Project CP GonzaloCardenalbrian ex oscillations

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1 Oscillations in an interneuron network

Oscillations are thought to play a critical role in coordinating brain activity and information transfer. One potential mechanism for their generation is via delayed synaptic coupling in neuronal networks.

In this exercise you will implement a network of inhibitory interneurons driven by excitatory Poisson spiking and examine its dynamics.

```
[1]: # !pip install brian2
```

```
[2]: import brian2 as b2
import numpy as np
import matplotlib.pyplot as plt
import scipy
```

1.1 Task 0: testing your Brian implementation for a single neuron

Implement a single conductance-based leaky integrate-and-fire neuron in Brian!

The membrane potential is described by the following ODE:

$$C\frac{d}{dt}V(t) = g_{\text{leak}}(E_{\text{rest}} - V(t)) + I_{\text{syn}}(t)$$
$$I_{\text{syn}}(t) = g_e(t) (E_e - V(t)) + g_i(t) (E_i - V(t))$$

If V exceeds a threshold V_t it is reset to V_r and remains fixed for a refractory period $\tau_{\rm ref}$.

The neuron can receive both excitatory and inhibitory input. For both types of synapses the postsynaptic conductance in response to a single presynaptic spike at time 0 should be modeled as a dual exponential:

$$g(t) = g_{\text{peak}} s \left[e^{-(t-\tau_l)/\tau_d} - e^{-(t-\tau_l)/\tau_r} \right] \Theta(t-\tau_l)$$

$$s = \left(\max \left(e^{-t/\tau_d} - e^{-t/\tau_r} \right) \right)^{-1} = \left(\left(\frac{\tau_d}{\tau_r} \right)^{\frac{\tau_r}{\tau_r - \tau_d}} - \left(\frac{\tau_d}{\tau_r} \right)^{\frac{\tau_d}{\tau_r - \tau_d}} \right)^{-1} \quad \text{normalizing factor}$$

where τ_l, τ_r, τ_d are the synaptic delay, rise time constant and decay time constant resp.

Analytically one can derive (*Bonus exercise) a set of ODEs describing g(t):

$$\begin{split} \dot{g}(t) &= \frac{-g + x}{\tau_d} \\ \dot{x}(t) &= -\frac{x}{\tau_r} \\ x(\tau_l) &= g_{\rm peak} s \frac{\tau_d - \tau_r}{\tau_r} \quad \text{initial value, } x(t) = 0 \text{ for } 0 < t < \tau_l \end{split}$$

- (a) Implement a single such neuron in Brian.
- (b) Test your implementation of the conductance: Using either an inhibitory or excitatory synapse, connect your neuron to a single presynaptic unit that sends a single input spike. Plot the resulting conductance, synaptic current and membrane potential. Compare the numerical conductance trace to the analytical expression for g(t) and check the peak conductances. Hints: Use Brian's "SpikeGeneratorGroup" (see tutorial). Identify the "on_pre" condition needed for the Brian synapse in the equations given above.

Use the following parameters:

$$g_{
m leak} = 10 \
m nS$$
 $C = 100 \
m pF$ $E_{
m rest} = -65 \
m mV$ $E_e = 0 \
m mV$ $V_t = -75 \
m mV$ $V_t = -52 \
m mV$ $V_r = -67 \
m mV$ $V_{
m ref} = 1 \
m ms$

Excitatory synapses:

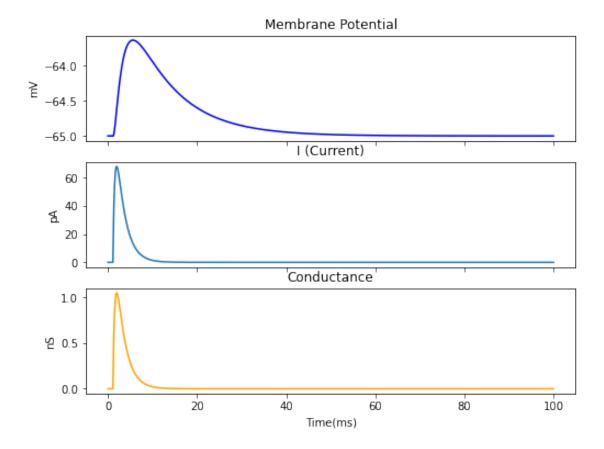
$$au_l = 1 ext{ ms}$$
 $au_r = 0.5 ext{ ms}$
 $au_d = 2 ext{ ms}$
 $ag_{ ext{peak}} = 1 ext{ nS}$

