

StarPep toolbox User Guide

StarPep Developer Team

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Preface

Welcome to the *StarPep toolbox* project GitHub repository. Here is where all the components of the project are developed, reviewed, and maintained.

About the Software

[StarPep toolbox](#) is a software for studying the antimicrobial peptides' (AMPs) chemical space with molecular network-based representations and similarity searching models. This application aims to contribute to peptide drug repurposing, development, and optimization.

This tool was developed as a Java desktop application that integrates the functionalities of several open-source projects. The graphical user interface was built on top of the [NetBeans Platform](#), using the Java SE Runtime Environment 8. The graph database structure was implemented with the [Neo4j](#) platform. Some visualization features and the calculation of network properties were based on [Gephi](#). The sequence alignment algorithms were implemented using the [BioJava](#) API.

The AMPs were collected from a large variety of biological data sources to be organized into an integrated graph database called [starPepDB](#), composed of 45.120 AMPs and their metadata. This integrated graph database is embedded in StarPep toolbox to enable end-user querying, filtering, visualizing, and analyzing the AMPs taking advantage of network-based representations.

The main features of StarPep toolbox are listed below:

- **AMPs' chemical space filtering:** obtain a subset of AMPs from the StarPepDB using their metadata (function, target pathogen, biological origin, chemical modifications, original database, and cross-referenced entries to PDB, PubMed, and UniProt).
- **Molecular descriptors:** calculate molecular descriptors of the AMPs by applying statistical and aggregation operators on physicochemical amino acid properties (e.g., net charge, isoelectric point, molecular weight, etc.).
- **Network Science:** build different types of networks (metadata, chemical space, and half-space proximal) and calculate global/local properties, centrality metrics, communities, etc.

- **Similarity searching:** create multi-query similarity searching models that can lead to the repurposing of AMPs with novel functional activities.

The Team

This project was developed by members and collaborators of the *Grupo de Medicina Molecular y Traslacional (MeM&T)* at Universidad San Francisco de Quito, which is lead by [Yovani Marrero-Ponce](#).

Contributing

We encourage your participation as a contributor in this project considering your interest, availability, or skill requirements. Detailed information about ways of collaborating on this project can be found in our [contributing guidelines](#).

License

...

Get in touch

If you want to report a problem or suggest an improvement, you should [open an issue](#) at this Github repository, and we can follow your questions or suggestions. But, you can also contact Yovani by emailing ymarrero77@yahoo.es.

1 Installing StarPep toolbox

The binary executable files for Windows, Mac, and Linux are available in this [link](#). You can download the zip distribution and extract it to a folder or use an installer for the application.

1.1 Hardware requirements

- **Memory (RAM):** A minimum of 4 GB is required, but we recommend 8GB or more.
- **Processors:** We recommend a multi-core processor due to the fact that the software has been implemented to enable parallel processing of computationally intensive tasks.
- **Hard Disk:** a minimum of 500 MB of free space is required.

1.2 Software requirements

- Java SE Runtime Environment 8.

Note

It does not work (yet) with versions of Java greater than 8.

1.3 Issues with java versions

StarPep toolbox **does not yet support** any version of Java > 8 . **The requirement is java 8.** If you have multiple Java versions installed on your system, please configure starPep toolbox to run on the supported one (Java 8). Find the `etc/starPep.conf` file in the installed directory and configure the `jdkhome="/path/to/jdk"` accordingly. The symbol “#” at the beginint of the line means that it is commented out, please remove it.

1.4 Increasing the memory heap size

You may increase the memory heap further if there is enough RAM available in your system (**recommended**). First, you have to switch to the directory where the application has been installed or extracted. Open the text file “starpep.conf” located under the etc folder. Once the file has been open, locate the default options setting and change the min/max heapsize values (-J-Xms or -J-Xmx). For instance, to increase the memory heap size from 4G to 8G, enter the value:

```
default_options="--branding starpep -J-Xms24m -J-Xmx8G"
```

Then save the text file `etc/starpep.conf` and run the application.

1.5 Running starPep toolbox

StarPep toolbox can be initiated by running the *bin* executable files located in the installed directory, or by clicking the application icon (if installed).



Figure 1.1: Loading screen of StartPep toolbox

2 Getting Started

2.1 Main view

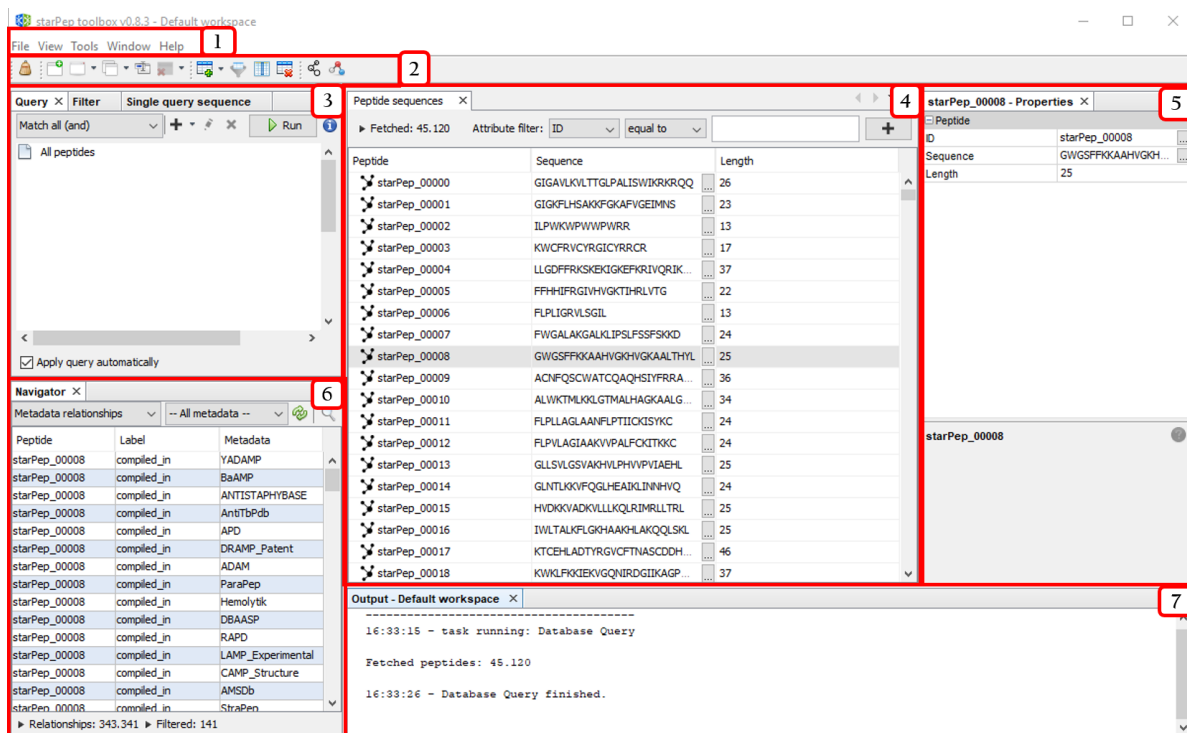


Figure 2.1: StartPep toolbox main window

1. Menu bar
2. Quick access bar
3. Tools panel
4. Central panel
5. Properties panel
6. Navigator panel
7. Output panel

i Note

The above windows panels may be opened from the **Window** option in the menu bar.

2.2 Menu bar

2.2.1 File

The following options are accessible from the **File** option:

i Note

Workspaces may be used to work with different data models: one per workspace.

- **File** :arrow_right: **New workspace**: Creates a new workspace.
- **File** :arrow_right: **Select workspace** :arrow_right: **[workspace_name]**: Switches to a new workspace.
- **File** :arrow_right: **Copy data to** :arrow_right: **New workspace**: Duplicates data model to a new workspace.
- **File** :arrow_right: **Rename current workspace**: Renames the current workspace.
- **File** :arrow_right: **Remove workspace**:
 - **Remove current workspace**: Removes the current workspace.
 - **Remove other workspaces**: Removes the other workspaces, and only remains the current workspace.
- **File** :arrow_right: **Clean project**: Removes all workspaces and sets the default workspace with the default data model.
- **File** :arrow_right: **Import** :arrow_right: **Peptide sequences (FASTA format)**: Imports peptide sequences into a new workspace.
- **File** :arrow_right: **Export**: Exports the following data
 - **Peptide sequences (FASTA format)**
 - **Molecular descriptors (CSV format)**
 - **Networks (GraphML format)**
 - **Metadata relationships (CSV format)**
- **File** :arrow_right: **Exit**: Shutdowns the program

2.2.2 View

The following options are accessible from the **View** option:

- **View** :arrow_right: **Toolbars**: Shows/hides a quick access bar.
 - **File**
 - **Workspace**
 - **Network**
 - **Molecular Descriptors**
- **View** :arrow_right: **Full Screen**: Switches to full screen.

2.2.3 Tools

The following options are accessible from the **Tools** option:

- **Tools** :arrow_right: **Peptide querying**: Opens/selects the query tab in the Tools panel.
- **Tools** :arrow_right: **Peptide search by**:
 - **Single Query sequence**: Opens/selects the single query tab in the Tools panel.
 - **Multiple Query sequences**: Opens/selects the multiple query tab in the Tools panel.
 - **Non-redundant set**: Opens/selects the non-redundant set tab in the Tools panel.
- **Tools** :arrow_right: **Peptide filtering**: Opens/selects the filter tab in the Tools panel.
- **Tools** :arrow_right: **Molecular features**:
 - **Extraction** :arrow_right: [**molecular descriptor option**]: Opens/selects the molecular descriptor tab in the Tools panel.
 - **Selection** :arrow_right: [**unsupervised feature selection**]: Opens/selects the unsupervised feature selection tab in the Tools panel.
 - **Explorer**: Opens the feature explorer window.
 - **Removing**: Opens the feature removing window.
- **Tools** :arrow_right: **Networks**:
 - **Metadata**: Opens the window to generate a metadata network.
 - **Similarity Network**: Opens/selects the chemical space tab in the Tools panel.
 - **Appearance**: Opens/selects the appearance tab in the Tools panel.
 - **Layout** :arrow_right: [**layout algorithm**]: Opens/selects the layout algorithm tab in the Tools panel.
 - **Clustering** :arrow_right: [**clustering algorithm**]: Opens/selects the clustering algorithm tab in the Tools panel.

- **Centrality** :arrow_right: **[measure]**: Opens/selects the centrality measure tab in the Tools panel.
- **Subnetwork mining** :arrow_right: **[graph-based algorithm]**: Opens/selects the graph-based algorithm tab in the Tools panel.
- **Tools** :arrow_right: **Options**: Displays the software configuration window.

2.2.4 Window

The following options are accessible from the **Window** option:

- **Peptide sequences**: Opens/selects the peptide sequences window in the center panel.
- **Network visualization**: Opens/selects the network visualization window in the center panel.
- **Properties**: Opens/selects the properties panel.
- **Navigator**: Opens/selects the navigator panel.
- **Output**: Opens/selects the output panel.
- **Configure Window** :arrow_right: **[options]**: Window settings
- **Reset Windows**
- **Close Window**
- **Close All Documents**
- **Close Other Documents**
- **Documents...**: Opens the Document management window.

2.2.5 Help

The **Help** :arrow_right: **About** opens the About information window.

2.3 Quick access bar

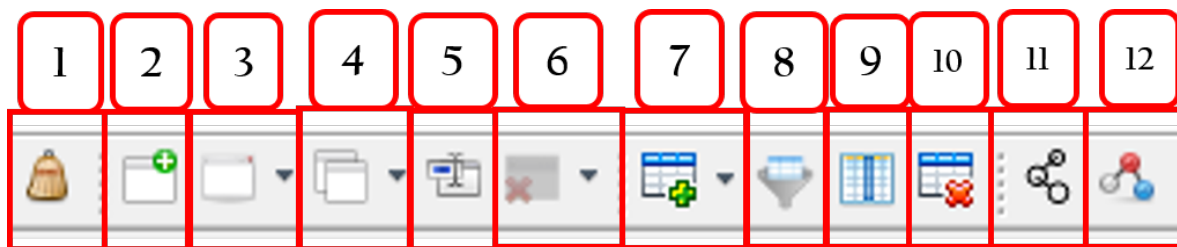


Figure 2.2: Quick access bar.

Note

These options may be shown/hidden from the menu entry: **View** :arrow_right: **Toolbars** :arrow_right: [options].

Shortcut to:

1. **File** :arrow_right: **Clean project**
2. **File** :arrow_right: **New workspace**
3. **File** :arrow_right: **Select workspace**
4. **File** :arrow_right: **Copy data to** :arrow_right: **New workspace**
5. **File** :arrow_right: **Rename current workspace**
6. **File** :arrow_right: **Remove workspace**
7. **Tools** :arrow_right: **Molecular features** :arrow_right: **Extraction**
8. **Tools** :arrow_right: **Molecular features** :arrow_right: **Selection**
9. **Tools** :arrow_right: **Molecular features** :arrow_right: **Explorer**
10. **Tools** :arrow_right: **Molecular features** :arrow_right: **Removing**
11. **Tools** :arrow_right: **Network** :arrow_right: **Metadata Network**
12. **Tools** :arrow_right: **Network** :arrow_right: **Similarity Network**

2.4 Tool panels: an overview

2.4.1 Query panel

This panel may be opened from **Tools** :arrow_right: **Peptide querying**.

Note

The recovered peptides are those linked to the specified metadata nodes.

1. Selects the joining condition for the query criteria: **Match all (and)** or **Match all (or)**.
2. Adds a new term (linked metadata) to the query.
3. Edits the query term selected.
4. Deletes the query term selected.
5. Runs the query.
6. List of current query terms.
7. Applies the query automatically with each change.

