engraulidae	<i>Lycengraulis figueiredoi</i>	LC	F
Engraulidae	<i>Lycengraulis grossidens</i>	LC	E
Engraulidae	<i>Lycengraulis limnichthys</i>	DD	E
Engraulidae	<i>Lycengraulis poeyi</i>	LC	E
Engraulidae	<i>Lycothrissa crocodilus</i>	LC	F
Engraulidae	<i>Papuengraulis micropinna</i>	DD	E
Engraulidae	<i>Pseudosetipinna haizhouensis</i>	DD	M
Engraulidae	<i>Pterengraulis atherinoides</i>	LC	E
Engraulidae	<i>Setipinna breviceps</i>	LC	E
Engraulidae	<i>Setipinna brevifilis</i>	DD	F
Engraulidae	<i>Setipinna melanochir</i>	DD	E
Engraulidae	<i>Setipinna paxtoni</i>	DD	M
Engraulidae	<i>Setipinna phasa</i>	LC	E
Engraulidae	<i>Setipinna taty</i>	LC	E
Engraulidae	<i>Setipinna tenuifilis</i>	DD	E
Engraulidae	<i>Setipinna wheeleri</i>	DD	F
Engraulidae	<i>Stolephorus advenus</i>	DD	M
Engraulidae	<i>Stolephorus andhraensis</i>	LC	E
Engraulidae	<i>Stolephorus apiensis</i>	LC	M
Engraulidae	<i>Stolephorus baganensis</i>	LC	M
Engraulidae	<i>Stolephorus brachycephalus</i>	LC	E
Engraulidae	<i>Stolephorus carpentariae</i>	LC	E
Engraulidae	<i>Stolephorus chinensis</i>	LC	E
Engraulidae	<i>Stolephorus commersonii</i>	LC	M
Engraulidae	<i>Stolephorus continentalis</i>	DD	M
Engraulidae	<i>Stolephorus dubiosus</i>	LC	E
Engraulidae	<i>Stolephorus holodon</i>	LC	E
Engraulidae	<i>Stolephorus indicus</i>	LC	E
Engraulidae	<i>Stolephorus insignis</i>	NE	M
Engraulidae	<i>Stolephorus insularis</i>	LC	E
Engraulidae	<i>Stolephorus multibranchus</i>	DD	M
Engraulidae	<i>Stolephorus nelsoni</i>	DD	E
Engraulidae	<i>Stolephorus oceanicus</i>	DD	M
Engraulidae	<i>Stolephorus pacificus</i>	DD	M
Engraulidae	<i>Stolephorus ronquilloi</i>	DD	E
Engraulidae	<i>Stolephorus shantungensis</i>	DD	E

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Engraulidae	<i>Stolephorus teguhi</i>	DD	E
Engraulidae	<i>Stolephorus tri</i>	LC	M
Engraulidae	<i>Stolephorus waitei</i>	DD	M
Engraulidae	<i>Thryssa adela</i>	DD	M
Engraulidae	<i>Thryssa aestuaria</i>	LC	E
Engraulidae	<i>Thryssa baelama</i>	LC	E
Engraulidae	<i>Thryssa brevicauda</i>	LC	E
Engraulidae	<i>Thryssa chefuensis</i>	DD	E
Engraulidae	<i>Thryssa dayi</i>	DD	M
Engraulidae	<i>Thryssa dussumieri</i>	LC	E
Engraulidae	<i>Thryssa encrasicholoides</i>	DD	M
Engraulidae	<i>Thryssa gautamiensis</i>	DD	E
Engraulidae	<i>Thryssa hamiltonii</i>	LC	E
Engraulidae	<i>Thryssa kammalensis</i>	DD	E
Engraulidae	<i>Thryssa kammalensooides</i>	DD	E
Engraulidae	<i>Thryssa malabarica</i>	DD	E
Engraulidae	<i>Thryssa marasriae</i>	LC	E
Engraulidae	<i>Thryssa mystax</i>	LC	E
Engraulidae	<i>Thryssa polybranchialis</i>	DD	M
Engraulidae	<i>Thryssa purava</i>	DD	M
Engraulidae	<i>Thryssa rastrosa</i>	EN B1ab(i,ii,iii,v)+2ab(i,ii,iii,v)	F
Engraulidae	<i>Thryssa scratchleyi</i>	DD	E
Engraulidae	<i>Thryssa setirostris</i>	LC	E
Engraulidae	<i>Thryssa spinidens</i>	DD	M
Engraulidae	<i>Thryssa stenosoma</i>	DD	M
Engraulidae	<i>Thryssa vitrirostris</i>	LC	E
Engraulidae	<i>Thryssa whiteheadi</i>	LC	M
Pristigasteridae	<i>Chirocentron bleekermanus</i>	LC	E
Pristigasteridae	<i>Ilisha africana</i>	LC	E
Pristigasteridae	<i>Ilisha amazonica</i>	LC	F
Pristigasteridae	<i>Ilisha compressa</i>	LC	M
Pristigasteridae	<i>Ilisha elongata</i>	LC	E
Pristigasteridae	<i>Ilisha filigera</i>	DD	E
Pristigasteridae	<i>Ilisha fuerthii</i>	LC	E
Pristigasteridae	<i>Ilisha kampeni</i>	LC	E
Pristigasteridae	<i>Ilisha lunula</i>	DD	E
Pristigasteridae	<i>Ilisha macrogaster</i>	DD	E
Pristigasteridae	<i>Ilisha megaloptera</i>	LC	E
Pristigasteridae	<i>Ilisha melastoma</i>	LC	E

