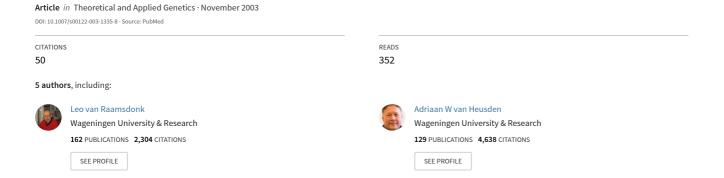
Biodiversity assessment based on cpDNA and crossability analysis in selected species of Allium subgenus Rhizirideum



L. W. D. van Raamsdonk · W. Ensink · A. W. van Heusden · M. Vrielink-van Ginkel · C. Kik

Biodiversity assessment based on cpDNA and crossability analysis in selected species of *Allium* subgenus *Rhizirideum*

Received: 20 August 2002 / Accepted: 19 April 2003 / Published online: 25 June 2003 © Springer-Verlag 2003

Abstract The chloroplast DNA diversity of 33 accessions belonging to 16 species of five sections in Allium subgenus Rhizirideum was studied by analysing the sequence of three fragments: the trnL-F intergenic spacer, the rps 16 intron and rbcL (rubisco large subunit). The three sections Cepa, Schoenoprasum and Rhizirideum, representing the majority of the included species, each possess a separate clade after phylogenetic analysis. Exceptions to this general rule are the placement of Allium pskemense (section Cepa) connected to Allium senescens (section Rhizirideum) and Alium roylei, taking an intermediate position between sections Cepa and Schoenoprasum. Both species were located in their own section after nuclear DNA analysis. A range of crossing experiments has been carried out. The different position of A. roylei when comparing cpDNA and nDNA diversity was not confirmed with the production of hybrid seeds after crossing A. roylei with species other than those of section Cepa. The different position of A. pskemense in the cpDNA and the nDNA tree can not be compared to its crossability, since only a few crossing experiments are reported for this species. The hypothesis that a shorter distance between two species in a cpDNA tree compared to their distance in a nDNA tree will indicate interfertility at a certain level, is neither confirmed nor rejected by the currently available results.

Keywords *Allium* subgenus *Rhizirideum* · *Allium cepa* · *Allium roylei* · Sequence analysis · trnL · rps16 · rbcL · Onion · Phylogeny reconstruction · cpDNA

Communicated by R. Hagemann

L. W. D. van Raamsdonk (💌) · W. Ensink · A. W. van Heusden · M. Vrielink-van Ginkel · C. Kik Plant Research International, Business Unit Genetics and Breeding, P.O. Box 16, 6700 AA Wageningen, The Netherlands

e-mail: Leo.vanraamsdonk@wur.nl

L. W. D. van Raamsdonk RIKILT Institute for Food Safety, P.O. Box 230, 6700 AE Wageningen, The Netherlands

Introduction

The genus Allium comprises approximately 700 species, arranged in four large subgenera with 100-300 species each (Allium, Rhizirideum, Amerallium and Melanocrommyum) and two much smaller subgenera (Bromatorrhiza and Caloscordon; Hanelt et al. 1992). The main cultivated species are found in the subgenera *Allium* (garlic, leek) and *Rhizirideum* (onion, shallot, bunching onion, chives, chinese chives), whereas a range of ornamentals originates from subgenus Melanocrommyum. Most research has been focused on the improvement of these cultivated species. Especially onion (Allium cepa), as the most important cultigen in the genus, received much attention with respect to the relationship with wild species (El-Gadi and Elkington 1977; Hanelt 1990; van Raamsdonk and de Vries 1992a) and to crossability (Saini and Davis 1967; McCollum 1971, 1974; Gonzalez and Ford-Lloyd 1987; van der Meer and de Vries 1990; van Raamsdonk et al. 1992; Keller et al. 1996). Diversity studies based on DNA variation have been carried out to study the position of A. cepa (Havey 1992; Wilkie et al. 1993; Bradeen and Havey 1995; Linne von Berg et al. 1996; Dubouzet et al. 1997; Mes et al. 1997, 1999; van Raamsdonk et al. 1997, 2000; Fritsch et al. 2001; Lilly and Havey 2001). Although several studies of selected sets of species of Allium subgenus Rhizirideum were based on chloroplast (cp) DNA, detailed sequence analyses of specified cpDNA fragments, or a comparison of cpDNA and nDNA based phylogenies of exactly the same set of species were rarely published (Friesen et al. 1999, 2000; Mes et al. 1999; Fritsch et al. 2001).

Chloroplast DNA is in general maternally inherited, as is the case in *Allium* (Havey 1995), and its lack of recombination makes it suitable for the study of hybridisation and introgression (Rieseberg and Soltis 1991; Rieseberg 1995). The importance of cpDNA for studying the aforementioned phenomena has recently been emphasized by Hollingworth et al. (1999). In *Allium* subgenus *Rhizirideum* the inheritance of nuclear (n) DNA variation in hybrids has been studied by means of

the Hybrid Distance approach (van Raamsdonk et al. 2000). A cpDNA study of the same set of species will give much more insight about events of hybrid speciation, will yield information on the crossability relationship to be expected and provides the opportunity to compare phylogenies based on biparentally and maternally inherited DNA.

In the present study, sequences of four cpDNA fragments will be used for studying the diversity in cpDNA subjected to cladistic analysis. The study includes equal amounts of accessions of the sections Cepa, Rhizirideum and Schoenoprasum in order to reach a balanced share in the resulting phylogeny. The use of more than one accession for most species of sections Rhizirideum and Schoenoprasum allows us to compare the infraspecific variation with the interspecific variation. Some species of section *Oreiprason* (three species), of section Petroprason (one) and of section Reticulatobulbosa (one) were added for comparison. Allium tuberosum will be used as an outgroup. The aim of the present study is: (1) to analyse the cpDNA diversity and unravel the maternally based relationships in Allium subgenus Rhizirideum, (2) to study the occurrence of introgression events and of hybrid speciation, and (3) to analyse whether a shorter distance between two species in a cpDNA tree compared to their distance in a nDNA tree will indicate interfertility.

Materials and methods

Plant material

The species and accessions included in the analysis are listed in the Appendix.

Vouchers are deposited in either the herbaria of Wageningen University (WAG), the Royal Botanic Gardens in Kew (K), the Institute für Pflanzengenetik und Kulturpflanzenforschung in Gatersleben (GAT), and the Plant Research international (formerly the Institute for Horticultural Plant Breeding, Wageningen, WAHO). All accessions labeled as "BG" originate from an originally wild population.

DNA-isolation

Genomic DNA was isolated from leaf material with a midi DNA preparation procedure (Beek et al. 1992) with some minor modifications: after hooking the DNA out of the isopropanol mixture, the DNA was washed overnight in 70% EtOH and 100 mM NH₄Ac, and resuspended in 200 μl or more-sterile TE (10 mM Tris-HCl pH = 8.0 and 1 mM EDTA). DNA concentrations were measured using a fluorometer. DNA could also be isolated with the mini prep DNA-isolation method: approximately 0.25 g of fresh leaf material was collected in Eppendorf tubes, frozen and ground in liquid nitrogen, and stored at -50 °C; 750 µl of DNAisolation buffer (IB) with Na₂S₂O₅ (3.8 g/l) was added to the leaf material. This mixture was incubated for 60 min at 65 °C with occasional inverting of the tubes [IB = lysis buffer:extraction buffer:sarkosyl (5% w/v) = 2.5:2.5:1; lysis buffer = 0.2 M Tris-HCl pH = 7.5, 0.05 M EDTA, 2 M NaCl, 2% w/v CTAB; extraction buffer = 0.35 M Sorbitol, 0.1 M Tris HCl pH = 7.5, 5 mM EDTA]. The DNA was further purified by adding 750 μ l of chloroform/ isoamylalcohol (24:1), inverting the tubes (10-20 times) and centrifuging for 5 min at 14,000 rpm. After transfer of the

Table 1 Primers used for the amplification of three cpDNA fragments

rps16-R2:	5'-TCGRGATCGAACATCAATTGCAAC
rps16-F:	5'-GTGGTAGAAAGCAACGTGCGACTT
rps16-1A:	5'-GGGGGGGCGAATTTAGGG
rbcL-2:	5'-GATATCTTGGCAAGCATTCCGAG
rbcL-4:	5'-CGATTAGCTRCTGCACCAGGYGC
trnL (UAA) 5' exon:	5'-CGAAATCGGTAGACGCTACG
trnL (UAA) 3' exon:	5'-GGGGATAGAGGACTTGAAC
trnL-F-e:	5'-GGTTCAAGTCCCTCTATCCC
trnL-F-f:	5'-ATTTGAACTGGTGACACGAG

supernatant to a new tube the DNA was precipitated by the addition of $400-\mu l$ isopropanol (-20 °C). The DNA could either be hooked out or it had to be pelleted for 5 min at 14,000 rpm. The DNA samples were washed once with 70% ethanol and re-suspended in $100 \ \mu l$ of TE.

cpDNA sequencing

Four sequences of the cpDNA genome were amplified by specific primers: the trnL-F intergenic spacer and the trnL exon (Taberlet et al. 1991), the rps 16 intron (Oxelman et al. 1997) and the rbcL (rubisco large subunit). Four rbcL primers were designed from three *Allium* sequences available in the EMBL databases. The best results were obtained with primers 2 and 4, amplifying a sequence of approximately 1,100 nucleotides. Primers are listed in Table 1.