Inhibitory synapses:

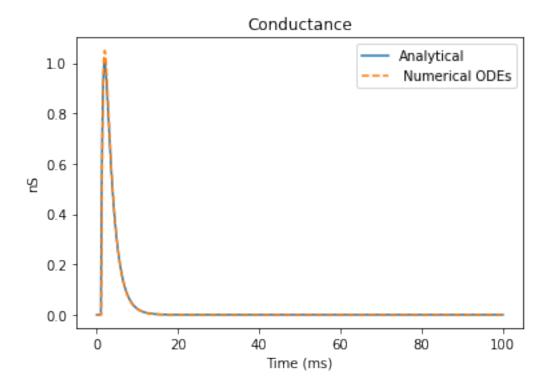
$$au_l = 1 ext{ ms}$$
 $au_r = 0.45 ext{ ms}$
 $au_d = 1.2 ext{ ms}$
 $au_{ ext{peak}} = 5 ext{ nS}$

```
[3]: #### a) implementing the neuron
     b2.start_scope()
     gleak = 10*b2.nS
     C = 100*b2.pF
     Em = -65*b2.mV
     Ee = 0*b2.mV
     Ei = -75*b2.mV
     Vt = -52*b2.mV
     Vr = -67*b2.mV
     tref = 1*b2.ms
     #excitatory parameters
     gepeak = 1*b2.nS
     tde = 2*b2.ms
     tre = 0.5*b2.ms
     tle = 1*b2.ms
     #inhibitory parameters
     gipeak = 5*b2.nS
     tdi = 1.2*b2.ms
     tri = 0.45*b2.ms
     tli = 1*b2.ms
     se = ((tde/tre)**(tre/(tre-tde))-(tde/tre)**(tde/(tre-tde)))
     si = ((tdi/tri)**(tri/(tri-tdi))-(tdi/tri)**(tdi/(tri-tdi)))
     eqs= '''
     dv/dt = (gleak*(Em - v) + Isyn)/C : volt (unless refractory)
     Isyn = (ge*(Ee -v) + gi*(Ei-v)) : amp
     dge/dt = (-ge + x_e)/tde : siemens
     dx_e/dt = -x_e/tre : siemens
     dgi/dt = (-gi + x_i)/tdi : siemens
     dx_i/dt = -x_i/tri : siemens
     #initial x(tl)
     J_e = gepeak * ((tde-tre)/tre)/se
     J_i =gipeak*((tdi-tri)/tri)/si
     x_e ='''
     x_e += J_e
     1.1.1
     x_i='''
     x_i += J_i
     T_{i},T_{i},T_{i}
```

```
G = b2.NeuronGroup(1, eqs, method='euler', threshold='v>Vt', reset ='v = Vr',
     →refractory=tref)
    G.v = Em
    #### b) Testing implementation of the conductance
    Gexc = b2.NeuronGroup(2, eqs, method='euler', threshold='v>Vt', reset ='v = u
     Gexc.v = Em
    #sending a single input spike
    times = [0*b2.ms]
    indices = np.zeros(len(times))
    SGG = b2.SpikeGeneratorGroup(1, indices, times)
    #Excitatory Synapse
    Sexc = b2.Synapses(SGG, Gexc, on_pre= x_e, delay= tle)
    Sexc.connect()
