

Resistance Gene-Guided Genome Mining: Serial Promoter Exchanges in *Aspergillus nidulans* Reveal the Biosynthetic Pathway for Fellutamide B, a Proteasome Inhibitor

Hsu-Hua Yeh,^{†,‡,○} Manmeet Ahuja,^{‡,▽,○} Yi-Ming Chiang,^{†,§} C. Elizabeth Oakley,[‡] Shauna Moore,[‡] Olivia Yoon,[‡] Heather Hajovsky,[‡] Jin-Woo Bok,^{||} Nancy P. Keller,^{||} Clay C. C. Wang,^{*,†,⊥} and Berl R. Oakley^{*,‡}

[†]Department of Pharmacology and Pharmaceutical Sciences, School of Pharmacy, University of Southern California, Los Angeles, California 90089, United States

[‡]Department of Molecular Biosciences, University of Kansas, Lawrence, Kansas 66045, United States

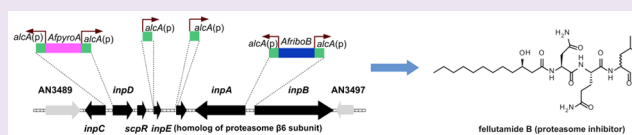
[§]Department of Pharmacy, Chia Nan University of Pharmacy and Science, Tainan City 71710, Taiwan

^{||}Department of Bacteriology and Department of Medical Microbiology and Immunology, University of Wisconsin, Madison, Wisconsin 53706, United States

[⊥]Department of Chemistry, University of Southern California, Dornsife College of Letters, Arts, and Sciences, Los Angeles, California 90089, United States

Supporting Information

ABSTRACT: Fungal genome projects are revealing thousands of cryptic secondary metabolism (SM) biosynthetic gene clusters that encode pathways that potentially produce valuable compounds. Heterologous expression systems should allow these clusters to be expressed and their products obtained, but approaches are needed to identify the most valuable target clusters. The *inp* cluster of *Aspergillus nidulans* contains a gene, *inpE*, that encodes a proteasome subunit, leading us to hypothesize that the *inp* cluster produces a proteasome inhibitor and *inpE* confers resistance to this compound. Previous efforts to express this cluster have failed, but by sequentially replacing the promoters of the genes of the cluster with a regulatable promoter, we have expressed them successfully. Expression reveals that the product of the *inp* cluster is the proteasome inhibitor fellutamide B, and our data allow us to propose a biosynthetic pathway for the compound. By deleting *inpE* and activating expression of the *inp* cluster, we demonstrate that *inpE* is required for resistance to internally produced fellutamide B. These data provide experimental validation for the hypothesis that some fungal SM clusters contain genes that encode resistant forms of the enzymes targeted by the compound produced by the cluster.



The sequencing of fungal genomes has revealed that fungi have vast, and only partially realized, potential to produce secondary metabolite (SM) natural products (NPs). For even well-studied fungi, the SMs that have been isolated and identified after decades of effort are vastly outnumbered by cryptic SM biosynthetic gene clusters (BGCs), encoding biosynthetic pathways, the products of which are currently unknown. For example, examination of the *Penicillium chrysogenum*,¹ *Aspergillus terreus*, *Streptomyces avermitilis*,² and *Saccharopolyspora erythraea*³ genomes suggests that only 10–20% of the SMs potentially generated from these organisms have been identified. The explosion in fungal genome sequencing such as the 1000 Fungal Genomes Project of the Department of Energy Joint Genomes Initiative (<http://1000.fungalgenomes.org/home/>), moreover, is revealing that many species of fungi have large numbers of heretofore unexplored BGCs. Collectively, these fungi harbor thousands of BGCs that undoubtedly produce many valuable NPs that can, in principle, be accessed. There is a strong need for methods, however, to

identify the clusters that are most likely to produce valuable products.

Although emphasis is usually focused on enzymatic genes, many BGCs contain other genes not obviously involved in product synthesis, including a category that may be thought of as self-protective and includes transporters, enzymes that detoxify the final product, and resistant versions of the protein targeted by the BGC (discussed in refs 4 and 5). The latter category has, in principle, the potential to be particularly useful in identifying the activity of the NP synthesized by the BGC. In fact, resistance gene-guided genome mining of BGCs has recently been demonstrated by Bradley Moore's group in the marine actinomycete *Salinispora*.⁶ In the study, they queried the genomes of 86 *Salinispora* strains for putative target modifying resistance genes associated with natural product BGCs. They

Received: March 7, 2016

Accepted: June 13, 2016

Published: June 13, 2016



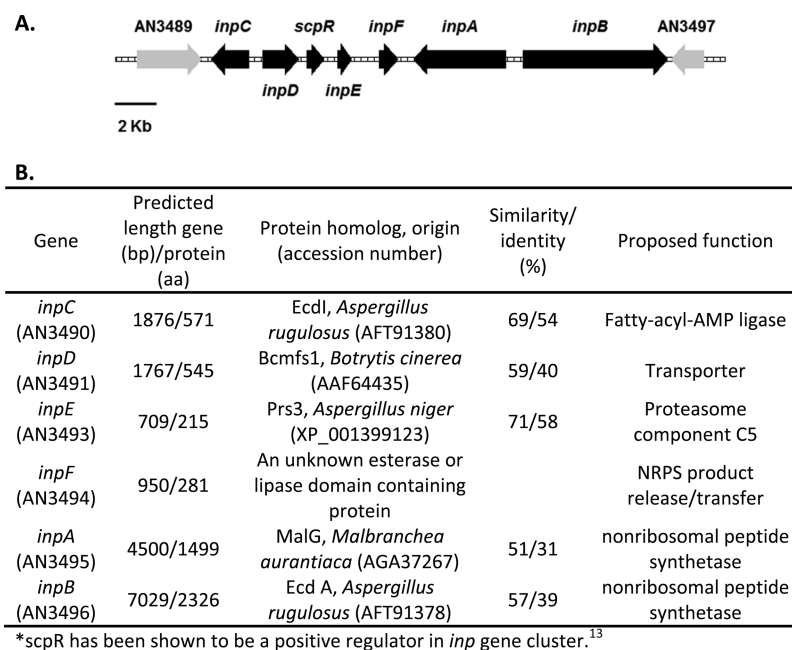


Figure 1. (A) Schematic representation of the *inp* cluster. Each arrow indicates the direction of transcription and relative sizes of the open reading frames (ORFs). (B) The deduced function of each ORF and the amino acid sequence similarity/identity, as compared with the homologues found in a BLAST search of the NCBI nonredundant protein database.

focused on BGCs linked to protein coding genes related to lipid transport and metabolism. Using this approach, they identified a ~22 kb PKS-NRPS hybrid gene cluster of 10 open reading frames from *S. pacifica* CNS-863. They heterologously expressed the gene cluster (renamed *tlm*) in the *S. coelicolor* M1152 host and structurally characterized the products as thiolactomycin and analogs which have been previously shown to be fatty acid synthase II inhibitors.

Links between BGCs and resistance genes have also been observed in filamentous fungi. Regueira *et al.*⁷ have noted several instances in which resistance is apparently mediated by a gene within the BGC that encodes a putative resistant allele of the enzyme inhibited by the SM produced by the cluster.^{8–10} These data led Regueira *et al.* to predict correctly that a particular SM biosynthetic cluster in *Penicillium brevicompactum* is responsible for the synthesis of mycophenolic acid, because the cluster contained a gene encoding IMP dehydrogenase, the target of mycophenolic acid. Other putative examples of embedded BGC resistant target genes include the methionine aminopeptidase encoded in the fumagillin gene cluster [ref 11 and our additional database analyses] and the HMG-CoA reductase gene found in the lovastatin/compactin BGC.^{8,10}

In examining the uncharacterized *Aspergillus nidulans* BGCs, we noted that one contains a gene (*inpE*) encoding a putative proteasome subunit, a protein with no obvious role in NP synthesis (Figure 1). This BGC has been designated the *inp* cluster.^{12,13} We hypothesized that *inpE* might mediate host resistance to the SM product of the *inp* gene cluster, and the fact that InpE has strong homology to the $\beta 6$ subunit of the proteasome, which is highly evolutionarily conserved (HomoGene: 2087, Figure S1), suggested that the product of the *inp* cluster might be a proteasome inhibitor effective against a wide range of organisms. Proteasome inhibitors have recently come to the fore as cancer chemotherapy agents.^{14,15} Bortezomib is a blockbuster drug, recently approved by the FDA, that is effective against multiple myeloma and other cancers. Carfilzomib is another proteasome inhibitor (based on a

natural product) that has recently been FDA approved for multiple myeloma, and other proteasome inhibitors are in clinical trials.¹⁵ We were, thus, particularly interested in determining the product of this BGC.