The PCR reaction mixture (total volume 25 μ l) contained 25 ng of template DNA, 25 ng of the primers (Table 1), 10 mM of each dNTP, 0.1 μl (0.5 units) of superTaq polymerase, 50 mM of KCl, 10 mM of Tris-HCl, 1.5 mM of MgCl₂ and 0.01% gelatine (pH = 8.3). The mixture was covered by two drops of mineral oil. PCR was performed with a thermal cycler (Perkin Elmer) as follows: 35 cycles at 94 °C for 1 min (denaturation), annealing for 2 min, from 35 °C to 72 °C with a ramp of 1 °C/5 s, 72 °C for 2 min (extension); and one final cycle at 72 °C for 10 min. Optimal annealing temperatures were 60 °C for the trnL-F intergenic spacer, 55 °C for the trnL exon, 50 °C for the rps 16 intron and 60 °C for the rbcL sequence. Electrophoresis was performed through 1.5% 1× TBE agarose gels, for 6 to 8 h at 4 V/cm, stained with Ethidium bromide. Small fragments of the gel containing the DNA fragment were cut out and the DNA was extracted from the gel using the Qiaex protocol (QIAGEN corporation). The DNA amount was measured and 150-400 ng of DNA, depending on the size of the fragment, was used for the sequencing reaction. The fragment was amplified with a sequence mix, containing fluorescence-labelled nucleotides, and one of the primers in a PCR reaction of 25 cycles. The amplified fragment was sequenced in an ABI sequencer.

Scoring of data

All sequences were aligned manually. The resulting raw-data matrix consisted of 369 nucleotide positions for the trnL-F intergenic spacer, 339 for the trnL exon, 861 for the rps 16 intron (between primers R2 and 1A) and 1,128 for the rbcL (between primers 2 and 4). All differences were scored as 0, 1, 2 etc. depending on the number of states per mutation site. A mutation could either consist of a single nucleotide (single-site mutation) or a deletion or duplication of a stretch of nucleotides (indel). The direction of all mutations was estimated on the basis of the situation found in the outgroup (*A. tuberosum*).

Data analysis

All analyses are based on a matrix with digits (0, 1, 2, etc.) indicating independent states of each character per accession. All

Table 2 Sequences for 33 accessions of *Allium* subgenus *Rhizirideum* of the trnL-F intergenic spacer. Mutations are indicated by capitals (nucleotide changes) or by lower-case letters (duplications or deletions) in the bottom rows of the table. Letters in the

sequences are given according to the IUB codes: A: adenosine, C: cytidine, G: guanosine, T: thymidine, R: A or G, Y: C or T, K: G or T, M: A or C, and n: any base

	1 10	20 30	40	50	60	70	80	90	100
Jumbo	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATAATTC	rcc t tttt	TTTTATAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	. Т
Stuttgarter	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATAATTC	rcc t tttt	TTTTATAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	
Polar	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATAATTC	CC TTTTT	TTTTATAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	т
95081 vav	CAGTAAAAAGCCCGTTT	FACTTCTTTACTATAATTC	CC T TTTT	TTTTATAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	. Т
206 osch	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATAATTC	CC TTTTT	TTTTATAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	. Т
103 gal	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATAATTC	CC TTTTT	TTTTATAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	. Т
207 pske	CAKTAnAAARSCCnYTT	PACTTCTTTACTATAATTCT	CC TT TTTT	TTTTATAATT A	TAAGTGGTTCA	AARAAAA	TGG'	TTCAAARAAAA	. T
102 fist	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATAATTC	CC TTTTT	TTTTATAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	. T
201 alta	חחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחח	וחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחחח	nnn nnnnn.	nnnnnTAAGT	GGTTCA	AAGAAAA		CAAAGAAAA	. Т
104 royl	CAGTAAAAAGCCCGTTT	TACTTCTTTACTATA A TTC	rcc tt tttt	TTTTATAAKT	GGTTCA	AARAAAA		CAAARAAAA	. Т
All 911 schoe	CAGTAAAAAGCCCGTTT	TACTTCTTTAMTATAATTC	rcc tt tttt	TTTATAATT	GGTTCA	AARAAAA		CAAATAAAA	-
93007 schoe		nAnTTCTTTACTATA A TTC				AAAAAA		CAAATAAAA	
Tax 1605 scho		TACCTCTTTAA <i>n</i> ATA A TTCT				AARAAAA		CAAATAAAA	
Tax 42 altync		TATTTTTTAnTATA A TTC			GGTTCA	AAGAAAA		CAAATAAAA	
93008 altync		FA <i>nn</i> TCTTTACTATA A TTC				AAG <i>n</i> AAA		CAAATAAAA	
Tax 536 karel	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATA A TTC	CC TT TTTT	TTTTATAATT	GGTTCA	AAAAAA		CAAATAAAA	. Т
All 1122 sen		TACTTCTTTACTATA A TTC				AARAAAA		TTCAAARAAAA	
89010 sen		PACTTCTTTACTATAATTC				AAAAAA		TTCAAAAAAAA	
93005 sen		PACCTCTTTACTATAATTC				AAGAAAA		TTCAAAGAAAA	
89008 nut		PACTTCTTTACTATAATTC				AAGAAAA		TTCAAAnAAAA	
89009 nut		PACTTCTTTACTATAATTC				AAAAAA		TTCAAAGAAAA	
89011 nut		FACTTCTWTAC <i>n</i> ACA A TTC				AAGAAAA		TTCAAAGAAAA	
93012 nut		nAnttctttactata a ttc		TTTTATAAGn		AAG <i>n</i> AAA		TTCAAAGAAAA	
Tax 256 ang	CAGTAAAAAGCCCGTTT	FACTTCTTTACTATAATTC	rcc t tttt	TTTTATAAGT	GGTTCA	AAGAAAA	TGG'	TTCAAAGAAAA	. Т
m 0005 1'	G2.GM2.2.2.2.GGGGGMMM				aammaa				т
Tax 2335 lin	CAGTAAAAAGCCCGTTT	PACTTCTTTACTATAATTC	ree Trrr	TTTTATAAGT	GGTTCA	AAAAAAA	TGG	TTCAAAnAAAA	. 1
All 1123 glob	CACTAAAAACCCCCTTTT	TACTTCTTTACTATAATTC	rcc m orono		CCTTC+		. m x = cmcc	TTCAAAnAAAA	CCTnnAACAT
Tax 632 saxa		TACTTCTTTACTATA A TTC.						TTCAAAGAAAA TTCAAAGAAAA	
Tax 750 hymen		TACTTCTTTACTATA A TTC TACTTCTnTAC G ATA A TTC		TTTTATAAGT		AAGAAAA CA. AAAGAAAA	ITANGIGG	CAAAGAAAA	
rax /50 nymen	CAGTAAAAAGCCCGTTT	TACTICIMIACGATAATIC	CACIIII	IIIIAIAAGI	GGIICA	MAGAAAA		CAAAGAAAA	. 1
Tax 94 oblig	CACTAAAAACCCCCTTTT	TACTTCTnTACTATCATnC	TCC T TTTTT	ጥጥጥል ጥል ልርጥ	CCTTC	. A A C A A A C A Z	TA A GTGG	TTCAAAGAAAA	СУУУСТУУТ
Tax 24 Obitq	CAGIAAAAGCCCGIII	TACTICIMIACIAI CATHE	100 11111	IIIIAIAAOI	GOTTCA	unomun ciu	innoido	11CMMGMIM	CAMMOMMAI
86001 tub	CAGTAAAAAGCCCGTTT	TACTTCTTTACTATA TTC	TCC TCTT	TTTTATAAGn	GGTTCA	AAGAAAA			T
89006 tub		TACTTCTTTACTAnA TTC		TTTTATAAGT	GGTTCA	AAGAAAA			T
89007 tub		FACTTCTTTACTATA TTC		TTTTATAAGT		AAGAAAA			T
Tax 1830 tub	CAGTAAAAAGCCCGTTT	TACTTCTTTACTATA TTC	TCC TCTT	TTTTATAAGT	GGTTCA	AAGAAAA			Т
		A B	С						
		a	bc	d		е	f	g	h

