# clusterProfiler: an R package for Statistical Analysis and Visualization of Functional Profiles for Genes and Gene Clusters

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#### 1 Introduction

In recently years, high-throughput experimental techniques such as microarray and mass spectrometry can identify many lists of genes and gene products. The most widely used strategy for high-throughput data analysis is to identify different gene clusters based on their expression profiles. Another commonly used approach is to annotate these genes to biological knowledge, such as Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG), and identify the statistically significantly enriched categories. These two different strategies were implemented in many bioconductor packages, such as *Mfuzz* and *BHC* for clustering analysis and *GOstats*(Falcon et al., 2007) for GO enrichment analysis.

After clustering analysis, researchers not only want to determine whether there is a common theme of a particular gene cluster, but also to compare the biological themes among gene clusters, which have different expression profiles. To bridge this gap, we designed *clusterProfiler*, for comparing functional profiles among gene clusters.

This document presents an introduction to the use of *clusterProfiler*, an R package for the analysis of lists of genes and gene clusters based on their GO annotation distribution or enrichment categories of GO and KEGG, and provides methods for visulization.

## 2 Quick start

The following lines provide a quick and simple example on the use of *clusterProfiler* to explore a set of genes and compare gene clusters.

The analysis proceeds as follows:

• First a sample dataset is loaded. This dataset contains 5 gene clusters.

```
> require(clusterProfiler)
> data(gcSample)
> gcSample
$C1
  [1] "23753" "57222" "5036" "5037" "10111" "10856" "6228"
  [8] "9361" "1537" "3376" "6124" "4175" "2539"
$C2
```

```
[1] "6629" "10291" "7094" "3843" "6611" "10399" "10576"
  [8] "4705" "5216" "6697" "5868" "80777" "1973" "1938"
 [15] "23450" "9343" "1917" "9520"
 $C3
  [1] "4905"
             "10383" "10953" "645958" "7280"
                                                  "10381"
              "5985" "23197" "290" "309"
  [7] "5869"
                                                  "10577"
 [13] "23071" "121504" "2495"
                                "653226" "84617"
 $C4
  [1] "51552" "8336" "302"
                             "5984" "50814" "8813" "871"
             "23344" "4134" "10262" "22919" "159"
 $C5
  [1] "11171" "8243"
                       "112464" "2194"
                                         "9318"
                                                  "79026"
  [7] "1654" "65003" "6240"
                                "3476"
                                         "6238"
                                                  "3836"
 [13] "4176" "1017" "249"
• Use groupGO for genes classification based on GO distribution at a specific level.
 > x <- groupGO(gene = gcSample[[1]], organism = "human",
 + ont = "CC", level = 2, readable = TRUE)
 > summary(x)
                  GOID
                                    Description Count
 GO:0005576 GO:0005576
                           extracellular region 1
 GO:0005623 GO:0005623
                                           cell
                                                   13
 GO:0019012 GO:0019012
                                         virion
                                                   0
 GO:0031974 GO:0031974 membrane-enclosed lumen
                                                    7
                       macromolecular complex
 GO:0032991 GO:0032991
                                                   6
 GO:0043226 GO:0043226
                                                  13
                                      organelle
 GO:0044421 GO:0044421 extracellular region part
                                                   1
 GO:0044422 GO:0044422
                                organelle part
 GO:0044423 GO:0044423
                                                  Ω
                                    virion part
 GO:0044456 GO:0044456
                                                   1
                                   synapse part
 GO:0044464 GO:0044464
                                      cell part
                                                   13
 GO:0045202 GO:0045202
                                        synapse
                                                    1
 GO:0055044 GO:0055044
                                       symplast
                                                    0
                                                                                Ge
 GO:0005576
 GO:0005623 SDF2L1/ERGIC1/PA2G4/PEBP1/RAD50/RUVBL2/RPS23/LONP1/CYC1/IARS/RPL4/MCM6/
 GO:0019012
                                             SDF2L1/PA2G4/RAD50/RUVBL2/LONP1/RPL4/
 GO:0031974
                                                    PA2G4/RAD50/RUVBL2/RPS23/RPL4/
 GO: 0032991
 GO:0043226 SDF2L1/ERGIC1/PA2G4/PEBP1/RAD50/RUVBL2/RPS23/LONP1/CYC1/IARS/RPL4/MCM6/
 GO:0044421
 GO:0044422
                 SDF2L1/ERGIC1/PA2G4/PEBP1/RAD50/RUVBL2/RPS23/LONP1/CYC1/RPL4/MCM6/
 GO:0044423
 GO:0044464 SDF2L1/ERGIC1/PA2G4/PEBP1/RAD50/RUVBL2/RPS23/LONP1/CYC1/IARS/RPL4/MCM6/
```