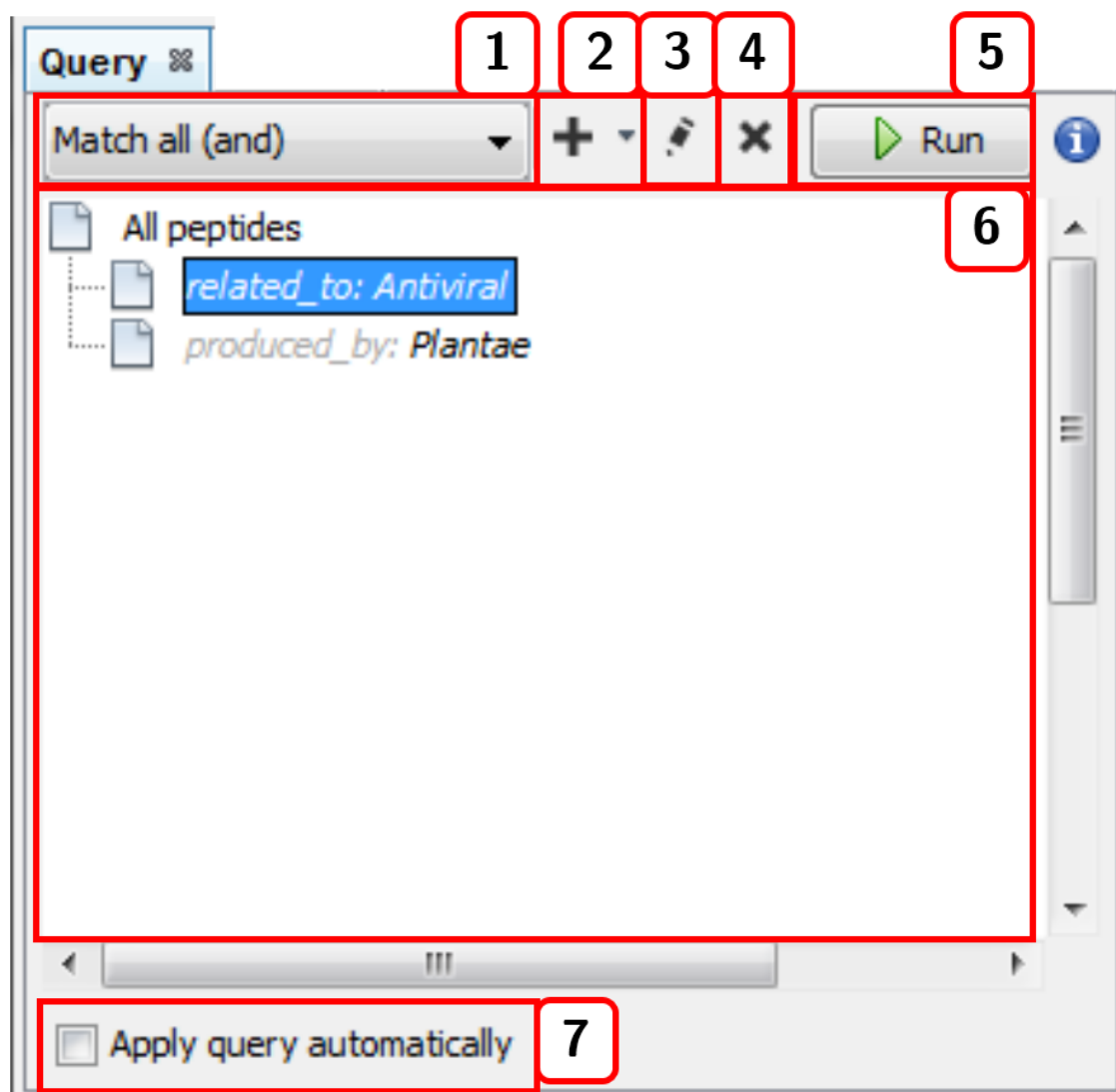


Figure 2.3: Query panel

2.4.2 Filter panel

This panel may be opened from **Tools** :arrow_right: **Peptide filtering**.

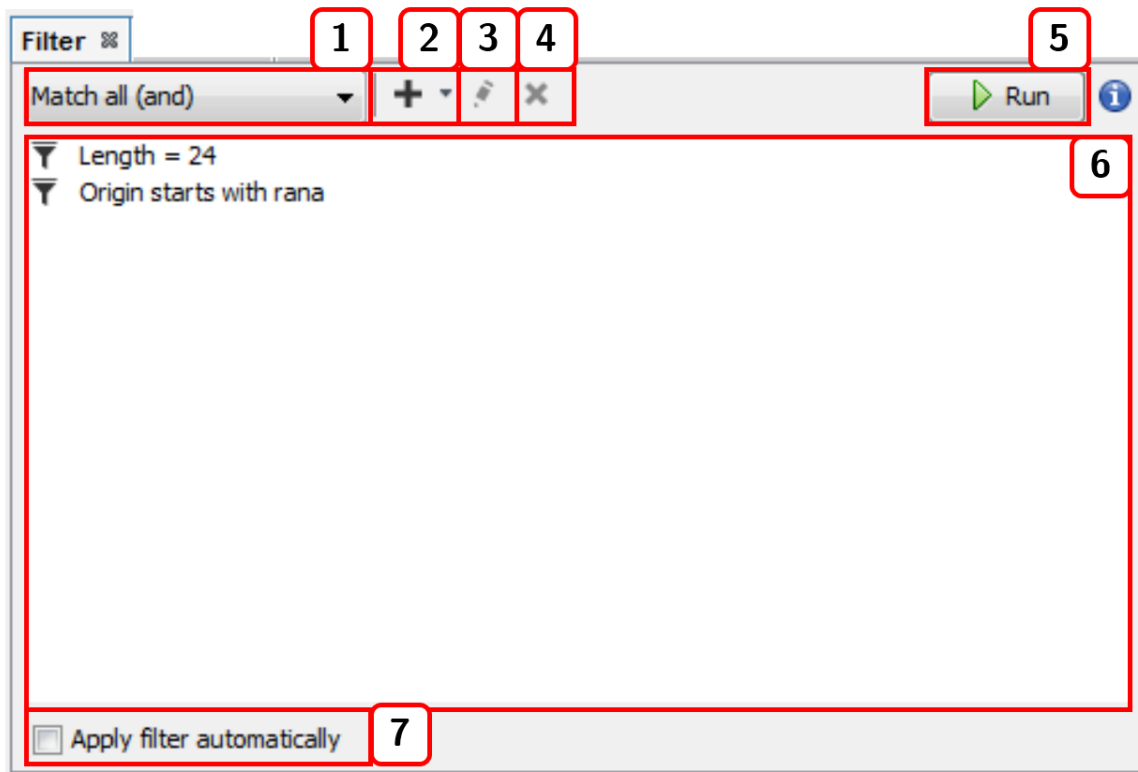


Figure 2.4: Filter panel

1. Selects the joining condition for the filter criteria: **Match all (and)** or **Match all (or)**.
2. Adds a new filter.
3. Edits the selected filter.
4. Deletes the selected filter.
5. Runs the filter.
6. List of current filters.
7. Applies the filter automatically with each change.

2.4.3 Sequence search

This panel can be opened from **Tools** :arrow_right: **Peptide search by** :arrow_right: [sequence search option]. For instance, **Single query sequence**:

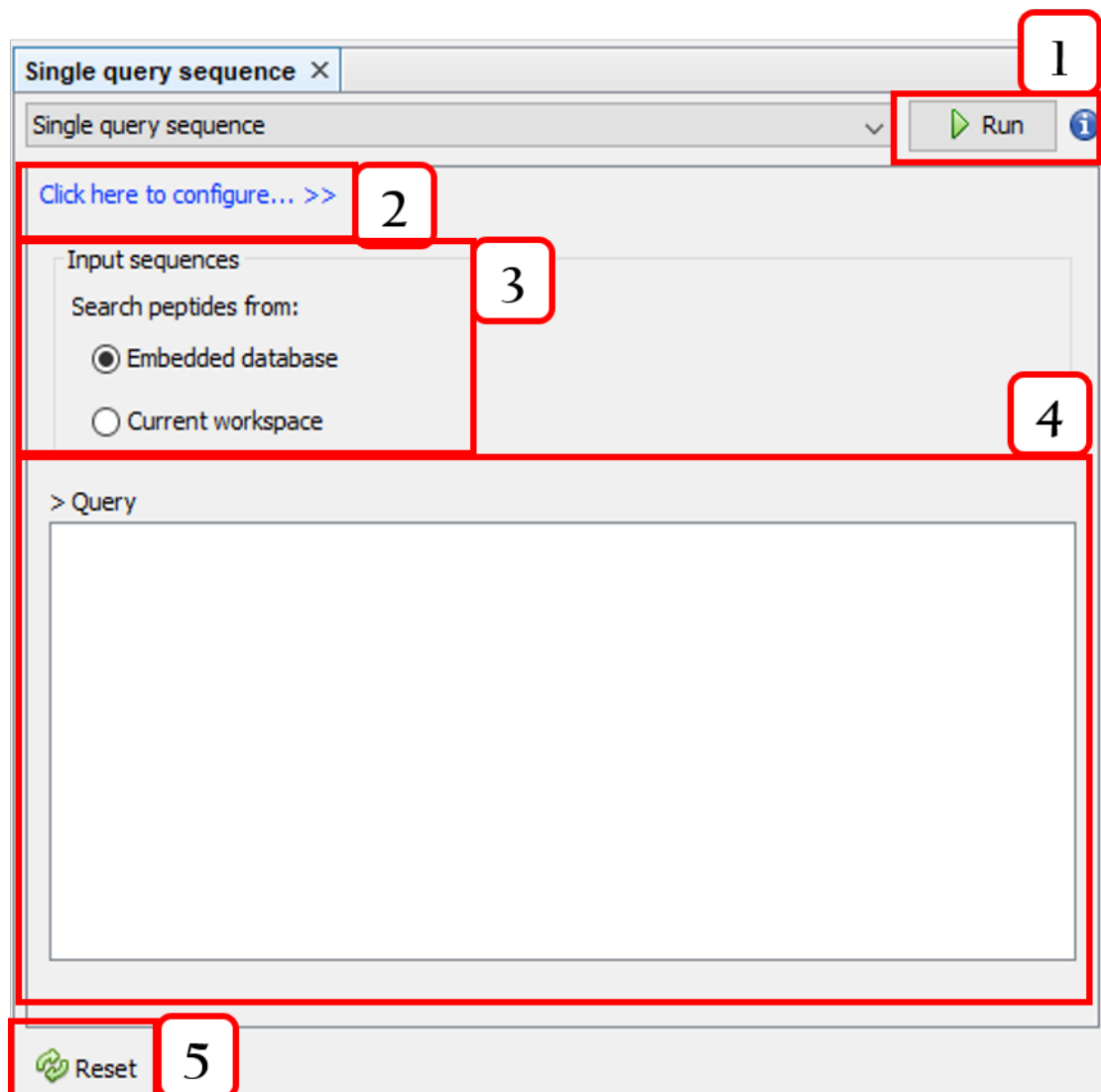


Figure 2.5: Single query sequence.

1. Runs the query.
2. Configures the sequence alignment.
3. Selects the target sequences.
4. Input sequence.
5. Resets the query.

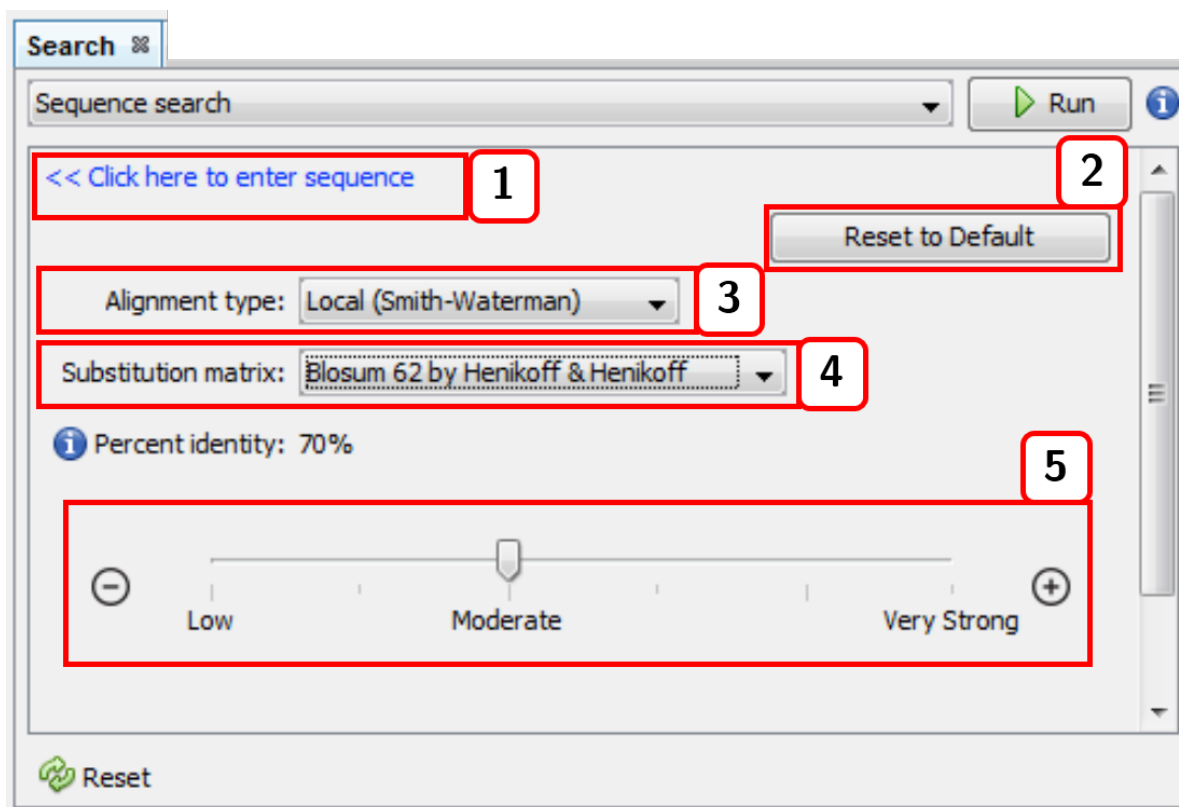


Figure 2.6: Sequence alignment settings.