characters were extracted from the original data matrix with nucleotide positions. The phylogenetic reconstruction was carried out using the PAUP software package 3.1 (Swofford 1993). Data were treated as unordered and equally weighted. Consensus trees were based on all most-parsimonious trees of equal length after a heuristic search of trees using the TBR and MULPARS options. Bootstrap analysis has been carried out with 500 replicates.

Hybridisation experiments

Flowers of the female parents were emasculated by hand prior to anthesis. Mature pollen was transferred from anthers of the pollen donor to the stigma of the female parent by means of a paint brush. All pollination was carried out in insect-free glasshouses. Ripe F1 seeds were harvested during summer and sown at late autumn. Seedlings were raised during winter in frost-free glasshouses. All presumed hybrids were checked on their hybrid background by using RAPDs.

Results

cpDNA diversity

Single nucleotide mutations or duplications/deletions were found in three of the four sequences investigated.

The trnL-F intergenic spacer showed 15 single nucleotide mutations, and 12 duplications and deletions of stretches longer than one nucleotide (Table 2). Only mutations that appeared to be unambiguous after checking the original sequence data are indicated and used for phylogenetic analysis. This spacer between the trnL intron and the 3' end of the trnF gene includes 369 nucleotide positions in Allium subgenus Rhizirideum. All species except A. tuberosum share a duplication of nine nucleotides at position 87 in Table 2 (CAAAnAAA). The representatives of sections Cepa and Schoenoprasum except for Allium pskemense share the absence of any further duplication in that region, whereas e.g. the species of section Rhizirideum share a further duplication of five nucleotides (TGGTT) in front of the first-mentioned duplication. It is unlikely that these two duplications arose independently, since independent duplications would imply that the five nucleotides TGGTT were shifted nine positions to the right after the duplication of the nine nucleotides. It is more likely to assume a duplication of 14 nucleotides in section Rhizirideum and at least one other duplication of nine nucleotides in the sections Cepa and Schoenoprasum. In this explanation it is not possible to

Table 2 (continued)

`	,									
	110	120	130	140	150	160	170	180		200
Jumbo	TCAATATCTTT									AGAAAGTCTTC
Stuttgarter	TCAATATCTTT									ARAAAGTCTTC
Polar	TCAATATCTTT									AGAAAGTCTTC
95081 vav	TCAATATCTTT									AGAAAGTCTTC
206 osch	TCAATATCTTT									AGAAAGTCTTC
103 gal	TCAATATCTTT									AGAAAGTCTTC
207 pske	TCAATATCTTT	CTTATTCATT	TTACTATTTC	CACAAATAGA	CCCAAAGTGAA	TTATT				ARAAAGYCTTC
102 fist	TCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACAAATAGA	CCCAAAG C G C A	TTATT				AGAAAGTCTTC
201 alta	TCAATATCTTT	CTTAT <i>n</i> CATT	TTACTCTTTC	CACAAATAGA	CCCAAAG C G C A	ATTATT	CACAGCCCATC	TCATTTTCCTT	Anwttcacar	AGAAAGTCTTC
104 royl	TCAATATCTTT	CTTATnCATT	TTACTCTTTC	ממבמבממים	CCAAAKT T AA	TTATT	CACAGCCCATC	TCATTTTCCTT	'ACATTCACAZ	ARAAAGTCTTC
101 1011	101111111111111111111111111111111111111	011111101111					0110110000111			
All 911 schoe	TCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACAAATAGA	CCCAAAGTGAA	TTATT	CACAGCCCATC	TCATTTTCCTT	'ACATTCACA	AGAAATTCTTC
93007 schoe	TCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACAAATAAA	CCCAAAnnGAA	TTATT	CACAGCCCATC	CTCATTTTCCTT	'ACATTCACA	ARAAA <i>nn</i> CTTC
Tax 1605 scho	TCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACAAATARA	CCCAAAGTGAA	TTATT	CACAGCCCATC	TCATTTTCCTT	'ACATTCACAA	AGAAATTCTTC
Tax 42 altync	nCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACAAATARA	CCCAAAGTGAA	TTATT	CACAGCCCATC	CTCATTTTCCTT	'ACATTCACA	AGAAATTCTTC
93008 altync	TCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACAAATAGA	CCCAAAGnGAA	TTATT	CACAGCCCATC	TCATTTTCCTT	'ACATTCACA	AGAAAG <i>n</i> ATTC
Tax 536 karel	TCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACAAATAAA	CCCAAATTGA	ATTATT	CACAnCCCATA	TCnTTTTCCTT	'ACnTTCnCAF	ARAAATTCT <i>n</i> C
All 1122 sen	TCAATATCTTT									ARAAATTCTTC
89010 sen	TCAATATCTTn									ARAAATTCT <i>n</i> C
93005 sen	TCAATATCTTT									AGAAAGTCTTY
89008 nut	TCAATATCTTT									AGAAAGTCTTC
89009 nut	TCAATATCTTT									AGAAAGTCTTC
89011 nut	TCAATATCTTT	CTTATTCATT	TTACTCTTTC	CACCAATAGA	CCCAAAGTGA	TTAATI				AGAAAGTCTTC
93012 nut	TCAATATCTTT									AGAAAGTCTTC
Tax 256 ang	TCAATATCTTT	CTTATTCATT'	TTACTCTTTC	CACAAATAGA	CCCAAAGTGA	TTATT	CACAGCCCATC	TCATTTTCCTT	'ACATTCACAA	AGAAAGTCTTC
Tax 2335 lin	TCAATATCTTT	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	ጥጥ እ ረጥረጥጥጥረ	77 (77 7 7 7 7 7 7 7 7 7 7	~~~	יחים עיחים	CACACCCCATC	THE ATTITUTE COTT	יאכאיידיכאכאז	AGAAAGTCTTC
1dx 2333 1111	TCAATATCTTT	CITATICATI	TIACICITIO	LACAMAIAGA	CCAAAGIGA	TITALI	CACAGCCCAIC	CICATITICCII	ACATICACA	MOMMOTOTIC
All 1123 glob	TnnATnTCTTT	CTTATTCATA	CTACTCTTnC	CTCAAATnnA	CCCCAAnTGA	ATAAnT nT	TCnYnSCCCATC	CTCATTTTCCTT	nCnTnCACA	AGAAAGTCTTC
Tax 632 saxa	TCAATATCTTT	CTTATTCATT'	TTACTCTTTC	CACAAATAGA	CCCAAAGTGA	A <i>n</i> AATT AT	TCACAGCCCATC	CTCATTTTCCTT	ACATTCACA	AGAAAGTCTTC
Tax 750 hymen	TCAATATCTTT	CTTATTCATT'	TTACTCTTTC	CACAAATAGA	CCCAAAGTGA	TTATT	CACAGYCCATO	TCATTTT A CTT	'ACATTCACAA	AGAAAGTCTTC
Tax 94 obliq	TCAATATCTTT	CTTATTCATT'	TTACTCTTTC	CACAAATAGA	CCCAAAGTGA	AGAATT AT	TCACAGCCCATC	CTCATTTTCCTT	'ACATTCACAA	Anaa r gtcttc
86001 tub	TCAATATCTTT	CTTATTCATT'	TTACTCTTTC	CACAAAT G GA	CCAAAGTGA	TTATT	CACAGCCCATC	CTCATTTTCCTT	'ACATTCACA	AGAAAGTCTTC
89006 tub	TCAATATCTTT									AGAAAGTCTTC
89007 tub	TCAATATCTTT									AGAAAGTCTTC
Tax 1830 tub	TCAATATCTTT									AGAAAGTCTTC
107 1030 CUD	ICAMINICITI	CITALICALI	1111010111	CAAA I GGA	CCAAAGIGA	** TUT I	CACAGCCCATC		TICHT I CHCAP	
				D	EFG			Н		I

prove whether the duplication in section *Cepa* is the same as found in section *Schoenoprasum* or not. There is at least one difference between these sections at the fifth nucleotide of the nine-nucleotide duplication (position 91 in Table 2: G or T, respectively). Duplications at position 75 and at 162–164 show synapomorphisms between species of section *Oreiprason* (*Allium globosum* and *Allium saxatile*) and a species of section *Petroprason* (*Allium obliquum*). A duplication of 25 nucleotides is found in the sequences of *A. cepa* and of *Allium vavilovii* at position 249–273 (Table 2).

