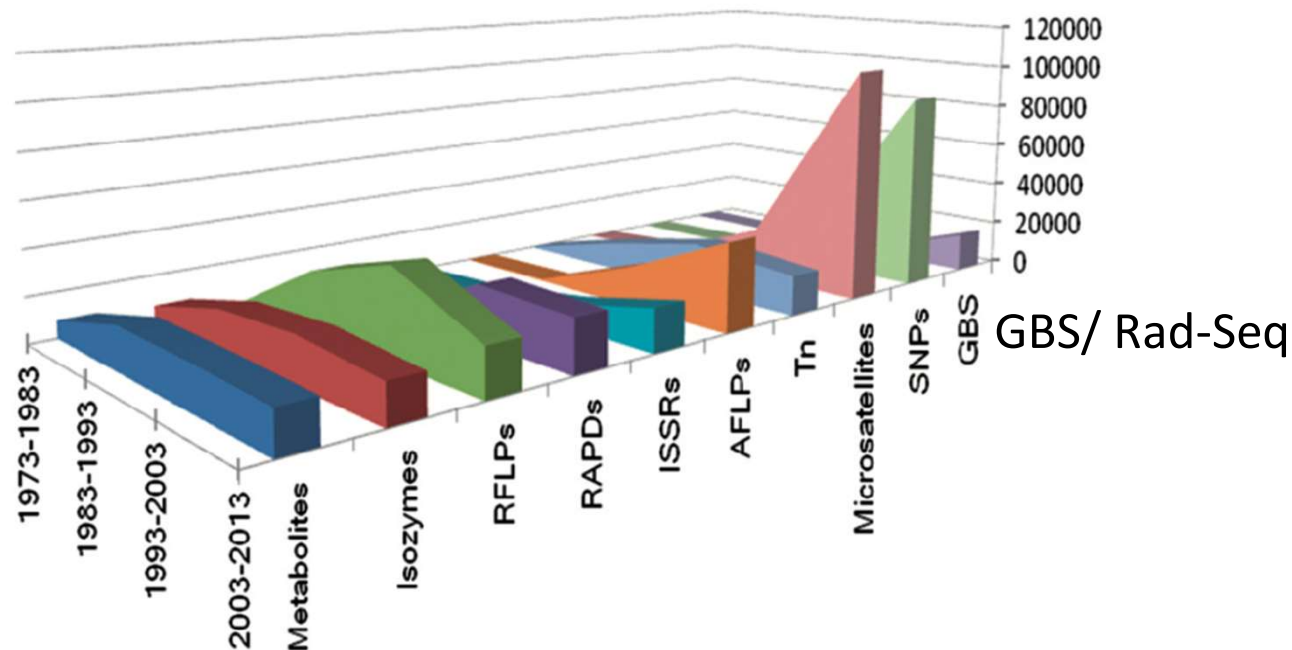


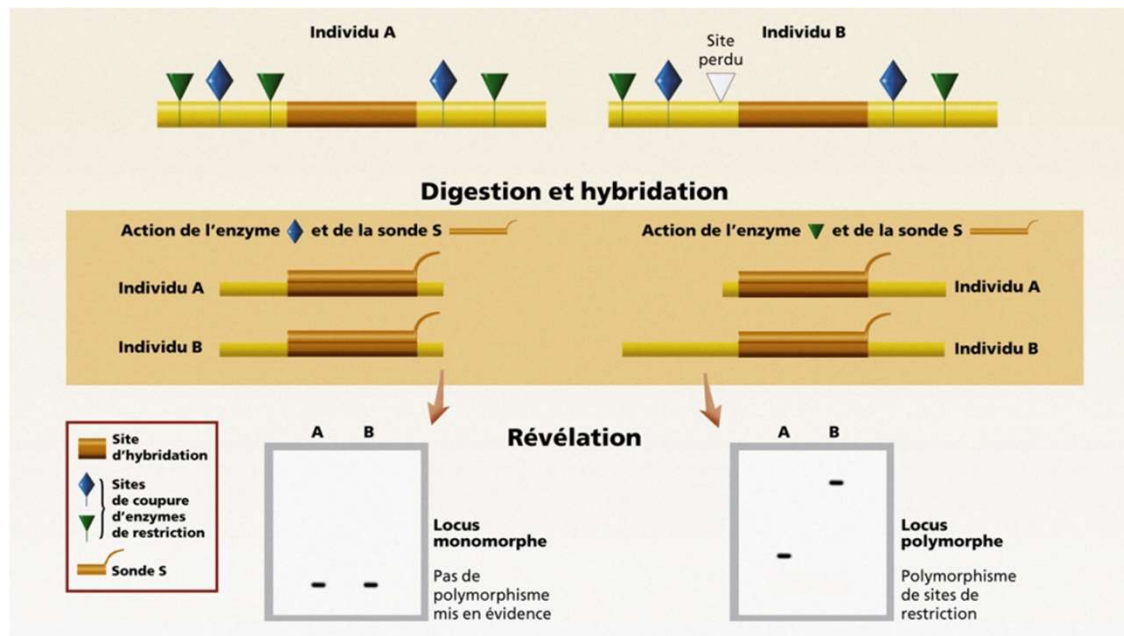
# “Restriction enzyme methods for reduced genomic representation”