    # State monitors
    M = b2.StateMonitor(Gexc, ('v', 'Isyn', 'ge'), record=True)
    #spikemon = b2.SpikeMonitor(G)
    #run
    print('Before v = %s' % Gexc.v[0])
    b2.run(100*b2.ms)
    print('After v = %s' % Gexc.v[0])
    Before v = -65. mV
    After v = -64.99987088 \text{ mV}
[4]: fig, ax = plt.subplots(3, sharex=True, figsize=(8,6))
    ax[0].plot(M.t/b2.ms, M.v[1]/b2.mV, 'b')
    ax[0].set(title = 'Membrane Potential', ylabel='mV')
    ax[1].plot(M.t/b2.ms, M.Isyn[1]/b2.pA)
    ax[1].set(title = 'I (Current)', ylabel='pA')
    ax[2].plot(M.t/b2.ms, M.ge[1]/b2.nS, 'orange')
    ax[2].set(title = 'Conductance', ylabel='nS', xlabel='Time(ms)');
```



```
[5]: def ge(t):
             neg_exp = lambda tau : np.exp(-(t*b2.ms-tle)/tau)
             return gepeak * (neg_exp(tde) - neg_exp(tre)) * np.heaviside(t*b2.ms_
     \rightarrow-tle, 0)/se
     time = np.linspace(0,100, 10000)
     plt.plot(time,ge(time)/b2.nS, label = 'Analytical')
     plt.plot(M.t/b2.ms, M.ge[1]/b2.ns, '--',label = ' Numerical ODEs', )
     plt.gca().set(title='Conductance', xlabel ='Time (ms)', ylabel='nS')
    plt.legend();
    WARNING
               /Users/gonuni/opt/anaconda3/envs/brian2/lib/python3.6/site-
    packages/brian2/units/fundamentalunits.py:1025: UserWarning: Unknown ufunc
    'heaviside' in __array_prepare__
      warn("Unknown ufunc '%s' in __array_prepare__" % uf.__name__)
     [py.warnings]
               /Users/gonuni/opt/anaconda3/envs/brian2/lib/python3.6/site-
    WARNING
    packages/brian2/units/fundamentalunits.py:1060: UserWarning: Unknown ufunc
    'heaviside' in __array_wrap__
      warn("Unknown ufunc '%s' in __array_wrap__" % uf.__name__)
     [py.warnings]
```