A pathway-specific regulatory gene, *scpR*, has been identified in the *inp* cluster,¹³ and studies by the Hertweck and Brakhage groups have shown that overexpression of *scpR* results in the expression of some of the genes in the *inp* gene cluster.¹³ However, the activation of the cluster in the *scpR* overexpression strain was insufficient to enable isolation of the product of the pathway. The final product of the pathway as well as pathway intermediates remained completely unknown.

A variety of approaches have been used to activate expression of fungal BGCs,¹⁶ but many BGCs have not yet responded to any of these approaches. We have developed a strategy in which we take advantage of high efficiency gene targeting in *A. nidulans*^{17,18} to simply replace the promoters of BGC genes with a regulatable promoter that allows us to activate transcription of each gene to high levels. We were able to use this approach to complete the identification of all the products released by nonreduced PKS genes in *A. nidulans*.¹⁹ These experiments involved replacing the promoters of only 1–3 genes per BGC, however, and many fungal BGCs contain several genes. We wished to determine if it is feasible to use the promoter replacement approach to elucidate larger BGCs. Since a limited number of selectable markers were available to us, we need to develop a method of recycling selectable markers for transformation to allow all the promoters of the BGC genes to be replaced.

In this study, we report the use of a serial promoter exchange approach, including a method for recycling a selectable marker, to replace the promoters of six genes of the *inp* cluster with the regulatable *alcA* promoter [*alcA*(p)]. By inducing expression of the genes, we were able to obtain production of the final product of this BGC, which we have determined to be fellutamide B. Fellutamide B is a proteasome inhibitor that is of particular interest because it induces nerve growth factor

Table 1. *A. nidulans* Strains Used in This Study^a

LO1362	<i>pyrG89, riboB2, pyroA4, nkuA::argB</i>
LO7692	<i>pyroA4, riboB2, pyrG89, nkuA::argB</i> , sterigmatocystin cluster (AN7804-AN7825)Δ, emericellamide cluster (AN2545-AN2549)Δ, asperfuranone cluster (AN1036-AN1029)Δ, monodictyphenone cluster (AN10023-AN10021)Δ, terrequinone cluster (AN8513-AN8520)Δ, austinol cluster part 1 (AN8379-AN8384)Δ
LO7858	<i>alcA(p)inpB-AfriboB-alcA(p)inpA</i> in LO7692
LO7860	<i>alcA(p)inpB-AfriboB-alcA(p)inpA, alcA(p)inpC-AfpyroA-alcA(p)inpD</i> in LO7692
LO7870	<i>alcA(p)inpB-AfriboB-alcA(p)inpA, alcA(p)inpC-AfpyroA-alcA(p)inpD, alcA(p)inpE</i> in LO7692. Note <i>AfpyrG</i> was removed by looping out.
LO7975	<i>pyrG89; pyroA4; riboB2; pyrG89; nkuA::argB</i> ; sterigmatocystin cluster (AN7804-AN7825)Δ; <i>yA::AtpyrG</i>
LO8080	<i>alcA(p)inpB-AfriboB-alcA(p)inpA, alcA(p)inpC-AfpyroA-alcA(p)inpD, alcA(p)inpF-AfpyrG</i> in LO7692
LO8085	<i>alcA(p)inpB-AfriboB-alcA(p)inpA, alcA(p)inpC-AfpyroA-alcA(p)inpD, alcA(p)inpE, alcA(p)inpF-AfpyrG</i> in LO7692.
LO9169	<i>pyroA4; riboB2; pyrG89; nkuA::argB; inp</i> cluster (AN3496-AN3490 inclusive):: <i>AtpyrG</i>
LO9173	<i>pyroA4; riboB2; pyrG89; nkuA::argB; inp</i> cluster (AN3496-AN3490 inclusive):: <i>AtpyrG</i> ; <i>yA::AN3493-AfriboB</i>
LO9222	<i>alcA(p)inpB-AfriboB-alcA(p)inpA, alcA(p)inpC-AfpyroA-alcA(p)inpD, alcA(p)inpF</i> in LO7692
LO9385, 86	<i>alcA(p)inpB-AfriboB-alcA(p)inpA, alcA(p)inpC-AfpyroA-alcA(p)inpD, alcA(p)inpF, inpE::AtpyrG</i> in LO7692
LO9467, 68, 69	<i>yA::AN3493-ptpA-res</i> in LO9385
LO10079, 80, 81	<i>alcA(p)inpA::AtpyrG</i> in LO9222
LO10083, 84, 85	<i>alcA(p)inpB::AtpyrG</i> in LO9222
LO10089, 90, 91	<i>alcA(p)inpC::AtpyrG</i> in LO9222
LO10093, 96, 97	<i>alcA(p)inpD::AtpyrG</i> in LO9222
LO10099, 100, 101	<i>alcA(p)inpF::AtpyrG</i> in LO9222

^aNote that all strains carry *veA1*.

synthesis.²⁰ By purifying and determining the structures of fellutamide B and the shunt product produced when subsets of the cluster genes are activated by promoter replacement, we are able to propose a biosynthetic pathway for fellutamide B. Through targeted gene deletions, we have found that, as we hypothesized, *inpE* is not required for fellutamide B production but confers resistance to internally produced fellutamide B. Our results not only provide the first identification and characterization of a fellutamide biosynthetic gene cluster, but they provide additional experimental verification that some fungal SM clusters contain genes encoding resistant alleles of the protein targeted by the SM biosynthesized by the cluster. Therefore, as with bacterial BGCs, we can also apply target-directed genome mining to fungal BGC systems.

RESULTS AND DISCUSSION

Determination of the Product of the *inp* Gene Cluster by Serial Promoter Replacement. The *inp* gene cluster contains two nonribosomal peptide synthetase (NRPS) genes, *inpA* (AN3495) and *inpB* (AN3496). *InpA* and *InpB* have the domain architecture CATR and TCATCAT, respectively.¹² C, A, and T refer to a condensation, adenylation, and thiolation domain, respectively.²¹ The R domain refers to a NAD binding domain that was proposed to release the NRP product via a reductive elimination mechanism. Since there are a total of three A domains, the metabolite associated with this cluster was proposed to be a tripeptide with a reduced C terminus.¹²

Our starting strain for this work was LO7692 (full genotypes of strains are listed in Table 1). We created this strain by deleting six secondary metabolism gene clusters (sterigmatocystin, emericellamide, asperfuranone, monodictyphenone, terrequinone, and one of the portions of the split cluster that encodes the austinol biosynthetic pathway). Deletion of these SM clusters reduces the SM background and facilitates identification of any new metabolite that might be produced

as a result of our promoter replacements. We first replaced the promoters of the two NRPS genes, *inpA* and *inpB* (Figure 2A). These genes are divergently transcribed, which allowed us to replace both promoters with *alcA(p)* in a single transformation using a strategy we have previously described,¹⁹ resulting in

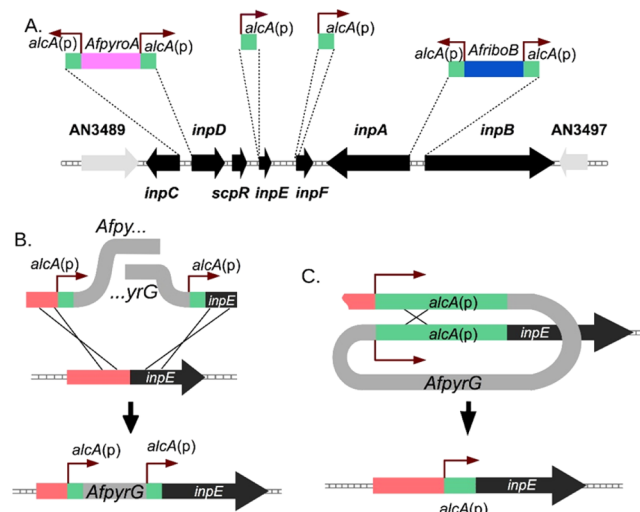


Figure 2. Promoter replacements and strategy. (A) A schematic diagram of the replacements made of the native promoters of the *inp* cluster. (B and C) Promoter replacement with marker recycling strategy used to replace the promoters of *inpE* and *inpF*. (B) The endogenous promoter is replaced with a fragment carrying *AfpyrG*, a marker that can be selected for and against, flanked by direct repeats of the *alcA* promoter [*alcA(p)*]. (C) Spontaneous recombination between the *alcA(p)* repeats results in excision of *AfpyrG*, which is selected for on 5-fluoroorotic acid, resulting in a single copy of *alcA(p)* driving expression of the target gene, *inpE*. Since *AfpyrG* is removed, it can be reused as a selectable marker.