1. Returns to the input sequence view.
2. Resets the alignment configuration.
3. Alignment type (local or global).
4. Substitution matrix.
5. Percent identity (default: 98%).

2.4.4 Molecular feature extraction

This option is accessible from the menu option **Tools** :arrow_right: **Molecular features** :arrow_right: **Extraction** :arrow_right: [molecular descriptor option]. For instance, All descriptors:

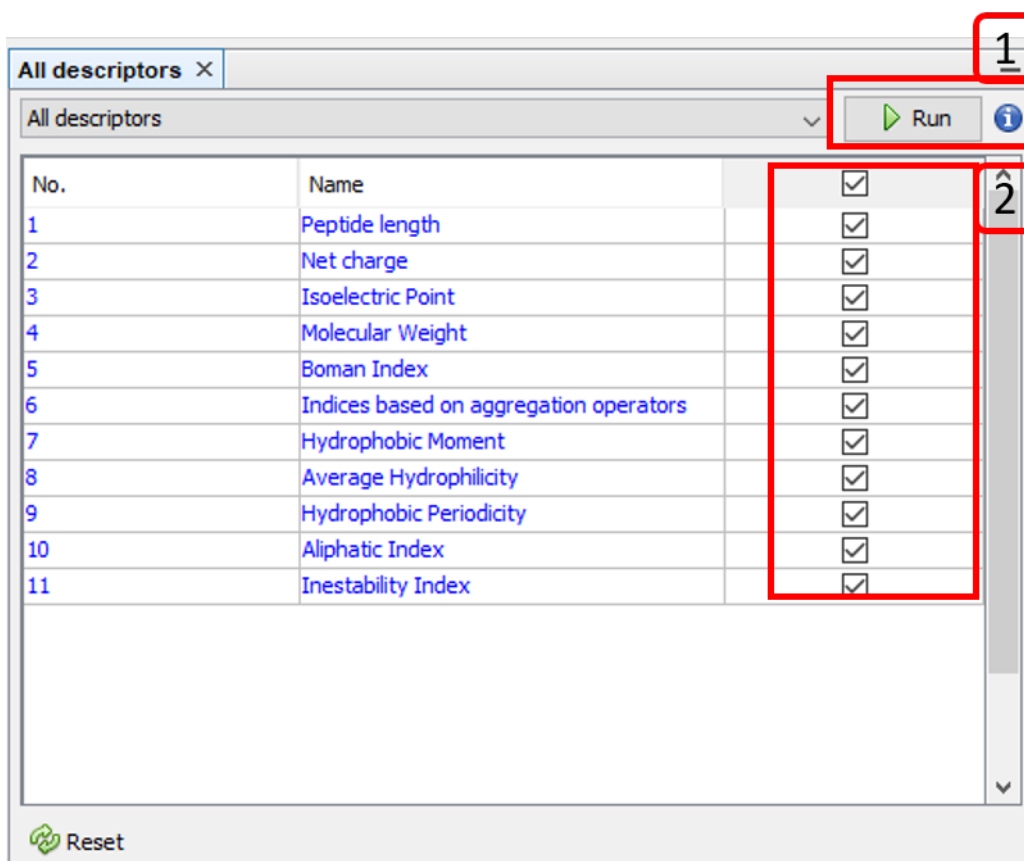


Figure 2.7: Calculating all molecular descriptors.

1. Runs the selected molecular descriptor algorithms.
2. Selects/Unselect molecular descriptor algorithms.

Note

The calculated molecular descriptors can be removed by accessing the menu options **Tools** :arrow_right: **Molecular features** :arrow_right: **Removing**.

Besides, calculated molecular features can be displayed in the columns list at the center panel (enabling molecular feature filtering). This option is accessible from the menu option **Tools** :arrow_right: **Molecular features** :arrow_right: **Explorer**:

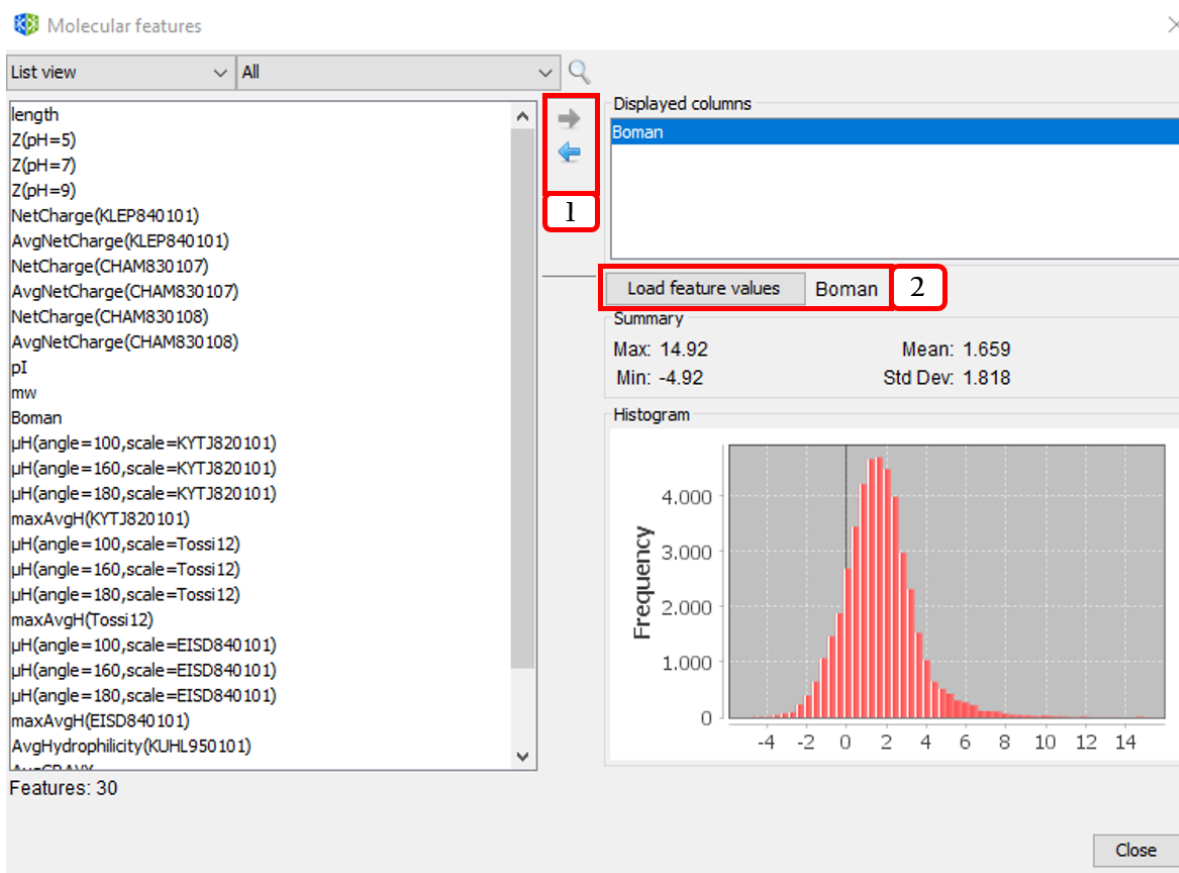


Figure 2.8: Adding molecular features (*Boman*) to the displayed columns list.

1. Adds/Removes molecular descriptors to/from the displayed columns list.
2. Visualizes the histogram and data summary (max, min, mean, and standard deviation of molecular feature values).

2.4.5 Molecular feature selection

This option is accessible from the menu option **Tools** :arrow_right: **Molecular features** :arrow_right: **Selection** :arrow_right: [**unsupervised feature selection**]. For instance, Filtering & subset optimization:

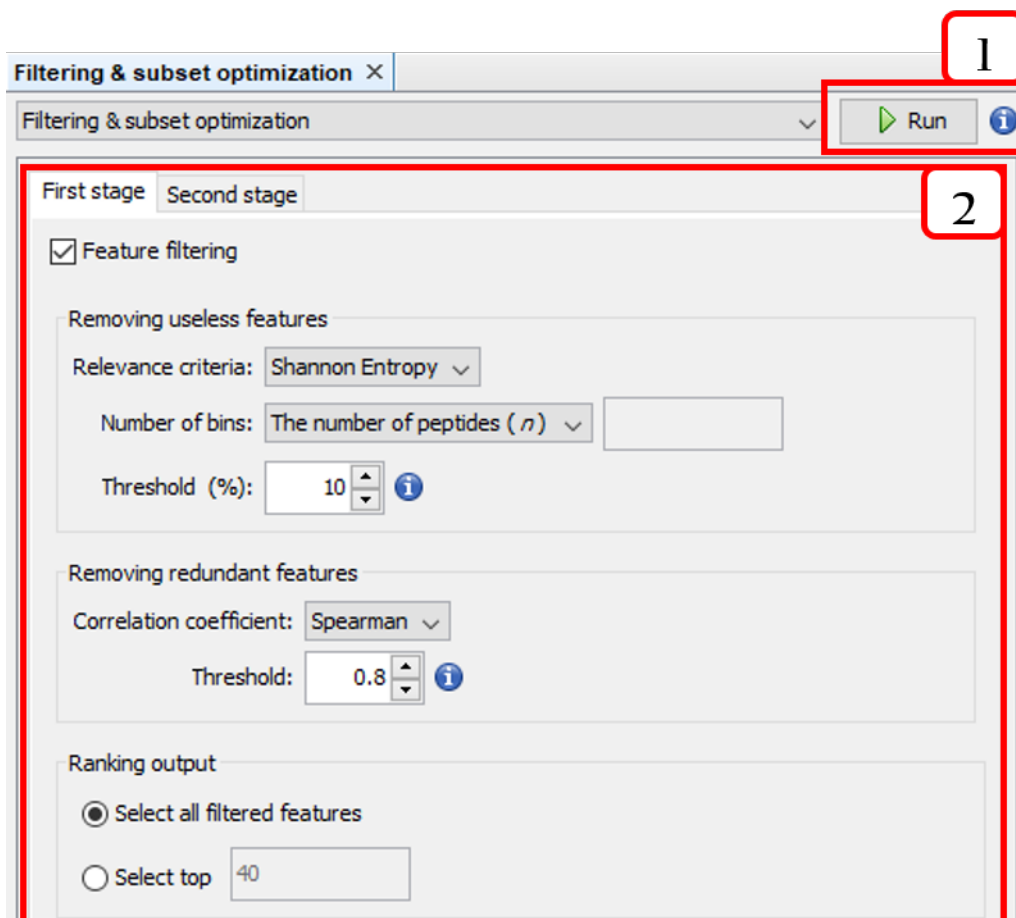


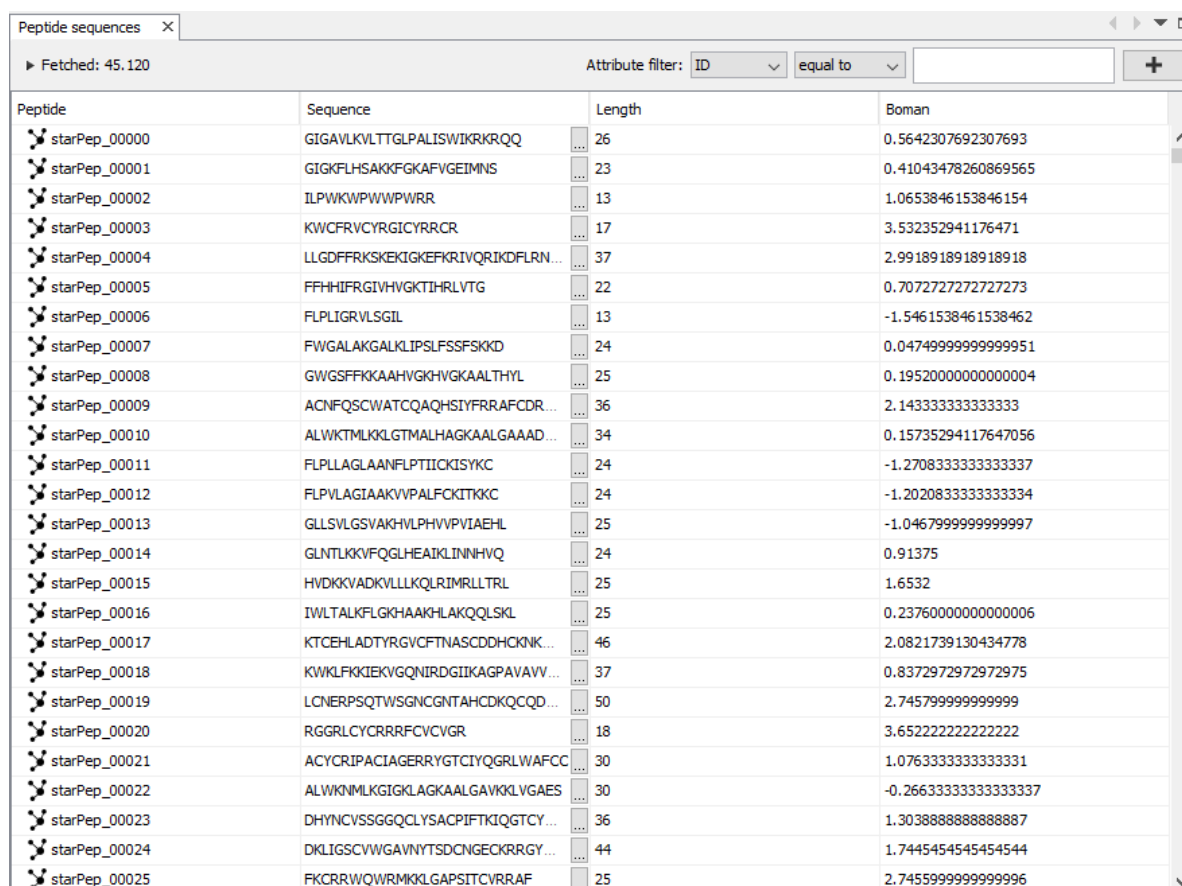
Figure 2.9: The two-stage unsupervised feature selection

1. Runs the two-stage unsupervised feature selection.
2. Configures the two-stage unsupervised feature selection.

2.5 Center panels

2.5.1 Peptide sequences window

This window is opened from **Window** :arrow_right: **Peptide sequences**. The Peptide sequences window shows the result of applying a query, filter, or search. The rows showed can also be filtered by attributes such as **ID**, **Sequence**, **Length**, or calculated features.