## Genotyping for Population Genomics (and shallow phylogenetics / phylogeography)



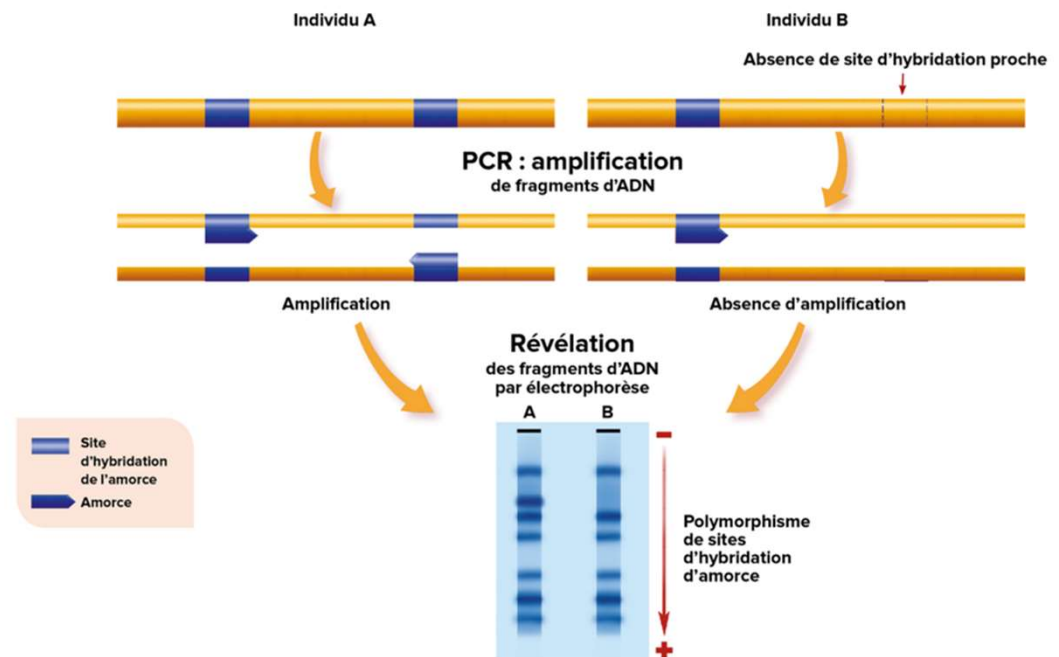
Popularity of different molecular marker systems over the past 50 years.

# Restriction enzyme methods



RFLP: *Restriction Fragments Length Polymorphism*

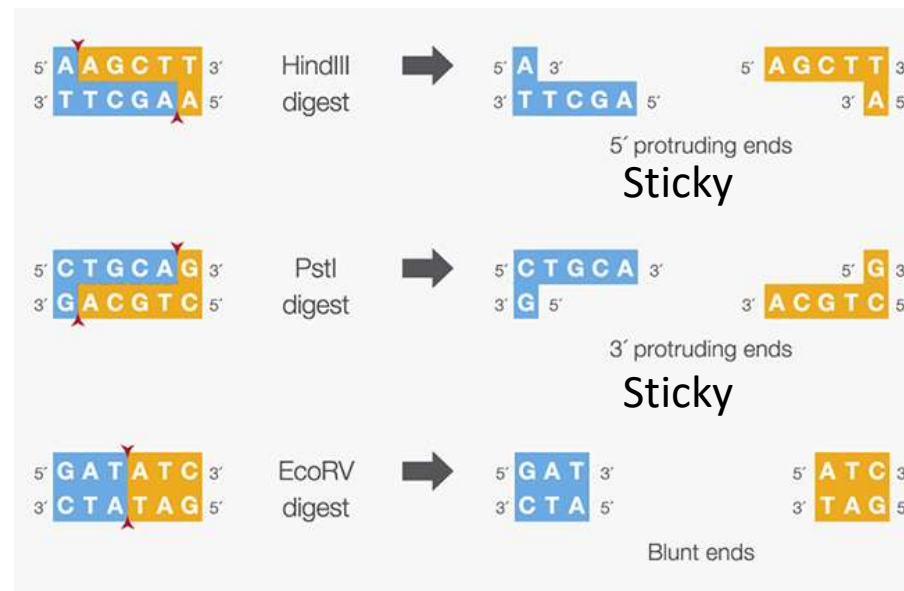
AFLP: *Amplification Fragment Length Polymorphism*



# Restriction enzyme methods: RAD-Seq and derived methods

RAD-Seq = Restriction Site-Associated DNA Sequencing

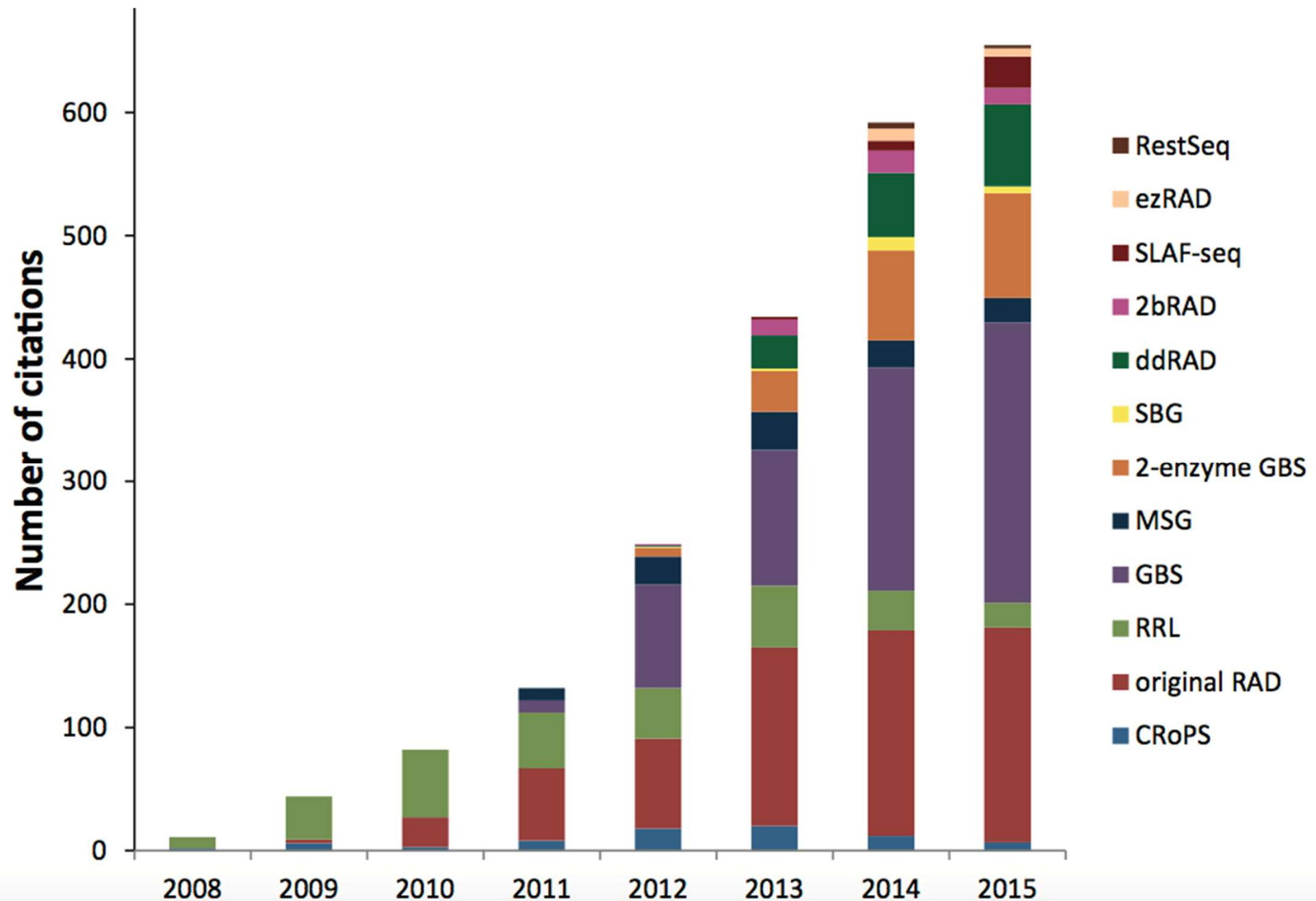
1- Genome complexity is reduced by cutting the DNA with restriction enzymes.