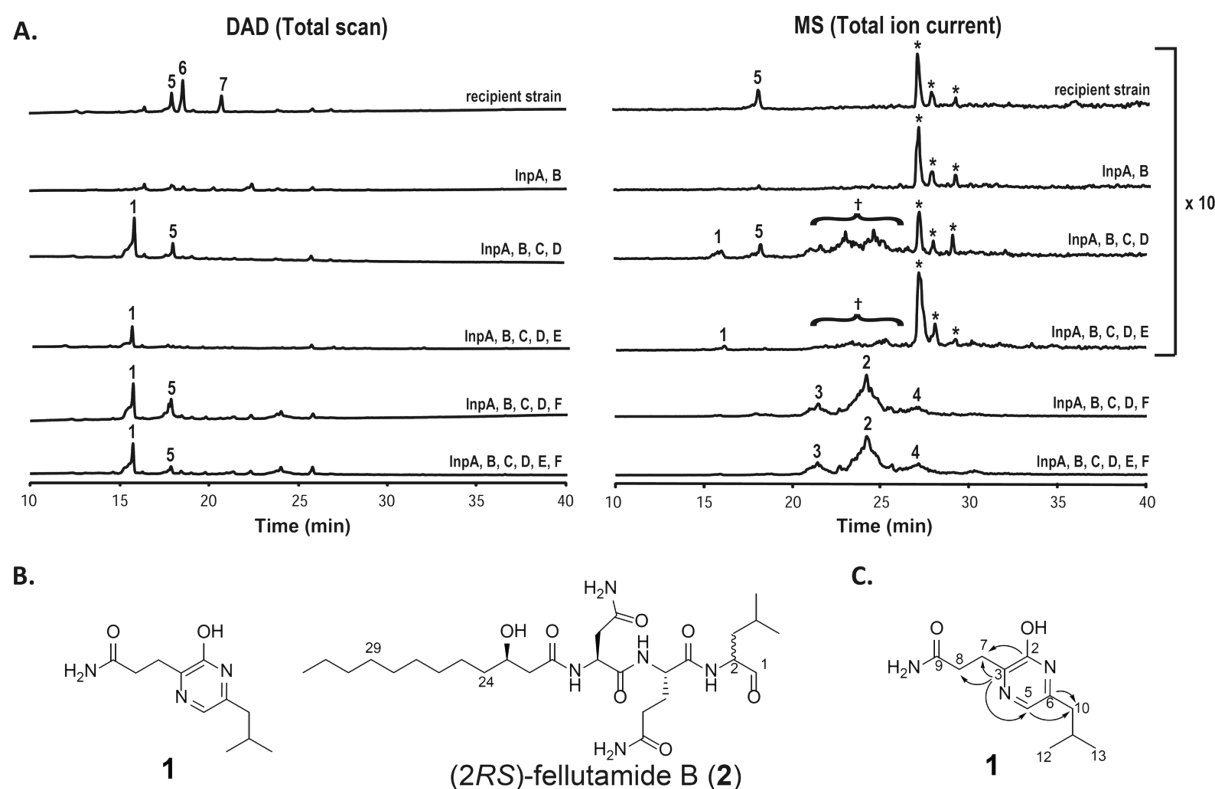


Figure 3. (A) DAD total scan and MS total ion current (TIC) HPLC profiles of culture media extracted from *A. nidulans* strains expressing *inp* cluster genes under the control of *alcA* promoter. The MS TIC profile of recipient; *InpA, B*; *InpA, B, C, D*; and *InpA, B, C, D, E* strains are magnified 10-fold. **1** is 2-hydroxy-5-isobutyl-3-propanamidylpyrazine. **2** is (2*RS*)-felltutamide B. **3** and **4** are felltutamide B derivatives with different lengths of the aliphatic side chains (see Figure S2). **5** is triacetylfulsarine C, and **6** is lumichrome. **7** has similar UV–Vis spectra with **6** and has 14 amu more than **6**, and therefore, **7** could be methyl lumichrome. (B) Chemical structures of compounds **1** and **2**. (C) Key long-range correlations (gHMBC) of compound **1**. *Unknown metabolites detected from MS detector. †Trace amount of felltutamides **2**, **3**, and **4** were detected by MS detector.

strain LO7858 in which *inpA* and *inpB* are inducible. Induction of expression of *inpA* and *inpB* did not lead to production of compounds detectable by diode array detector (DAD) or mass spectrometer (MS; Figure 3A). This result suggested that expression of additional gene(s) in the cluster might be necessary for the production of the SM. In the earlier study using *scpR* overexpression, the *inpC* and *inpD* genes were shown to be coregulated with *inpA* and *inpB*, which indicates that they are part of the same SM cluster.¹³ *InpC* encodes a putative acyl-AMP ligase that might synthesize an AMP substrate as a starter unit.²² *InpD* encodes a putative transmembrane transporter that might facilitate the transportation of the substrates or metabolites synthesized by the pathway. Since *inpC* and *D* are also divergently transcribed, we replaced the promoters of *inpC* and *D* with *alcA*(p) in an *inpA* and *B* promoter replacement strain (Figure 2A) creating strain LO7860 in which *inpA, B, C*, and *D* are all inducible. Simultaneous induction of expression of *inpA, B, C*, and *D* did lead to production of a new DAD and MS detectable peak (compound **1**, Figure 3A).

Compound **1** was purified from a 1 L culture, and its structure was elucidated by spectroscopic methods. The molecular formula of compound **1** was found to be C₁₁H₁₇N₃O₂ by means of HRESIMS and ¹³C NMR experiments, representing five indices of hydrogen deficiency (IHD). The ¹H, ¹³C, gHMQC, and COSY NMR data of **1** revealed it contained isobutyl [δ_{H} 2.34 (2H, d, *J* = 7.6 Hz, H₂-10), 1.93 (1H, m, H-11), and 0.93 (6H, d, *J* = 6.4 Hz, H₃-12 and H₃-13);

and δ_{C} 40.3 (C-10), 29.7 (C-11), and 22.6 (C-12 and C-13)] and propanamidyl [δ_{H} 2.99 (2H, t, *J* = 7.2 Hz, H₂-7) and 2.61 (2H, t, *J* = 7.2 Hz, H₂-8); and δ_{C} 29.4 (C-7), 32.8 (C-8), and 178.3 (C-9)] moieties (Table S1). Subtracting isobutyl and propanamidyl moieties from the molecular formula of **1**, the remaining C₄H₂N₂O represents four IHD, indicating a C₄N₂ heterocyclic aromatic ring. Compounds containing a pyrazine heterocycle are known in nature and have been thought to derive from condensation of two amino acids.²³ In gHMBC spectra, long-range ¹³C–¹H correlations between two olefinic carbons [δ_{C} 158.9 (C-2) and 156.3 (C-3)] and H₂-7 as well as between carbons [δ_{C} 140.7 (C-6) and 123.9 (C-5)] and H₂-10 established that compound **1** does contain a pyrazine moiety (Figure 3C). Moreover, long-range C-3/H₂-8 and C-3/H₂-5 gHMBC correlations revealed that compound **1** is 2-hydroxy-5-isobutyl-3-propanamidylpyrazine. In addition to compound **1**, we identified several MS detectable minor peaks eluted from 20 to 25 min. However, we were unable to characterize their structures due to the low yield of those metabolites (Figure 3A).