Comparing the numerical computed conductance to the analytical (g(t)) we can see both achieve peak conductance as 1 as expected from the value gpeak = 1nS we set up. If we observe close numerical peak conductance goes a bit higher than 1 but this difference is negligible

1.2 Task 1: Setting up the network

Create a network of 200 of the above described units. Introduce inhibitory synapses between the units with a connection probability of 20% (i.e. the units are inhibitory interneurons).

Adding excitatory drive: Create a second population of 8000 pyramidal cells that fire Poisson spike trains at a rate of 6Hz each (Hint: Brian "PoissonGroup"). Add excitatory synapses from this pyramidal cell population onto your interneuron network with a connection probability of 10%. Comprehension question: What is the total rate of excitatory input that a single interneuron receives on average?

Add State/Spike/PopulationRate Monitors as you see fit (see also task 2). Wrap all your code into one reusable function!

```
[12]: #Now we make our network fuction for next tasks
def network(Ninter, Npir, frates, runtime):
    b2.start_scope()

gleak = 10*b2.nS
    C = 100*b2.pF
    Em = -65*b2.mV
```

```
Ee = 0*b2.mV
   Ei = -75*b2.mV
   Vt = -52*b2.mV
   Vr = -67*b2.mV
   tref = 1*b2.ms
   #excitatory parameters
   gepeak = 1*b2.nS
   tde = 2*b2.ms
   tre = 0.5*b2.ms
   tle = 1*b2.ms
   #inhibitory parameters
   gipeak = 5*b2.nS
   tdi = 1.2*b2.ms
   tri = 0.45*b2.ms
   tli = 1*b2.ms
   se = ((tde/tre)**(tre/(tre-tde))-(tde/tre)**(tde/(tre-tde)))
   si = ((tdi/tri)**(tri/(tri-tdi))-(tdi/tri)**(tdi/(tri-tdi)))
   eqs= '''
   dv/dt = (gleak*(Em - v) + Isyn)/C : volt (unless refractory)
   Isyn = (ge*(Ee -v) + gi*(Ei-v)) : amp
   dge/dt = (-ge + x_e)/tde : siemens
   dx_e/dt = -x_e/tre : siemens
   dgi/dt = (-gi + x_i)/tdi : siemens
   dx_i/dt = -x_i/tri : siemens
   #initial x(tl)
   J_e = gepeak * ((tde-tre)/tre)/se
   J_i =gipeak*((tdi-tri)/tri)/si
   x_e = 111
   x_e += J_e
   1.1.1
   x_i=
   x_i += J_i
#Interneurons
   Gint = b2.NeuronGroup(Ninter, eqs, method='euler', threshold='v>Vt', reset⊔
\rightarrow = 'v = Vr', refractory=tref)
   Gint.v = Em
```

```
#Pyramidal Poisson Group
    Gpir = b2.PoissonGroup(Npir, rates=frates*b2.Hz)

#Interneuron Synapses
    Sint = b2.Synapses(Gint, Gint, on_pre= x_i, delay= tle)
    Sint.connect(p = 0.2)

#Pyramidal cells to Interneuon Synapses
    Spyr = b2.Synapses(Gpir, Gint, on_pre = x_e, delay= tle)
    Spyr.connect(p = 0.1)

# State monitors
    M = b2.StateMonitor(Gint, ('v','Isyn', 'ge', 'gi'), record=True)
    spikemon = b2.SpikeMonitor(Gint)
    ratemon = b2.PopulationRateMonitor(Gint)

#run
    b2.run(runtime*b2.ms)

return M, spikemon, ratemon, runtime, frates
```

The total rate of excitatory input that a single interneuron receives is:

$$6Hz(rate)*0.1(connectivity)*\frac{8000(pyramidalneurons)}{200(interneurons)}=24Hz$$

1.3 Task 2: Simulating and visualizing the network activity

Run your network simulation (pick a reasonable simulation time).

- (a) Show the interneuron network's spiking activity and population rate over time. In addition plot the membrane potential of one or two example interneurons.
- (b) How are the firing rates of individual interneurons distributed?
- (c) Quantify the regularity of interneuron firing using the coefficient of variation of interspike-intervals.

(Bonus) Simulate your network for at least 1050 ms (the longer the better). Use the function get_PSD(signal, dt) given below to calculate the power spectral density of the population rate and plot it. Hint: leave the keyword arguments of the function as they are, remember to provide the simulation time step in SECONDS!

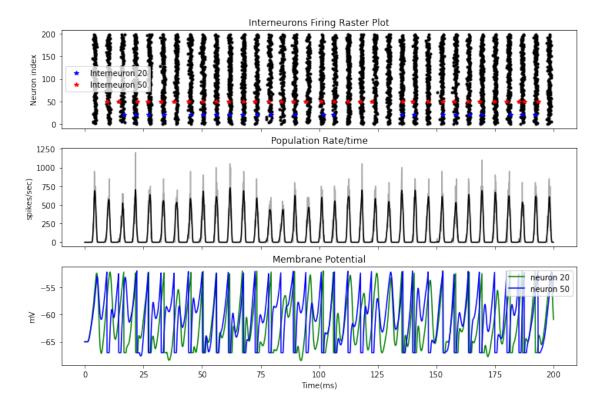
Summarize your results to describe the network state that you observe.

```
[13]: statemet, spikenet, ratemet, runtime, frate6 = network(200, 8000, 6, 200)
```