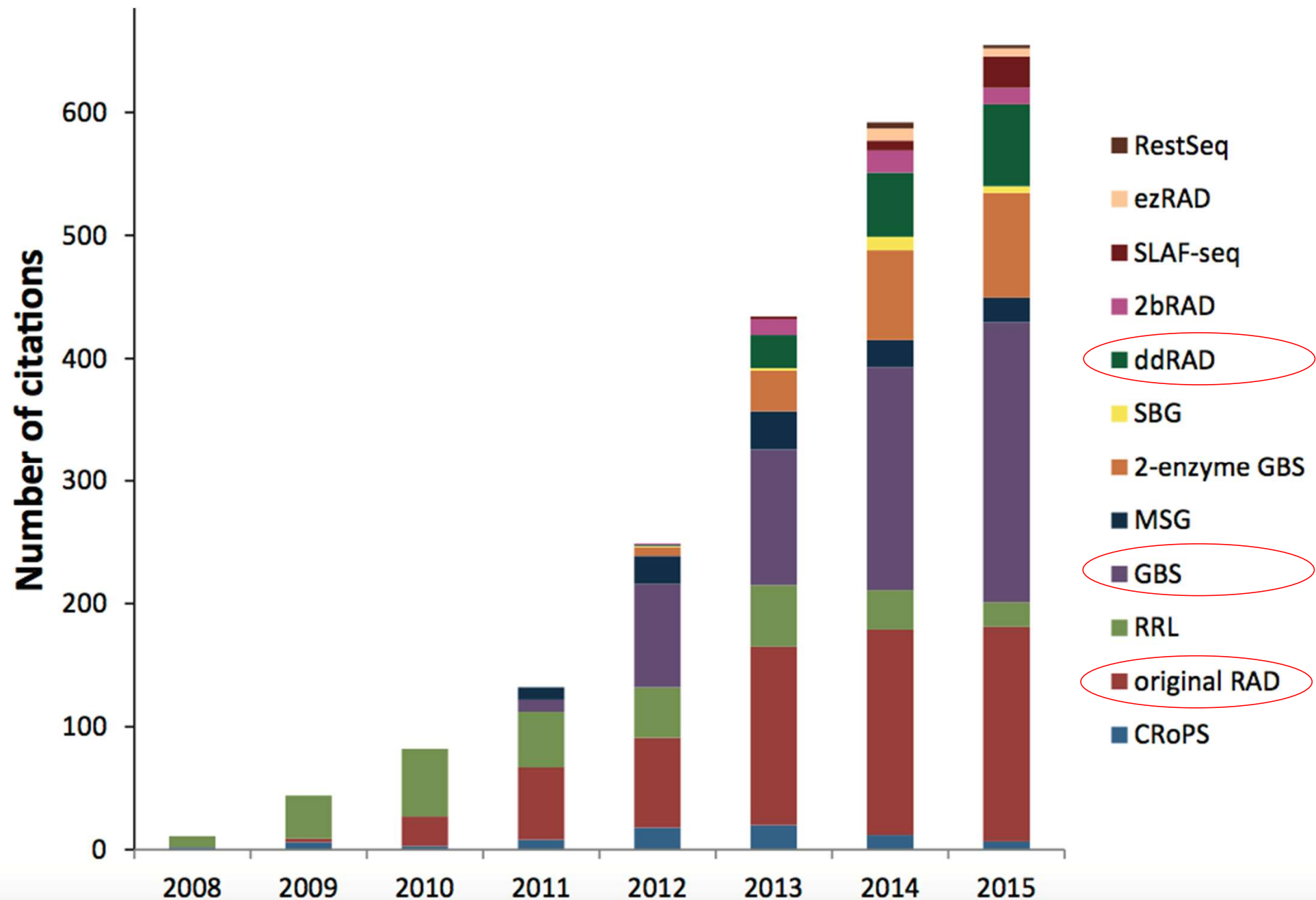
2- Only sequences associated with restriction sites are sequenced