Р

GO:0045202

GO:0055044

• Use enrichGO for GO enrichment analysis.

```
> y <- enrichGO(gene = gcSample[[2]], organism = "human",
+ ont = "MF", pvalueCutoff = 0.01, readable = TRUE)</pre>
```

• Use enrichKEGG for KEGG pathway enrichment analysis.

```
> z <- enrichKEGG(gene = gcSample[[3]], organism = "human",
    pvalueCutoff = 0.05, readable = TRUE)
> summary(z)
     pathwayID
                                           Description
                 Pathogenic Escherichia coli infection
05130 hsa05130
04145 hsa04145
                                             Phagosome
04540 hsa04540
                                           Gap junction
04962 hsa04962 Vasopressin-regulated water reabsorption
04614 hsa04614
                              Renin-angiotensin system
     GeneRatio BgRatio
                              pvalue
                                          gvalue
05130
        4/17 58/5894 1.826892e-05 0.0002115348
04145
         5/17 156/5894 5.827611e-05 0.0003373880
04540
         4/17 90/5894 1.039489e-04 0.0004012064
          2/17 44/5894 6.898981e-03 0.0199707355
04962
          1/17 17/5894 4.798133e-02 0.1111146614
04614
                              geneID Count
05130
           TUBB2C/TUBB2A/TUBB3/TUBB6
04145 TUBB2C/TUBB2A/TUBB3/RAB5B/TUBB6
04540
           TUBB2C/TUBB2A/TUBB3/TUBB6
04962
                           NSF/RAB5B
04614
                               ANPEP
```

The input parameters of *gene* is a vector of entrez genes or ORF IDs (for yeast), and *organism* must be one of "human", "mouse", and "yeast", according to the gene IDs. For GO analysis, *ont* must be assigned to one of "BP", "MF", and "CC" for biological process, molecular function and cellular component, respectively. In groupGO, the *level* specify the GO level for gene projection. In enrichment analysis, the *pvalueCutoff* is to restrict the result based on their pvalues. Consider multiple testing, qvalues are also provided, for estimating FDR. The *readable* is a logical parameter, if TRUE, the gene IDs will map to gene symbols.

In addition, these results can be visualized by our plot function. For example:

```
> plot(x, title = "CC Ontology Classification, level 2",
+ font.size = 12)
> plot(z, title = "KEGG Enrichment")
```

• Gene clusters can be compared by *compareCluster*, and plotted by bar chart or dot chart.

