

# Package

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**Type** Package

**Title** CUTENESS: An R package to cluster gene expression by contour plots

**Version** 0.1.0

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**Description** The CUTENESS package allows to analyse toxicogenomics gene expression dataset with multiple dose levels and time-points. It allows to identify the expression patterns with respect to both variables and to cluster genes accordingly. It also identify enriched pathways/go terms that are associated to each cluster.

**Depends** R (>= 3.4),

stats,  
utils,  
AnnotationDbi,  
gProfileR,  
gtools,  
reshape,  
plotly,  
clv,  
gplots,  
org.Hs.eg.db,  
org.Mm.eg.db,  
org.Rn.eg.db,  
xlsx

**License** GPL (>= 3)

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 6.1.1

**Suggests** knitr,  
rmarkdown,  
testthat

**VignetteBuilder** knitr

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build_items_list	<i>This function computes the venn diagram of the genes associated to time, dose or their interaction</i>
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## Description

This function computes the venn diagram of the genes associated to time, dose or their interaction

## Usage

```
build_items_list(PvalMat, p.val.th = 0.01)
```

## Arguments

PvalMat	matrix with pvalue associated to the dose, timepoint and the dose*timepoint effect that is the output of the compute_anova_dose_time function
p.val.th	is the threshold at which p.values are considered significant. Default = 0.01

## Value

a list containing the genes in each position of the venn diagram

## Examples

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data, dose_index = 2, time_point_index = 3)
ItemsList = build_items_list(PvalMat)
```

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clustering_summary	<i>This function evaluate the summary index for a clustering results by taking into account the within and between variability</i>
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**Description**

This function evaluate the summary index for a clustering results by taking into account the within and between variability

**Usage**

```
clustering_summary(DB, cluster)
```

**Arguments**

DB	your matrix dataset
cluster	is a numeric vector of clustering results

**Value**

a vector of evaluation indexes

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compute_anova_dose_time	<i>This function computes a two way anova between dose and time for the expression value of every genes</i>
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**Description**

This function computes a two way anova between dose and time for the expression value of every genes

**Usage**

```
compute_anova_dose_time(exp_data, pheno_data, dose_index, time_point_index)
```

**Arguments**

exp_data	is the expression matrix with genes on the rows and samples on the columns
pheno_data	is a dataframe with phenodata informations. Samples are on the rows. The columns should include the dose and time point information.
dose_index	numeric value specifying the column of the phenodata table containing the doses
time_point_index	numeric value specifying the column of the phenodata table containing the time points

**Value**

a matrix with pvalue associated to the dose, timepoint and the dose\*timepoint effect

## Examples

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data, dose_index = 2, time_point_index = 3)
```

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compute_enrichment	<i>This function perform enrichment of the genes in each cluster</i>
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## Description

This function perform enrichment of the genes in each cluster

## Usage

```
compute_enrichment(optimal_clustering, corrType = "fdr",
  type_enrich = "KEGG", org_enrich = "rnorvegicus", pth = 0.05,
  sig = FALSE, mis = 0, only_annotated = FALSE)
```

## Arguments

optimal_clustering	vector of final clustering
corrType	string specifying the algorithm used for determining the significance threshold, one of gSCS, fdr, bonferroni. Default: fdr
type_enrich	string specifying the enrichment type. Default = KEGG
org_enrich	string specifying the organism. Default = rnorvegicus
pth	numeric value specifying the pvalue threshold. Default = 0.05
sig	whether all or only statistically significant results should be returned
mis	minimum size of functional category, smaller categories are excluded
only_annotated	statistical domain size, one of "annotated", "known"

## Value

a list with the enriched pathways for each cluster of genes

## Examples

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data, dose_index = 2, time_point_index = 3)
ItemsList = build_items_list(PvalMat)
responsive_genes = unique(c(unlist(ItemsList$Dose), unlist(ItemsList$Time), unlist(ItemsList$`Dose:Time:Dose`)))
contour_res = create_contour(exp_data, pheno_data, responsive_genes, dose_index = 2, time_point_index = 3, grid = 10)
hls_res = hls_genes_clustering(contour_res$GenesMap, nClust = c(5, 10, 15, 20, 25), method = "pearson", hls.method = "hls")
clpr = create_prototypes(clust_res = hls_res, summaryMat = hls_res$summaryMat, contour_res = contour_res)
enrRes = compute_enrichment(clpr$optcl, corrType = "fdr", type_enrich = "KEGG", org_enrich = "rnorvegicus", pth = 0.05)
```

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convert_genes	<i>This function convert genes identifiers</i>
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**Description**

This function convert genes identifiers

**Usage**