FAMILY	SPECIES NAME	GLOBAL CATEGORY & CRITERIA	SYSTEM
Pristigasteridae	<i>Ilisha novacula</i>	LC	F
Pristigasteridae	<i>Ilisha obfuscata</i>	DD	M
Pristigasteridae	<i>Ilisha pristigastroides</i>	DD	E
Pristigasteridae	<i>Ilisha sirishai</i>	DD	M
Pristigasteridae	<i>Ilisha striatula</i>	DD	E
Pristigasteridae	<i>Neopisthopterus cubanus</i>	VU B2ab(i,ii,iii)	E
Pristigasteridae	<i>Neopisthopterus tropicus</i>	LC	E
Pristigasteridae	<i>Odontognathus compressus</i>	LC	E
Pristigasteridae	<i>Odontognathus mucronatus</i>	LC	E
Pristigasteridae	<i>Odontognathus panamensis</i>	LC	E
Pristigasteridae	<i>Opisthopterus dovii</i>	LC	E
Pristigasteridae	<i>Opisthopterus effulgens</i>	DD	E
Pristigasteridae	<i>Opisthopterus equatorialis</i>	LC	M
Pristigasteridae	<i>Opisthopterus macrops</i>	LC	M
Pristigasteridae	<i>Opisthopterus tardoore</i>	LC	E
Pristigasteridae	<i>Opisthopterus valenciennesi</i>	DD	E
Pristigasteridae	<i>Pellona castelnaeana</i>	LC	E
Pristigasteridae	<i>Pellona dayi</i>	DD	M
Pristigasteridae	<i>Pellona ditchela</i>	LC	E
Pristigasteridae	<i>Pellona flavipinnis</i>	LC	F
Pristigasteridae	<i>Pellona harroweri</i>	LC	E
Pristigasteridae	<i>Pristigaster cayana</i>	LC	F
Pristigasteridae	<i>Pristigaster whiteheadi</i>	LC	F
Pristigasteridae	<i>Raconda russeliana</i>	LC	E
Sundasangidae	<i>Sundasang malletti</i>	DD	F
Sundasangidae	<i>Sundasang megalops</i>	DD	F
Sundasangidae	<i>Sundasang mekongensis</i>	LC	F
Sundasangidae	<i>Sundasang mesops</i>	DD	F
Sundasangidae	<i>Sundasang microps</i>	DD	F
Sundasangidae	<i>Sundasang platyrhynchus</i>	DD	F
Sundasangidae	<i>Sundasang praecox</i>	LC	F

APPENDIX B

IUCN RED LIST METHODS AND DATA USE

To supplement the Red List methods described in Chapter 2, further information on important terminology, threat classifications, distribution mapping methodology and the estimation of declines used in the Red List assessments and the thesis are outlined. Further information regarding the uncertainty within the Red List assessments is also expressed below.