Since compound **1** is a dipeptide but not a tripeptide as predicted by the two NRPS domain architectures,¹² we suspected that **1** is not the final metabolite of the *inp* biosynthetic pathway and that, in addition to *inpA, B, C*, and *D*, expression of other genes of the *inp* gene cluster might be necessary for producing the *bona fide* final metabolite. *InpE* encodes a putative proteasome component, as mentioned, and the predicted product of *inpF* is an unknown protein that

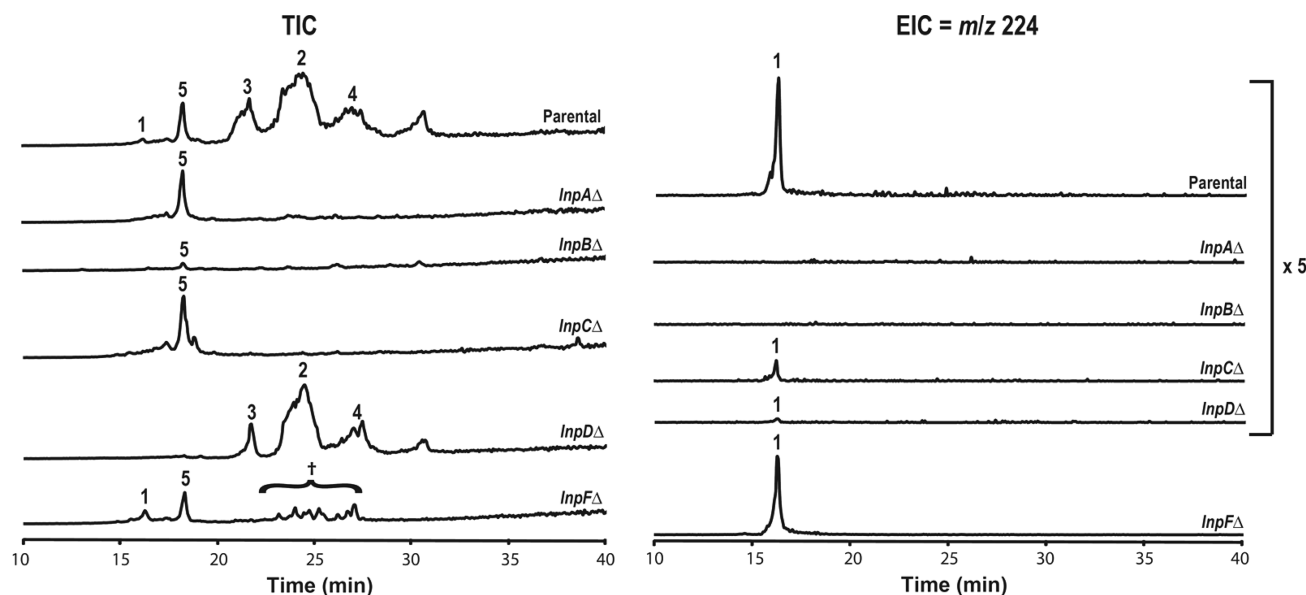


Figure 4. Total ion current (TIC) and extracted ion chromatography (EIC) for compound 1 (EIC = m/z 224) of culture media extracted from *A. nidulans* strains producing fellutamides (parental, LO9222), and individual *inp* gene deletion strains cluster genes under the fellutamide producing condition. The EIC profile of parental and *InpA*, *B*, *C*, and *D* deletion strains are magnified 5 fold. †Trace amount of felllutamides 2, 3, and 4 were detected by MS detector.

contains an esterase-lipase domain. Although activation of transcription of *inpE* and *F* was not addressed in the previous *scpR* overexpression work,¹³ microarray transcription data from *A. nidulans* grown under various culture conditions has indicated that *inpA–F* and *scpR* are part of the same SM gene cluster.²⁴ Because *inpE* and *F* are not divergently transcribed, we replaced the promoters of *inpE* and *inpF* individually in an *inpA*, *B*, *C*, and *D* promoter replacement strain. These replacements required the use of a system for recycling the *Afp_{pyrG}* selectable marker (Figure 2B and C). Satisfyingly, the *inpA*, *B*, *C*, *D*, and *F* promoter replacement strain (LO8080) produced several broad MS detectable peaks upon induction, but the *inpA*, *B*, *C*, *D*, and *E* promoter replacement strain (LO7870) did not (Figure 3A). The parent ion of each broad peak has a mass difference of 28 amu, and all have related MS/MS fragmentation patterns (Figure S2). These results suggest that the broad peaks are related metabolites synthesized by a single biosynthetic pathway. These metabolites are barely detectable by DAD, and therefore they likely contain chromophores with low molar absorptivity, or they are produced at a very low level. We were able to purify the major MS detectable metabolite (compound 2) from a large-scale culture. We found that the majority of compound 2 is in the hyphae, and we were able to isolate >20 mg of compound 2 from 1 L of culture. Compound 2 has a molecular formula $C_{27}H_{49}N_5O_7$, obtained from its HRESIMS. Searching for compounds with the molecular formula $C_{27}H_{49}N_5O_7$ in the SciFinder chemical database revealed a lipotriptide, fellutamide B, that has a reported MS/MS fragmentation similar to compound 2 (Figure S2).²⁵ Fellutamide B (Figure 3B) was first isolated by Shigemori *et al.* from the fungus *Penicillium fellutanum*, and the absolute configuration of C-2 was determined to be a mixture of *R* and *S* forms.²⁵ Recently, Wu *et al.* isolated optically pure (2*S*)-fellutamide B from a marine-derived *Penicillium purpurogenum* G59.²⁶ Comparison of the ^{13}C spectral data of (2*S*)-fellutamide B published by Wu *et al.* with those of compound 2 isolated by us revealed that

they had almost identical ^{13}C chemical shifts (Table S2), except that in the ^{13}C NMR spectrum of compound 2, several ^{13}C NMR signals are either broad or split, especially for those carbons near C-2. This suggested that compound 2 that we isolated is (2*RS*)-fellutamide B. Peaks 3 and 4 were determined to be fellutamide B derivatives, with different lengths of aliphatic side chains, from their MS/MS fragmentation data (Figure S2). Recently, fellutamides were detected from a histone deacetylase, RpdA, a knock-down *A. nidulans* mutant, although the production of fellutamides was not tied to a particular SM gene cluster.²⁷ Fellutamide B has been shown to bind the β -subunits of the 20S proteasome and potently inhibit proteasome catalytic activity.²⁰ We verified that fellutamide B (2) purified from *A. nidulans* strongly inhibits the activity of mammalian proteasomes *in vitro* (Figure S3), and fractions containing fellutamide B derivatives also exhibited strong proteasomal inhibition.

A complete synthesis for fellutamide B has been published.^{28,29} Our promoter-replaced strain, however, produces >20 mg/L of fellutamide B with extremely low materials costs. Although there are costs associated with fermentation and purification, it may prove cost-effective to produce fellutamides in *A. nidulans* promoter-replaced strains, and there is likely scope for further strain improvement and improvement in production conditions to increase yields.

To determine if the gene *inpE* is involved in a biotransformation step in the fellutamide biosynthesis pathway, we constructed strain LO8085 in which all six genes, *inpA–F*, were placed under the control of *alcA*(p). The HPLC profile of this strain (Figure 3A) was similar to that of the strain expressing *inpA*, *B*, *C*, *D*, and *F*, suggesting that *inpE* is not involved in a biotransformation step. Our data reveal that the *inp* cluster is responsible for the biosynthesis of fellutamides.