Peptide	Sequence	Length	Boman
starPep_00000	GIGAVLKVLTTGLPALISWIKRKRQQ	26	0.5642307692307693
starPep_00001	GIGKFLHSAKFGKAFVGEIMNS	23	0.41043478260869565
starPep_00002	ILPWKWPWWPWRR	13	1.0653846153846154
starPep_00003	KWCFRVCYRGICYRRCR	17	3.532352941176471
starPep_00004	LLGDFFRKSKEKIGKEFKRIVQRIKDFLRN...	37	2.9918918918918918
starPep_00005	FFHHIFRGIVHVGKTIHRLVTG	22	0.7072727272727273
starPep_00006	FLPLIGRVLSGIL	13	-1.5461538461538462
starPep_00007	FWGALAKGALKLIPSLFSSFSKID	24	0.047499999999999951
starPep_00008	GWGSFFKAAHVGVGKAALHTYL	25	0.19520000000000004
starPep_00009	ACNFQSCWATCQAQHSIYFRRAFCDR...	36	2.1433333333333333
starPep_00010	ALWKTMLKLGTMALHAGKAALGAAAD...	34	0.15735294117647056
starPep_00011	FLPLLAGLAANFLPTIICKISYKC	24	-1.2708333333333337
starPep_00012	FLPVLAGIAAKVVPALFCKITKCC	24	-1.2020833333333334
starPep_00013	GLLSVLGSVAKHVLPVHPVIAEHL	25	-1.0467999999999997
starPep_00014	GLNTLKKVFQGLHEAIKLINNIVQ	24	0.91375
starPep_00015	HVDKKVADKVLKQLRIMRLTRL	25	1.6532
starPep_00016	IWLTALKFLGKHAAKHLAQQLSKL	25	0.23760000000000006
starPep_00017	KTCEHLADTYRGVCFNASCDDHCKNK...	46	2.0821739130434778
starPep_00018	KWKLFKQIEKVGQNIKAGPAVAVV...	37	0.8372972972972975
starPep_00019	LCNERPSQTVSGNCGNTAHCDKQCQD...	50	2.7457999999999999
starPep_00020	RGGRLCYCRRRFCVGVGR	18	3.6522222222222222
starPep_00021	ACYCRIPACIAGERRYGTCTYQGRWAFCC	30	1.0763333333333331
starPep_00022	ALWKNMLKIGKLAGKAALGAVKLVGAES	30	-0.2663333333333337
starPep_00023	DHYNCVSSGGQCLYSACPIFTKIQTCTY...	36	1.3038888888888887
starPep_00024	DKLIGSCVWGAVNYSDCNGECKRRGY...	44	1.7445454545454544
starPep_00025	FKCRRWQWRMKKLGAPISITCVRRAF	25	2.7455999999999996

Figure 2.10: Peptide sequences window

2.5.2 Network visualization window

This window is opened from **Window** :arrow_right: **Network Visualization**. It consists of two views: **Scene** and **Preview**. The **Scene** view allows to customize some visual properties of the network such as background color, zoom, position, and individual colors for edges and nodes. The options highlighted in Fig. 2.11 are the following:

1. Switch background.
2. Zoom options.
3. Selector. It allows to change the node diameter of the cursor while selecting nodes.
4. Additional options. It allows to enable or disable the options **Autoselect neighbors** and **Show peptide labels** (we recommend disabling the latter in order to render clearer graphs in metadata network analysis).
5. More advanced sizing and coloring options for nodes. By pressing **More.../Less...**, the options are shown/hidden.
6. Network rendering area.
7. Node label options. The first one allows to show/hide the node labels. The second one brings three options to modify node label size: **Fixed**, **Scale size**, and **Node size**. This option **Node size** is handy for adjusting the label size proportionally to the node size. The third one modify the label color options. There are three choices: **Unique**, **Object**, and **Text**.
8. Node label font properties.
9. Node label size.
10. Two edges options. The first one shows/hides edges. The second one enables edges to have the attached node color.
11. Edge thickness.
12. Shows/Hides edge labels.
13. Edge label font properties.
14. Edge label size.

i Note

When you right-click the mouse on the scene view, a context menu is displayed.

The **Preview** view shows the rendered the graph according to the calculated layout and all the configurations. Attractive networks may be rendered in this other view. To update the drawing, press the **Refresh** button.

2.6 Navigator panel

This panel is opened from **Window** :arrow_right: **Navigator**. The navigator changes between the **Metadata relationships** and **Graph table** options according to whether the **Peptide sequences** or **Network visualization** window is active.

On the one hand, in the **Metadata relationships** view, the user can seek metadata nodes. Right-click on a row will show a context menu to select or center nodes on the graph, as well as the **Properties** window for the relationship.

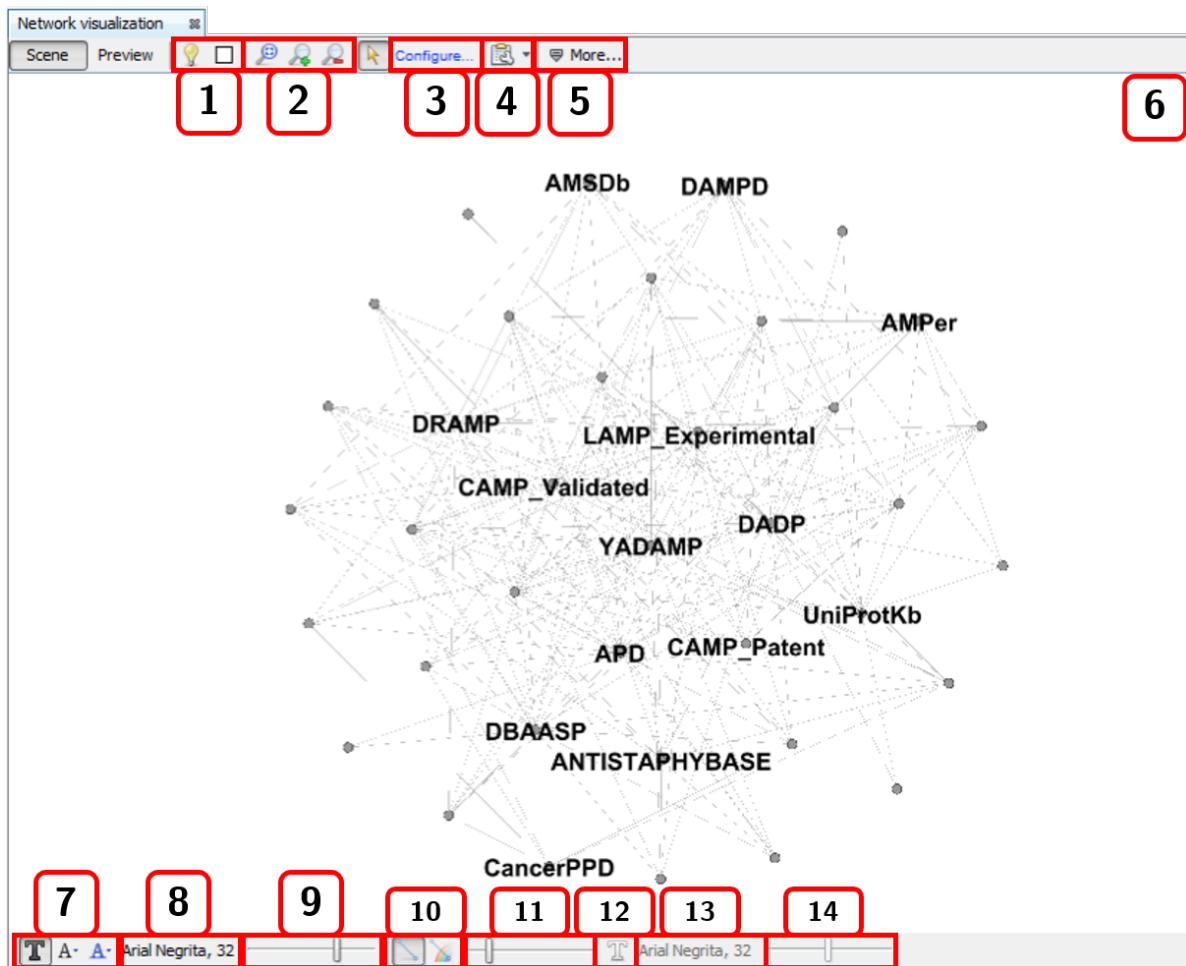


Figure 2.11: Network visualization window: Scene view.

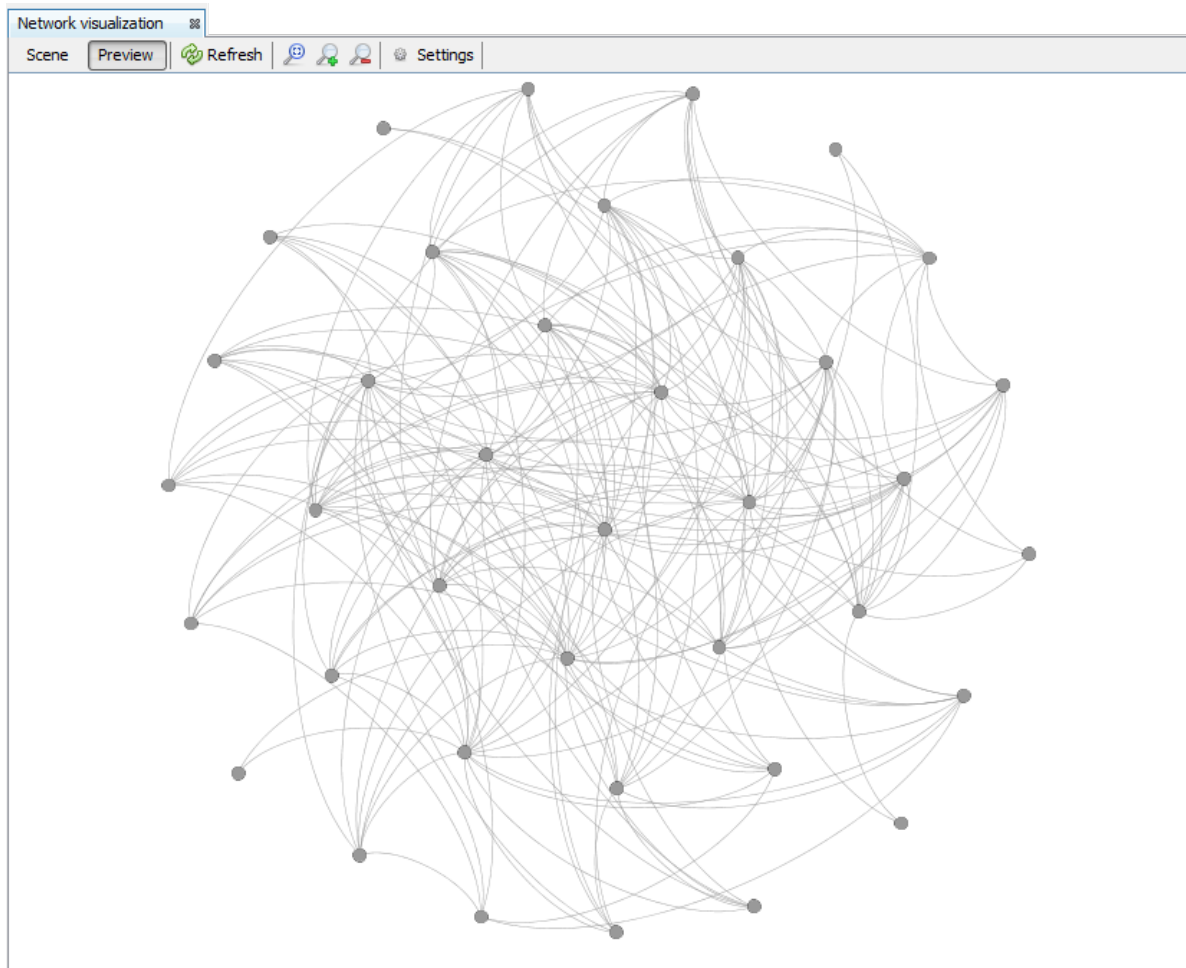
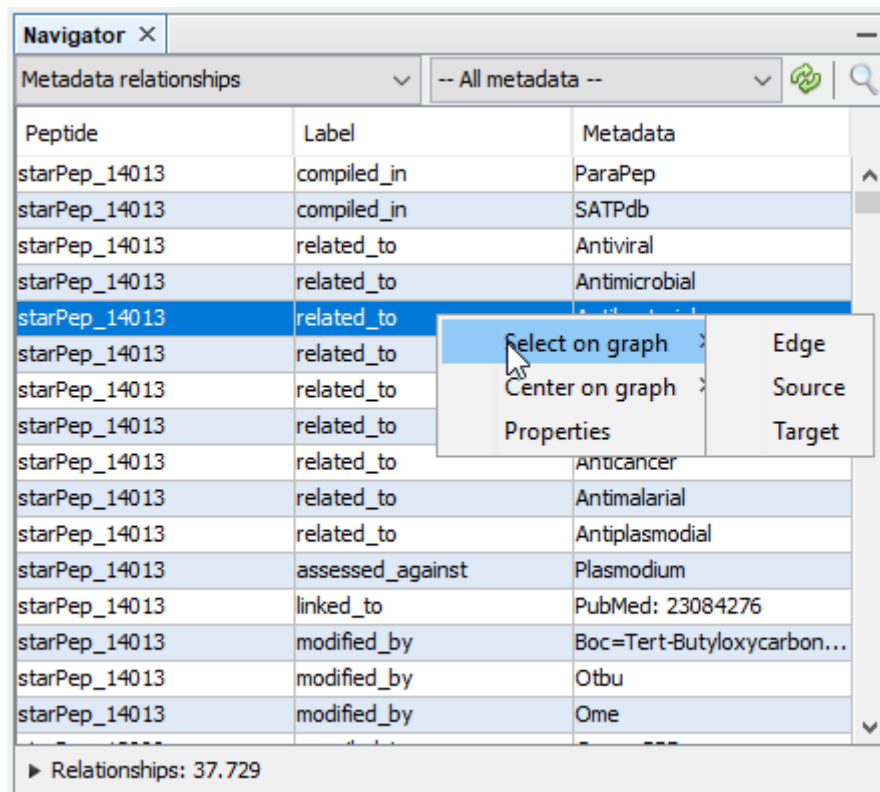