3- There is a whole series of methods derived from the initial protocol

# Restriction enzyme methods: RAD-Seq and derived methods



# Restriction enzyme methods: RAD-Seq and derived methods



# Restriction enzyme methods: RAD-Seq and derived methods

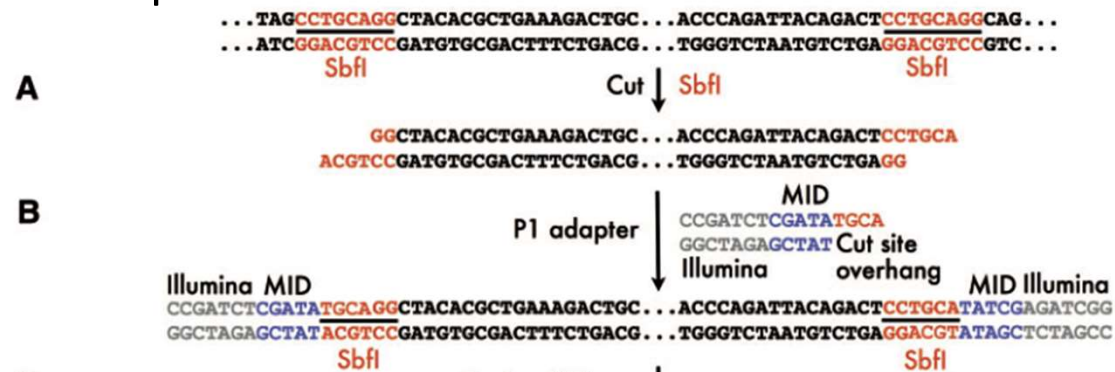
## The process of RADSeq

A



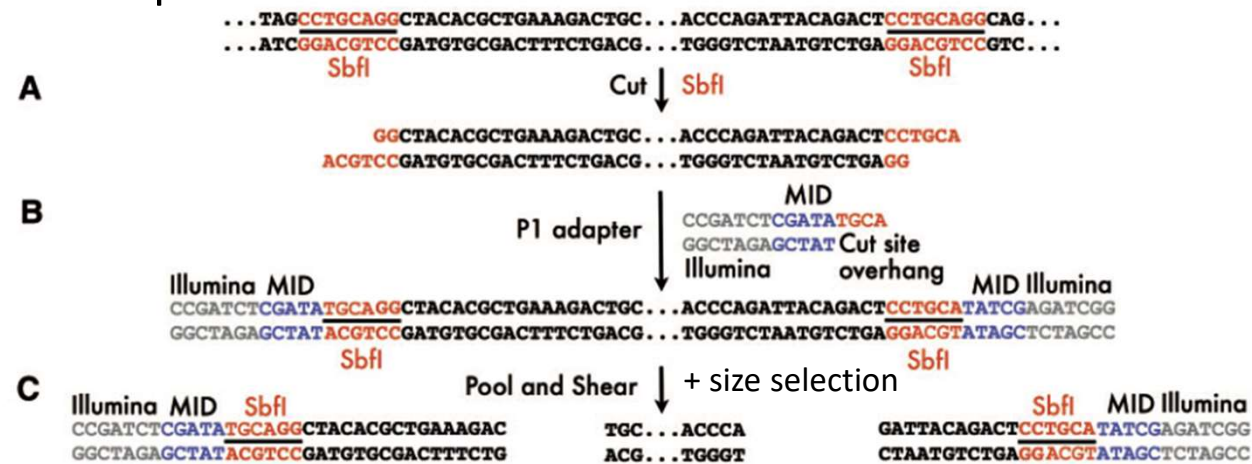
# Restriction enzyme methods: RAD-Seq and derived methods

