# BIOS 7720: Applied Functional Data Analysis

Lecture 11: Function on Scalar Regression (cont)

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#### **FoSR**

$$egin{aligned} & \mathsf{y}_i(s) = f_0(s) + f_1(s) \mathsf{x}_i + b_i(s) + \epsilon_i(s) \ & b_i(s) \sim \mathsf{GP}(0, \mathbf{\Sigma}_b) \ & \epsilon_i(s) \stackrel{\mathsf{iid}}{\sim} \mathcal{N}(0, \sigma_\epsilon^2) \end{aligned}$$

- Last time we saw how to estimate using an iterative procedure
- Applied to the NHANES data

#### FoSR

- Iterative procedure:
  - Estimate the working model under independence
  - Fit fpca to the residuals
  - Extract the eigenfunctions
  - Fit the "weak" oracle model
  - Repeat 1-4 as necessary

Consider the model

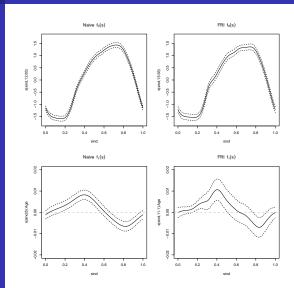
$$\mathsf{IAC}_i(s) = f_0(s) + f_1(s)\mathsf{Age}_i + b_i(s) + \epsilon_i(s)$$

• Problem: data size is huge

```
df <- readr::read_rds(here::here("data","data_processed","NHANES_AC_processed.rds"))
## extract the PA data
1X <- log(1+as.matrix(df[,paste0("MIN",1:1440)]))</pre>
1X[is.na(1X)] <- 0
N <- nrow(1X)
## bin the data into 60 minute intervals
tlen <- 60
nt <- ceiling(1440/tlen)
inx cols <- split(1:1440, rep(1:nt, each=tlen)[1:1440])
1X_bin <- vapply(inx_cols, function(x) rowMeans(1X[,x], na.rm=TRUE), numeric(N))</pre>
## get subject average curves
inx_rows <- split(1:N, factor(df$SEQN, levels=unique(df$SEQN)))
1X_bin_ind <- t(vapply(inx_rows, function(x) colMeans(1X_bin[x,], na.rm=TRUE), numeric(nt)))</pre>
nid <- nrow(1X_bin_ind)
# get a data frame for model fitting
sind <- seq(0,1,len=nt)
df_fit <-
  data.frame(lAC=as.vector(t(lX_bin_ind)),
             sind = rep(sind, nid),
             SEQN = rep(unique(df$SEQN), each=nt)) %>%
  left_join(dplyr::select(df[!duplicated(df$SEQN),], SEQN, Age), by="SEQN") %>%
  mutate(id = factor(SEQN)) %>%
  filter(!is.na(Age))
```

```
## subset the data to just 500 participants
set.seed(10110)
nid_samp <- 500
id_samp <- sample(unique(df_fit$id), size=nid_samp, replace=FALSE)</pre>
df_fit_sub <- subset(df_fit, id %in% id_samp)</pre>
## fit the naive model
fit_naive <- bam(1AC ~ s(sind, bs="cc",k=20) + s(sind, by=Age, bs="cc",k=20),
                 method="fREML",data=df_fit_sub, discrete=TRUE)
## extract the resindals
resid_mat <- matrix(fit_naive$residuals,
                    nid_samp, nt,byrow=TRUE)
## fit fpca
fpca_fit <- fpca.face(resid_mat, knots=15)</pre>
## add in eigenfunctiosn
for(k in 1:length(fpca_fit$evalues)){
    df_fit_sub[[paste0("Phi",k)]] <- rep(fpca_fit$efunctions[,k],nid_samp)</pre>
```

- Plot the estimated  $\hat{f}_0(s)$  and  $\hat{f}_1(s)$  from the naive and FRI fits
- Compare the shapes and CIs
- Do these results make sense?



- Problem: functional random intercept models computationally expensive
  - RAM
  - Time
- One possible solution
  - Choose smoothing parameter(s) for fixed effects using leave-several-out CV MSE estimated using the naive model
  - Estimate standard errors by bootstrapping curves
    - ullet Fix  $\lambda$  at the value obtained from using CV MSE
    - ullet Estimate  $\lambda$  at each stage

#### In-Class Exercise

- Obtain optimal smoothing parameters for the varying coefficient model using leave-one-function out CV MSE (you may use a relatively coarse grid for  $\log(\lambda)$  for faster computation time). If you find your implementation is too slow for whatever reason, you can implement leave-several-function out CV MSE instead.
- ② You may fix  $\lambda_{f_0}$  at it's estimated value from the FRI fit (otherwise you'd need to do a bivariate grid search)
- Given the smoothing parameter you found, estimate  $SE(\hat{f}_1(s))$  by bootstrapping curves (you may use a relatively small number of bootstrap samples, 50-100)
- Plot the results, compare to the naive and FRI model results

### In-Class Exercise: Some Helpful Code

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```
## get indices for the leave-function(s)-out CV
# get ungiue participant IDs
uid <- unique(df_fit_sub$SEQN)
nid <- length(uid)
# number of folds to split the participants
nfolds <- 5
# set the seed
set.seed(1010)
# create a vector of fold indicators
fold_vec <- sample(rep(1:nfolds, ceiling(nid/nfolds))[1:nid])</pre>
# combine with IDs
df_folds <- data.frame(SEQN=uid, fold=sample(1:nfolds))</pre>
# merge with the data frame for fitting
df_fit_sub <-
 left_join(df_fit_sub, df_folds, by="SEQN")
# create a list with each element as the vector of row indices
# associated with each fold
inx_ls <- split(1:nrow(df_fit_sub), df_fit_sub$fold)</pre>
```

#### In-Class Exercise: Some Helpful Code

```
## set up grid search (here do a coarse search for computation time)
nlambda <- 25
loglambda <- seg(5,20,len=nlambda)
lambda <- exp(loglambda)
## set up progress bar
pb <- txtProgressBar(min=0,max=nlambda*nfolds,style=3)
inx pb <- 1
## loop over candidate smoothing parameters
mse <- rep(NA, nlambda)
## get smoothing parameters from the FRI model
sp fri <- fit fri$sp[1:2]
for(1 in 1:nlambda){
 ## loop over "folds"
 mse_1 <- rep(NA, nfolds)
 for(i in 1:nfolds){
   # get the training and test data
   inx_li <- inx_ls[[i]]
   df_train <- df_fit_sub[-inx_li,]
   df_test <- df_fit_sub[inx_li,]
   # fit the model using the training data
   fit_li <- bam(1AC ~ s(sind, bs="cc",k=20,sp=sp_fri[1]) +
                         s(sind, by=Age, bs="cc",k=20,sp=lambda[1]),
                    data=df train)
    # predict on the test data
   fhat_test <- predict(fit_li, newdata=df_test, type='response')</pre>
   mse_1[i] <- mean((fhat_test-df_test$1AC)^2)
   ## update progress bar
   setTxtProgressBar(pb, inx_pb)
   inx_pb <- inx_pb + 1
  # average over the folds for this candidate smoothing parameter
 mse[1] <- mean(mse 1)
```