Red List terminology

Within the scope of the Red List methodology, specific definitions are used. The term population refers to the total number of individuals of a species throughout its global distribution, while population size is the total number of mature individuals (e.g., those capable of reproduction). Both terms, population and population size, are required for criteria A, C and D (IUCN Standards and Petitions Subcommittee, 2017). Generation length is applicable to criteria A, C1 and E, and is the average age of parents of the current cohort (i.e., recruited individuals in the population) and serves as a measure of the turnover rate of breeding individuals within the population (IUCN Standards and Petitions Subcommittee, 2017). The pre-disturbance generation length was used to account for potential variation under threat such as exploitation (IUCN Standards and Petitions Subcommittee, 2017). Declines must be calculated over a period of time equal to three generation lengths or ten years, whichever is longer (IUCN, 2012). The equation used in the assessments to calculate generation length is as follows:

Generation Length =

Age at first reproduction + (age at last reproduction – age at first reproduction) / 2

Location defines a geographically or ecologically distinct area where a single threatening event can rapidly impact all individuals of the taxon present and is necessary for the application of criteria B and D (IUCN Standards and Petitions Subcommittee, 2017). The known geographic extent of a species is quantitatively expressed in two ways: Extent of Occurrence (EOO) used for criteria A and B and Area of Occupancy (AOO) used for criteria A, B and D (IUCN Standard and Petitions Subcommittee, 2017). The EOO is defined by the smallest, continuous imaginary boundary that can be drawn around the area where the species is known, inferred or suspected to be present. It is also referred to as the ‘minimum convex polygon’ and represents the degree to which threatening factors are spatially spread across a taxon’s geographic range. The AOO is the area within the species’ EOO that is actually occupied, accounting for the fact that the EOO likely contains unoccupied or unsuitable habitat. A 2x2 km grid is used to standardize estimates of AOO (IUCN, 2017).

Threat classifications

Major threats used in analyses follow the hierarchal threat schematic provided by the IUCN Red List. Major threats were coded within a species assessment only if confirmation of impact on the species or locality within its range exists. Many clupeiform species have limited data available regarding their conservation status. Therefore, for threats that were only suspected to impact a species, the threat was neither coded within the species assessment, nor included in the analyses.

Major threats known to impact clupeiforms include biological resource use (n = 106), pollution (n = 47), natural system modifications (n = 42), climate change (n = 23), invasive species and diseases (n = 15), energy production (n = 7), residential or commercial development (n = 5), human intrusions (n = 1) and transportation service corridors (n = 1). Within these major threats, sub-threats were also coded to specify the source of the major threat (IUCN, 2019). For clupeiforms, biological resource was coded for species impacted by bycatch, subsistence, artisanal, recreational, commercial and industrial exploitation. Pollution as a major threat is sourced from agricultural, domestic, industrial and/or military effluents but also includes sedimentation. Large and small dams as well as water management/use (e.g., water abstraction) are included under natural system modifications. Climate change is broken down into specific impacts, which include droughts, habitat shifting and temperature extremes. The invasive species and diseases category include both native, and non-native problematic species or diseases. Energy production exclusively refers to impacts from mining and quarrying for this taxa. Threats known to impact five or less species but ultimately may disturb critical habitats include residential and commercial development (e.g., commercial, industrial or housing development projects), transportation corridors (e.g., shipping lanes) and human intrusions which stem from recreational activities. For further detailed information regarding IUCN Red List threat schemes, see the IUCN Red List website (www.iucnredlist.org).

Distribution maps

A species-specific distribution map is a depiction of a taxon's native geographic range or limits of distribution and can be helpful in communicating and/or addressing conservation

planning. These maps are used for visualization and spatial analyses and can also be used in different types of analyses that can identify gaps in knowledge and conservation priority areas by, for example, highlighting areas with a high number of threatened or Data Deficient species. However, the polygons neither depict the potential spread of extinction risk nor do they represent that a species is uniformly distributed throughout. They can be used to support the estimate of AOO or EOO, but do not represent either parameter.

Estimates of decline

Time series data of spawning stock biomass (SSB), catch-per-unit effort (CPUE), total landings reported to the FAO (FAO, 2016) and reconstructed catches (Pauly and Zeller, 2016a), where available, were used as indices of abundance to estimate population decline. If available, estimated biomass (e.g., SSB) from fishery stock assessments took priority over other data types, such as landings, when calculating declines. Fishery-dependent data (e.g., reported landings or reconstructed catches) were reported to the species, genus or family level.

Uncertainty within Red List data

Data were often pieced together from various sources to determine the species' conservation status; it is understood that there is inherent uncertainty within the available data and thus, the resulting conclusions. Uncertainty may arise from factors including natural variation, vagueness of terms and definitions or measurement error (Akçakaya et al., 2000; IUCN, 2012) and can be managed by using parameter estimates from expert knowledge and data to produce a range of plausible categories (Mace et al., 2008; IUCN, 2012; Collen et al., 2016; IUCN, 2017). The level of uncertainty within the data was expressed using the terms

observed, estimated, inferred or suspected, following guidelines defined by the Red List (IUCN Standards and Petitions Subcommittee, 2017).