Figure 2.12: Network visualization window: Preview view.

i Note

If a peptide sequence is selected in the center panel, only the related metadata are shown in this navigator panel. Click on the **Refresh** button to show all.



Peptide	Label	Metadata
starPep_14013	compiled_in	ParaPep
starPep_14013	compiled_in	SATPdb
starPep_14013	related_to	Antiviral
starPep_14013	related_to	Antimicrobial
starPep_14013	related_to	Anticancer
starPep_14013	related_to	Antimalarial
starPep_14013	related_to	Antiplasmodial
starPep_14013	assessed_against	Plasmodium
starPep_14013	linked_to	PubMed: 23084276
starPep_14013	modified_by	Boc=Tert-Butyloxycarbon...
starPep_14013	modified_by	Otbu
starPep_14013	modified_by	Ome

► Relationships: 37.729

Context menu options:

- Select on graph
- Center on graph
- Properties
- Edge
- Source
- Target

Figure 2.13: Navigator for the Peptide sequences window

On the other hand, in the **Graph table** view, the user can switch the view from nodes table to edges table, and also customize the columns (such as network measures) shown in the data grid. These data tables can be exported to an external text file (CSV format). There is also a context menu that is accessed via right-click on any row.

Navigator ×		
Graph table ▾		
Node Edge 🔍 Columns... 📄		
Name	Degree	Label
starPep_00000	22	Peptide
HIPdb	14	Database
ADAM	335	Database
DRAMP_Patent	130	Database
StraPep	60	Database
AntiTbPdb	54	Database
RAPD	27	Database
CAMP_Patent	27	Database
NeuroPep	4	Database
YADAMP	273	Database
ParaPep	67	Database
DBAASP	1,218	Database
CAMP_Validated	257	Database
AVPdb	22	Database
BaAMP	32	Database
LAMP_Patent	13	Database
APD	350	Database
▶ Nodes: 1.863 ▶ Edges: 6.547		

Figure 2.14: Navigator for the Network visualization window

3 Working with networks

3.1 Metadata network

The construction of metadata network is accessible from the menu option **Tools** :arrow_right: **Networks** :arrow_right: **Metadata Network**.

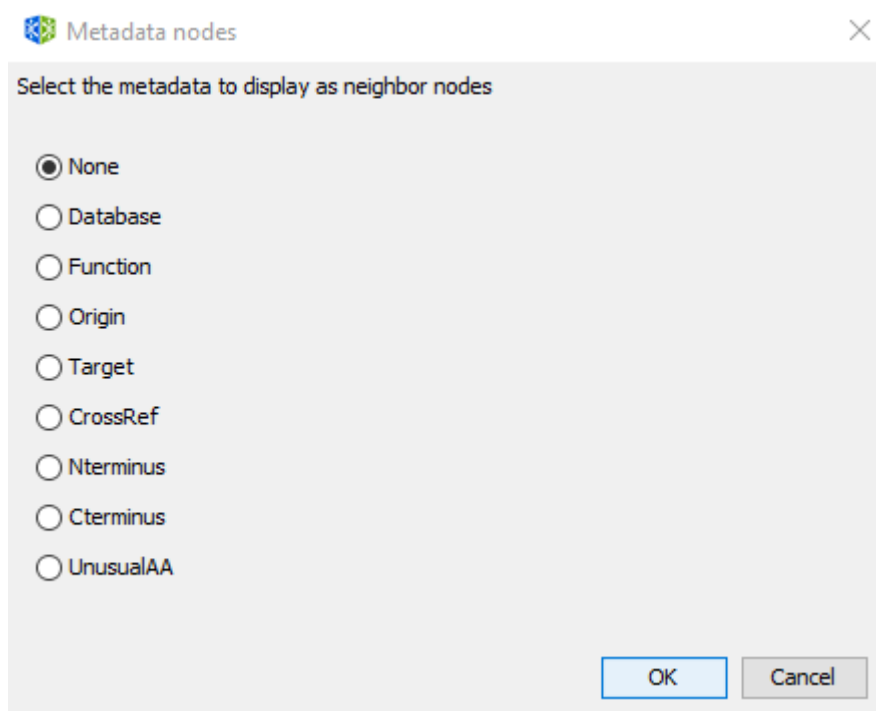


Figure 3.1: Options for metadata network

In metadata networks, nodes representing metadata are connected to nodes representing peptides by the following relationships:

Table 3.1: Metadata node names and relationships in starPepDB.

Metadata node	Relationship
Origin	<i>produced_by</i>
Target	<i>assessed_against</i>
Function	<i>related_to</i>
Database	<i>compiled_in</i>
Crossref	<i>linked_to</i>
Nterminus	<i>modified_by</i>
UnusualAA	<i>constituted_by</i>
Cterminus	<i>modified_by</i>
Subcategory of another node	<i>is_a</i>

3.2 Similarity network

The construction of similarity network is accessible from the menu option **Tools** :arrow_right: **Networks** :arrow_right: **Similarity Network**. To create a similarity network, we first recommended to configure the workflow using the **Configuration Wizard** and then press the button **Run**.

1. Runs the workflow for building the similarity network.
2. Opens Configuration Wizard (Sect.3.2.1).
3. Changes between Nodes and Edges tabs.
4. Applies PCA coordinates changes.
5. Selects X and Y axis for PCA coordinates.
6. PCA results panel.

1. **Similarity threshold selector:** After changing the value, it is necessary to press **Apply**.
2. **Network Density plot:** Helps to decide a similarity threshold.

3.2.1 Configuration wizard

This section will show the **configuration wizard** for mapping and visualizing the Chemical Space.

3.2.1.1 Wizard Step 1: Input sequences

To remove redundant sequences, press **Yes (recommended)**. Then, you can choose between local or global alignment, multiple substitution matrices, and a identity threshold.

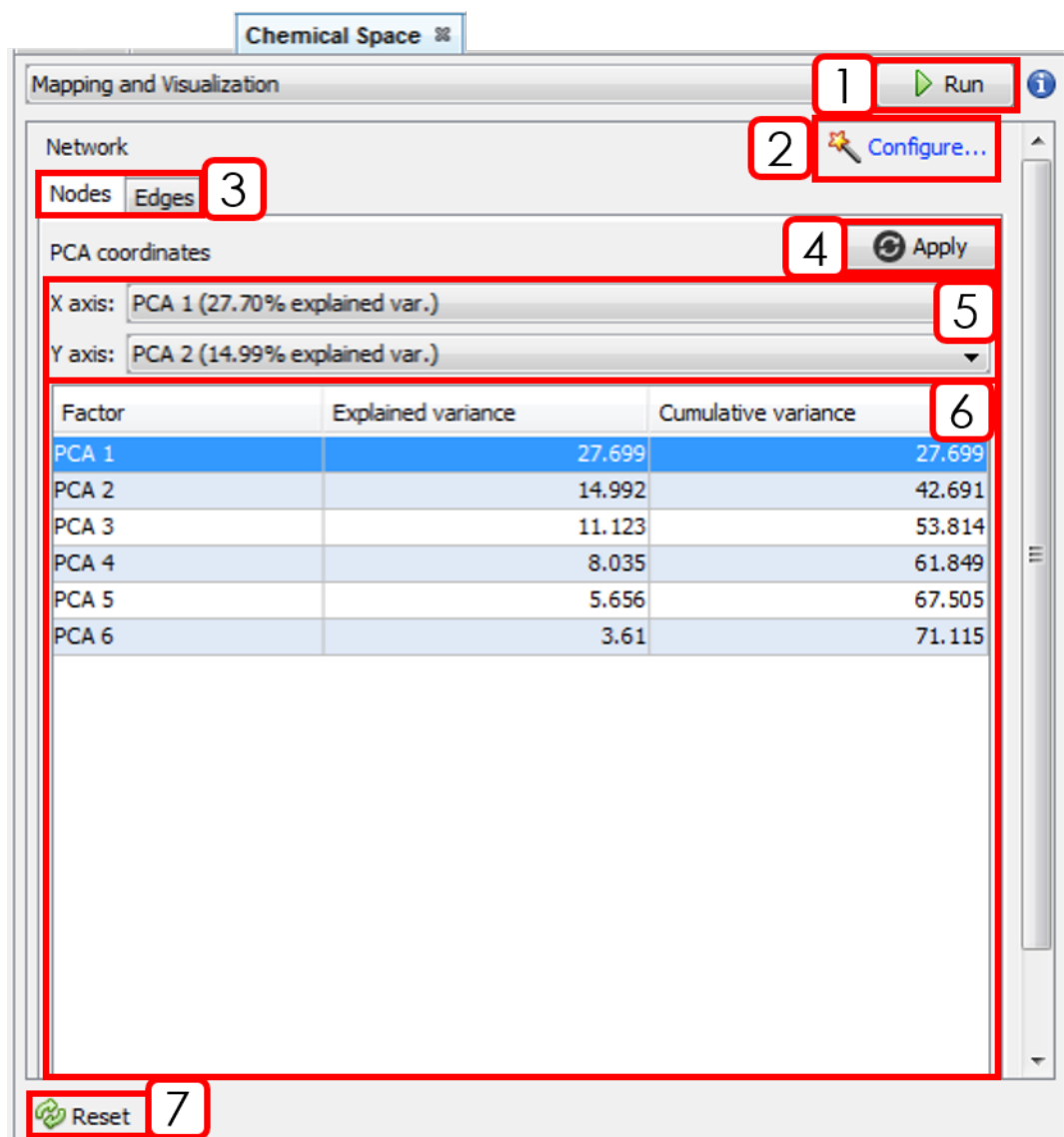


Figure 3.2: Chemical Space window (Nodes tab).

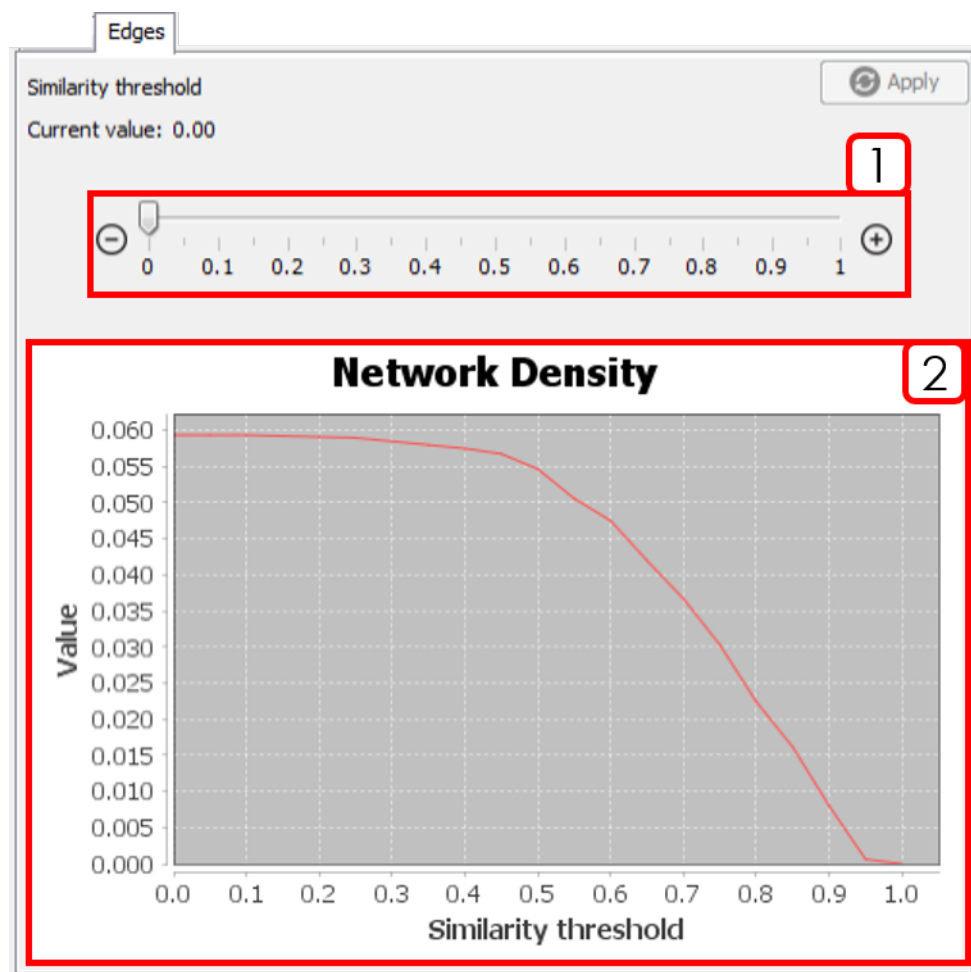


Figure 3.3: Chemical Space window (Edges tab).