The intron of the rps16 gene sequenced with the R2 primer showed good results, but the reverse amplification with the F primer stopped at a region with nine G nucleotides. A new primer (20-mer) was designed covering the G region, which successfully amplified the remaining part of the rps16 intron sequence. This part, 175 nucleotides shorter than the part of the intron enclosed by R2 and F, showed seven single-site mutations and 15 duplications or deletions (sequences not shown). A duplication of 43 nucleotides is found in the sequences of A. cepa and of A. vavilovii, which is supported by the duplication of 25 nucleotides in the sequences of the trnL-F intergenic spacer of the same species. Agarose-gel electrophoresis showed that the intron of the rps16 gene is present in a large and a small copy, whereas Allium

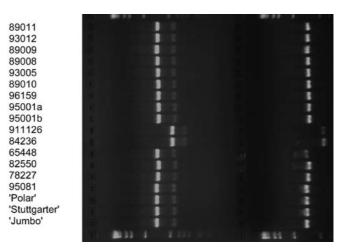


Fig. 1 Results of amplification of the rps16 intron for a range of species. *Left*: amplification at 55 °C, *right*: amplification at 60 °C. Note the shorter fragment in *A. fistulosum* and *A. altaicum*

fistulosum and Allium altaicum show two copies of shorter size (Fig. 1). Sequence analysis showed that the main but smaller copy in these two species is 160 nucleotides shorter than the main copy in the other species. The short copy of Allium schoenoprasum accession 96152 possesses the same deletion of 160 nucleo-

Table 2 (continued)

(1111	210 220	230	240	250	260	270	280	290	300
Jumbo	TTTTTGAAAATCGGA								
Stuttgarter	TTTTTGAAAATCGGA	AAAATTAGGGAA'							
Polar	TTTTTGAAAATCGGA	AAAATTAGGGAA'							
95081 vav	TTTTTGAAAATCGGA	AAAATTAGGGAA'							
206 osch	TTTTTGAAAATCGGA	AAAATTAGGGAA'							TTGTGTCACCG
103 gal	TTTTTGAAAATCGGA	AAAATTAGGGAA'							TTGTGTCACCG
207 pske	TTTTTGAAAATCGAA	AAAATTAGGGAA'							TTGTGTCACCG
102 fist	TTTTTGAAAATCGGA	AAAATTAGGGAA'							TTGTGTCACCG
201 alta	TTTTTGAAAATCGGA	AAAATTAGGGAA'							TTGTGTCACCG
201 4164	TTTTTGAAAATCGGA	AAAA11AOOOAA	IAGCICAIIIIGI			GIAC	DAGCAGAGGC	IIGAAAAICC.	ITGIGICACCG
104 royl	TTTTTGAAAAT CGG A	AAAATTAGGGAA	TAGCTCAKTTGT	Γ		GTAA	AAGCAAAGGC'	TTGAAAATCC:	FTGTGTCACCG
All 911 schoe	TTTTTGAAAAT CGG A	AAAATTAGGGAA'	TAGCTCAKTTGT	י		GTA	AGCAAAGGC'	TTGAAAATCC	TTGTGTCACCG
93007 schoe	TTTTTGAAAAT CGG A	AAAATTAGGGAA'							TTGTnnCACCG
Tax 1605 scho	TTTTTGAAAATCGGA	AAAATTAGGGAA'							TTGTTTCACCG
Tax 42 altync	TTTTTGAAAATCGGA	AAAATTAGGGAA'							TTGTTTCACCG
93008 altync	TTTTTGAAAAT CGG A	AAAATTAGGGAA'							TTGTGTCACCG
Tax 536 karel	TTTTTnAAAnTCGGA	AAAATTAGGGAA'							TTnTTTCnCCG
ran 550 narer	***************************************			•					
All 1122 sen	TTTTT <i>n</i> AAAAT CG AA	AAAATTAGGGAA'	TACCTCAKTTGT	C		GTAA	AAGCAAAGGC'	TTGAAAATCC'	TTGTGTCACCG
89010 sen	TTTTTnAAAAT CG AA	AAAATTAGGGAA'	TACCTCATTT <i>n</i> T	C		TTAA	Ancaaaggc'	TTnAAAATCC	TTnTTTCACCG
93005 sen	TTTTTGAAAAT CG AA	AAAATTAGGGAA'	TAGCTCAGTTGT	r		GTAC	GAGCA <i>n</i> AGGC'	TTGAAAATCC	TTGTGTCACCG
89008 nut	TTTTTGAAAAT CG AA	AAAATTAGGGAA'	TAGCTCAGTTGT	r		GTA	AGCAAAGGC'	TTGAAAATCC	TTGTGTCACCG
89009 nut	TTTTTGAAAAT CG AA	AAAATTAGGGAA'	TAGCTCAGTTGT	C C		GT <i>nI</i>	Ancaaaggc'	TTGAAAATCC	TTGTGTCACCG
89011 nut	TTTTTGAAAATCGAA	AAAATTAGGGAA'	TATCTCAGTTGT	C		GTA	AAGCAGAGGC"	TTGAAAATCC:	TTGTGTCTCCG
93012 nut	TTTTTGAAAAT CG AA	AAAATTAGGGAA'	TAGCTCAGTTGT	r		GTA	agca <i>n</i> aggc'	TTGAAAATCC:	TTGTGTCACCG
Tax 256 ang	TTTTTGAAAAT CG AA	AAAATTAGGGAA'	TAGCTCAGTTGT	Г		GTA	agcaaaggc'	TTGAAAATCC	TTGTGTCACCG
3									
Tax 2335 lin	TTTTTGAAAAT CG AA	AAAATTAGGGAA'	TAGCTCAGTTGT	r		GTAA	AAGCAAAGGC'	TTGAAAATCC'	TTGTGTCACCG
All 1123 glob	TTTTTGAAAAT CG AA	TnTATTAGGAAA	nR nCTCATTTG1	r		GTnr	ากกกกกกกกกก	TCGAA <i>nn</i> TCC'	TCnnnnnnnn
Tax 632 saxa	TTTTTGAAAATCGAA	AAAATTAGGAAA'							TCGTGTCnCCG
Tax 750 hymen	TTTTTGAAAATCGAA	AAAATTAGGGAA'							TTGTGTCACCG
Tan 750 Hymen	111110111111100111	10000101				0111	110011011000	110/11111111111111111111111111111111111	11010101000
Tax 94 obliq	TTTTTGAAAAT C AAA	AAAATTAGGGAA'	TAGCTCAGTTGT	Г		GTA	nAnCAnAGGC'	TTGAAAATCC'	TCGTGTCACCG
86001 tub	TTTTTGAAAAT CG AA	አ አ አ አጥጥአ ሮርር ኣ ኣ ነ	TACCTCACTTC	г		Cum a C	מא כי כי א כי א כי כי כי	דידיי א א א א ידייי	TTGTGTCACCG
89001 tub		AAAA11AGGGAA 'GAAAATTAGGGAA							TTGTGTCACCG
89006 tub 89007 tub		G AAAATTAGGGAA'							TTGTGTCACCG
		G AAAATTAGGGAA' G AAAATTAGGGAA'							TTGTGTCACCG
Tax 1830 tub	TITTIGAATGT AAC	GAAAATTAGGGAA	I AGC I CAG I TG	L		GTAC	JAGCAGAGGC	I I GAAAATCC.	1 1 G T G T C A C C G
	J KL	М							

1

tides. A duplication of seven nucleotides is erratically found in a range of different species. This set of seven nucleotides is included at one end of the already mentioned duplication of 43 nucleotides in A. cepa and A. vavilovii. The seven-nucleotide duplication is found at five locations in the tree of Fig. 2 (character 31), which indicates that it apparently does not influence the structure of the tree. It either might have arisen simultaneously in several events or it can be the remnant of a basic, larger duplication, that vanished in most species for its larger part during evolution. The large subunit of the rubisco gene (rbcL) is very conservative, and at the level of sections and species hardly any mutations were found. Four single-site mutations were recovered. The trnL exon with a size of 339 base pairs did not show any polymorphisms between the accessions included in the dataset.