It is worth pointing out that with our use of *alcA*(p) to drive expression of several genes, there is potential for recombination between the *alcA*(p) copies that would loop out some of the genes, resulting in a loss of fellutamide production. In fact, we

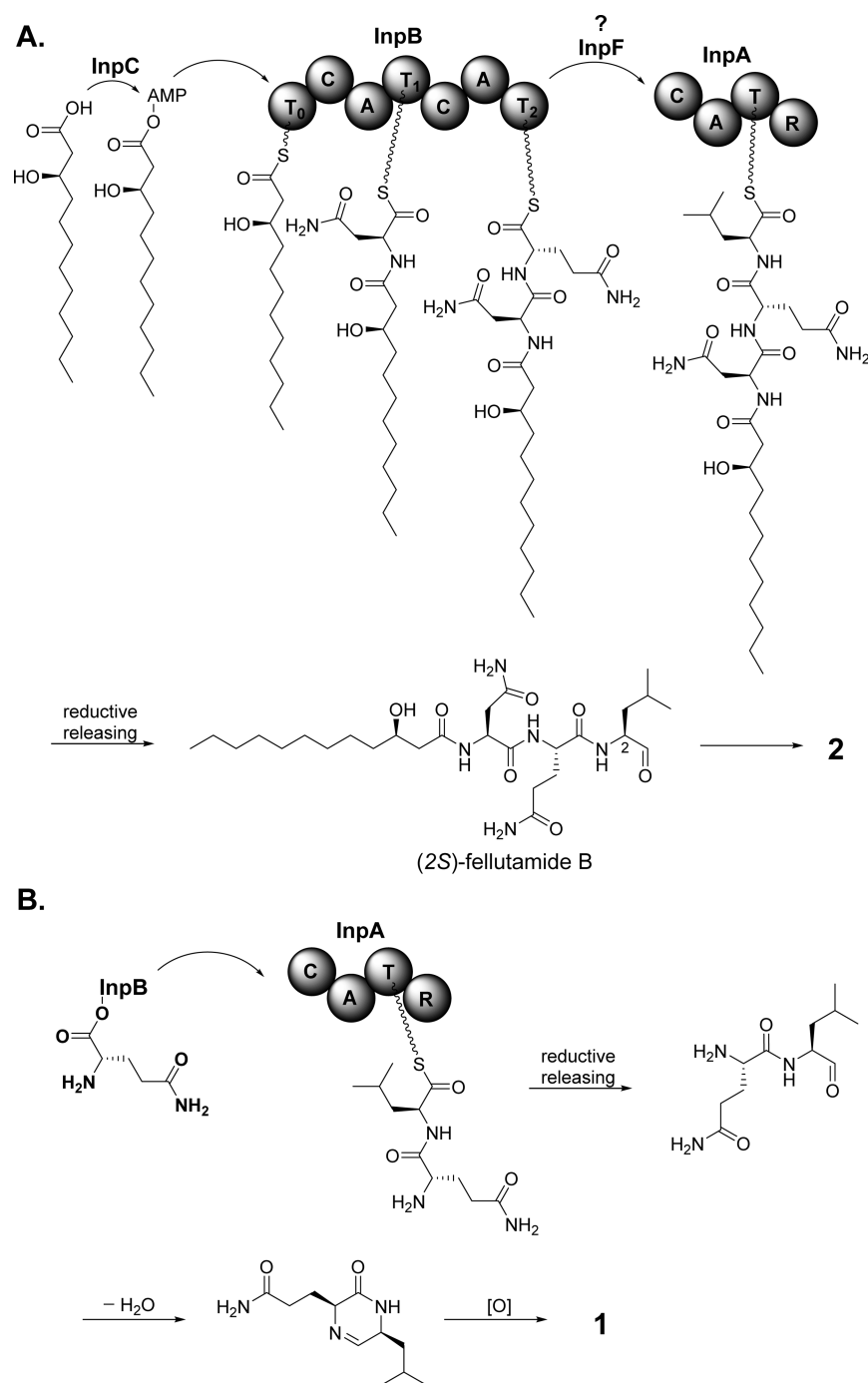


Figure 5. (A) Proposed biosynthesis pathway for (2RS)-fellutamide B (2) and (B) 2-hydroxy-5-isobutyl-3-propanamidylpyrazine (1).

use recombination between repeated *alcA*(p) sequences in our marker recycling procedure (Figure 2C), although in this instance we use 5-FOA to impose a strong selection for recombinants in which *Afp_{pyrG}* has been looped out. In practice, we have not encountered any lack of stability with our fellutamide producing strains. Most possible loop outs would result in the loss of a selectable marker, which is selected against by growth in minimal medium. The version of *alcA*(p) we use is only 400 bp in length, and this may minimize recombination. It is also worth pointing out that although our approach worked very efficiently, additional approaches using CRISPR/Cas9 technology have been reported recently.³⁰

These approaches facilitate marker recycling, and they may provide a useful alternative approach.

To gain additional understanding of the fellutamide biosynthetic pathway, we augmented the data we obtained from our promoter replacement strains by individually deleting *inpA*, *inpB*, *inpC*, *inpD*, and *inpF* in strain LO9222. In LO9222, each of the *inp* genes is under control of the *alcA* promoter except for *inpE*, which is under control of its native promoter. By inducing expression and determining the compounds that accumulate, we hoped to elucidate the biosynthetic functions of the proteins encoded by each gene. Results are shown in Figure 4. As anticipated, deletion of *inpA*, *inpB*, or *inpC* eliminated the production of fellutamides. Deletion of *inpF* greatly diminished

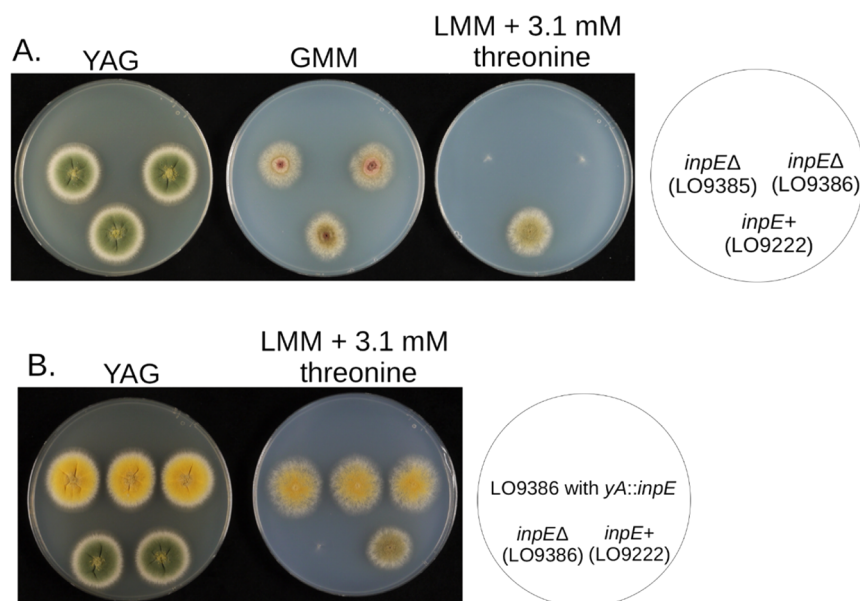


Figure 6. Induction of fellutamide production inhibits growth if *inpE* is deleted. (A) Two *inpE* deletant transformants and their parent, LO9222. On complete medium (YAG) the *alcA* promoter [*alcA*(p)] is strongly repressed and all strains grow well. All strains grow reasonably well on glucose minimal medium (GMM), which represses *alcA*(p) slightly less completely. On lactose minimal medium (LMM) with 3.1 mM threonine, fellutamide production is induced and growth of the *inpE* deletion strains is strongly inhibited. (B) Adding the *inpE* gene back by inserting it at the *yA* (yellow spore color) locus restores growth in strains producing fellutamide. The top three colonies are transformants (LO9467, LO9468, LO9469) of the *inpE*Δ strain, LO9386, in which *inpE* has been inserted at *yA*. They grow at least as well as the parental strain LO9222 when fellutamide production is induced on LMM + threonine. Uridine (10 mM) and uracil (1 g/L) were added to all media to fully supplement the *pyrG89* mutation that is uncomplemented in some strains.

the production of fellutamides. The fellutamides were produced, however, in the *inpD* deletion strain so *inpD*, which putatively encodes a transporter, is not required for fellutamide production. Both of the NRPS genes, *inpA* or *inpB*, were required for production of the cyclodipeptide **1** whereas, although not required, deletion of *inpC* or *inpD* diminished production of **1**. Deletion of *inpF* does not interfere with production of **1**, suggesting that *inpF* is not required for the production of **1**.

The overexpression experiments and individual gene knock out experiments in aggregate enabled us to propose a highly speculative biosynthetic pathway for fellutamide B (**2**; Figure 5A). A homologue of InpC, EcdI, has been shown to catalyze lipoinitiation of EcdA, an NRPS involved in echinocandin B biosynthesis.²² Therefore, we propose that in the first step of fellutamide B biosynthesis InpC activates 3-hydroxydodecanoic acid to generate 3-hydroxydodecanoyl-AMP that is then loaded onto the T₀ domain of InpB. The 3-hydroxydodecanoyl-S-phosphopantetheinyl-T₀ is sequentially extended with L-Asn and L-Gln by the two CAT modules of InpB. The linear lipodipeptide from InpB is then transferred onto InpA for the addition of the third amino acid, L-Leu. Reductive releasing of the lipotriptide by the R domain of InpA produces (2*S*)-fellutamide B. Tautomerization of C-2, which might occur in our culture conditions, led to the formation of (2*RS*)-fellutamide B (**2**). It should be noted that there is no fatty acid synthase or highly reduced PKS (HRPKS) gene in or near the *inp* gene cluster. Therefore, the 3-hydroxy fatty acid side chain might be derived from primary metabolism. Indeed, feeding commercial available decanoic-*d*₁₉ acid to the fellutamide producing strain led to the production of 3-*d*₁₈ (Figure S4). Neither 2-*d*₁₉ nor 4-*d*₁₉ was detected in the feeding experiment, suggesting that beta-oxidation catabolic processes

in mitochondria could be the main source of the 3-hydroxy fatty acid.