APPENDIX C

LIST OF ALL SPECIES WITH KNOWN THREATS USED IN CAP ANALYSIS

List of all 144 clupeiform species with known threats alphabetical by family and then by species name. Threats are coded with 1 if impacted by the threat and 0 if not impacted. The primary habitat system (SYS) is listed; F = Freshwater, M = Marine, and E = Euryhaline, which include estuarine species as well as anadromous species. The global IUCN Red List categories (RL CAT) are also listed; EC = elevated conservation concern and include all threatened (Critically Endangered, Endangered and Vulnerable) and Near Threatened species, LC = Least Concern, DD = Data Deficient.

FAMILY	SPECIES NAME	SYS	RL CAT	EXPLOITATION	CLIMATE CHANGE	ENERGY PRODUCTION & MINING	HUMAN DISTURBANCE	INVASIVE SPECIES	NATURAL SYSTEM MODIFICATIONS	POLLUTION	DEVELOPMENT	SERVICE CORRIDORS
Clupeidae	<i>Alosa aestivalis</i>	E	EC	0	0	0	0	1	1	1	0	0
Clupeidae	<i>Alosa alabamae</i>	E	EC	1	1	0	1	0	1	1	0	1
Clupeidae	<i>Alosa alosa</i>	E	LC	1	0	1	0	0	1	1	0	0
Clupeidae	<i>Alosa braschnikowi</i>	E	DD	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Alosa fallax</i>	E	LC	1	0	0	0	0	1	1	0	0
Clupeidae	<i>Alosa immaculata</i>	E	EC	1	0	0	0	0	1	0	0	0
Clupeidae	<i>Alosa kessleri</i>	E	LC	1	0	0	0	0	1	0	0	0
Clupeidae	<i>Alosa killarnensis</i>	F	EC	0	0	0	0	1	0	1	0	0
Clupeidae	<i>Alosa macedonica</i>	F	EC	1	1	0	0	0	0	1	0	0
Clupeidae	<i>Alosa maeotica</i>	E	LC	0	0	0	0	0	1	0	0	0
Clupeidae	<i>Alosa pseudoharengus</i>	E	LC	0	0	0	0	0	1	0	0	0

FAMILY	SPECIES NAME	SYS	RL CAT	EXPLOITATION	CLIMATE CHANGE	ENERGY PRODUCTION & MINING	HUMAN DISTURBANCE	INVASIVE SPECIES	NATURAL SYSTEM MODIFICATIONS	POLLUTION	DEVELOPMENT	SERVICE CORRIDORS
Clupeidae	<i>Alosa sapidissima</i>	E	LC	1	0	0	0	0	1	1	0	0
Clupeidae	<i>Alosa saposchnikowii</i>	E	DD	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Alosa sphaerocephala</i>	E	LC	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Alosa tanaica</i>	E	LC	0	1	0	0	0	1	1	0	0
Clupeidae	<i>Alosa vistonica</i>	F	EC	1	1	0	0	0	0	1	0	0
Clupeidae	<i>Alosa volgensis</i>	E	EC	1	0	0	0	0	1	0	0	0
Clupeidae	<i>Anodontostoma chacunda</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Brevoortia gunteri</i>	M	LC	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Brevoortia patronus</i>	E	LC	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Brevoortia tyrannus</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Chirocentrus dorab</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Clupea harengus</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Clupea pallasii</i>	M	DD	1	1	0	0	1	0	0	0	0
Clupeidae	<i>Clupeichthys bleekeri</i>	F	EC	0	0	0	0	0	1	1	0	0
Clupeidae	<i>Clupeoides papuensis</i>	F	DD	0	0	1	0	0	0	1	0	0
Clupeidae	<i>Clupeoides venulosus</i>	F	EC	0	0	1	0	0	0	1	0	0
Clupeidae	<i>Clupeonella abrau</i>	F	EC	0	0	0	0	1	1	0	0	0
Clupeidae	<i>Clupeonella engrauliformis</i>	M	EC	1	1	0	0	1	0	0	0	0
Clupeidae	<i>Clupeonella grimmi</i>	M	EC	1	1	0	0	1	0	0	0	0
Clupeidae	<i>Clupeonella muhlisi</i>	F	EC	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Corica laciniata</i>	F	DD	0	0	0	0	0	1	0	0	0
Clupeidae	<i>Dayella malabarica</i>	E	LC	0	0	0	0	1	0	1	0	0
Clupeidae	<i>Dorosoma cepedianum</i>	E	LC	1	0	0	0	0	1	0	0	0
Clupeidae	<i>Dorosoma chavesi</i>	F	EC	0	0	0	0	0	1	0	0	0