Phylogeny

The phylogenetic analysis of the dataset including 53 mutations for 33 accessions resulted in 52 most-parsimonious trees of 78 steps with a consistency index (CI) =

0.805 (CI_{uninformative} = 0.754) and a retention index (RI) = 0.915. One of the most-parsimonious trees is shown in Fig. 2. Section *Cepa* is basically divided in three clades. One consists of Allium galanthum, a second one of A. cepa and A. vavilovii, the last clade combines A. fistulosum/A. altaicum with Allium oschaninii. The species Allium pskemense of section Cepa is connected to the branch of one of the accessions Allium senescens (section Rhizirideum), but it shares some characters with certain branches of section Cepa, e.g. character 31 and 32b (Fig. 2). A. obliquum shares two duplications of three and seven nucleotides respectively in the trnL-F intergenic spacer with A. globosum and A. saxatile. A duplication of seven nucleotides in the rps16 intron is shared by A. saxatile and Allium hymenorrhizum. The 80% majority rule consensus tree based on the 52 most-parsimonious trees is shown in Fig. 3. The polytomies are caused by the synapomorphisms as indicated by the tree in Fig. 2. Sections Cepa and Schoenoprasum are closely connected, together with Allium roylei. The species of section Rhizirideum do not share a synapomorphism unambiguously, although some synapomorphisms are indicated in the individual tree of Fig. 2. Sections Petroprason and Oreiprason are not clearly separated. Bootstrap analysis

Table 2 (continued)

Tubic = (continue	<i>(</i> 4)						
	310	320	330	340	350	360	370
Jumbo					rggtagagca g a		
Stuttgarter					rggtaganca g a		
Polar					rggta <i>n</i> agca g a		
95081 vav					rggtagagca g a		
206 osch	GTTCAAATT'	TGGCTCCTGA	ATTGTTGGGA	TAGCTCA <i>n</i> TT	rggtagagca g a	AGGACTGAAAA?	ſС
103 gal	GTTCAAATT'	TGACTCCTGA	ATTGTTGGGA	FAGCTCAGT7	rggtagagca g a	AGGACTGAAAA?	ГC
207 pske	GTTCAAATT'	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGTT	rggtaragca g a	AGGACTGAAAA?	ГC
102 fist	GTTCAAATT'	TGG G TCCTGA	ATTGTTGGGA	TAGCTCAGTT	rgg <i>n</i> agagca g a	AGGACTGAAAA?	ГC
201b alta	GTTCAAATT'	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGTT	rggtagagca g a	AGGACTGAAAA	ГC
104 royl	GTTCAAATT.	nGGCTCCTGA	ATTGTTGGGA	TAGCTCAKTT	rggtaaagcaa <i>i</i>	AGGACTGAAAA'	ГC
All 911 schoe					rggtaaagcaa <i>i</i>		
93007 schoe					ΓGGTAA <i>nnnnn</i>		
Tax 1605 scho					rggtaaascaa <i>i</i>		
Tax 42 altync	GTTCAAATT'	TGGCTCCTG	ATTGTTGGGA	TAGCTCAKTT	rggtaaagcaaa	AGGACTGAAAA'	ГC
93008 altync					rggtagagca <i>ni</i>		
Tax 536 karel	GTTCAAATT'	TGGCTCCTGA	ATTGTTGGGA	TASCTCA <i>n</i> T7	rggtaaa <i>n</i> caaa	AGGACTGAAAA.	ГC
All 1122 sen	GTTCAAATT	TGGCTCCTGA	ATTGTTGGGA	TACCTCATT	rggtaaa <i>n</i> caa <i>i</i>	AGGACTGAAA <i>n</i> T	ГС
89010 sen	GTTCAAATT'	TGGCTCCT <i>nP</i>	ATT <i>n</i> TTGGGA	TAnCTCATT	rggtaaa <i>n</i> caa <i>i</i>	AGGACTGAAAA?	ГC
93005 sen	GTTCAAATT'	TGGCTCCTG	ATTGTTGGGA	TAGCTCAGTT	rggta <i>n</i> agca <i>ni</i>	AGGACTGAAAA?	ГС
89008 nut	GTTCAAATT'	TGGCTCCTG	ATTGTTGGGA	TAGCTCAGTT	rggta <i>n</i> agcaa	AGGACTGAAAA	ГC
89009 nut	GTTCAAATT'	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAnTT	rggta <i>n</i> agcaa <i>i</i>	AGGACTGAAAA	ГC
89011 nut	GTTCAAATT'	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGTT	rggtanagca <i>ni</i>	AGGACTGAAAA	ГC
93012 nut	GTTCAAATT'	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGTT	rggtagagc <i>nni</i>	าททททททททท	nn
Tax 256 ang	GTTCAAATT	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGTT	rggta <i>n</i> agca <i>ni</i>	AGGACTGAAAA	ГC
Tax 2335 lin	COMO A A TOM	maaamaama <i>i</i>	3 TTTCTTTCCC3	T N C C TT C N C TT C	rggta <i>n</i> agcaa <i>i</i>		T.C
1ax 2335 1111	GIICAAAII	1GGC1CC1GF	AIIGIIGGA	IAGCICAGI.	IGGIAHAGCAAA	AGGAC I GAAAA.	i C
All 1123 glob	nnnnnnnn	nnnnnnnn	ากกกกกกกกกกก	וחחחחחחחחו	ากกกกกกกกกกา	ากกกกกกกกกก	nn
Tax 632 saxa	GTTCAAATT	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGTT	rggtaga <i>n</i> caa <i>i</i>	AGG <i>nn</i> TGAAAA	ГC
Tax 750 hymen	GTTCAAATT	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGT	rggtanagca <i>ni</i>	AGGACTGAAAA	ГC
Tax 94 obliq	GTTCAAATT	TGGGTCCTGA	ATTGTTGGGA	TAGCTCAGT	rggnanagcan <i>i</i>	AGGA <i>n</i> TGAAAA	rc
	011011111						
86001 tub	GTTCAAATT	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGT	rggtagagca g a	AGGACTGAAAA'	ГC
89006 tub	GTTCAAATT	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGTT	rggtagagca g a	AGGACTGAAAA	ГC
89007 tub	GTTCAAATT	TGGCTCCTG	ATTGTTGGGA	TAGCTCAGT	rggtaga <i>nc</i> a <i>n</i>	AGGACTGAAAA'	ГC
Tax 1830 tub	GTTCAAATT	TGGCTCCTGA	ATTGTTGGGA	TAGCTCAGT	rggtagagca g a	AGGACTGAAAA'	ГC

showed weak support to some clades as indicated in Fig. 3. Although the separate position of *A. hymenor-rhizum*, the clade with *Allium pskemenselA*. *senescens* and the position of *A. galanthum* at the root of the Cepa clade are weakly supported, these positions are all supported by more than 80% of the most-parsimonious trees. The final, conclusive tree is exclusively based on branches supported after bootstrap analysis (Fig. 4b).

Crossability analysis

Twenty one crosses are carried out in 16 combinations between eight species. The results are listed in Table 3. No hybrid seeds have been obtained in the current crossability program, which might be caused by the fact that almost all crosses involve combinations of species belonging to different sections.