## The process of RADSeq



# Restriction enzyme methods: RAD-Seq and derived methods

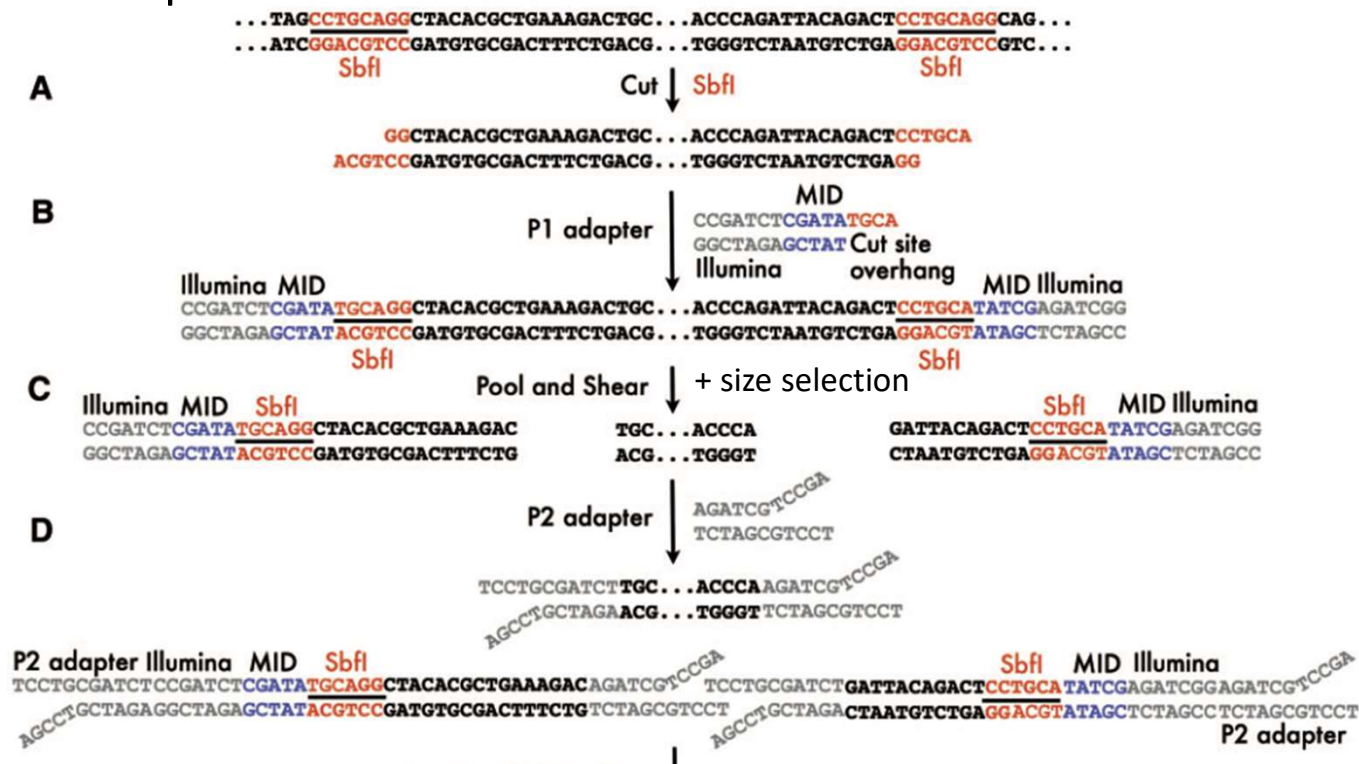
## The process of RADSeq





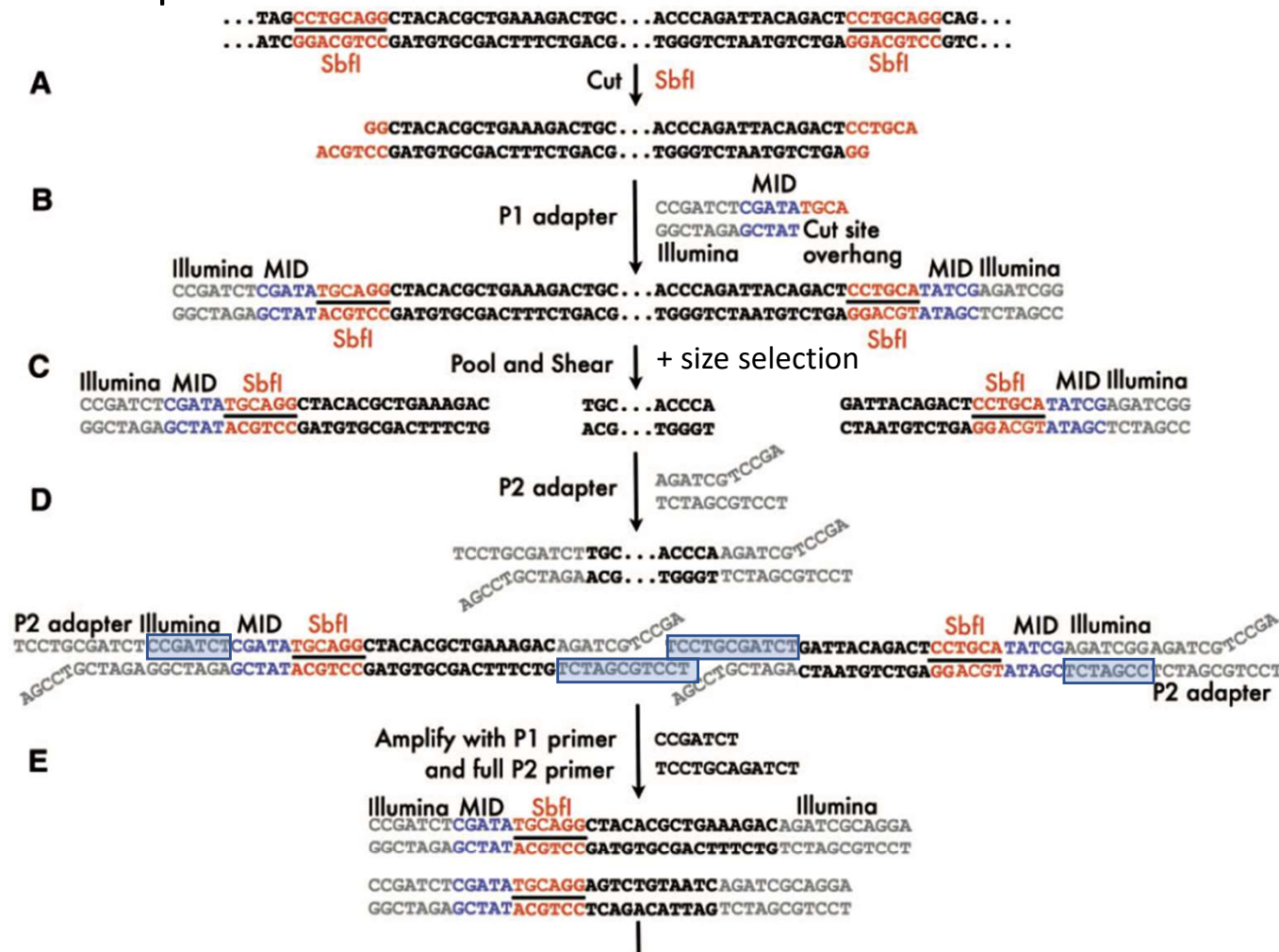
# Restriction enzyme methods: RAD-Seq and derived methods