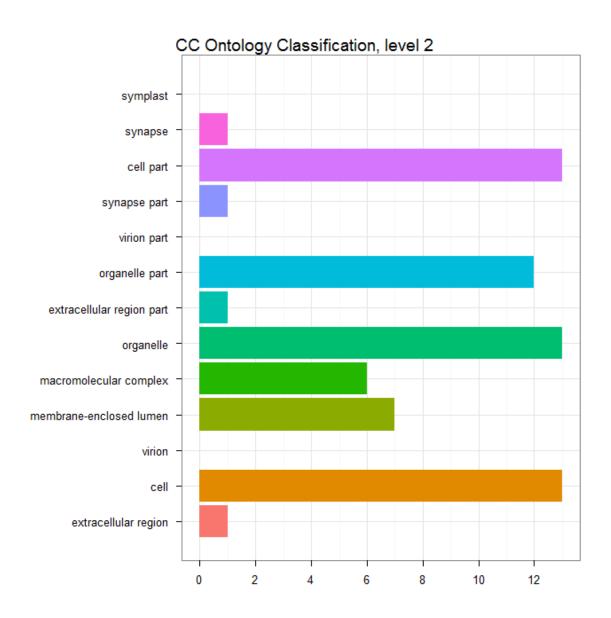


Figure 1: Example of gene classification

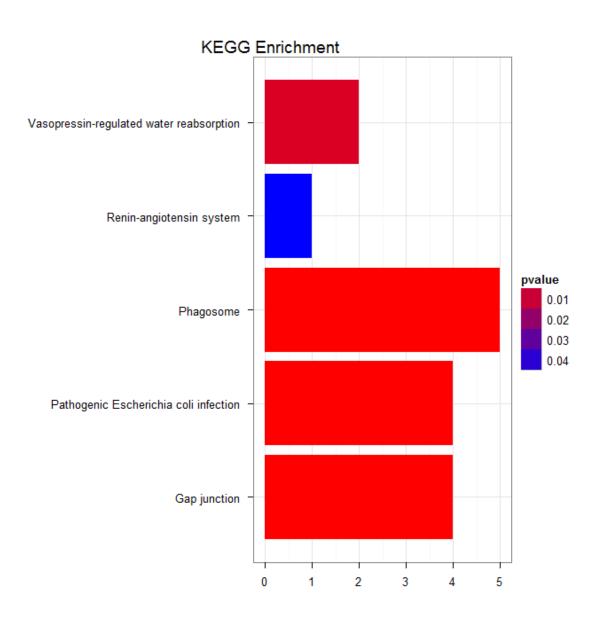


Figure 2: Example of KEGG enrichment analysis

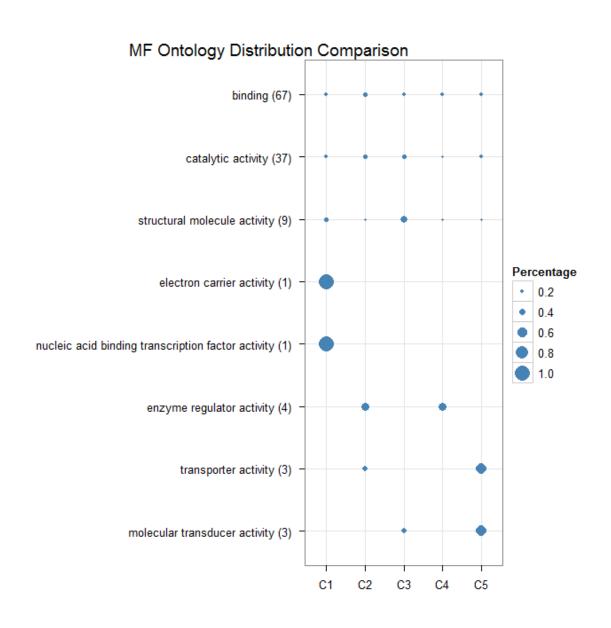


Figure 3: Example of comparing MF ontology distribution using dotplot

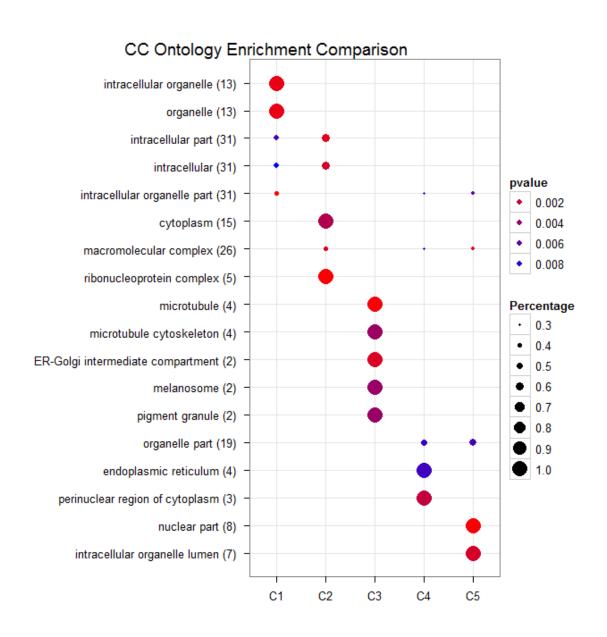


Figure 4: Example of comparing CC ontology enrichment using dot chart

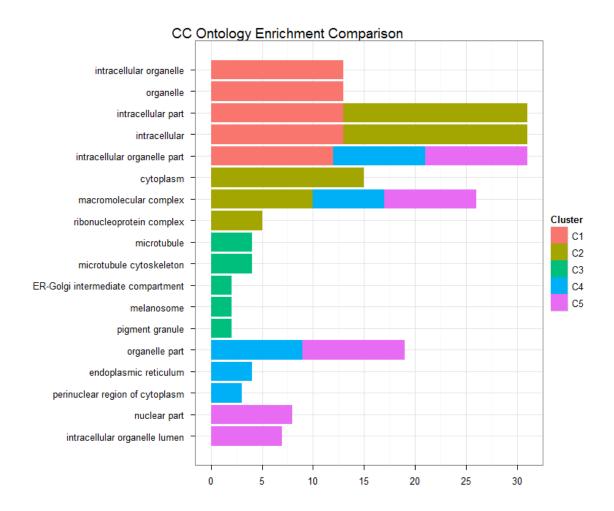


Figure 5: Example of comparing CC ontology enrichment using bar chart

```
> plot(yy, title = "CC Ontology Enrichment Comparison",
+ type = "bar", by = "count")

> zz <- compareCluster(gcSample, fun = enrichKEGG,
+ organism = "human", pvalueCutoff = 0.05)

> plot(zz, title = "KEGG Pathway Enrichment Comparison")
```

By default, only top 5 categories of each cluster was plotted. User can changes the parameter *limit* to specify how many categories of each cluster to be plotted, and if *limit* set to NULL, the whole result will be plotted. By default, the dot sizes were based on their corresponding row percentage, and user can set the parameter *by* to "count" to make the comparison based on gene counts.