Table 3 Results of crossing experiments between species of *Allium* subgenus *Rhizirideum*

0

Parents	Flowers	Seeds
Crossing Experiments in 1997		
roylei × senescens All 1122	10	0
roylei × altyncolicum Tax 42	13	0
roylei × karelinii Tax 536	7	0
roylei × tuberosum 86001	18	0
roylei × obliquum Tax 94	16	0
roylei × schoenoprasum Tax 1605	16	0
tuberosum 86001 × karelinii Tax 536	18	0
tuberosum 86001 × roylei	15	0
tuberosum 86001 self poll.	40	43
Crossing experiments in 1998		
cepa 96211 × senescens All 1122	120	0
cepa 96211 × schoenoprasum Tax 1605	140	0
cepa 96211 × tuberosum 86001	250	0
roylei 95001 × altyncolicum Tax 42	53	0
roylei 95001 × schoenoprasum Tax 1605	46	0
roylei 95001 × tuberosum 86001	51	0
roylei 95001 self poll.	120	99
senescens All 1122 × cepa 96030	60	0
senescens All 1122 × roylei 95001	73	0
senescens All 1122 × tuberosum 86001	78	0
altyncolicum Tax 42 × senescens All 1122	37	0
tuberosum 86001 × senescens All 1122	41	0
tuberosum 86001 × schoenoprasum Tax 1605	35	0
tuberosum 86001 × altyncolicum Tax 42	67	0

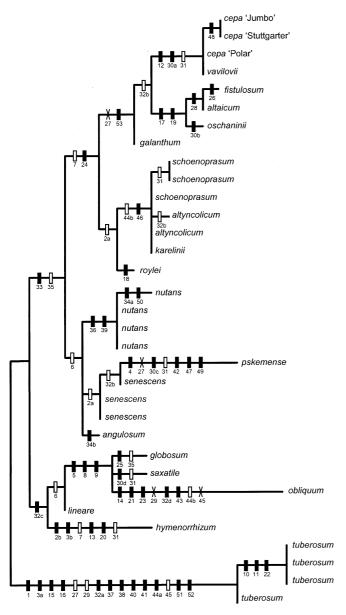


Fig. 2 One of the 52 most-parsimonious trees of 78 steps-long of representatives of *Allium* subgenus *Rhizirideum* (CI = 0.808; CI uninformative = 0.756; RI = 0.911). The *indices* of the bars indicate the position of the mutation: 1–12: deletions/duplications in the trnL-F intergenic spacer; 13–27: single-site mutations in the trnL-F intergenic spacer; 28–42: deletions/duplications in the rps16 intron; 43–49: single-site mutations in the rps16 intron; 50–53: single-site mutations in the rubisco large subunit

Discussion

Phylogeny

The division of the included species in clades is generally in agreement with the accepted taxonomic division based on morphology, cytogenetics or chemical compounds (El-Gadi and Elkington 1977; Hanelt 1990; Hanelt et al.

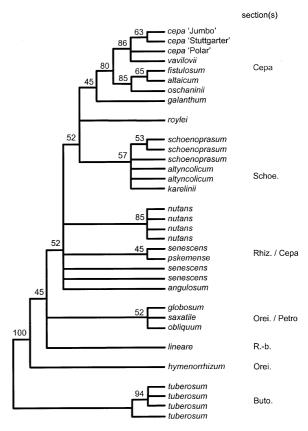
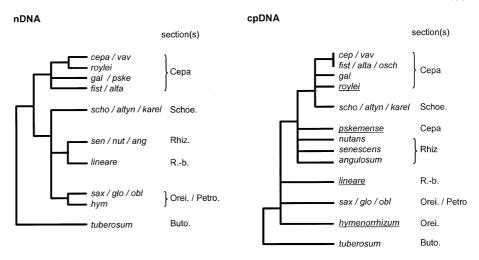


Fig. 3 Eighty percent majority rule consensus tree of 52 mostparsimonious trees showing the evolutionary position of 33 accessions of *Allium* subgenus *Rhizirideum*. Bootstraps values are indicated at the branches. Section names are abbreviated: *Cepa*: section *Cepa*; *Rhiz*: section *Rhizirideum*; *Schoe*: section *Schoenoprasum*; *Orei*: section *Oreiprason*; *Petro*: section *Petroprason*; *R-b*: section *Reticulato-bulbosa*; *Buto*: section *Butomissa*

1992; van Raamsdonk and de Vries 1992a; Hanelt and Fritsch 1994). The combination of species of sections *Oreiprason* and *Petroprason* in one cluster is also found by Dubouzet et al. (1997). This position is supported by the close resemblance of the species of these two sections in morphology and geographical distribution (Hanelt et al. 1992).

All species included with more than one accession (i.e. belonging to sections *Schoenoprasum* and *Rhizirideum*) appear to be monophyletic; or else due to a lack of discriminating characters monophyly was neither confirmed nor rejected (*A. cepalA. vavilovii* and *A. schoenoprasum/Allium altyncolicum*). The same conclusion has been reached for all wild species of section *Cepa*, but cultigens were intermixed with their immediate ancestor (van Raamsdonk et al. 1997). Monophyly of species was also found after nDNA analysis except for one accession of *A. senescens* that was placed in one clade with the only accession of *Allium angulosum* (van Raamsdonk et al. 2000). These results agree with the results of Dubouzet et

Fig. 4 Comparison of the nDNA phylogeny (van Raamsdonk et al. 2000) with the cpDNA phylogeny (current results) of a set of species of *Allium* subgenus *Rhizirideum*



al. (1997) where all accessions per species were placed in the same cluster.

The three alliances in section Cepa as proposed by Hanelt (1990) cannot completely be recognised in the current results (Figs. 2 and 3). The three groups, i.e. A. fistulosum and A. altaicum, A. galanthum and A. pskemense, and A. cepa, A. vavilovii and A. oschaninii, do arise after morphological analysis (van Raamsdonk and de Vries 1992a), and after analysis of nDNA variation (Bradeen and Havey 1995; van Raamsdonk et al. 2000). The species with a deviating position are A. oschaninii and A. pskemense. The position of A. oschaninii close to A. fistulosum and A. altaicum is in concordance with chromosome C-banding patterns (de Vries and Jongerius 1988), but disagrees with other data (van Raamsdonk 1992b). A distant position of A. oschaninii was also found after cpDNA analysis (Havey 1992; Lilly and Havey 2001). The current results support the main division between the clade of A. cepa/A. vavilovii and A. fistulosum/A. altaicum ("subsection Phyllodolon"). The distinction of the A. galanthum/A. pskemense alliance is not supported by the consensus tree since A. pskemense possess an A. senescens-like cpDNA type with some modifications and one notable reversal that is shared with other members of section *Cepa* (Fig. 2).

Each of the four main subgenera of the genus *Allium* posses a separate cluster after phenetic analysis of cpDNA diversity in the study of Linne von Berg et al. (1996). Species of sections *Cepa* and *Schoenoprasum* are located in the same cluster in their results, whereas the two species included in their study belonging to section *Rhizirideum* were placed at a larger distance. This seems to be in concordance with the present results, but different data sets can not be reliably compared by using different approaches, i.e. phylogenetic and phenetic analyses.

A phylogenetic analysis of section *Cepa*, including *A. roylei*, was carried out by Havey (1992). *A. oschaninii* and *A. pskemense* were placed at the root of the clade of section *Cepa*. Nothing can be said about the relationship between *A. pskemense* and any representative of section

Rhizirideum, since none of the latter were included in his analysis.

It is most important for a proper comparison of differently based phylogenies that the same set of material is used in the datasets to be compared (e.g. Sanderson and Donoghue 1989). Such a comparison was carried out using the cpDNA dataset of Havey (1992) and the RAPDnDNA dataset of van Raamsdonk et al. (1997) for the species exclusively belonging to section Cepa. There appeared to be a striking correlation between the cpDNA tree (Havey 1992) and the crossability dendrogram (van Raamsdonk et al. 1992), but differences in the level of crossability should be correlated with differences in species position between nDNA and cpDNA phylogenies, because of the different modes of inheritance of these types of DNA (van Raamsdonk et al. 1997). In order to extend a view on the crossability of A. cepa and A. roylei beyond the border of section Cepa, it is important to have different phylogenies available based on the same set of material.