Both heterologous expression and individual gene knock out experiments have demonstrated that InpF, a putative esterase or lipase domain containing protein is also involved in the biosynthesis of fellutamides. We propose that InpF is involved in the release and transfer of the lipodipeptide from InpB to InpA. Fellutamides are detected at a very low level when InpF is not expressed, suggesting that the transfer of the lipodipeptide from InpB to InpA is slow and that InpF catalyzes the transfer, leading to the high-level production of fellutamides.

Our individual knock out experiments show that both InpA and InpB are required for the biosynthesis of compound **1**. We propose that the C-terminal A domain in InpB recognizes and activates Gln, and the A domain in InpA recognizes and activates Leu. The Glu-Leu dipeptide is generated by InpA via condensation of Gln and Leu (Figure 5B). Reductive elimination of the Gln-Leu dipeptide followed by intramolecular Schiff base formation and oxidation leads to **1**. Since compound **1** was detected in the InpF deletant strain and InpF expression does not lead to the high production of **1**, the transfer of activated Glu to InpA is likely to be InpF independent. Our individual knock out experiments show that both InpC and InpD are not necessary for, but are involved in, the biosynthesis of **1**. However, our data do not allow us to deduce the precise roles of InpC and InpD in the biosynthesis of **1**.

InpE Confers Resistance to Internally Produced Fellutamides. The predicted product of *inpE* (AN3493) is a strong homologue of the conserved β6 proteasome subunit (Figure S1), and as mentioned, we hypothesized that *inpE* confers resistance to the product of the *inp* cluster, fellutamide B. If InpE confers fellutamide resistance, deletion of *inpE* might

confer heightened sensitivity. To test this possibility, we deleted the entire *Inp* cluster (AN3490-AN3496) by replacing it with the *Aspergillus terreus* *pyrG* gene (*AtpyrG*) in strain LO1362 creating strain LO9169. In this construction, *inpE*/AN3493 is eliminated from the genome. Anticipating that deletion of *inpE* might increase fellutamide sensitivity, we created a control strain by reinserting *inpE* at the *yA* (yellow conidia) locus in LO9169, creating strain LO9173. In broth dilution tests,³¹ LO9169, LO9173, and LO7975, a control strain with an intact *inp* cluster, grew at all concentrations of fellutamide B tested (0.5 to 256 $\mu\text{g/mL}$, at which concentration it precipitated out of solution, data not shown). These data indicate that *A. nidulans* is highly resistant to externally applied fellutamide B and that the mechanism of resistance does not require *inpE*. It is possible that the compound simply is not taken up by *A. nidulans* and does not penetrate the cell wall.

We noted, however, that in our experiments the vast majority of fellutamide B accumulates within hyphae, and we reasoned that *A. nidulans* must have a mechanism for counteracting the toxicity of internally produced fellutamide B. To test this hypothesis, we deleted *inpE* in strain LO9222 in which *inpA–D* and *inpF* are under control of *alcA*(p). We designated the resulting deletants LO9385 and LO9386. We tested these *inpE* deletants under repressing conditions that prevent transcription from *alcA*(p). Under these conditions, no fellutamide B will be produced, and any effect of *inpE* Δ would be independent of fellutamide B production. As shown in Figure 6A, *inpE* Δ transformants grew normally under *alcA*(p) repressing conditions, indicating that *inpE* does not have a function important for growth in the absence of fellutamide B production. We next tested growth of these *inpE* Δ strains on media that induce transcription from *alcA*(p), resulting in fellutamide B production. As shown in Figure 6A, growth was strongly inhibited in the *inpE* Δ strains, revealing that *inpE* confers resistance to internally produced fellutamide B. As a control, we genetically added back the *inpE* gene to one of the *inpE* Δ strains, LO9386, by inserting a copy of *inpE*, under control of its normal promoter, at the *yA* locus. The reinserted *inpE* gene restored growth when fellutamide B production was induced (Figure 6B), confirming that *inpE* confers resistance to internally produced fellutamide B.

These data raise a question of why induction of fellutamide production in strains in which *inpE* is under control of its normal promoter is not harmful. If fellutamide is produced at high levels in cells and *inpE* was not expressed, the prediction is that the fellutamide would be toxic to the cell. Transcription data on *inpE* available at the *Aspergillus* genome database (www.aspergillusgenome.org) and microarray data in the supplemental data set for ref 24 appear to provide an answer. They reveal that *inpE* is constitutively expressed. It is not coregulated with the other genes of the *Inp* cluster, and the fellutamide-resistant proteasome subunit it encodes is expressed even when the remainder of the genes of the *Inp* cluster are not expressed at significant levels. *InpD*, a putative transporter may contribute to fellutamide resistance as well. Deletion of *inpD* and simultaneous induction of *inpA*, *inpB*, *inpC*, and *inpF* results in a small reduction in growth relative to the parental strain when *inpD* is intact (Figure S5). The reduction in growth is much less than is the case if *inpE* is deleted and fellutamide production is induced, however (compare Figure 6 with Figure S5), so *inpE* appears to be the principal resistance mechanism.

The *A. nidulans* genome contains another strong $\beta 6$ homologue, AN5784. We deleted AN5784 in LO9222 (leaving

inpE intact) to determine if it had any role in resistance to internally produced fellutamide. Deletion of AN5784 reduced conidiation and slightly reduced growth but did not confer altered sensitivity to internally produced fellutamide (Figure S6). The simplest interpretation of these data is that AN5784 encodes a fellutamide B-sensitive $\beta 6$ subunit and *inpE* encodes a fellutamide B-resistant subunit. *InpE* confers resistance to internally produced fellutamide B and is functional enough to support growth in the absence of AN5784, but growth and conidiation are not normal (i.e., the proteasome is not fully functional if AN5784 is deleted). AN5784 fully supports normal growth but does not confer fellutamide B resistance.

Conclusions. Our data reveal that the fellutamide B cluster in *A. nidulans* carries a proteasome subunit that confers resistance to fellutamide B, while another homologous proteasome subunit is present elsewhere in the organism. These data provide experimental confirmation for the hypothesis that in some instances fungal SM clusters carry within them genes that encode resistant alleles of the protein targeted by the compound produced by the cluster. The hallmarks of such clusters are that they contain a gene predicted to encode a protein involved in an essential function unrelated to SM biosynthesis and a homologue of that gene is present elsewhere in the genome. Our result is in agreement with Bradley Moore's observation in bacterial SM clusters and demonstrates that the resistance-gene-guided genome mining approach can be broadly applied to both fungal and bacterial BGCs.⁶ Our data also reveal that obtaining products from cryptic SM clusters by serial promoter replacement is feasible and practical and can result in yields that may be commercially useful. Finally, an additional benefit of the serial promoter replacement approach is that we have obtained a shunt product from strains in which the promoters of some of the genes of the *inp* cluster have been replaced.

EXPERIMENTAL SECTION

Strains. Strains used in this study are listed in Table 1. All strains carry the *veA1* mutation. Strain FGSC4 used in Figure S5 was obtained from the Fungal Genetics Stock Center in the 1970s. It is wild-type except that it carries the *veA1* mutation, as do the vast majority of laboratory strains. The FGSC has changed its designation for *Aspergillus nidulans* strains by adding an "A" after FGSC and before the number. The FGSC currently carries a wild-type *A. nidulans* strain designated FGSCA4. This strain is wild-type at the *veA* locus. Although the strain sequenced in the *A. nidulans* genome project is usually called FGSCA4, it is actually FGSC4, the strain obtained by the N. R. Morris lab from the FGSC in the 1970s. This is why the *veA* allele in the sequenced genome is the *veA1* allele. The DNA sequenced in the genome project was isolated from FGSC4 by the Oakley lab.