We chose "percentage" as default parameter to represent the sizes of dots, since some categories may contain a large number of genes, and make the dot sizes of those small categories too small to compare. To provide the full information, we also provide number of identified genes in each category (numbers in parentheses), as shown in Figure 1. If the dot sizes were based on "count", the parentheses will not shown.

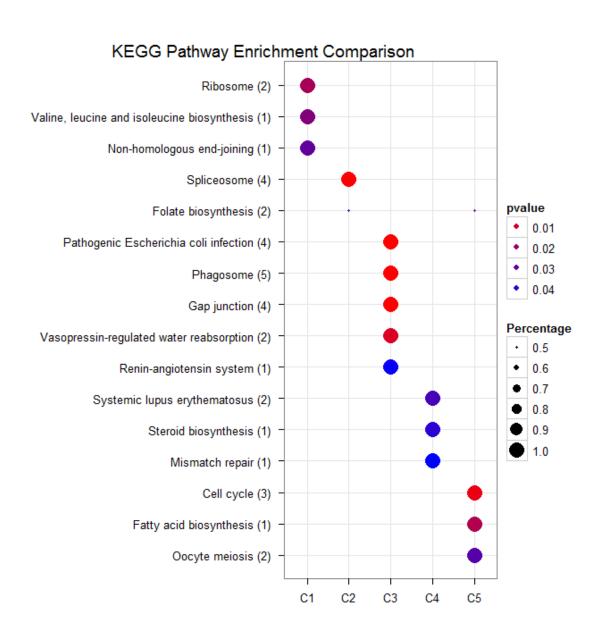


Figure 6: Example of comparing KEGG enrichment among gene clusters

The p-values indicate that which categories are more likely to have biological meanings. The dots in the image are color-encoded based on their corresponding p-values. Color gradient ranging from red to blue correspond to in order of increasing p-values. red indicate low p-values (high enrichment), and blue indicate high p-values (low enrichment). P-values were filtered out by the threshold giving by parameter *pvalueCutoff*.