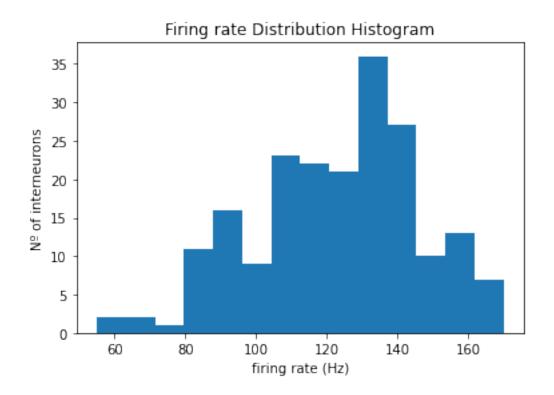
WARNING "dt" is an internal variable of group "poissongroup", but also exists in the run namespace with the value 0.0001. The internal variable will be used. [brian2.groups.group.Group.resolve.resolution conflict]

```
[14]: # we define a plotting fuction as we are going to repeat the task in task 3
     def plotting(statenet, spikenet, ratenet, runtime, frates):
         fig, ax = plt.subplots(3, sharex=True, figsize=(12,8))
         ax[0].plot(spikenet.t/b2.ms, spikenet.i, '.k')
         ax[0].plot(spikenet.t[spikenet.i==20]/b2.ms, spikenet.i[spikenet.i==20],__
      ax[0].plot(spikenet.t[spikenet.i==50]/b2.ms, spikenet.i[spikenet.i==50],__
      →'*r', label = 'Interneuron 50')
         ax[0].set(title = 'Interneurons Firing Raster Plot', ylabel='Neuron index')
         ax[0].legend()
         ax[1].plot(ratenet.t/b2.ms, ratenet.rate/b2.Hz, 'darkgrey')
         ax[1].plot(ratenet.t/b2.ms, ratenet.smooth_rate(width=.2*b2.ms)/b2.Hz, 'k')
         ax[1].set(title = 'Population Rate/time', ylabel='spikes/sec)')
         ax[2].plot(statenet.t/b2.ms, statenet.v[20]/b2.mV, 'g', label= 'neuron 20')
         ax[2].plot(statenet.t/b2.ms, statenet.v[50]/b2.mV, 'b', label= 'neuron 50')
         ax[2].set(title = 'Membrane Potential', ylabel='mV', xlabel='Time(ms)');
         ax[2].legend()
         plt.suptitle(str(frates) + 'Hz Frequency', size = 15)
     sixHzFRnt = plotting(statenet, spikenet, ratenet, runtime, frate6)
```

6Hz Frequency



```
[37]: def firing_rates_dis(spikenet, runtime, nbins):
          #plotting an histogram for visualize interneuron's firing rate distribution
          INfr = spikenet.count/runtime
          plt.hist(INfr*1000 , bins = nbins)
          plt.gca().set(title = 'Firing rate Distribution Histogram', xlabel= 'firing⊔
       →rate (Hz)', ylabel='Nº of interneurons')
          plt.show()
          #coefficient of variation of interspike-intervals
          spikeintervals = 1/INfr
          spikeintervalslen = spikeintervals.shape[0]
          for i in np.arange(spikeintervalslen):
              if spikeintervals[i-1] == float("inf"):
                  spikeintervals[i-1] = 1e-12
          cv = np.std(spikeintervals)/np.mean(spikeintervals)
          print("The mean of interneurons FR is " +str(np.mean(INfr*1000))+"Hz")
          print("The coefficient of variation of interspike-intervals is " + str(np.
       \rightarrowround(cv,2)))
          return cv
      cvSINt= firing_rates_dis(spikenet, runtime, 14)
```



The mean of interneurons FR is 123.15Hz
The coefficient of variation of interspike-intervals is 0.23

We can see the firing rate distribution of our interneuron network above.

```
[56]: statenetlong, spikenetlong, ratenetlong, runtimelong, frate6long = network(200, ⊔ →8000, 6, 1050)
```

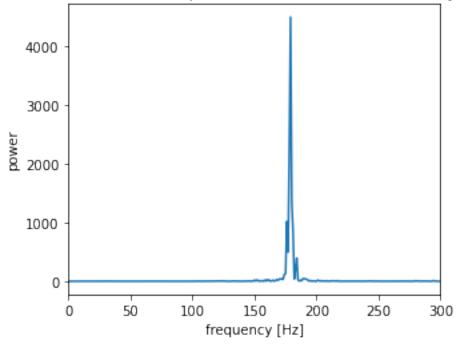
```
[61]: def get_fft(signal, dt):
    """
    returns scipy fast fourier transform with correction for the time step dt
    dt: [sec]
    """
    return dt*scipy.fft.fft(signal)

def get_PSD(signal, dt, df=1, offset=0.05, subtract_mean=True, eps=0.1):
    """
    Computes the power spectral density of the signal
    Input:
        signal: [Hz] typically the population rate
        dt: [sec] simulation time step
        df: [Hz] desired frequency resolution of the PSD
        offset: [sec] signal in the time interval [0, offset] is removed before
        →doing the Fourier transform.
```