FAMILY	SPECIES NAME	SYS	RL CAT	EXPLOITATION	CLIMATE CHANGE	ENERGY PRODUCTION & MINING	HUMAN DISTURBANCE	INVASIVE SPECIES	NATURAL SYSTEM MODIFICATIONS	POLLUTION	DEVELOPMENT	SERVICE CORRIDORS
Clupeidae	<i>Dorosoma petenense</i>	E	LC	0	1	0	0	1	0	0	0	0
Clupeidae	<i>Ethmalosa fimbriata</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Ethmidium maculatum</i>	M	DD	1	1	0	0	0	0	0	0	0
Clupeidae	<i>Etrumeus acuminatus</i>	M	LC	1	1	0	0	0	0	0	0	0
Clupeidae	<i>Etrumeus whiteheadi</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Gilchristella aestuaria</i>	E	LC	0	0	0	0	0	1	0	0	0
Clupeidae	<i>Gonialosa whiteheadi</i>	E	DD	0	0	0	0	0	1	0	0	0
Clupeidae	<i>Gudusia chapra</i>	F	LC	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Harengula clupeola</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Harengula humeralis</i>	E	LC	0	0	0	0	0	0	1	0	0
Clupeidae	<i>Harengula jaguana</i>	M	LC	0	0	0	0	0	0	1	0	0
Clupeidae	<i>Jenkinsia lamprotaenia</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Jenkinsia majua</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Konosirus punctatus</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Laeviscutella dekimpei</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Lile gracilis</i>	E	LC	0	1	0	0	0	0	0	0	0
Clupeidae	<i>Lile piquitinga</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Limnothrissa miodon</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Microthrissa minuta</i>	F	EC	1	0	1	0	0	1	0	0	0
Clupeidae	<i>Microthrissa royauxi</i>	F	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Minyclupeoides dentibranchialis</i>	E	LC	0	0	0	0	0	1	0	0	0
Clupeidae	<i>Nannothrissa parva</i>	F	LC	1	0	0	0	0	0	0	0	0

FAMILY	SPECIES NAME	SYS	RL CAT	EXPLOITATION	CLIMATE CHANGE	ENERGY PRODUCTION & MINING	HUMAN DISTURBANCE	INVASIVE SPECIES	NATURAL SYSTEM MODIFICATIONS	POLLUTION	DEVELOPMENT	SERVICE CORRIDORS
Clupeidae	<i>Nannothrissa stewarti</i>	F	EC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Nematalosa come</i>	M	LC	1	0	0	0	0	1	1	1	0
Clupeidae	<i>Nematalosa erebi</i>	F	LC	1	0	0	0	0	1	0	0	0
Clupeidae	<i>Nematalosa japonica</i>	M	DD	1	0	0	0	0	1	0	1	0
Clupeidae	<i>Nematalosa nasus</i>	E	LC	1	0	0	0	0	1	0	0	0
Clupeidae	<i>Odaxothrissa ansorgii</i>	F	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Odaxothrissa mento</i>	F	LC	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Opisthonema berlangai</i>	M	EC	0	1	0	0	0	0	0	0	0
Clupeidae	<i>Opisthonema libertate</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Opisthonema medirastre</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Opisthonema oglinum</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Opisthopecterus effulgens</i>	E	DD	0	1	0	0	0	0	0	0	0
Clupeidae	<i>Pellonula leonensis</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Pellonula vorax</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Poecilothrissa centralis</i>	F	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Poecilothrissa moeruensis</i>	F	EC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Potamalosia richmondia</i>	E	EC	0	0	0	0	0	1	1	0	0
Clupeidae	<i>Potamothrissa obtusirostris</i>	F	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Sardina pilchardus</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Sardinella aurita</i>	M	LC	1	0	0	0	0	0	0	0	0