Chemical Space Mapping and Visualization

Steps

- 1. Input sequences**
2. Feature extraction
3. Feature selection
4. (Dis)similarity metric
5. Network model

Input sequences

Select input peptides from:

☐ Embedded database

☒ Current workspace

Do you want to remove redundant sequences?

☐ No

☒ Yes

Alignment type: Local (Smith-Waterman)

Substitution matrix: Blosum 62 by Henikoff & Henikoff

Percent identity: 98%

Very Low Low Moderate Strong

< Back Next > Finish Cancel Help

Figure 3.4: Wizard Step 1: Input sequences.

3.2.1.2 Wizard Step 2: Feature extraction

If you already calculated a set of molecular descriptors, you can select the first option and press **Next**. If not, select the new descriptors to be calculated.

Steps

1. Training compounds
- 2. Feature extraction**
3. Feature selection
4. Distance function
5. Full/HSP network

Feature extraction

What do you want to do?

☒ Use the available set of molecular descriptors

☐ Compute new set of molecular descriptors

No.	Name	<input checked="" type="checkbox"/>
1	Length	<input checked="" type="checkbox"/>
2	Net charge	<input checked="" type="checkbox"/>
3	Isoelectric Point	<input checked="" type="checkbox"/>
4	Molecular Weight	<input checked="" type="checkbox"/>
5	Boman Index	<input checked="" type="checkbox"/>
6	Indices based on aggregation operators	<input checked="" type="checkbox"/>
7	Hydrophobic Moment	<input checked="" type="checkbox"/>
8	Average Hydrophilicity	<input checked="" type="checkbox"/>
9	Hydrophobic Periodicity	<input checked="" type="checkbox"/>
10	Aliphatic Index	<input checked="" type="checkbox"/>
11	Instability Index	<input checked="" type="checkbox"/>

< Back Next > Finish Cancel Help

Figure 3.5: Wizard Step 2: Feature extraction

3.2.1.3 Wizard Step 3: Feature selection

If you plan to use all available descriptors, select the first option, and press **Next**. If not, select and configure the **two-stage unsupervised feature selection** method.

Chemical Space Mapping and Visualization

Steps

1. Input sequences
2. Feature extraction
3. **Feature selection**
4. (Dis)similarity metric
5. Network model

Feature selection

What do you want to do?

☐ Use all features

☒ Unsupervised feature selection

First stage | Second stage

☒ Feature filtering

Removing useless features

Relevance criteria: Shannon Entropy

Number of bins: The number of peptides (n)

Threshold (%): 10

Removing redundant features

Correlation coefficient: Spearman

Threshold: 0,8

Ranking output

☒ Select all filtered features

☐ Select top 40

< Back Next > Finish Cancel Help

Figure 3.6: Wizard Step 3: Feature selection.

3.2.1.4 Wizard Step 4: Distance function.

Select the desired **distance function** and the **standardization/normalization** for the calculated descriptors.

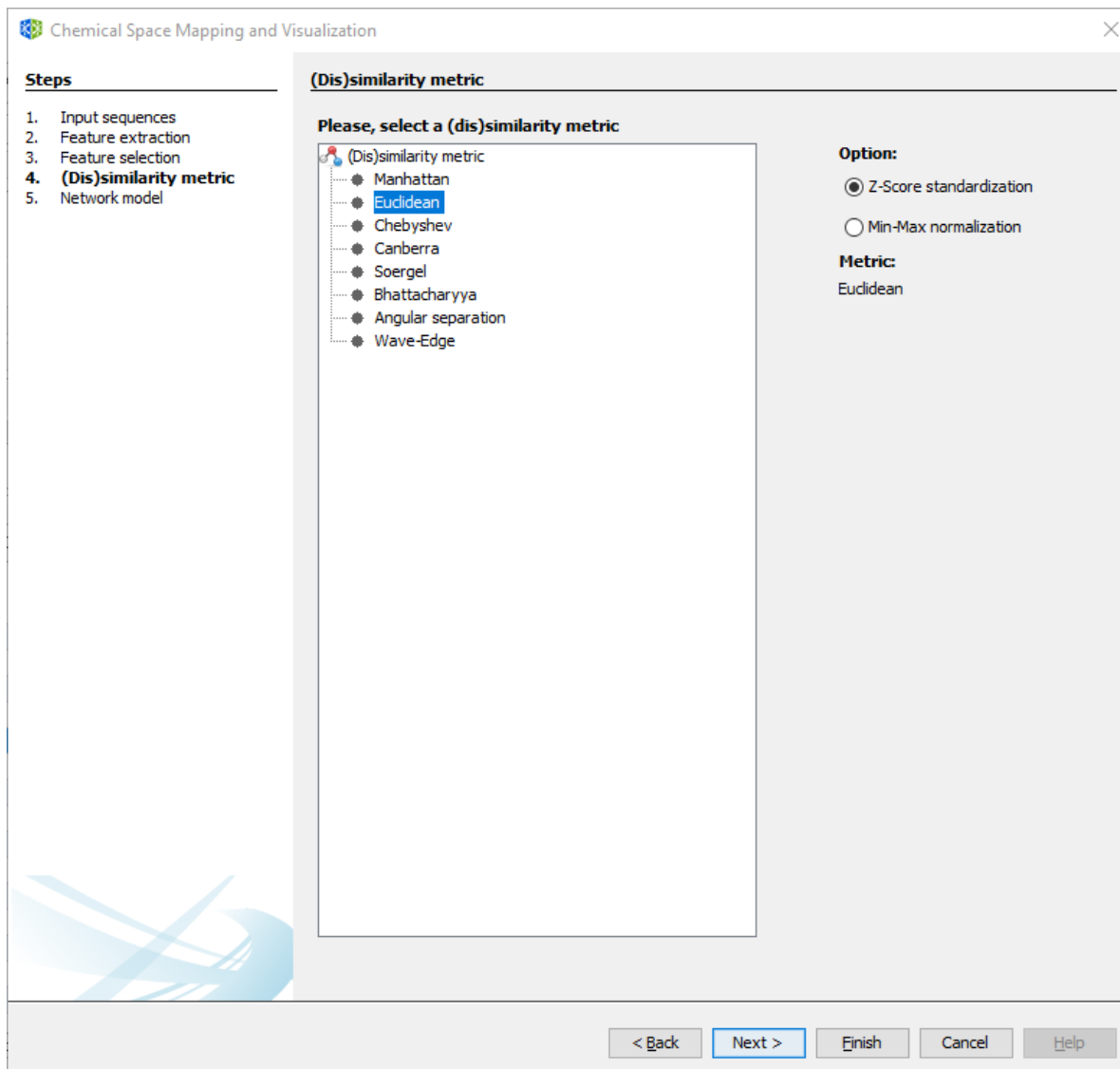


Figure 3.7: Wizard Step 4: Distance function.

3.2.1.5 Wizard Step 5: Network model

For generating a network model, select between the **Half-Space Proximal Network** or the traditional **Chemical Space Network/Similarity Network** (not recommended for large datasets). For more details, please refer to [the methodological paper](#).

Note

The position of nodes may be determined by the first two principal components of descriptor space. However, **layout algorithms** are recommended for a better rearrangement of nodes.

3.3 Network model options

After creating the network model, the following options are available.

1. Positioning nodes.
2. Adding/removing similarity edges.
3. Embedding new peptides. When new peptides are projected, a network model will be opened into a new workspace.

3.4 Layout algorithms

A layout algorithm option may be opened from **Tools** :arrow_right: **Network** :arrow_right: **Layout** :arrow_right: [layout option]. The main graph layouts available are Fruchterman Reingold, ForceAtlas 2, Yifan Hu Proportional, and Random Layout. Any layout result could be adjusted using the options Rotate, Contraction, Expansion, Noverlap, and Label Adjust.

3.5 Appearance

This panel is opened from **Tools** :arrow_right: **Network** :arrow_right: **Appearance**.

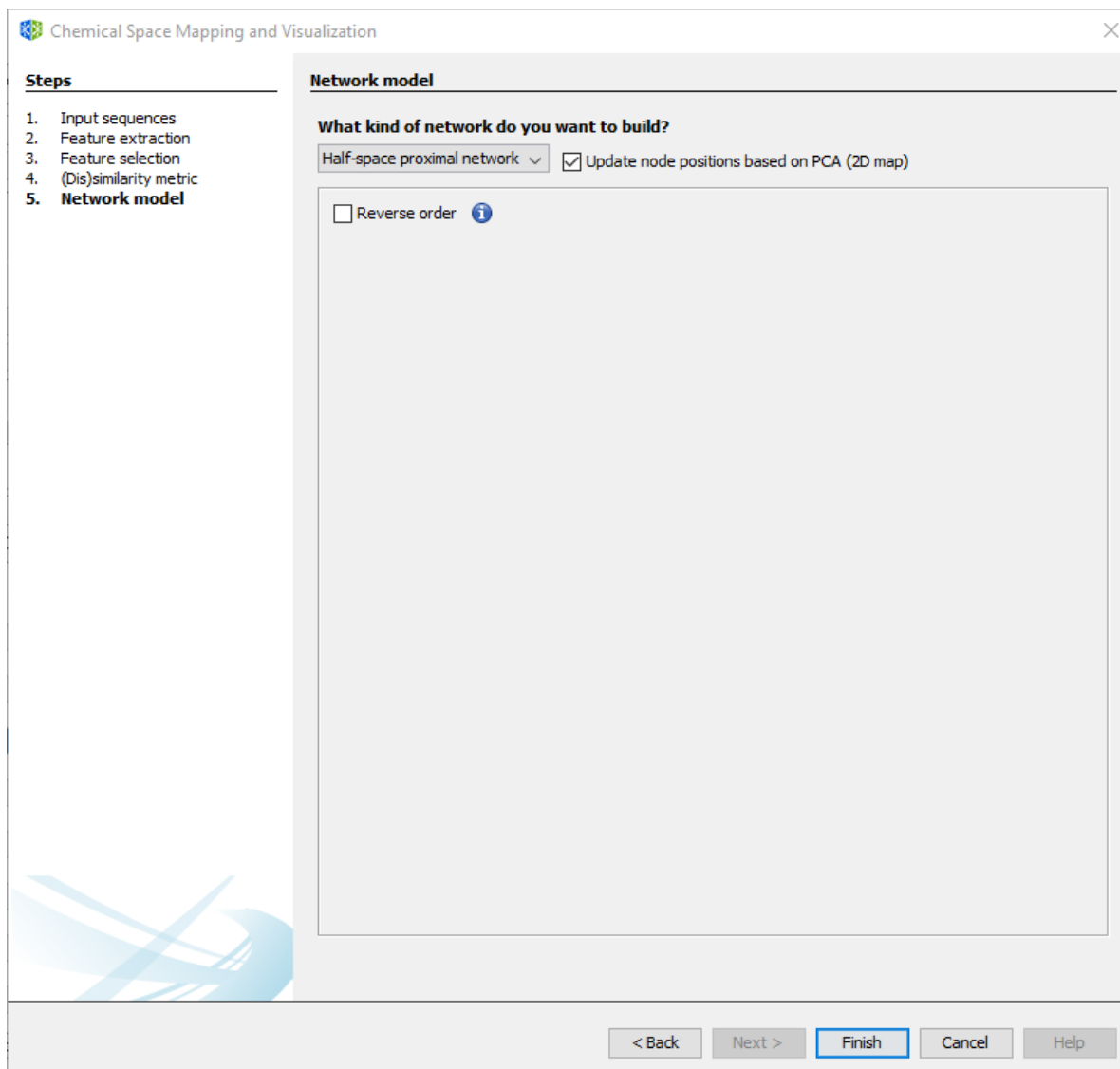


Figure 3.8: Wizard Step 5: Network model.

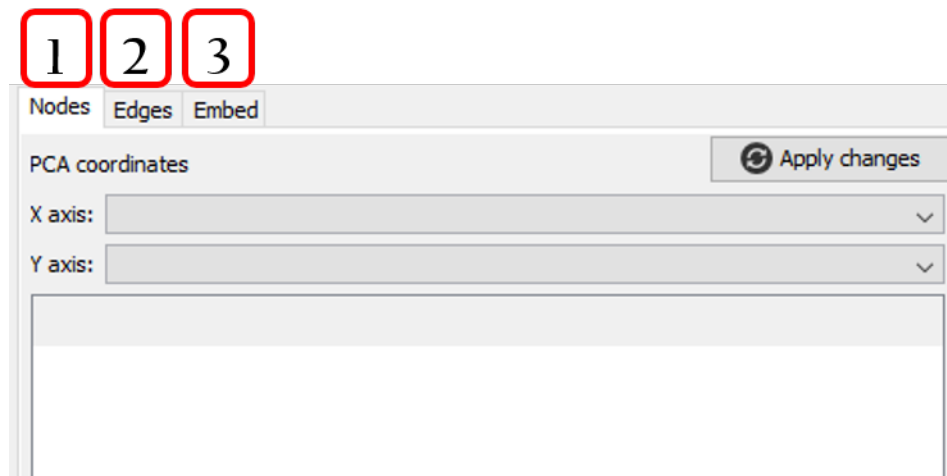


Figure 3.9: Network model options.