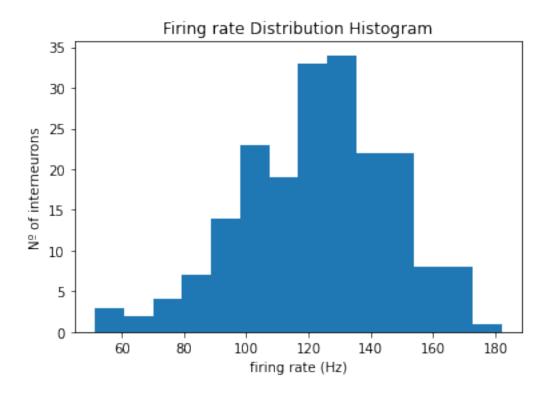
```
Use this parameter to ignore the initial transient signals of \Box
\hookrightarrow the simulation.
       subtract_mean (bool): If true, the mean value of the signal is_
\hookrightarrow subtracted.
   Returns:
       freqs: range of frequencies for which psd is given (resolution: df)
             averaged power spectral density for the frequency range given by \Box
\hookrightarrow freqs
   nnn
   f_{max} = np.around(1/(2 * dt)) # maximal frequency of PSD
   N_init = int(offset/dt) # offset in array steps
   N snippet = int(2 * f max / df) # required length of one snippet for
→resolution df (in array length)
   N signal = len(signal)
   T_snippet = N_snippet*dt # length of snippets in units of time [seconds]
   k = N_signal//N_snippet # see how many snippets we can average given the
\rightarrow signal length
   N_required = k * N_snippet + N_init
   # crop the signal if necessary
   if not k or (N_signal < N_required):</pre>
       raise ValueError('Signal length not sufficient for PSD calculation, u
→simulate for at least {:.2f}seconds!'.format(T_snippet+offset))
   if N signal > N required:
       print("PSD: drop samples")
       signal = signal[:N required]
   signal = signal[N_init:]
   print("\n Now using {} out of {:.2f} signal seconds for PSD calculation ⊔
\rightarrow (df={}Hz, k={})".format(len(signal)*dt, N_signal*dt, df, k))
   average signal = np.mean(signal)
   D = np.var(signal)/2
   if subtract mean:
       signal = signal - average_signal
   signal = signal.reshape(k, N_snippet) # reshape into one row per snippet⊔
\hookrightarrow (k)
   psd_snippetwise = np.abs(get_fft(signal, dt))**2
   psd = np.mean(psd_snippetwise, axis=0)
   # normalize
   psd /= T_snippet
   # crop
   psd = psd[:int(N_snippet/2)]
   freqs = np.arange(0, f_max, df)
   # sanity check
```

Now using 1.0 out of 1.05 signal seconds for PSD calculation (df=1Hz, k=1)





```
[58]: cvSINt= firing_rates_dis(spikenetlong, runtimelong, 14)
```



The mean of interneurons FR is 122.88571428571427Hz
The coefficient of variation of interspike-intervals is 0.24

Summery With 6Hz of excitatory input from the Poisson Group the interneuron network exhibits synchronized oscillations with a firing rate mean around 120 Hz in the longer simulation. We can confirm at this FR(6Hz) the interneurons act synchronously. However the frequency interval of ripples are in 140-220Hz and here the network is exhibiting a fast-gamma episode (90-140Hz). Moreover, in the longer simulation the FR mean stablises in a lower value. This phenomena could be representing the IFA (intraripple frequency accommodation that the paper talks about). This coefficient of variation of the inter-spike interval is directly related with this asyncronic behaviour explining its higher value in longer simulations.