## The process of RADSeq



# Restriction enzyme methods: RAD-Seq and derived methods

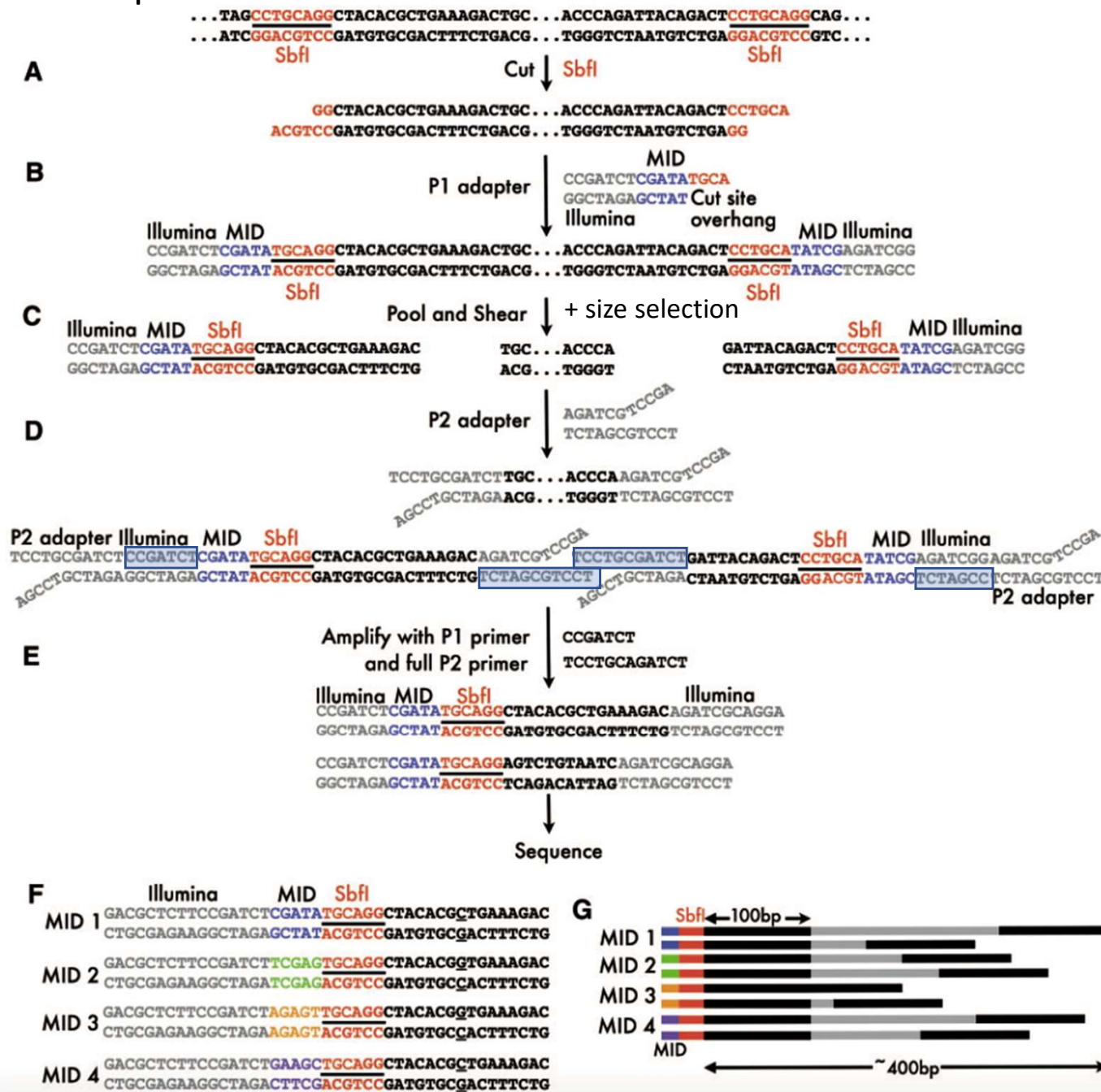
## The process of RADSeq



# Restriction enzyme methods: RAD-Seq and derived methods

# The process of RADSeq

Davey et al. 2011

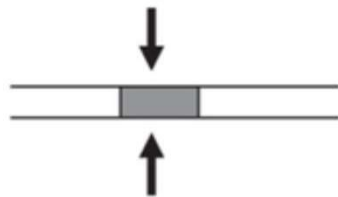


# Restriction enzyme methods: RAD-Seq and derived methods

## 1- Sequencing of fragments adjacent to single restriction enzyme cut sites:

- **original RADseq** (Baird et al. 2008)
- **2bRAD** (Wang et al. 2012) => type IIB restriction endonucleases

Type IIP homodimer



'Orthodox' IIP enzymes (e.g. EcoRI, EcoRV) cut at the recognition site

Type IIB



Type IIB require two recognition sites and cut on the outside (e.g. BpII)

# Restriction enzyme methods: RAD-Seq and derived methods

1- Sequencing of fragments adjacent to single restriction enzyme cut sites:

- **original RADseq** (Baird et al. 2008)
- **2bRAD** (Wang et al. 2012) => type IIb restriction endonucleases

2- Sequencing of fragments flanked by two restriction enzyme cut sites

A- Single enzyme, indirect size selection

- **GBS – genotyping by sequencing** (Elshire et al. 2011)
- **SBG – sequence-based genotyping** (Truong et al. 2012)

B- Double enzyme, indirect size selection

- **CRoPS – complexity reduction of polymorphic sequences** (Orsouw et al. 2007)

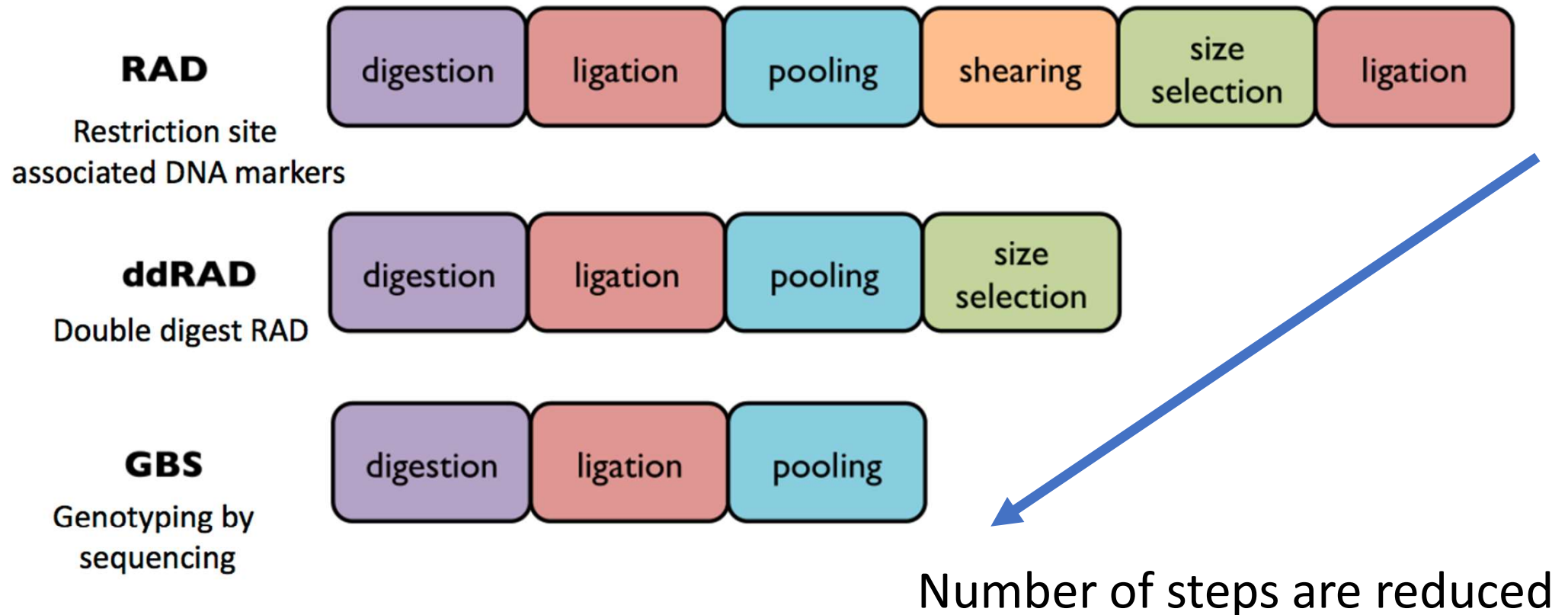
C- Single enzyme, direct size selection

- **RRLs – reduced representation libraries** (van Tassel et al. 2008)
- **MSG – multiplexed shotgun genotyping** (Andolfatto et al. 2011)
- **ezRAD** (Toonen et al. 2013)