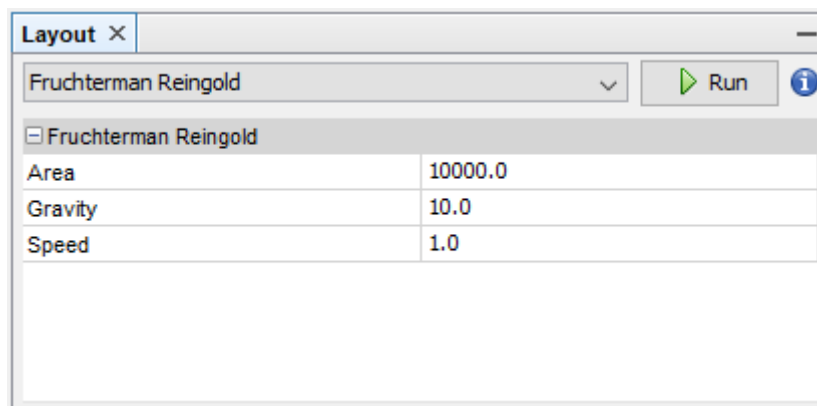
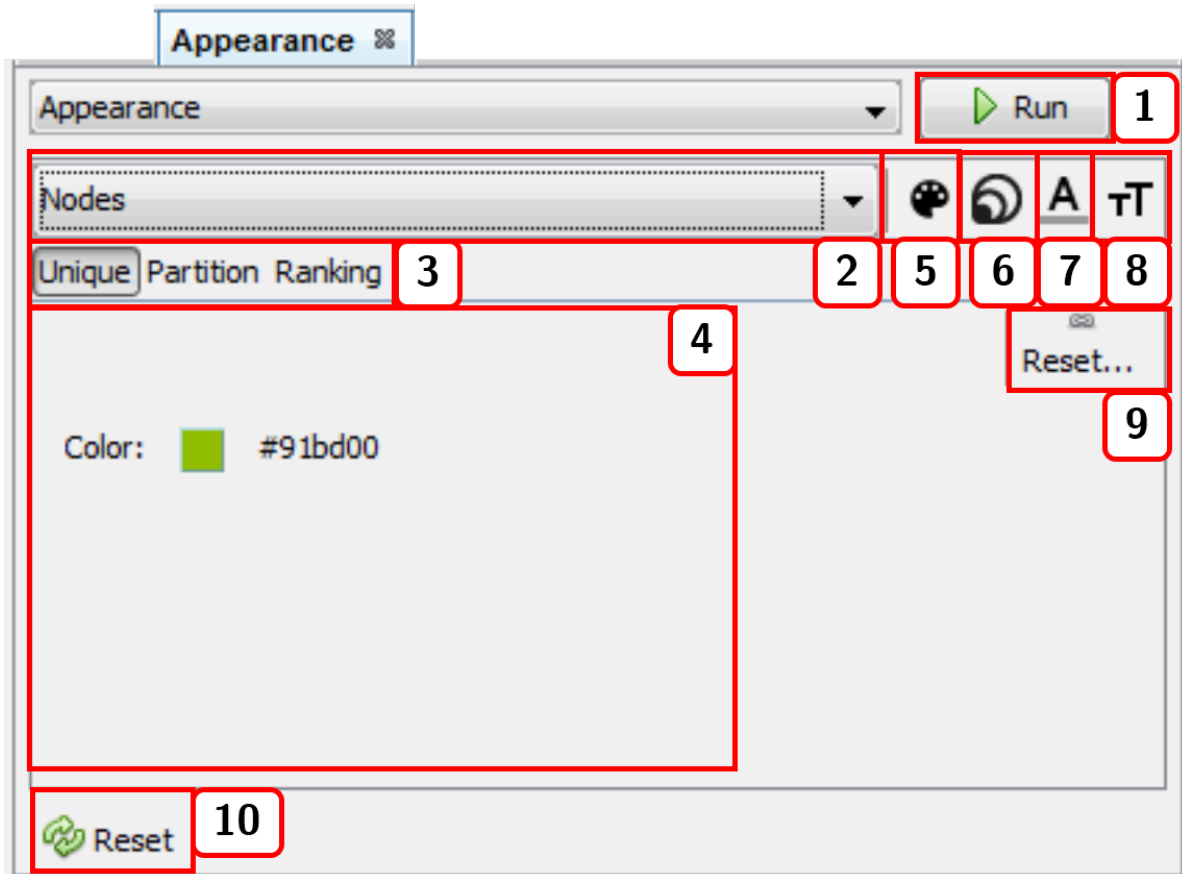


Figure 3.10: option.



1. Runs the appearance customization of either nodes or edges. If the **Preview** window is active in the **Network visualization** window, you need to press the button **Refresh** from mentioned window to update the network view.
2. Selects the elements (either **Nodes** or **Edges**) whose appearance is to be changed.
3. Applies configuration via **Unique**, **Partition**, or **Ranking** functions. For nodes, the calculated measures are available as **Partition** or **Ranking** options.
4. Modifiable configurations panel. For color options, you need to press and drag the cursor to the desired color, or press right-click to open the color window.
5. Changes the color of either **Nodes** or **Edges** (if edges are not taking the color of attached nodes, see sect. 2.5.2).
6. Changes **Nodes** size (this option only applies to nodes).
7. Changes label color of either or **Nodes** or **Edges**.
8. Changes label size of either or **Nodes** or **Edges**.
9. Resets current options.
10. Resets customization to the default appearance.

3.6 Clustering

A clustering panel may be opened from **Tools** :arrow_right: **Network** :arrow_right: **Clustering** :arrow_right: [clustering algorithm]. For instance, k-means:

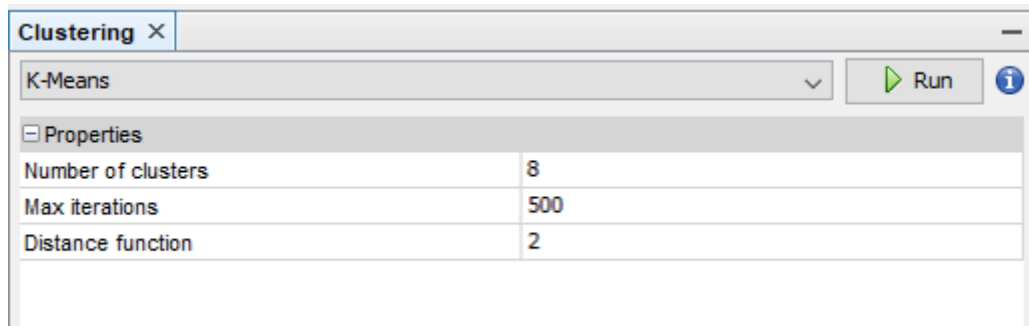


Figure 3.11: k-means clustering option.

i Note

After running the clustering algorithm, you may visualize the network structure in **Tools** :arrow_right: **Network** :arrow_right: **Appearance** :arrow_right: **Nodes** :arrow_right: **Partition**.

3.7 Centrality

A centrality panel may be opened from **Tools** :arrow_right: **Network** :arrow_right: **Centrality** :arrow_right: [measure option]. For instance, Betweenness Centrality:

i Note

After running the centrality measure, you may customize the appearance of nodes according to the centrality values in **Tools** :arrow_right: **Network** :arrow_right: **Appearance** :arrow_right: **Nodes** :arrow_right: **Ranking**.

3.8 Case study

In this case study, we will try to answer the following questions for a given sequence of interest.

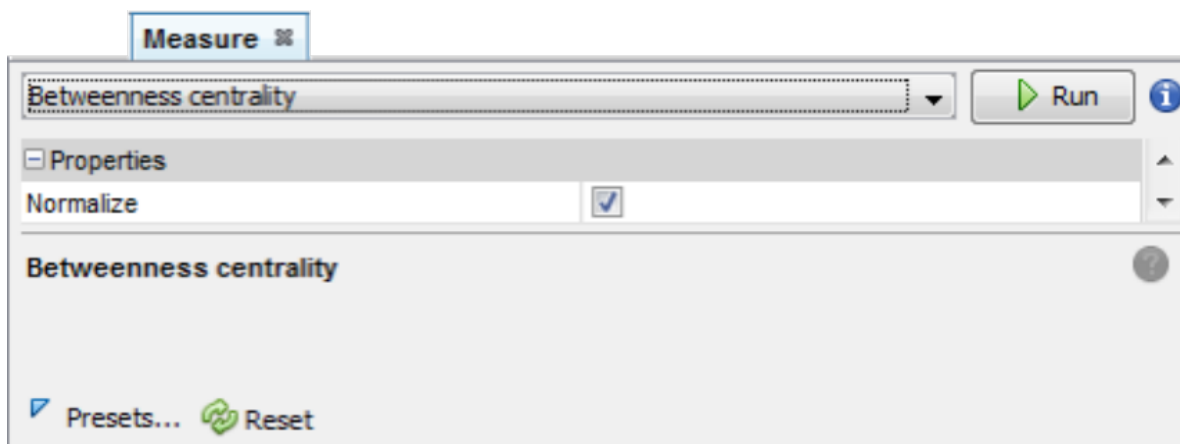


Figure 3.12: Betweenness Centrality option.

```
>Example sequence
FLPAIVGAAGQFLPKIFCAISKKC
```

3.8.1 Which biological database holds peptides similar to the sequence of interest?

Step 1: Opens the Search panel with the commands **Tools** :arrow_right: **Peptide search by** :arrow_right: **Single query sequence**. Types the query sequence in the input field, configures the sequence alignment at 70% of sequence identity, and press **Run**. This search should return 25 peptide sequences and 595 metadata relationships.

Step 2: Creates the metadata network by selecting the option **Database**.

Step 3: In the graph table view of **Navigator** window, select the option **Columns...**, then mark **Degree** and click **OK**.

We can sort the graph table by node Degree by clicking the Degree column 2 times, and now we can observe that the database **SATPdb** contains the most similar sequences to the query sequence.

3.8.2 What are the biological functions of peptides similar to the sequence of interest?

Follow the **Step 1** of the previous example.

Step 2: Creates the metadata network by selecting the option **Function**.