1.4 Task 3: Varying the excitatory drive

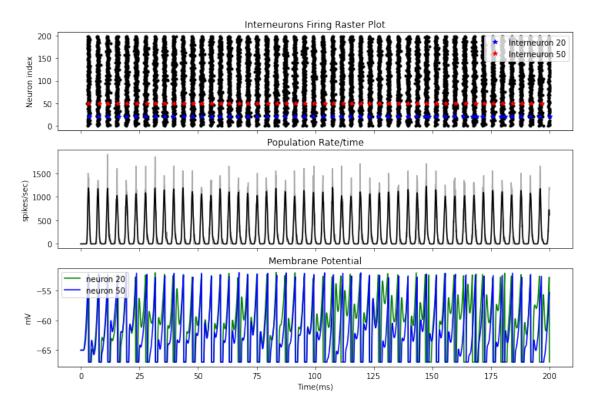
Repeat task 2 but change the Poisson firing rates of the excitatory input population to

- (a) 16 Hz
- (b) 2 Hz

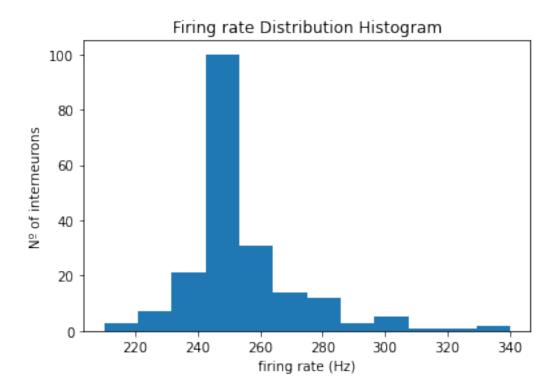
How do the network dynamics change?

```
[42]: #a)
statenet16, spikenet16, ratenet16, runtime, frate16 = network(200, 8000, 16, □
→200)
```

16Hz Frequency



[49]: cvSINt16= firing_rates_dis(spikenet16, runtime, 12)

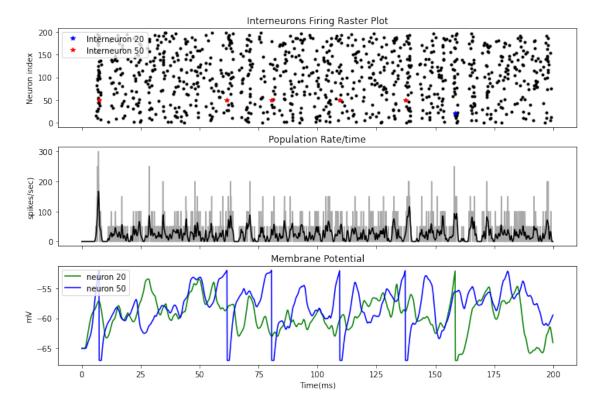


The mean of interneurons FR is 253.2Hz
The coefficient of variation of interspike-intervals is 0.07

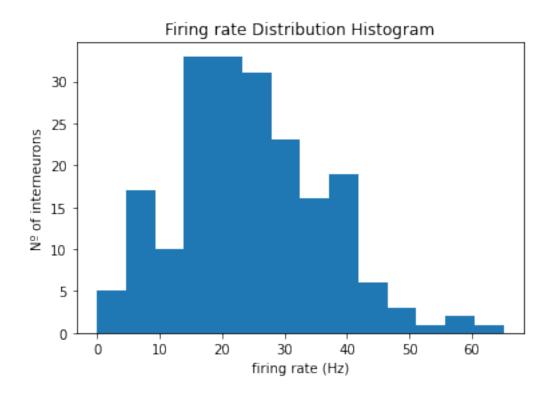
How do the network dynamics change? For a higher FR of the excitatory input (16Hz) the interneuron network exhibits fully synchronized oscillations corresponding with hippocampus ripples. The FR mean is exceeding the 240Hz higher bound of the ripples frequency interval the paper mention (140-220 Hz), smaller excitatory input rate is sufficent for achiving these ripples. Moreover, the cv of the interspike-intervals reduces due to this higher synchronicity.

```
[24]: #b)
statenet2, spikenet2, ratenet2, runtime, frate2 = network(200, 8000, 2, 200)
twoHzFRnt = plotting(statenet2, spikenet2, ratenet2, runtime, frate2)
```

2Hz Frequency



[46]: cvSINt2= firing_rates_dis(spikenet2, runtime, 14)