```
convert_genes(organism = "hsapiens", GList, annType = "SYMBOL")
```

**Arguments**

organism	a string specifying the organism under analysis
GList	a list of genes identifier
annType	string specifying the wanted gene identifier

**Value**

a list with the converted genes identifiers

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create_contour	<i>This function fits a 3D regression model for every gene in the dataset and creates an N x N contour plot</i>
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**Description**

This function fits a 3D regression model for every gene in the dataset and creates an N x N contour plot

**Usage**

```
create_contour(exp_data, pheno_data, responsive_genes, dose_index,
               time_point_index, gridSize = 50)
```

**Arguments**

exp_data	is the expression matrix with genes on the rows and samples on the columns
pheno_data	is a dataframe with phenodata informations. Samples are on the rows. The columns should include the dose and time point information.
responsive_genes	responsive_genes character vector with the genes statistically significant for the two-way anova
dose_index	numeric value specifying the column of the phenodata table containing the doses
time_point_index	numeric value specifying the column of the phenodata table containing the time points
gridSize	numeric value specifying size of the z-grid

**Value**

a list with list with estimated contour objects, 3D fitted objects, fitting statistics and feature values for time and dose

GenesMap	a matrix with the z-maps computed for each gene
RPGenes	a list with the 3D fitted objects
Statis	a matrix with the fitting statistics: PValue,Adj.R.Square,RMSE
DFList	a list with the data used for the fitting

**Examples**

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data,dose_index = 2,time_point_index = 3)
ItemsList = build_items_list(PvalMat)
responsive_genes = unique(c(unlist(ItemsList$Dose), unlist(ItemsList$Time), unlist(ItemsList$`Dose:Time:Dose:Time`)))
contour_res = create_contour(exp_data, pheno_data, responsive_genes,dose_index = 2,time_point_index = 3 ,grid = 100)
```

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create_prototypes	<i>this function create the cluster prototypes as the mean values of the z maps of all the genes in the cluster</i>
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---

**Description**

this function create the cluster prototypes as the mean values of the z maps of all the genes in the cluster

**Usage**

```
create_prototypes(clust_res, summaryMat, contour_res)
```

**Arguments**

clust_res	the clustering results object given in input by the function hls_genes_clustering
summaryMat	the matrix with the summary statistics computed for the different k values
contour_res	a list with the contours object returned in output by the create_contour function

**Value**

a list with the contour objects for each clustering prototype and the vector of the optimal clustering

meanXYZ	a list the contour object for each prototype
optcl	a vector with the optimal clustering

## Examples

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data, dose_index = 2, time_point_index = 3)
ItemsList = build_items_list(PvalMat)
responsive_genes = unique(c(unlist(ItemsList$Dose), unlist(ItemsList$Time), unlist(ItemsList$`Dose:Time:Dose`)))
contour_res = create_contour(exp_data, pheno_data, responsive_genes, dose_index = 2, time_point_index = 3, grid_size = 100)
hls_res = hls_genes_clustering(contour_res$GenesMap, nClust = c(5, 10, 15, 20, 25), method = "pearson", hls_method = "clpr")
clpr = create_prototypes(clust_res = hls_res, summaryMat = hls_res$summaryMat, contour_res = contour_res)
```

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enrich

*This function perform enrichment of the genes in each cluster*


---

## Description

This function perform enrichment of the genes in each cluster

## Usage

```
enrich(x, type, org, pval, adjust_method, sig = FALSE, mis = 0,
       only_annotated = TRUE)
```

## Arguments

x	a dataframe with the gene names on the first column
type	string specifying the enrichment type. Default = KEGG
org	string specifying the organism. Default = rnorvegicus
pval	numeric value specifying the pvalue threshold. Default = 0.05
adjust_method	string specifying the algorithm used for determining the significance threshold, one of gSCS, fdr, bonferroni. Default: fdr
sig	whether all or only statistically significant results should be returned
mis	minimum size of functional category, smaller categories are excluded
only_annotated	statistical domain size, one of "annotated", "known"

## Value

a list with the enriched pathways for each cluster of genes

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findCenter	<i>This function select cluster prototypes. It select the prototype as the more correlated with the other elements in the cluster</i>
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---

### Description

This function select cluster prototypes. It select the prototype as the more correlated with the other elements in the cluster

### Usage

```
findCenter(DB, clust_vector)
```

### Arguments

DB	your matrix dataset
clust_vector	a numeric vector of clustering results

### Value

a matrix of prototypes

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hls_genes_clustering	<i>It clusters the contour plots by also estimating the ideal number of clusters</i>
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---

### Description

It clusters the contour plots by also estimating the ideal number of clusters

### Usage