Media. For induction of *alcA* on solid medium, we used a lactose minimal medium (10 g/L lactose, 6 g/L NaNO_3 , 0.52 g/L KCl, 0.52 g/L $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 1.52 g/L KH_2PO_4 , 15 g/L agar, and 1 mL/L trace element solution³² (Vishniac and Santer, 1957)) supplemented with threonine at the concentrations indicated in the figure legends. Threonine is an inducer of *alcA*(p) but is not a substrate for the alcohol dehydrogenase encoded by *alcA*.³³ Repressing solid media were YAG (5 g/L yeast extract, 20 g/L D-glucose, 15 g/L agar and 0.8 mL/L trace element solution)³² or glucose minimal medium (GMM is 10 g/L D-glucose, 6 g/L NaNO_3 , 0.52 g/L KCl, 0.52 g/L $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 1.52 g/L KH_2PO_4 , 15 g/L agar, and 1 mL/L trace element solution).³²

Molecular Genetic Manipulations. Transforming fragments were generated by fusion PCR essentially as described.^{18,34} To reduce the frequency of PCR-induced mutations, we used high-fidelity DNA polymerases KOD DNA polymerase (EMD Biosciences), Phusion DNA polymerase (New England Biolabs or Thermo), or Q5 Hot Start

High-Fidelity 2X Master Mix (New England Biolabs). For KOD and Phusion DNA polymerases, the annealing temperature was $T_m + 2\text{ }^{\circ}\text{C}$ and $T_m + 3\text{ }^{\circ}\text{C}$ for Q5 with T_m calculated by the formula $T_m = 2(A + T) + 4(G + C)$. The extension rate was 30 s/kilobase of expected product.

Production of protoplasts and transformations were carried out as previously described^{18,34} except that Vintase Pro (Novo) was used at a final concentration of 100 mg mL⁻¹ for protoplasting.³⁴ Dual promoter exchanges were carried out as previously described.¹⁹ Cluster deletions to create strain LO7692 were carried out as previously described using the loop-out approach.³⁵ Selection for excision of fragments carrying *AfpYrG* on 5-FOA was as previously described.³⁶ Diagnostic PCR was carried out as described using Platinum Taq DNA polymerase (Invitrogen) or OneTaq Hot Start Quickload (New England Biolabs).³⁴ The strategy for deletion of individual *inp* cluster genes is shown in Figure S7. *Imp* genes along with the *alcA* promoter driving their expression were replaced with the *Aspergillus terreus pyrG* gene (*AtpyrG*).

Broth microdilution tests for inhibition of growth by fellutamide B were carried out according to the CLSI reference method for filamentous fungi.³¹ RPMI 1640 medium was supplemented with glutamine, 2% glucose, and any supplements required to supplement nutritional markers in the strains.

Fermentation and LC/MS Analysis. For *alcA*(p) induction, 3×10^7 spores were grown in 30 mL of liquid LMM medium [15 g/L lactose, 6g/L NaNO₃, 0.52g/L KCl, 0.52g/L MgSO₄·7H₂O, 1.52g/L KH₂PO₄, 1 mL/L trace elements solution, supplemented with riboflavin (2.5 mg/L), pyridoxine (0.5 mg/L), or uridine (10 mM) and uracil (1 g/L) when necessary] in 125 mL flasks at 37 °C with shaking at 180 rpm. Cyclopentanone was added to the medium at a final concentration of 10 mM 18 h after inoculation. Culture medium was collected 48 h after cyclopentanone induction by filtration and extracted twice with the same volume of EtOAc. EtOAc extract was then evaporated by TurboVap LV (Caliper LifeSciences). The crude extracts were redissolved in 0.5 mL of DMSO:MeOH (1:4) and 10 μL was injected for LC-DAD-MS analysis as described previously.³⁷ Details for large-scale purifications of compounds for each strain and compound identification are given in the Supporting Information as are spectral data. For the isotope feeding experiment, 1 mg of decanoic-*d*₁₉ acid (Cambridge Isotope Laboratories) in 5 μL of DMSO was added into liquid culture at 0 h (the time of spore inoculation) and 18 h (the time of cyclopentanone induction). Culture medium was collected and analyzed as described above.

Proteasome Inhibition Assays. Proteasome activity was assayed with a Millipore 20S Proteasome Activity Assay kit (EMD Millipore catalog # APT280) following the instructions provided with the kit. The assay was performed with proteasomes that were supplied with the kit or, more usually, human 20 proteasomes (BostonBiochem/R & D Systems catalog # E365).

■ ASSOCIATED CONTENT

■ Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acschembio.6b00213.

Detailed experimental procedures and additional tables and figures (PDF)

■ AUTHOR INFORMATION

Corresponding Authors

*E-mail: clayw@usc.edu.

*E-mail: boakley@ku.edu.

Present Addresses

#Drug Discovery and Development Center, Chia Nan University of Pharmacy and Science, Tainan City 71710, Taiwan

[▽]Industrial Biotechnology Division, Reliance Technology Group, Reliance Industries Limited, Reliance Corporate Park, Thane Belapur Road, Ghansoli, Navi Mumbai 400701, India

Author Contributions

[○]These authors contributed equally to this work.

Funding

The authors declare no competing financial interest.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

Supported by grant PO1-GM084077 from the National Institute of General Medical Sciences, by the H. L. Snyder Medical Foundation, and by the Irving Johnson Fund of the University of Kansas Endowment.

■ REFERENCES

- (1) van den Berg, M. A., Albang, R., Albermann, K., Badger, J. H., Daran, J. M., Driessen, A. J., Garcia-Estrada, C., Fedorova, N. D., Harris, D. M., Heijne, W. H., Joardar, V., Kiel, J. A., Kovalchuk, A., Martin, J. F., Nierman, W. C., Nijland, J. G., Pronk, J. T., Roubos, J. A., van der Klei, I. J., van Peij, N. N., Veenhuis, M., von Dohren, H., Wagner, C., Wortman, J., and Bovenberg, R. A. (2008) Genome sequencing and analysis of the filamentous fungus *Penicillium chrysogenum*. *Nat. Biotechnol.* 26, 1161–1168.
- (2) Omura, S., Ikeda, H., Ishikawa, J., Hanamoto, A., Takahashi, C., Shinose, M., Takahashi, Y., Horikawa, H., Nakazawa, H., Osonoe, T., Kikuchi, H., Shiba, T., Sakaki, Y., and Hattori, M. (2001) Genome sequence of an industrial microorganism *Streptomyces avermitilis*: deducing the ability of producing secondary metabolites. *Proc. Natl. Acad. Sci. U. S. A.* 98, 12215–12220.
- (3) Oliynyk, M., Samborsky, M., Lester, J. B., Mironenko, T., Scott, N., Dickens, S., Haydock, S. F., and Leadlay, P. F. (2007) Complete genome sequence of the erythromycin-producing bacterium *Saccharopolyspora erythraea* NRRL23338. *Nat. Biotechnol.* 25, 447–453.
- (4) Cimermanic, P., Medema, M. H., Claesen, J., Kurita, K., Wieland Brown, L. C., Mavrommatis, K., Pati, A., Godfrey, P. A., Koehrsen, M., Clardy, J., Birren, B. W., Takano, E., Sali, A., Lington, R. G., and Fischbach, M. A. (2014) Insights into secondary metabolism from a global analysis of prokaryotic biosynthetic gene clusters. *Cell* 158, 412–421.
- (5) Keller, N. P. (2015) Translating biosynthetic gene clusters into fungal armor and weaponry. *Nat. Chem. Biol.* 11, 671–677.
- (6) Tang, X., Li, J., Millan-Aguinaga, N., Zhang, J. J., O'Neill, E. C., Ugalde, J. A., Jensen, P. R., Mantovani, S. M., and Moore, B. S. (2015) Identification of Thiotetronic Acid Antibiotic Biosynthetic Pathways by Target-directed Genome Mining. *ACS Chem. Biol.* 10, 2841–2849.
- (7) Regueira, T. B., Kildegaard, K. R., Hansen, B. G., Mortensen, U. H., Hertweck, C., and Nielsen, J. (2011) Molecular basis for Mycophenolic Acid Biosynthesis of *Penicillium brevicompactum*. *Appl. Environ. Microbiol.* 77, 3035–3043.
- (8) Kennedy, J., Auclair, K., Kendrew, S. G., Park, C., Vederas, J. C., and Hutchinson, C. R. (1999) Modulation of polyketide synthase activity by accessory proteins during lovastatin biosynthesis. *Science* 284, 1368–1372.
- (9) Lowther, W. T., McMillen, D. A., Orville, A. M., and Matthews, B. W. (1998) The anti-angiogenic agent fumagillin covalently modifies a conserved active-site histidine in the *Escherichia coli* methionine aminopeptidase. *Proc. Natl. Acad. Sci. U. S. A.* 95, 12153–12157.
- (10) Abe, Y., Suzuki, T., Mizuno, T., Ono, C., Iwamoto, K., Hosobuchi, M., and Yoshikawa, H. (2002) Effect of increased dosage of the ML-236B (compactin) biosynthetic gene cluster on ML-236B production in *Penicillium citrinum*. *Mol. Genet. Genomics* 268, 130–137.
- (11) Wiemann, P., Guo, C. J., Palmer, J. M., Sekonyela, R., Wang, C. C., and Keller, N. P. (2013) Prototype of an intertwined secondary-

metabolite supercluster. *Proc. Natl. Acad. Sci. U. S. A.* 110, 17065–17070.