D- Double enzyme, direct size selection

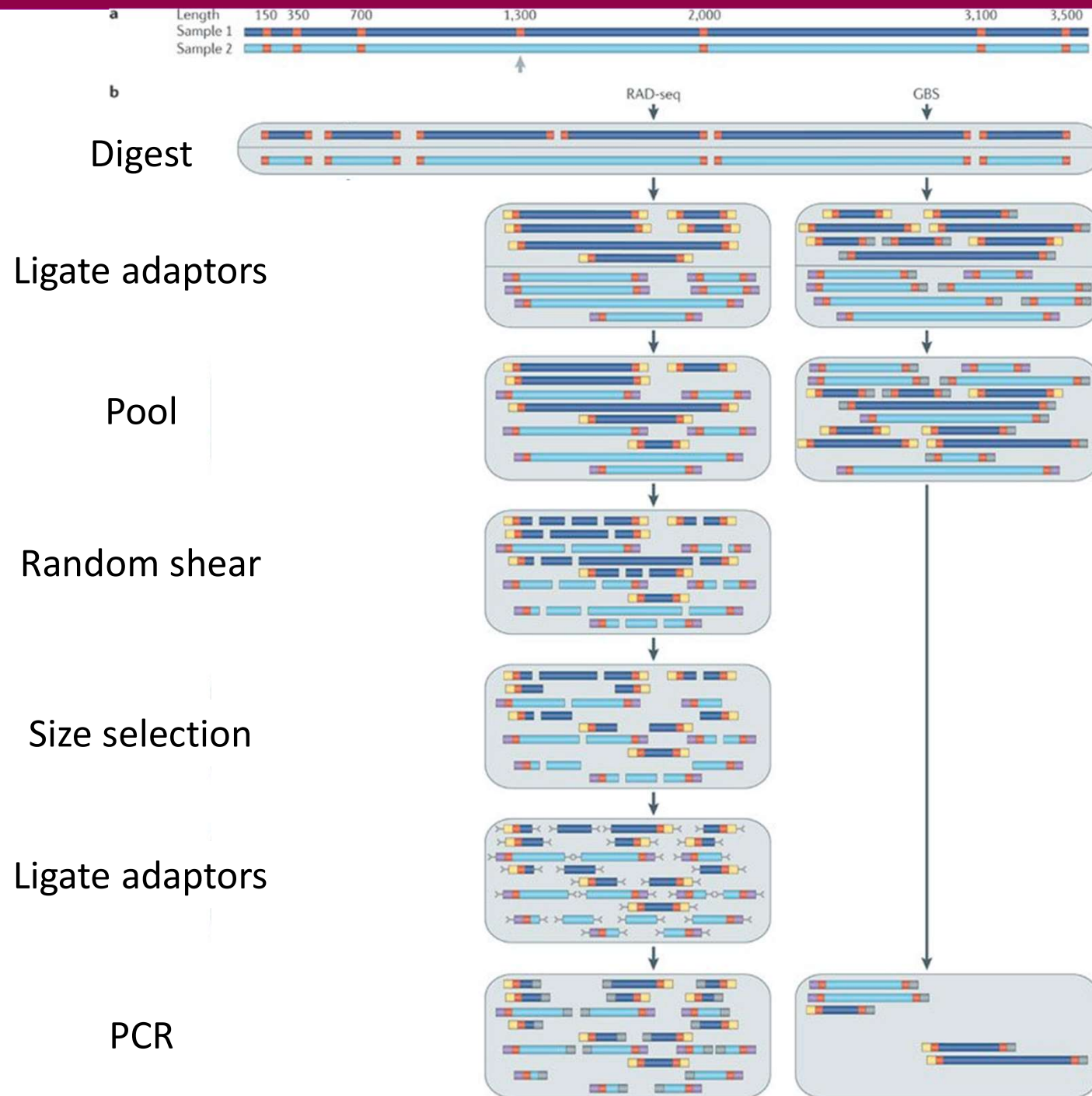
- **ddRAD – double-digest RAD** (Peterson et al. 2012)

# Restriction enzyme methods: RAD-Seq and derived methods





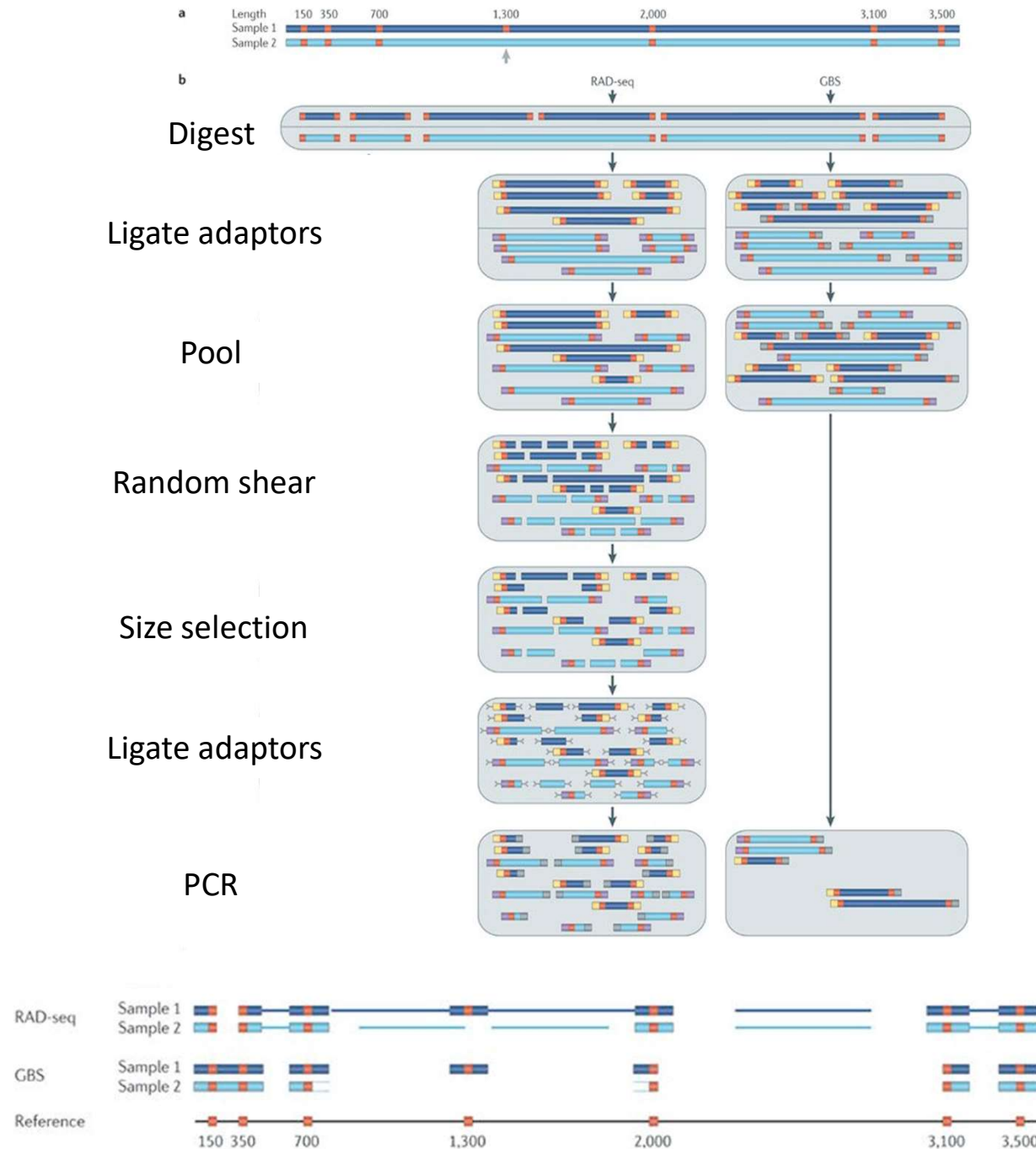
# Restriction enzyme methods: RAD-Seq and derived methods