FAMILY	SPECIES NAME	SYS	RL CAT	EXPLOITATION	CLIMATE CHANGE	ENERGY PRODUCTION & MINING	HUMAN DISTURBANCE	INVASIVE SPECIES	NATURAL SYSTEM MODIFICATIONS	POLLUTION	DEVELOPMENT	SERVICE CORRIDORS
Clupeidae	<i>Sardinella brasiliensis</i>	E	DD	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Sardinella lemuru</i>	M	EC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Sardinella longiceps</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Sardinella maderensis</i>	M	EC	1	1	0	0	0	0	1	0	0
Clupeidae	<i>Sardinella rouxi</i>	M	DD	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Sardinella tawilis</i>	F	EC	1	0	0	0	1	1	1	0	0
Clupeidae	<i>Sardinella zunasi</i>	M	LC	1	0	0	0	1	0	0	0	0
Clupeidae	<i>Sardinops sagax</i>	M	LC	1	1	0	0	0	0	0	0	0
Clupeidae	<i>Sauvagella madagascariensis</i>	E	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Sauvagella robusta</i>	F	EC	1	0	0	0	1	1	0	0	0
Clupeidae	<i>Setipinna phasa</i>	E	LC	0	0	0	0	0	0	1	0	0
Clupeidae	<i>Setipinna tenuifilis</i>	E	DD	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Sierrathrissa leonensis</i>	F	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Spratelloides delicatulus</i>	M	LC	1	0	0	0	0	0	0	0	0
Clupeidae	<i>Spratellomorpha bianalis</i>	E	DD	1	0	0	0	1	0	0	0	0
Clupeidae	<i>Sprattus sprattus</i>	E	LC	1	0	0	0	0	1	1	0	0
Clupeidae	<i>Stolothrissa tanganicae</i>	F	LC	1	0	0	0	0	0	1	0	0
Clupeidae	<i>Strangomera bentincki</i>	M	LC	1	1	0	0	0	0	0	0	0
Clupeidae	<i>Tenuالosa ilisha</i>	E	LC	1	0	0	0	0	1	1	0	0
Clupeidae	<i>Tenuالosa macrura</i>	E	EC	1	0	0	0	0	0	1	1	0
Clupeidae	<i>Tenuالosa reevesii</i>	E	DD	1	0	0	0	0	1	1	0	0
Clupeidae	<i>Tenuالosa thibaudeaui</i>	F	EC	1	0	0	0	0	1	0	0	0
Clupeidae	<i>Tenuالosa toli</i>	E	EC	1	0	0	0	0	0	1	0	0

FAMILY	SPECIES NAME	SYS	RL CAT	EXPLOITATION	CLIMATE CHANGE	ENERGY PRODUCTION & MINING	HUMAN DISTURBANCE	INVASIVE SPECIES	NATURAL SYSTEM MODIFICATIONS	POLLUTION	DEVELOPMENT	SERVICE CORRIDORS
Clupeidae	<i>Thrattidion noctivagus</i>	F	DD	0	0	0	0	0	1	0	0	0
Denticipitidae	<i>Denticeps clupeoides</i>	F	EC	0	0	1	0	0	0	1	1	0
Engraulidae	<i>Anchoa analis</i>	E	DD	0	0	0	0	0	0	1	0	0
Engraulidae	<i>Anchoa belizensis</i>	F	LC	1	0	0	0	1	0	0	0	0
Engraulidae	<i>Anchoa chamensis</i>	M	DD	0	1	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa choerostoma</i>	M	EC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa delicatissima</i>	E	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa eigenmannia</i>	M	LC	1	1	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa helleri</i>	M	LC	0	1	0	0	0	1	0	0	0
Engraulidae	<i>Anchoa mundeoloides</i>	E	LC	0	1	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa panamensis</i>	E	LC	1	1	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa scofieldi</i>	E	LC	0	1	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa spinifer</i>	E	LC	0	0	0	0	0	0	1	0	0
Engraulidae	<i>Anchoa starksi</i>	E	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Anchoa tricolor</i>	E	LC	1	0	0	0	0	1	1	0	0
Engraulidae	<i>Anchovia surinamensis</i>	E	LC	0	0	0	0	0	1	0	0	0
Engraulidae	<i>Anchoviella lepidentostole</i>	E	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Cetengraulis mysticetus</i>	M	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Coilia grayii</i>	E	LC	1	0	0	0	0	0	1	0	0
Engraulidae	<i>Coilia lindmani</i>	E	LC	1	0	0	0	0	0	1	0	0
Engraulidae	<i>Coilia mystus</i>	E	EC	1	0	0	0	0	1	1	0	0
Engraulidae	<i>Coilia nasus</i>	E	EC	1	0	0	0	0	1	1	0	0
Engraulidae	<i>Coilia neglecta</i>	E	LC	1	0	0	0	0	0	0	0	0