We also provide q-values, which were calculated by *qvalue*, for user to control false discovery rate. FDR control is necessary since enrichment analysis carrying out hundreds, if not thousands, of tests.

• Use compareCluster as a general function for comparing gene clusters of other kind of gene-ontology associations.

clusterProfiler can cooperates with other R package for comparing other ontology analyzed results.

For example, compareCluster can call function provided by *DOSE* to compare gene-disease associations among gene clusters. Comparing gene clusters in the context of disease is a critical step in translating molecular findings from microarray, proteomics and other high-throughput methods into clinical relevance.

### **3 Session Information**

The version number of R and packages loaded for generating the vignette were:

```
R version 2.13.0 (2011-04-13)
Platform: i686-pc-linux-qnu (32-bit)
locale:
 [1] LC_CTYPE=en_US.UTF-8
                              LC_NUMERIC=C
 [3] LC_TIME=en_US.UTF-8
                              LC_COLLATE=C
 [5] LC_MONETARY=C
                              LC MESSAGES=en US.UTF-8
 [7] LC_PAPER=en_US.UTF-8 LC_NAME=C
 [9] LC ADDRESS=C
                              LC TELEPHONE=C
[11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
attached base packages:
[1] grid stats
                      graphics grDevices utils
[6] datasets methods
                       base
other attached packages:
[1] clusterProfiler_1.1.22 RSQLite_0.9-4
[5] proto_0.3-9.1
                         ggplot2_0.8.9
                        reshape_0.8.4
[7] plyr_1.5.1
loaded via a namespace (and not attached):
 [1] AnnotationDbi_1.14.0 Biobase_2.12.0
 [3] GO.db_2.5.0
                       KEGG.db_2.5.0
 [5] org.Hs.eg.db_2.5.0 org.Mm.eg.db_2.5.0
 [7] org.Sc.sgd.db_2.5.0 qvalue_1.26.0
                       tools_2.13.0
 [9] tcltk_2.13.0
```

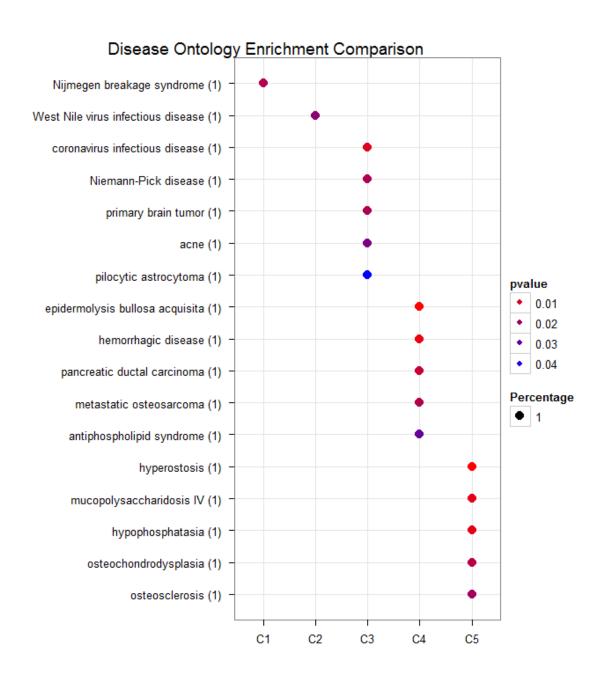


Figure 7: Example of comparing disease ontology enrichment among gene clusters

## References

S. Falcon, , and R. Gentleman. Using gostats to test gene lists for go term association. *Bioinformatics*, 23: 257–258, 2007.