Comparison of nDNA and cpDNA phylogenies and the analysis of introgression events

The set of species used in the current cpDNA study is identical to the set of species as used in the nDNA study of van Raamsdonk et al. (2000) except for A. oschaninii, due to the fact that most probably the pre-amplification of the nDNA of A. oschaninii did not work out well. As in the nDNA consensus tree (van Raamsdonk et al. 2000), sections Cepa, Schoenoprasum and Rhizirideum each possess a separate position in the tree (Figs. 3 and 4). The clade with A. globosum, A. saxatile and A. obliquum, combining species of sections *Oreiprason* and *Petropra*son, is also recognisable in both trees (Fig. 4). The relationships at the level of sections is not congruent between the nDNA and cpDNA-based phylogenies. For instance, the relatively close relationship between the sections Cepa and Schoenoprasum after cpDNA analysis (current results: Fig. 3) was not found after nDNA analysis (van Raamsdonk et al. 2000). The relative distances between the different sections in the nDNA tree were almost equal to each other, as was shown in the dendrogram after phenetic analysis (van Raamsdonk et al. 2000: Fig. 4).

The main differences between the cpDNA and the nDNA-based phylogenies (Fig. 4) are the remote positions of A. roylei, A. pskemense, A. lineare and A. hymenorrhizum. It is clear that A. pskemense possesses the A. senescens-like cpDNA, notwithstanding the fact that A. pskemense shares two mutations with accessions of the section Cepa. Considering the situation that the cpDNA genome is not recombined during gametogenesis. some sort of chloroplast-capture occurred during the evolution of A. pskemense, followed by a re-establishment of a nDNA composition that fits with those of the species of section Cepa. The same conclusion might be applied to the situation of A. roylei at the root of section Schoenoprasum, since no sharing of mutations is found with representatives of the clade of section Cepa (Fig. 2). The differences between section Cepa and section Schoenoprasum, together with A. roylei, however, are too weak to be supported by bootstrap analysis (Fig. 3). The same situation might explain the position of A. lineare and A. hymenorrhizum, but their position in the final cpDNA tree is not supported after bootstrap analysis (Fig. 4b). The relatively separated position of A. hymenorrhizum found after cpDNA as well as nDNA analysis, is comparable to the results of Dubouzet et al. (1997).

Several other analyses are based on cpDNA as well as nDNA. These studies exclude most species of section *Cepa* (Mes et al. 1999; Friesen et al. 2000) or are exclusively based on species of section *Cepa* without an outgroup from outside the section (Friesen et al. 1999). A correlation between haplotypes and geographic distribution has been found in several species of *Allium*, indicating either parallel adaptation or gene flow. Incongruence between datasets could not be tested for the structure of the ITS dataset (Mes et al. 1999). The nDNA and the cpDNA phylogenetic trees in the study of Friesen et al. (2000) differ only for the position of *Allium kingdonii*. Hybridisation as well as sample errors are presumed as a cause (Friesen et al. 2000).

Crossability and the prediction of interfertility

The currently presented crossability study is the first one that covers a range of species belonging to different sections of the subgenus *Rhizirideum* without using artificial pollination methods. Earlier studies focused primarily on the combination of *A. cepa* with other species. Hybrid plants with *A. galanthum*, *A. fistulosum*, *A. vavilovii* and *A. roylei* were obtained, whereas crosses of *A. cepa* with *A. pskemense*, *A. oschaninii*, *A. angulosum*, *A. senescens* and *A. schoenoprasum* failed (Saini and Davis 1967; McCollum 1971, 1974; Gonzales and Ford-Lloyd 1987; van Raamsdonk et al. 1992). Hybrids between *A. cepa* and a range of species are being

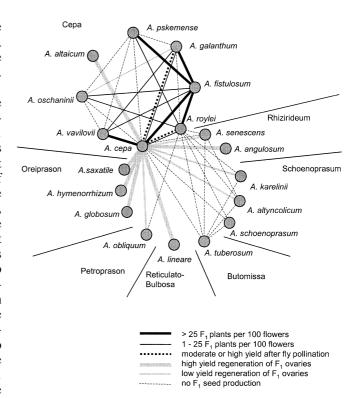


Fig. 5 Overview of the currently known crossability results between species of sections *Cepa, Rhizirideum, Schoenoprasum, Oreiprason, Petroprason, Reticulato-bulbosa* and *Butomissa*

produced by means of embryo-rescue techniques (Keller et al. 1996). All the currently known results are summarised in Fig. 5.

The hypothesis is proposed that a shorter distance between two species in a cpDNA tree compared to their distance in a nDNA tree will predict inter-fertility at a certain level. On the other hand, no conclusion can be drawn from a larger distance in the cpDNA tree (van Raamsdonk et al. 1997). This hypothesis is based on the situation that chloroplast capture, i.e. a short distance in a cpDNA tree, is the result of a certain level of effective inter-fertility. Introgression between species as basic mechanism can result in replacement of the cytoplasmic DNA of the recipient parent, whereas only a small portion of donor nDNA will finally be included in the recipient gene pool. The differences between the effect of introgression of nuclear and cytoplasmic-types of DNA are due to the absence of recombination in the latter (van Raamsdonk et al. 1997; compare Rieseberg and Brunsfeld 1992). The present comparison of nDNA- and cpDNAbased phylogenies with the same set of species and accessions allows a detailed analysis of the differences between bi-parentally and maternally inherited genomes, and can be used for the prediction of inter-fertility. Validation of the hypothesis depends on the availability of crossing results. The position of A. roylei (nDNA: section Cepa) close to section Schoenoprasum in the cpDNA phylogeny should indicate a certain level of crossability with representatives of this section, but the currently

available crossing results neither support not reject this assumption (Fig. 5). Band sharing of the nDNA profiles of A. roylei with those of other species revealed a larger percentage of bands shared with representatives of section Rhizirideum than of section Schoenoprasum (van Raamsdonk et al. 2000). Most probably a considerable secondary evolution after the putative hybridisation and introgression event took place in A. roylei. Therefore, the effect of parallel evolution can not completely be ruled out. A. roylei is close to an A. cepa accession with the S-type cytoplasm (Lilly and Havey 2001). This CMS onion might have been originated after introduction of an alien cytoplasm from a A. roylei-like plant relating to some representative of section Schoenoprasum. The position of A. pskemense close to A. senescens in the cpDNA tree might also predict some interfertility between this species and *A. senescens*. However, this prediction can not be approved by any crossability result of *A. pskemense* with a representative of section *Rhizirideum* (Fig. 5). Further crossability results are necessary in order to find supportive evidence for the hypothesis that a shorter distance between two species in a cpDNA tree compared to their distance in a nDNA tree might indicate a certain level of inter-fertility.

Acknowledgements We thank our late colleague Dr. J.H. Sandbrink from Plant Research International for critically reading this manuscript and Dr. W. Koopman for assisting in the analyses. Dr. J. Keller (IPK, Gatersleben, Germany) kindly provided a range of different accessions from the Gatersleben collection. This research was supported by the EC-FAIR Programme (FAIR CT95-465) and by the Dutch Ministry of Agriculture, Nature Management and Fisheries.

Appendix: list of plant material used

Name	Source and origin	CPRO ref. no.
Section Cepa		
A. cepa 'Jumbo'		
'Stuttgarter'		
'Polar'		96147
A. vavilovii	Grown from 83010: H.B.Chorog, wild origin	95081
A. oschaninii	H. B. Budapest Grown from 79147: USDA Beltsville	78227 82550
A. galanthum A. pskemense	H. B. Alma-Ata	65448
A. fistulosum	Grown from 76201: H. B. Odessa	84236
A. altaicum	Grown from 70201. II. B. Odessa	911126
A. roylei	Grown from 79150: USDA Beltsville C 502)111 2 0
	originally from India	95001
Section Schoenoprasum		
A. schoenoprasum	IPK Gatersleben s.n.	93007
1	IPK: All 911	96158
	IPK: Tax 1605	96152
A. altyncolicum	IPK: Tax 42	96150
	IPK Gatersleben s.n.	93008
A. karelinii	IPK: Tax 536	96154
Section Rhizirideum		
A. senescens	All Union Res. Inst. Veg. USSR	89010
	IPK Gatersleben s.n.	93005
	IPK: All 1122	96159
A. nutans	All Union Res. Inst. Veg. USSR	89008
	All Union Res. Inst. Veg. USSR	89009 89011
	All Union Res. Inst. Veg. USSR IPK Gatersleben s.n.	93012
A. angulosum	IPK: Tax 256	96149
Section Oreiprason	11 K. Tux 250	70117
A. globosum	IPK: All 1123	96151
A. saxatile	IPK: Tax 632	96157
A. hymenorrhizum	IPK: Tax 750	96153
Section Petroprason		
A. obliquum	IPK: Tax 94	96156
Section Reticulato-bulbosa		
A. lineare	IPK: Tax 2335	96155
Section Butomissa		
A. tuberosum	Origin: Thailand	
	All Union Res. Inst. Veg. USSR	89006
	All Union Res. Inst. Veg. USSR	89007
	IPK: Tax 1830	96148