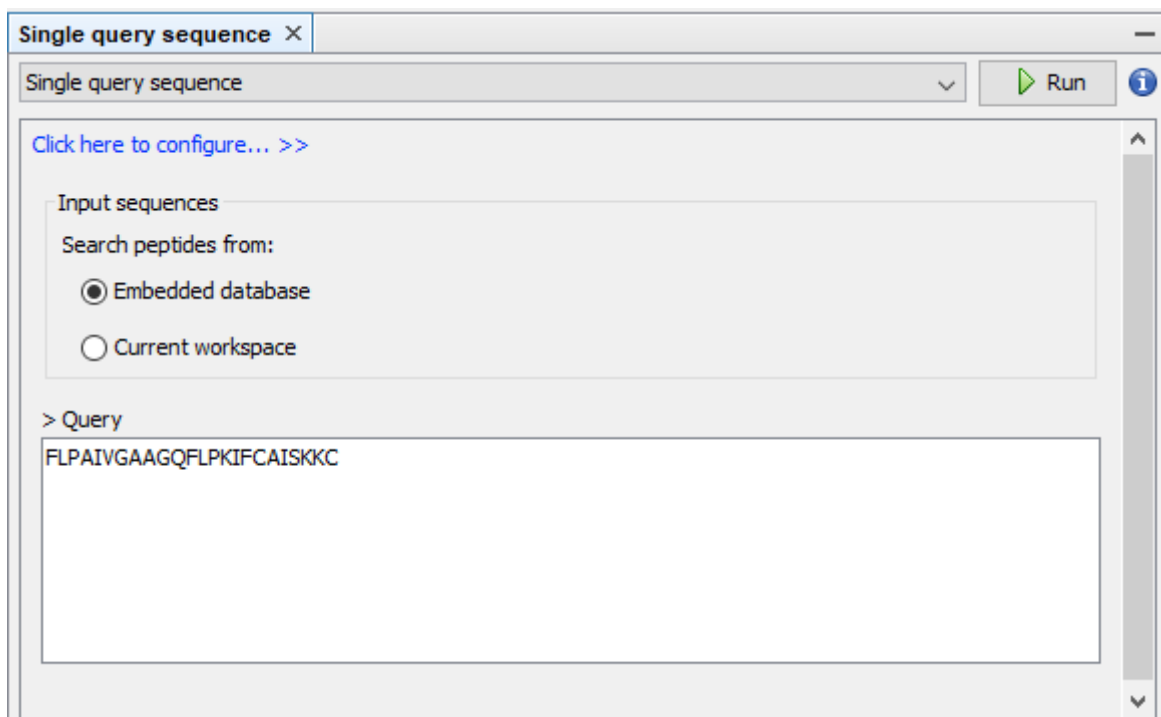


Figure 3.13: Sequence search panel

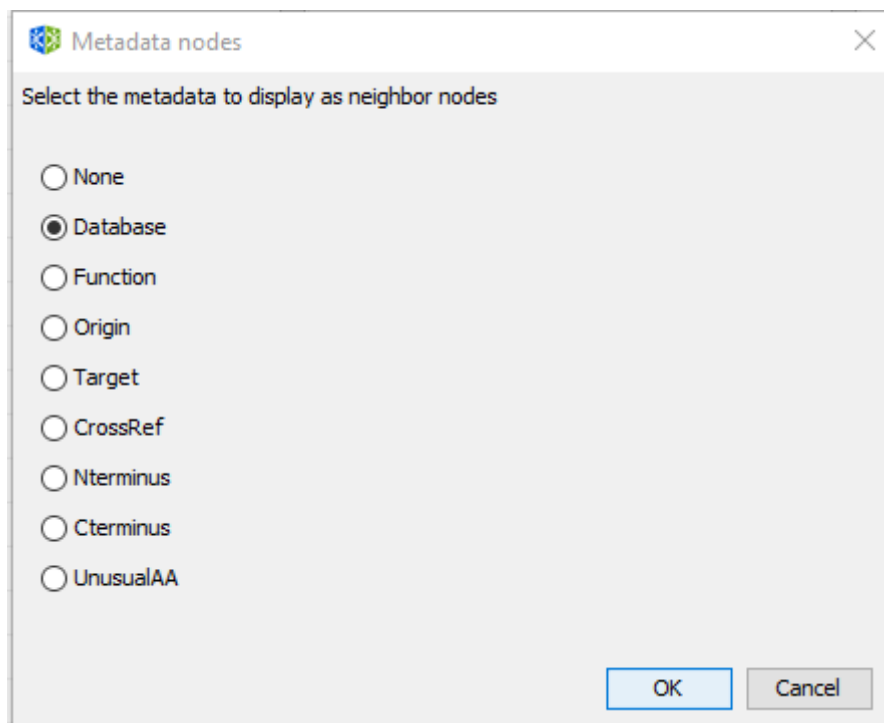


Figure 3.14: Options for metadata network

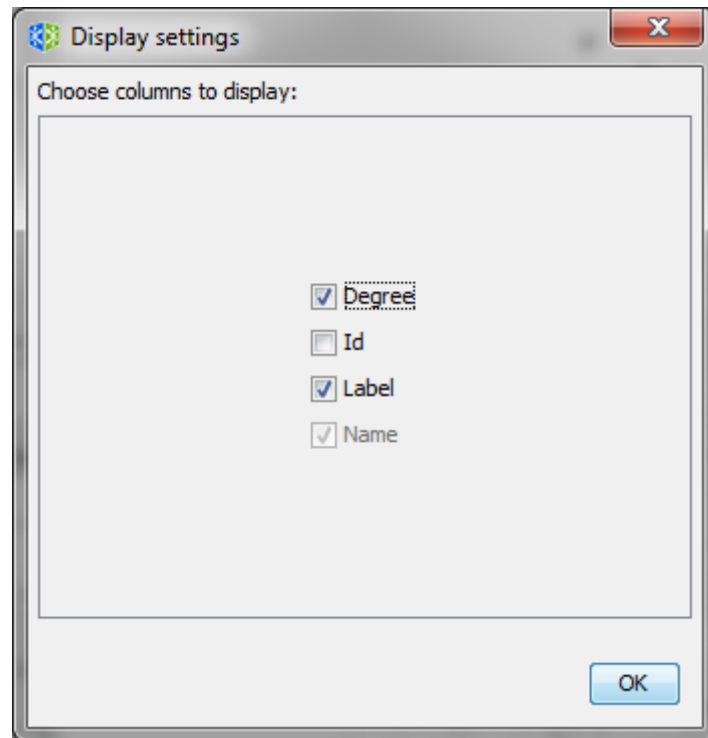
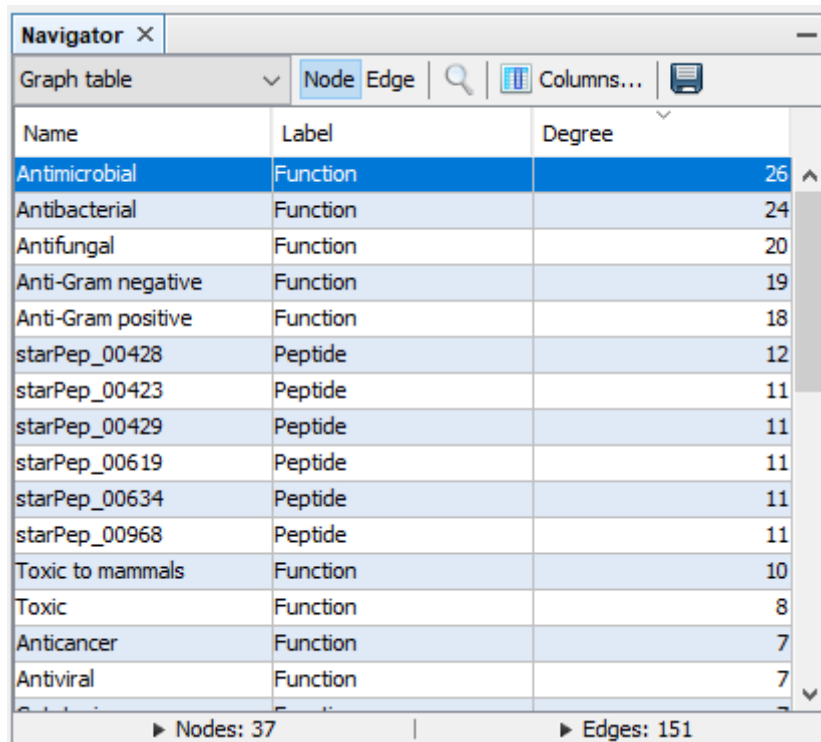


Figure 3.15: Display settings

Navigator X		
Graph table		
Node Edge		
Name	Label	Degree
SATPdb	Database	21
DADP	Database	20
YADAMP	Database	19
APD	Database	19
DRAMP_General	Database	19
dbAMP	Database	19
ADAM	Database	18
DBAASP	Database	18
CAMP_Validated	Database	18
LAMP_Experimental	Database	18
starPep_00026	Peptide	17
starPep_00027	Peptide	17
starPep_00028	Peptide	17
starPep_00029	Peptide	17
starPep_00030	Peptide	17
Nodes: 44 Edges: 268		

Figure 3.16: Graph table

Step 3: In the graph table view of Navigator window, select the option **Columns...**, then mark **Degree** and click OK.



Name	Label	Degree
Antimicrobial	Function	26
Antibacterial	Function	24
Antifungal	Function	20
Anti-Gram negative	Function	19
Anti-Gram positive	Function	18
starPep_00428	Peptide	12
starPep_00423	Peptide	11
starPep_00429	Peptide	11
starPep_00619	Peptide	11
starPep_00634	Peptide	11
starPep_00968	Peptide	11
Toxic to mammals	Function	10
Toxic	Function	8
Anticancer	Function	7
Antiviral	Function	7

Nodes: 37 | Edges: 151

Figure 3.17: Graph table

Step 4: In the **Network** visualization window, select the following options.

1. Shows node labels.
2. Disables the option **Show peptide labels**.
3. Modifies the label size to **Node size**.

In the **Appearance** panel (see sect. 3.5), customizes the appearance of nodes for sizing and coloring nodes according to the degree measure.

1. In the **Nodes** view, select **Node size** :arrow_right: **Ranking** :arrow_right: **Degree**. Set min and max sizes to 5 and 100 respectively, select the interpolator **Bezier**, select the second predefined spline and press Run.
2. In the **Nodes** view, customizes **Node color** :arrow_right: **Ranking** :arrow_right: **Degree**, and press Run.

Run the **Tools** :arrow_right: **Network** :arrow_right: **Layout** :arrow_right: **Fruchterman Reingold** about 10s and then press stop. The result may be similar to the following network:

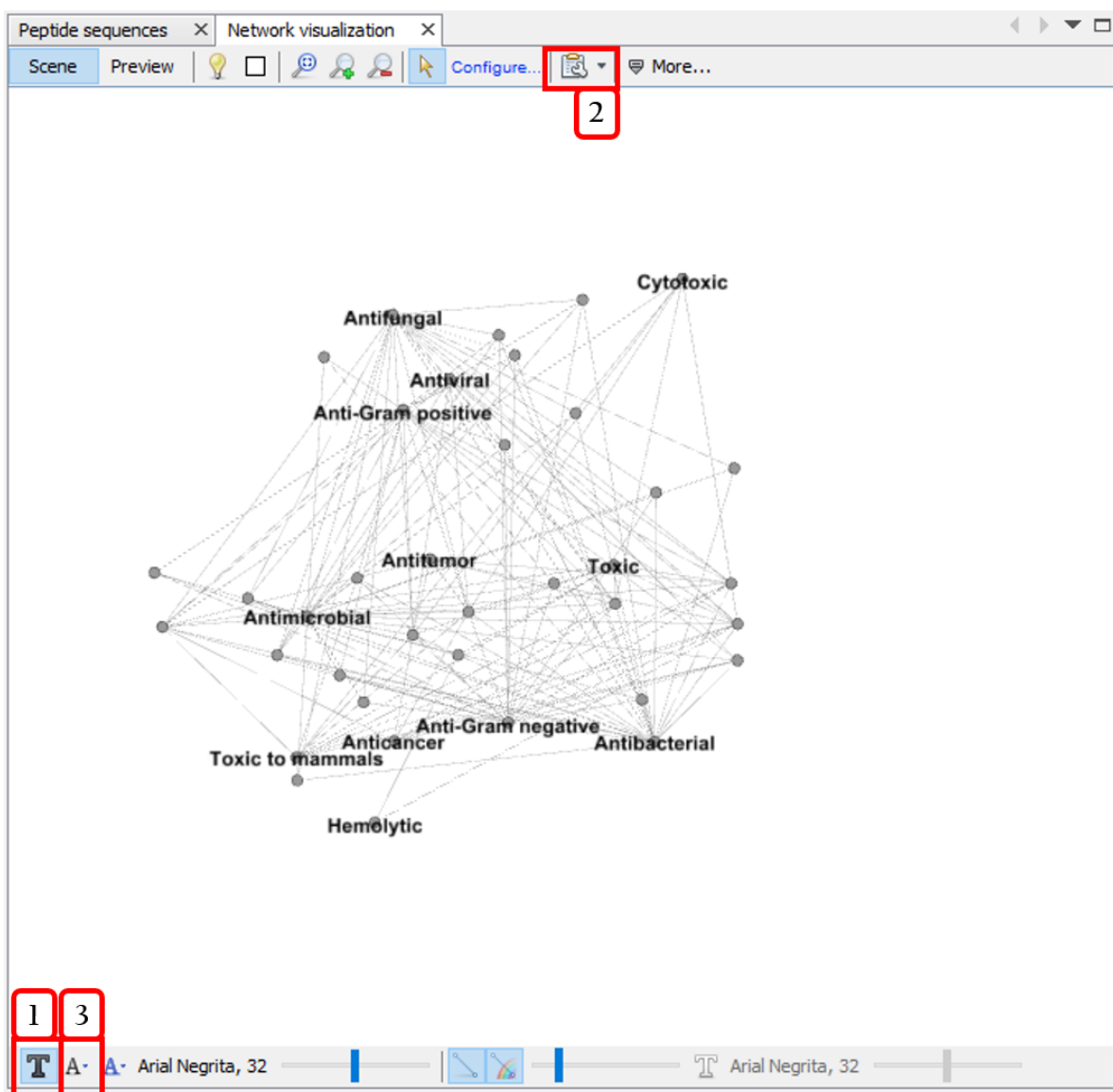


Figure 3.18: Network visualization

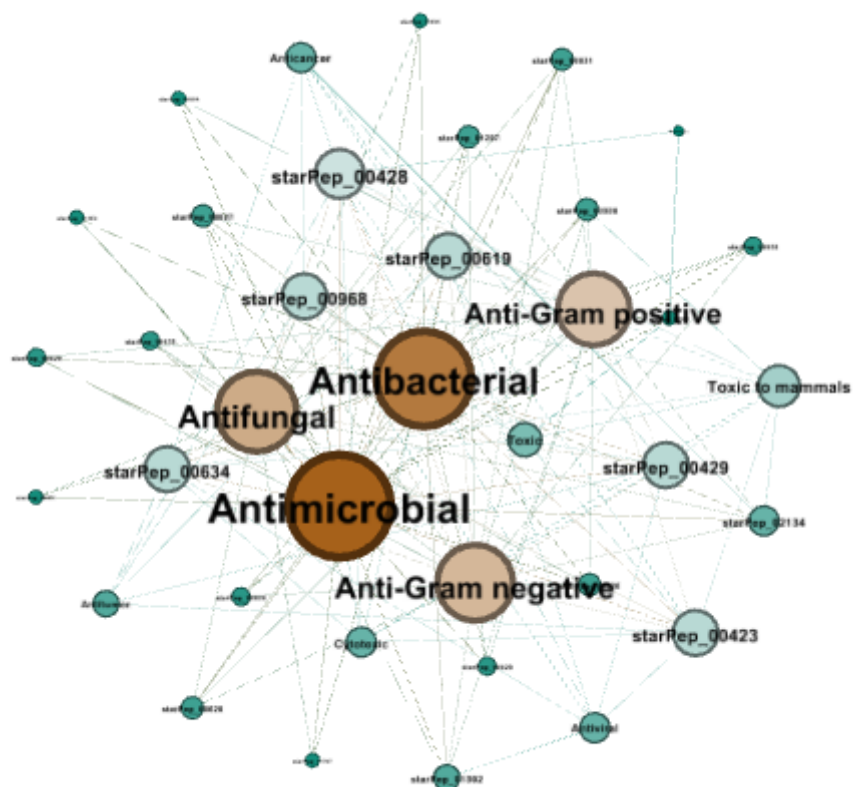


Figure 3.19: Metadata network