```
hls_genes_clustering(GenesMap, nClust = c(5, 10, 25, 50, 75, 100, 125,
  150, 175, 200, 250, 300), method = "pearson", hls.method = "ward")
```

### Arguments

GenesMap	a matrix with the z-maps computed with the function create_contour
nClust	vector of putative numbers used to determine number of clusters. Default = c(5,10,25,50,75,100,125,150,175,200,250,300)
method	string specifying the correlation method. Default = "pearson"
hls.method	string specifying method for the hierarchical clustering. Default = "ward"

### Value

a list with the clustering results and statistics for the optimal number of clusters

hls_res	a list containing the clustering results, the clustering vector and the centers for each k value given in input
summaryMat	a matrix with summary statistic for each k value given in input
clusterList	a list with the clustering vectors



## Examples

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data,dose_index = 2,time_point_index = 3)
ItemsList = build_items_list(PvalMat)
responsive_genes = unique(c(unlist(ItemsList$Dose),unlist(ItemsList$Time),unlist(ItemsList$`Dose:Time:Dose`)))
contour_res = create_contour(exp_data, pheno_data, responsive_genes,dose_index = 2,time_point_index =3 ,grid = 100)
hls_res = hls_genes_clustering(contour_res$GenesMap, nClust = c(5,10,15,20,25), method="pearson", hls.method="hls")
```

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plot3d	<i>This function plots the fitted 3d surface for the expression value of a gene</i>
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---

## Description

This function plots the fitted 3d surface for the expression value of a gene

## Usage

```
plot3d(toPlot = list(x, y, z), DF)
```

## Arguments

toPlot	is a list containing the predicted value for the x, y and z axis
DF	is the data frame containing the information for the samples used in the fitting process

## Value

a plotly object

## Examples

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data,dose_index = 2,time_point_index = 3)
ItemsList = build_items_list(PvalMat)
responsive_genes = unique(c(unlist(ItemsList$Dose),unlist(ItemsList$Time),unlist(ItemsList$`Dose:Time:Dose`)))
contour_res = create_contour(exp_data, pheno_data, responsive_genes,dose_index = 2,time_point_index =3 ,grid = 100)
plot3d(toPlot = contour_res$RPGenes[["Pdk4"]],DF = contour_res$DFList[["Pdk4"]])
```

---

plot\_clusters\_prototypes

*this function plots the clusters prototype*

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### Description

this function plots the clusters prototype

### Usage

```
plot_clusters_prototypes(meanXYZ, nR = 2)
```

### Arguments

meanXYZ	a list the contour object for each prototype computed with the function create_prototypes
nR	the number of rows to use in the plot

### Examples

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data, dose_index = 2, time_point_index = 3)
ItemsList = build_items_list(PvalMat)
responsive_genes = unique(c(unlist(ItemsList$Dose), unlist(ItemsList$Time),
                           unlist(ItemsList$`Dose:Time:DoseTime`),
                           unlist(ItemsList$`Dose:Time`)))
contour_res = create_contour(exp_data, pheno_data, responsive_genes, \cr
                             dose_index = 2, time_point_index = 3, gridSize = 50)
hls_res = hls_genes_clustering(contour_res$GenesMap, nClust = c(5, 10, 15, 20, 25), \cr
                              method = "pearson", hls.method = "ward")

clpr = create_prototypes(clust_res = hls_res, summaryMat = hls_res$summaryMat, contour_res)
plot_clusters_prototypes(clpr$meanXYZ, nR = 2)
```

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write\_xlsx\_for\_funmappone

*This function create an excel file with the same format of the input need by the FunMappOne tool*

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### Description

This function create an excel file with the same format of the input need by the FunMappOne tool

### Usage

```
write_xlsx_for_funmappone(optimal_clustering,
                          filePath = "../contour_clustering/gene_clustering.xlsx")
```

**Arguments**

`optimal_clustering` is a numeric vector with the clustering result for every gene

`filePath` is a string specifying the path of the xlsx file

**Examples**

```
data("WY14643")
exp_data = WY14643$exp_data
pheno_data = WY14643$pheno_data
PvalMat = compute_anova_dose_time(exp_data, pheno_data, dose_index = 2, time_point_index = 3)
ItemsList = build_items_list(PvalMat)
responsive_genes = unique(c(unlist(ItemsList$Dose), unlist(ItemsList$Time), unlist(ItemsList$`Dose:Time:Dose`)))
contour_res = create_contour(exp_data, pheno_data, responsive_genes, dose_index = 2, time_point_index = 3, grid = 100)
hls_res = hls_genes_clustering(contour_res$GenesMap, nClust = c(5, 10, 15, 20, 25), method = "pearson", hls.method = "hls")
enrRes = compute_enrichment(clpr$optcl, corrType = "fdr", type_enrich = "KEGG", org_enrich = "rnorvegicus", pth = 0.05)
write_xlsx_for_funmappone(clpr$optcl, filePath = "../contour_clustering/gene_clustering.xlsx")
```

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