References

- Beek JG van der, Verkerk R, Zabel P, Lindhout P (1992) Mapping strategy for resistance genes in tomato based on RFLPs between cultivars: Cf9 (resistance to *Cladosporum fulvum*) on chromosome 1. Theor Appl Genet 84:106–112
- Bradeen JM, Havey MJ (1995) Randomly amplified polymorphic DNA in bulb onion and its use to assess inbred integrity. J Am Soc Hort Sci 120:752–758
- Dubouzet JG, Shinoda K, Murata N (1997) Phylogeny of *Allium* L. subgenus *Rhizirideum* (G. Don ex Koch) Wendelbo according to dot-blot hybridization with randomly amplified DNA probes. Theor Appl Genet 95:1223–1228
- El-Gadi A, Elkington TT (1977) Numerical taxonomic studies on species in *Allium* subgenus *Rhizirideum*. NewsPhytol 79:183–201
- Friesen N, Pollner S, Bachmann K, Blattner FR (1999) RAPDs and noncoding chloroplast DNA reveal a single origin of the cultivated *Allium fistulosum* from *A. altaicum* (Alliaceae). Am J Bot 86:554–562
- Friesen N, Fritsch R, Pollner S, Blattner FR (2000) Molecular and morphological evidence for an origin of the aberrant genusd *Milula* within Himalayan species of *Allium* (Alliaceae). Mol Phylog Evol 17:209–218
- Fritsch R, Matin F, Klaas M (2001) *Allium vavilovii* M. Popov et Vved. and a new Iranian species are the closest among the known relatives of the common onion *A. cepa* L. (Alliaceae). Genet Res Crop Evol 48:401–408
- Gonzalez LG, Ford-Lloyd BV (1987) Facilitation of wide-crossing through embryo rescue and pollen storage in interspecific hybridization of cultivated *Allium* species. Plant Breed 98:318– 322
- Hanelt P (1990) Taxonomy, evolution and history. In: Rabinowitch
 HD, Brewster JL (eds) Onion and allied crops, volume I.
 Botany, physiology and genetics. CRC press, Boca Raton,
 Florida, pp 1–26
- Hanelt P, Fritsch R (1994) Notes on some infrageneric taxa in *Allium* L. Kew Bull 49:559–564
- Hanelt P, Schultze-Motel J, Fritsch R, Kruse J, Maass HI, Ohle H, Pistrick K (1992) Infrageneric grouping of *Allium* the Gatersleben approach. In: Hanelt P, Hammer K, Knüpffer H (eds) The genus *Allium* taxonomic problems and genetic resources. Institut für Pflanzengenetik und Kulturpflanzenforschung, Gatersleben, Germany, pp 107–123
- Havey MJ (1992) Restriction enzyme analysis of the chloroplast and nuclear 45S ribosomal DNA of *Allium* sections *Cepa* and *Phyllodolon* (Alliaceae). Plant Syst Evol 183:17–31
- Havey MJ (1995) Cytoplasmatic determinations using the polymerase chain reaction to aid in the extraction of maintainer lines from open-pollinated populations of onion. Theor Appl Genet 90:263–268
- Hollingworth ML, Bailey JP, Hollingworth PM, Ferris C (1999) Chloroplast DNA variation and hybridization between invasive populations of Japanese knotweed and giant knotweed (*Fallopia*, Polygonaceae). Bot J Linnean Soc 129:139–154
- Keller ERJ, Schubert I, Fuchs J, Meister A (1996) Interspecific crosses of onion with distant *Allium* species and characterization of the presumed hybrids by means of flow cytometry, karyotype analysis and genomic in situ hybridization. Theor Appl Genet 92:417–424
- Lilly JW, Havey MJ (2001) Sequence analysis of a chloroplast intergenic spacer for phylogenetic estimates in *Allium* section *Cepa* and a PCR-based polymorphism detecting mixtures of

- male-fertile and male-sterile cytoplasmic onion. Theor Appl Genet 102:78–82
- Linne von Berg G, Samoylov A, Klaas M, Hanelt P (1996) Chloroplast DNA restriction analysis and the infrageneric grouping of *Allium* (Alliaceae). Plant Syst Evol 200:253–261
- McCollum GD (1971) Sterility of some interspecific *Allium* hybrids, J Am Soc Hort Sci 96:359–362
- McCollum GD (1974) Chromosome behavior and sterility of hybrids between common onion, *Allium cepa*, and the related wild *A. oschaninii*. Euphytica 23:699–709
- Mes THM, Friesen N, Fritsch RM, Klaas M, Bachmann K (1997) Criteria for sampling in *Allium* based on chloroplast DNA PCR-RFLPs. Syst Bot 22:701–712
- Mes THM, Fritsch RM, Pollner S, Bachmann K (1999) Evolution of the chloroplast genome and polymorphic ITS regions in *Allium* subg. *Melanocrommyum*. Genome 42:1–12
- Meer QP van der, Vries JN de (1990) An interspecific cross between *Allium roylei* Stearn and *Allium cepa* L., and its backcross to *A. cepa*. Euphytica 47:29–31
- Oxelman B, Liden M, Berglund D (1997) Chloroplast rps16 intron phylogeny of the tribe *Sileneae* (Caryophyllaceae). Plant Syst Evol 206:393–410
- Raamsdonk LWD van, Vries T de (1992a) Systematics and phylogeny of *Allium cepa* L. and allies. In: Hanelt P, Hammer K, Knüpffer H (eds) The genus *Allium* taxonomic problems and genetic resources. Institut für Pflanzengenetik und Kulturpflanzenforschung, Gatersleben, Germany, pp 265–270
- Raamsdonk LWD van, Vries T de (1992b) Biosystematic studies in *Allium* L. section *Cepa*. Bot J Linnean Soc 109:131–143
- Raamsdonk LWD van, Wietsma WA, Vries JN de (1992) Crossing experiments in Allium L. section Cepa. Bot J Linnean Soc 109:293–303
- Raamsdonk LWD van, Smiech MP, Sandbrink JM (1997) Introgression explains incongruency between nuclear and chloroplast DNA-based phylogenies in *Allium* section *Cepa*. Bot J Linnean Soc 123:91–108
- Raamsdonk LWD van, Vrielink-van Ginkel M, Kik C (2000) Phylogeny reconstruction and hybrid analysis in *Allium* subgenus *Rhizirideum*. Theor Appl Genet 100:1000–1009
- Rieseberg LH (1995) The role of hybridization in evolution: old wine in new skins. Am J Bot 82:944–953
- Rieseberg LH, Brunsfeld SJ (1992) Molecular evidence and plant introgression. In: Soltis PS, Soltis DE, Doyle JJ (eds) Molecular systematics of plants. Chapman and Hall, New York, pp 151–176
- Rieseberg LH, Soltis DE (1991) Phylogenetic consequences of cytoplasmic gene flow in plants. Evol Trends Plants 5:65–84
- Saini SS, Davis GN (1967) Compatibility in some *Allium* species. Proc Am Soc Hort Sci 91:401–409
- Sanderson MJ, Donoghue MJ (1989) Patterns of variation in levels of homoplasy. Evolution 43:1781–1795
- Swofford D (1993) Phylogenetic analysis using parsimony, version
- Taberlet P, Gielly L, Pautou G, Bouvet J (1991) Universal primers for amplification of three non-coding regions of chloroplast DNA. Plant Mol Biol 17:1105–1109
- Vries JN de, Jongerius MC (1988) Interstitial C-bands on the chromosomes of *Allium* species from the section *Cepa*. Proc 4th Eucarpia Allium Symp, Wellesbourne, United Kingdom
- Wilkie SE, Isaac PG, Slater RJ (1993) Random amplified polymorphic DNA (RAPD) markers for genetic analysis in *Allium*. Theor Appl Genet 86:497–504