FAMILY	SPECIES NAME	SYS	RL CAT	EXPLOITATION	CLIMATE CHANGE	ENERGY PRODUCTION & MINING	HUMAN DISTURBANCE	INVASIVE SPECIES	NATURAL SYSTEM MODIFICATIONS	POLLUTION	DEVELOPMENT	SERVICE CORRIDORS
Engraulidae	<i>Coilia ramcarati</i>	E	DD	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Encrasicholina punctifer</i>	M	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Engraulis anchoita</i>	M	EC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Engraulis encrasicolus</i>	E	LC	1	0	0	0	1	0	0	0	0
Engraulidae	<i>Engraulis japonicus</i>	M	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Engraulis mordax</i>	M	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Engraulis ringens</i>	M	LC	1	1	0	0	0	0	0	0	0
Engraulidae	<i>Lycengraulis grossidens</i>	E	LC	0	0	0	0	0	0	1	0	0
Engraulidae	<i>Pterengraulis atherinoides</i>	E	LC	0	0	0	0	0	1	0	0	0
Engraulidae	<i>Stolephorus commersonnii</i>	M	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Stolephorus ronquilloi</i>	E	DD	1	0	0	0	0	1	1	0	0
Engraulidae	<i>Thryssa mystax</i>	E	LC	1	0	0	0	0	0	0	0	0
Engraulidae	<i>Thryssa rastrosa</i>	F	EC	0	0	1	0	1	0	0	0	0
Engraulidae	<i>Thryssa scratchleyi</i>	E	DD	1	0	1	0	0	0	1	0	0
Engraulidae	<i>Thryssa vitrirostris</i>	E	LC	1	0	0	0	0	0	0	0	0
Pristigasteridae	<i>Ilisha africana</i>	E	LC	1	0	0	0	0	0	0	0	0
Pristigasteridae	<i>Ilisha elongata</i>	E	LC	1	0	0	0	0	1	0	0	0
Pristigasteridae	<i>Ilisha novacula</i>	F	LC	0	0	0	0	0	1	0	0	0
Pristigasteridae	<i>Neoopisthopterus cubanus</i>	E	EC	0	0	0	0	0	0	1	1	0
Pristigasteridae	<i>Pristigaster whiteheadi</i>	F	LC	0	0	0	0	0	1	0	0	0
Pristigasteridae	<i>Raconda russeliana</i>	E	LC	1	0	0	0	0	0	0	0	0

APPENDIX D

LIST OF ALL SPECIES OF ELEVATED CONSERVATION CONCERN USED IN CAP ANALYSIS

List of the 33 clupeiform species assessed as threatened (Critically Endangered, Endangered and Vulnerable) or Near Threatened alphabetical by family and then by species name. The primary habitat system is coded with 0's and 1's for marine and freshwater species; euryhaline species are characterized by a 1 in both columns. Habitat refers to whether a species is a habitat generalist, coded with a 1 or a specialist, coded with a 0. The maximum known standard length (MaxSL) in centimeters, number of countries a species is known to be distributed within (COO) as a proxy for geographic distribution and the number of other clupeiforms within an individual species range as a proxy for relative diversity are listed. The number impacting a species and the global categories are also listed: CR = Critically Endangered, EN = Endangered, VU = Vulnerable, NT = Near Threatened, LC = Least Concern, DD = Data Deficient.

FAMILY	SPECIES NAME	GLOBAL CATEGORY	MARINE	FRESHWATER	HABITAT	MAXSL	COO	DIVERSITY	# OF THREATS
Clupeidae	<i>Alosa aestivalis</i>	VU	1	1	1	35.0	2	27	3
Clupeidae	<i>Alosa alabamae</i>	NT	1	1	1	51.0	1	20	6
Clupeidae	<i>Alosa immaculata</i>	VU	1	1	1	37.0	8	4	2
Clupeidae	<i>Alosa killarnensis</i>	CR	0	1	0	20.0	1	0	2
Clupeidae	<i>Alosa macedonica</i>	VU	0	1	0	35.0	1	0	3
Clupeidae	<i>Alosa vistonica</i>	CR	0	1	0	17.0	1	0	3
Clupeidae	<i>Alosa volgensis</i>	EN	1	1	1	35.0	5	13	2
Clupeidae	<i>Clupeichthys bleekeri</i>	VU	0	1	0	6.0	1	7	2
Clupeidae	<i>Clupeoides venulosus</i>	VU	0	1	0	9.0	2	6	2
Clupeidae	<i>Clupeonella abrau</i>	CR	0	1	0	9.5	1	0	2
Clupeidae	<i>Clupeonella engrauliformis</i>	EN	1	0	0	15.5	3	36	3