(12) von Dohren, H. (2009) A survey of nonribosomal peptide synthetase (NRPS) genes in *Aspergillus nidulans*. *Fungal Genet. Biol.* 46 (Suppl 1), S45–S52.

(13) Bergmann, S., Funk, A. N., Scherlach, K., Schroeckh, V., Shelest, E., Horn, U., Hertweck, C., and Brakhage, A. A. (2010) Activation of a silent fungal polyketide biosynthesis pathway through regulatory cross talk with a cryptic nonribosomal peptide synthetase gene cluster. *Appl. Environ. Microbiol.* 76, 8143–8149.

(14) Goldberg, A. L. (2012) Development of proteasome inhibitors as research tools and cancer drugs. *J. Cell Biol.* 199, 583–588.

(15) Kisselev, A. F., van der Linden, W. A., and Overkleeft, H. S. (2012) Proteasome inhibitors: an expanding army attacking a unique target. *Chem. Biol.* 19, 99–115.

(16) Chiang, Y. M., Chang, S. L., Oakley, B. R., and Wang, C. C. (2011) Recent advances in awakening silent biosynthetic gene clusters and linking orphan clusters to natural products in microorganisms. *Curr. Opin. Chem. Biol.* 15, 137–143.

(17) Nayak, T., Szewczyk, E., Oakley, C. E., Osmani, A., Ukil, L., Murray, S. L., Hynes, M. J., Osmani, S. A., and Oakley, B. R. (2006) A versatile and efficient gene-targeting system for *Aspergillus nidulans*. *Genetics* 172, 1557–1566.

(18) Szewczyk, E., Nayak, T., Oakley, C. E., Edgerton, H., Xiong, Y., Taheri-Talesh, N., Osmani, S. A., and Oakley, B. R. (2006) Fusion PCR and gene targeting in *Aspergillus nidulans*. *Nat. Protoc.* 1, 3111–3120.

(19) Ahuja, M., Chiang, Y. M., Chang, S. L., Praseuth, M. B., Entwistle, R., Sanchez, J. F., Lo, H. C., Yeh, H. H., Oakley, B. R., and Wang, C. C. (2012) Illuminating the diversity of aromatic polyketide synthases in *Aspergillus nidulans*. *J. Am. Chem. Soc.* 134, 8212–8221.

(20) Hines, J., Groll, M., Fahnestock, M., and Crews, C. M. (2008) Proteasome inhibition by fellutamide B induces nerve growth factor synthesis. *Chem. Biol.* 15, 501–512.

(21) Fischbach, M. A., and Walsh, C. T. (2006) Assembly-line enzymology for polyketide and nonribosomal peptide antibiotics: logic, machinery, and mechanisms. *Chem. Rev.* 106, 3468–3496.

(22) Cacho, R. A., Jiang, W., Chooi, Y. H., Walsh, C. T., and Tang, Y. (2012) Identification and characterization of the echinocandin B biosynthetic gene cluster from *Emericella rugulosa* NRRL 11440. *J. Am. Chem. Soc.* 134, 16781–16790.

(23) Badrinarayanan, S., and Sperry, J. (2011) Biomimetic synthesis of 2,5-bis(indol-3-ylmethyl)pyrazine via intermolecular amino aldehyde cyclization. *Synlett* 16, 2339–2342.

(24) Andersen, M. R., Nielsen, J. B., Klitgaard, A., Petersen, L. M., Zachariassen, M., Hansen, T. J., Blicher, L. H., Gottfredsen, C. H., Larsen, T. O., Nielsen, K. F., and Mortensen, U. H. (2013) Accurate prediction of secondary metabolite gene clusters in filamentous fungi. *Proc. Natl. Acad. Sci. U. S. A.* 110, E99–107.

(25) Shigemori, H., Wakuri, S., Yazawa, K., Nakamura, T., Sasaki, T., and Kobayashi, J. (1991) Fellutamide-A and fellutamide-B, cytotoxic peptides from a marine fish-possessing fungus *Penicillium-fellutanum*. *Tetrahedron* 47, 8529–8534.

(26) Wu, C. J., Li, C. W., and Cui, C. B. (2014) Seven new and two known lipopeptides as well as five known polyketides: the activated production of silent metabolites in a marine-derived fungus by chemical mutagenesis strategy using diethyl sulphate. *Mar. Drugs* 12, 1815–1838.

(27) Albright, J. C., Henke, M. T., Soukup, A. A., McClure, R. A., Thomson, R. J., Keller, N. P., and Kelleher, N. L. (2015) Large-scale metabolomics reveals a complex response of *Aspergillus nidulans* to epigenetic perturbation. *ACS Chem. Biol.* 10, 1535–1541.

(28) Schneekloth, J. S., Jr., Sanders, J. L., Hines, J., and Crews, C. M. (2006) Neurotrophic peptide aldehydes: solid phase synthesis of fellutamide B and a simplified analog. *Bioorg. Med. Chem. Lett.* 16, 3855–3858.

(29) Giltrap, A. M., Cergol, K. M., Pang, A., Britton, W. J., and Payne, R. J. (2013) Total synthesis of fellutamide B and deoxy-fellutamides B, C, and D. *Mar. Drugs* 11, 2382–2397.

(30) Nødvig, C. S., Nielsen, J. B., Kogle, M. E., and Mortensen, U. H. (2015) A CRISPR-Cas9 system for genetic engineering of filamentous fungi. *PLoS One* 10, e0133085.

(31) (2008) CLSI. *Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi; Approved Standard*, 2nd ed., Clinical and Laboratory Standards Institute, Wayne, PA.

(32) Vishniac, W., and Santer, M. (1957) The thiobacilli. *Bacteriol. Rev.* 21, 195–213.

(33) Creaser, E. H., Porter, R. L., Britt, K. A., Pateman, A. J., and Doy, C. H. (1985) Purification and preliminary characterization of alcohol dehydrogenase from *Aspergillus nidulans*. *Biochem. J.* 225, 449–454.

(34) Oakley, C. E., Edgerton-Morgan, H., and Oakley, B. R. (2012) Tools for manipulation of secondary metabolism pathways: rapid promoter replacements and gene deletions in *Aspergillus nidulans*. *Methods Mol. Biol.* 944, 143–161.

(35) Chiang, Y. M., Oakley, C. E., Ahuja, M., Entwistle, R., Schultz, A., Chang, S. L., Sung, C. T., Wang, C. C., and Oakley, B. R. (2013) An efficient system for heterologous expression of secondary metabolite genes in *Aspergillus nidulans*. *J. Am. Chem. Soc.* 135, 7720–7731.

(36) Dunne, P. W., and Oakley, B. R. (1988) Mitotic gene conversion, reciprocal recombination and gene replacement at the benA, beta-tubulin, locus of *Aspergillus nidulans*. *Mol. Gen. Genet.* 213, 339–345.

(37) Bok, J. W., Chiang, Y. M., Szewczyk, E., Reyes-Dominguez, Y., Davidson, A. D., Sanchez, J. F., Lo, H. C., Watanabe, K., Strauss, J., Oakley, B. R., Wang, C. C., and Keller, N. P. (2009) Chromatin-level regulation of biosynthetic gene clusters. *Nat. Chem. Biol.* 5, 462–464.