WARNING /Users/gonuni/opt/anaconda3/envs/brian2/lib/python3.6/site-packages/ipykernel_launcher.py:8: RuntimeWarning: divide by zero encountered in true_divide

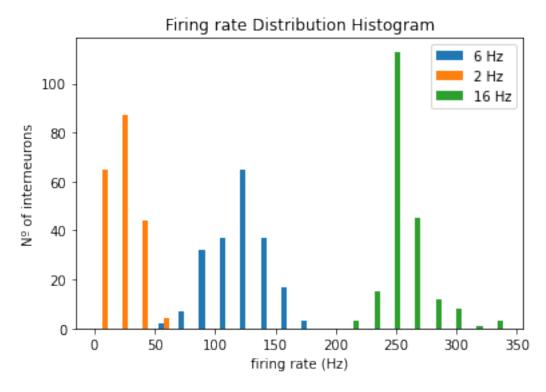
[py.warnings]

The mean of interneurons FR is 23.925Hz
The coefficient of variation of interspike-intervals is 0.84

How do the network dynamics change? For low FR of the excitatory input (2Hz) the interneuron network exhibits sparsel synchronized oscillations. A CV > 0.5 corresponds to a high-noise state of the network with units fire irregularly.

The amount of excitatory drive can modulate the oscillatory state of our interneuron network. At low input rates, the network expresses sparsely synchronized oscillations. With high input rates, the network transits towards a fully synchronized regime. Therefore, we can conclude synchronization depends on the excitatory input rate.

```
[44]: spikenetjoin = spikenet, spikenet2, spikenet16
Ninter = 200
frates = ['6 Hz', '2 Hz', '16 Hz']
def firing_rates_dis_join(spikenet, runtime, nbins, Ninter, frates):
    #plotting an histogram for visualize interneuron's firing rate distribution
    INfrjoin = []*Ninter*len(spikenet)
    for i in range(len(spikenet)):
```



Here we can observe the distribution of the 3 simulations with different excitatory input rate

1.5 Task 4: Reading

Read the paper by Donoso et al. (2018). It is a long paper, so you can focus mainly on the introduction and the first 3 pages of the Results and Figure 1. What kinds of oscillations in what brain area does this model attempt to explain? What is the potential functional role of these oscillations? Later on in the paper, how do the authors use the model to understand the effect of GABA modulators?

1.5.1 Paper comprehension and questions

The paper focus in explaning the relaying mechanism of Hippocampal ripples for memory consolidation.

There is two possible mechanisms one direct via Schaffer collaterals providing input from CA3 to CA1 in the Hippocampus and another indirect via local pyramidal cells in CA1. The direct leads to ripples and the indirect to the fast-gamma episodes both performed by the same interneuron network. Ripples are generated during deep sleep for hippocampus-dependent memory consolidation while gamma oscillations occur during locomotor activities at which sensory information is temporarily stored in the hippocampus.

Regarding the effect of GABA modulators and the in silico simulations with the model, authors wanted to check the "in vivo" observations of the GABA modulators on the ripple frequency; "The oscillation frequency of ripples in vitro is remarkably resistant to drugs that alter the time constant and peak conductance of GABAergic synaptic transmission". For that they simulate the action of the drug by modifying the peak conductance and the decay time constant of inhibitory conductances. Authors concluded that the effect of simulated GABAergic modulators confirmed the experimental results. Their model explained why drugs that induce an increase in decay time constant of GABAergic synaptic transmission can reduce ripple duration and interneuron firing rates without affecting ripple frequency.

1.6 Bonus:

Derive analytically the ODEs that you were given above for the numerical integration of the conductance (see Task 0).

Hint: Start by taking the derivative of the analytical expression for q(t) given a single spike.

1.7 References

Donoso, Schmitz, Maier, Kempter, "Hippocampal ripple oscillations and inhibition-first network models: Frequency dynamics and response to GABA modulators", J Neurosci, 2018

Brunel, Hakim, "Fast Global Oscillations in Networks of Integrate-and-Fire Neurons with Low Firing Rates", Neural Comput, 1999