FAMILY	SPECIES NAME	GLOBAL CATEGORY	MARINE	FRESHWATER	HABITAT	MAXSL	COO	DIVERSITY	# OF THREATS
Clupeidae	<i>Clupeonella grimmi</i>	EN	1	0	1	14.5	3	10	3
Clupeidae	<i>Clupeonella muhlisi</i>	EN	0	1	0	6.0	1	0	1
Clupeidae	<i>Dorosoma chavesi</i>	NT	0	1	1	18.0	2	0	1
Clupeidae	<i>Microthrissa minuta</i>	VU	0	1	1	3.5	1	12	3
Clupeidae	<i>Nannothrissa stewarti</i>	EN	0	1	0	2.3	1	1	1
Clupeidae	<i>Opisthonema berlangai</i>	VU	1	0	0	26.0	1	8	1
Clupeidae	<i>Poecilothrissa moeruensis</i>	VU	0	1	0	3.5	2	1	1
Clupeidae	<i>Potamalosa richmondia</i>	NT	1	1	1	32.0	1	13	2
Clupeidae	<i>Sardinella lemuru</i>	NT	1	0	1	23.0	10	102	1
Clupeidae	<i>Sardinella maderensis</i>	VU	1	0	1	30.0	44	32	3
Clupeidae	<i>Sardinella tawilis</i>	EN	0	1	0	13.6	1	0	4
Clupeidae	<i>Sauvagella robusta</i>	EN	0	1	1	4.7	1	0	3
Clupeidae	<i>Tenualosa macrura</i>	NT	1	1	1	52.0	2	59	3
Clupeidae	<i>Tenualosa thibaudeaui</i>	VU	0	1	1	26.0	4	8	2
Clupeidae	<i>Tenualosa toli</i>	VU	1	1	0	50.0	1	48	2
Denticipitidae	<i>Denticeps clupeoides</i>	VU	0	1	0	13.0	3	4	3
Engraulidae	<i>Anchoa choerostoma</i>	EN	1	0	1	7.5	1	4	1
Engraulidae	<i>Coilia mystus</i>	EN	1	1	1	20.0	4	55	3
Engraulidae	<i>Coilia nasus</i>	EN	1	1	1	41.0	4	51	3
Engraulidae	<i>Engraulis anchoita</i>	NT	1	0	0	17.0	3	27	1
Engraulidae	<i>Thryssa rastrosa</i>	EN	0	1	0	11.6	1	5	2
Pristigasteridae	<i>Neopisthopterus cubanus</i>	VU	1	1	1	9.0	1	20	2

VITA

Tiffany L. Birge

Department of Biological Sciences

Old Dominion University

Norfolk, Virginia 23529

EDUCATION

Bachelor of Biological Sciences (Marine Biology), Old Dominion University, Norfolk, Virginia. 2016.

Associate of Science, Tidewater Community College, Virginia Beach, Virginia. 2014.

PROFESSIONAL EXPERIENCE

Teaching Assistant, August 2017 – Present, Ichthyology Lab, Old Dominion University, Norfolk, Virginia

Research Assistant, August 2016 – Present, International Union for Conservation of Nature's Global Species Programme Marine Biodiversity Unit, Old Dominion University, Norfolk, Virginia

Phytoplankton Laboratory and Field Technician Intern, May 2016 – August 2016, Old Dominion University, Norfolk, Virginia

THESIS-RELATED PRESENTATIONS

March 2019. Oral presentation: The fast life of forage fishes: conservation status of the world's clupeiforms (Order Clupeiformes). Biology Graduate Student Organization Spring Symposium, Old Dominion University

March 2017. Poster presentation: The global extinction Risk of forage fishes of the order Clupeiformes. 31st Annual Meeting of the American Fisheries Society – Tidewater Chapter Meeting, Virginia Beach, Virginia