Davey et al. 2011

# Restriction enzyme methods: RAD-Seq and derived methods

Davey et al. 2011



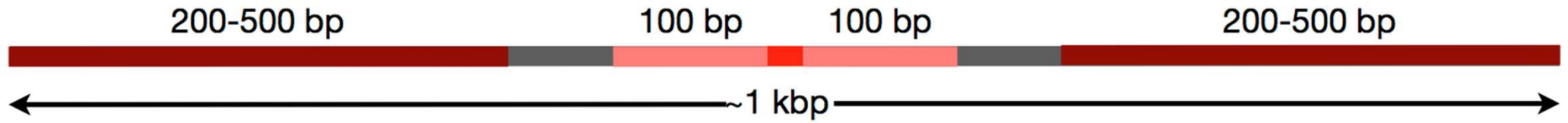


# Restriction enzyme methods: RAD-Seq and derived methods

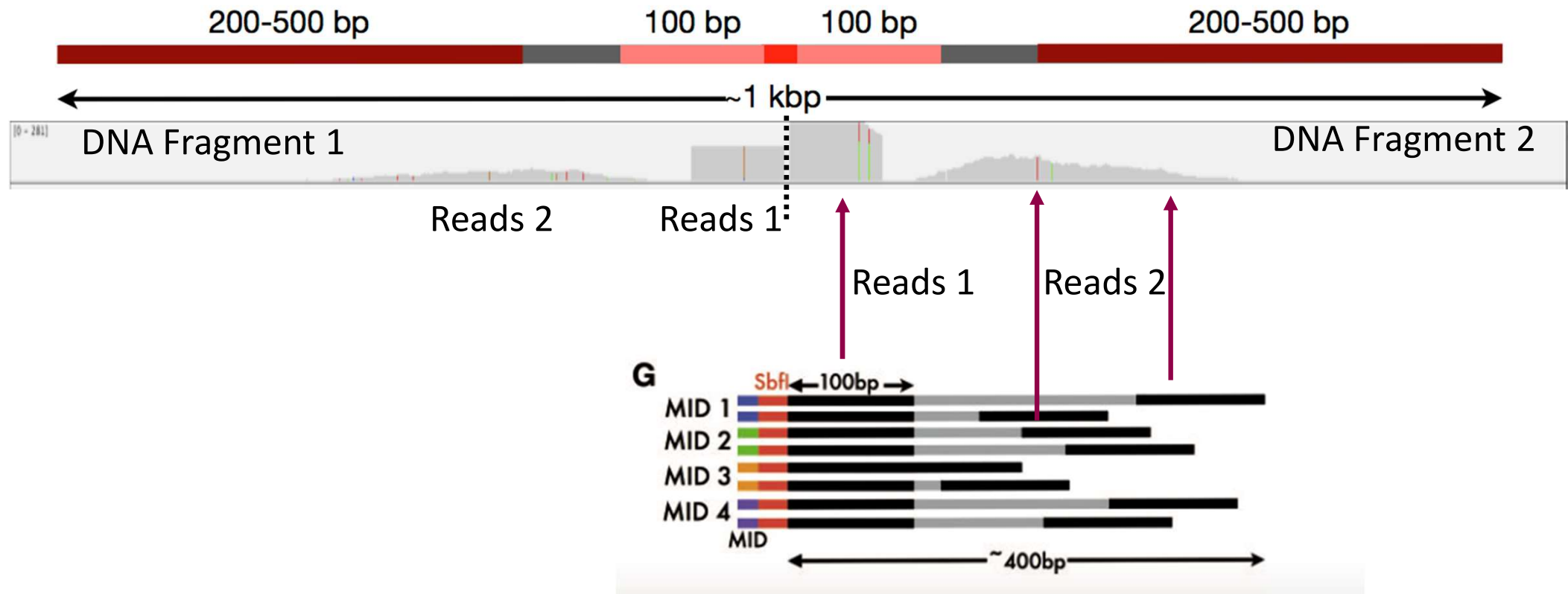
## Methods comparison

	Original RAD	2bRAD	GBS	ddRAD
<b>Options for tailoring number of loci</b>	Change restriction enzyme	Change restriction enzyme	Change restriction enzyme	Change restriction enzyme or size selection window
<b>Number of loci per 1 Mb of genome size*</b>	30–500	50–1,000	5–40	0.3–200
<b>Length of loci</b>	≤1kb if building contigs; otherwise ≤300 bp <sup>‡</sup>	33–36 bp	<300 bp <sup>‡</sup>	≤300 bp <sup>‡</sup>
<b>Cost per barcoded or indexed sample</b>	Low	Low	Low	Low
<b>Effort per barcoded or indexed sample<sup>§</sup></b>	Medium	Low	Low	Low
<b>Use of proprietary kit</b>	No	No	No	No
<b>Identification of PCR duplicates</b>	With paired-end sequencing	No	With degenerate barcodes	With degenerate barcodes
<b>Specialized equipment needed</b>	Sonicator	None	None	Pippin Prep <sup>  </sup>
<b>Suitability for large or complex genomes<sup>  </sup></b>	Good	Poor	Moderate	Good
<b>Suitability for <i>de novo</i> locus identification (no reference genome)<sup>#</sup></b>	Good	Poor	Moderate	Moderate
<b>Available from commercial companies</b>	Yes	No	Yes	Yes

# RAD-Seq bioinformatics



# RAD-Seq bioinformatics

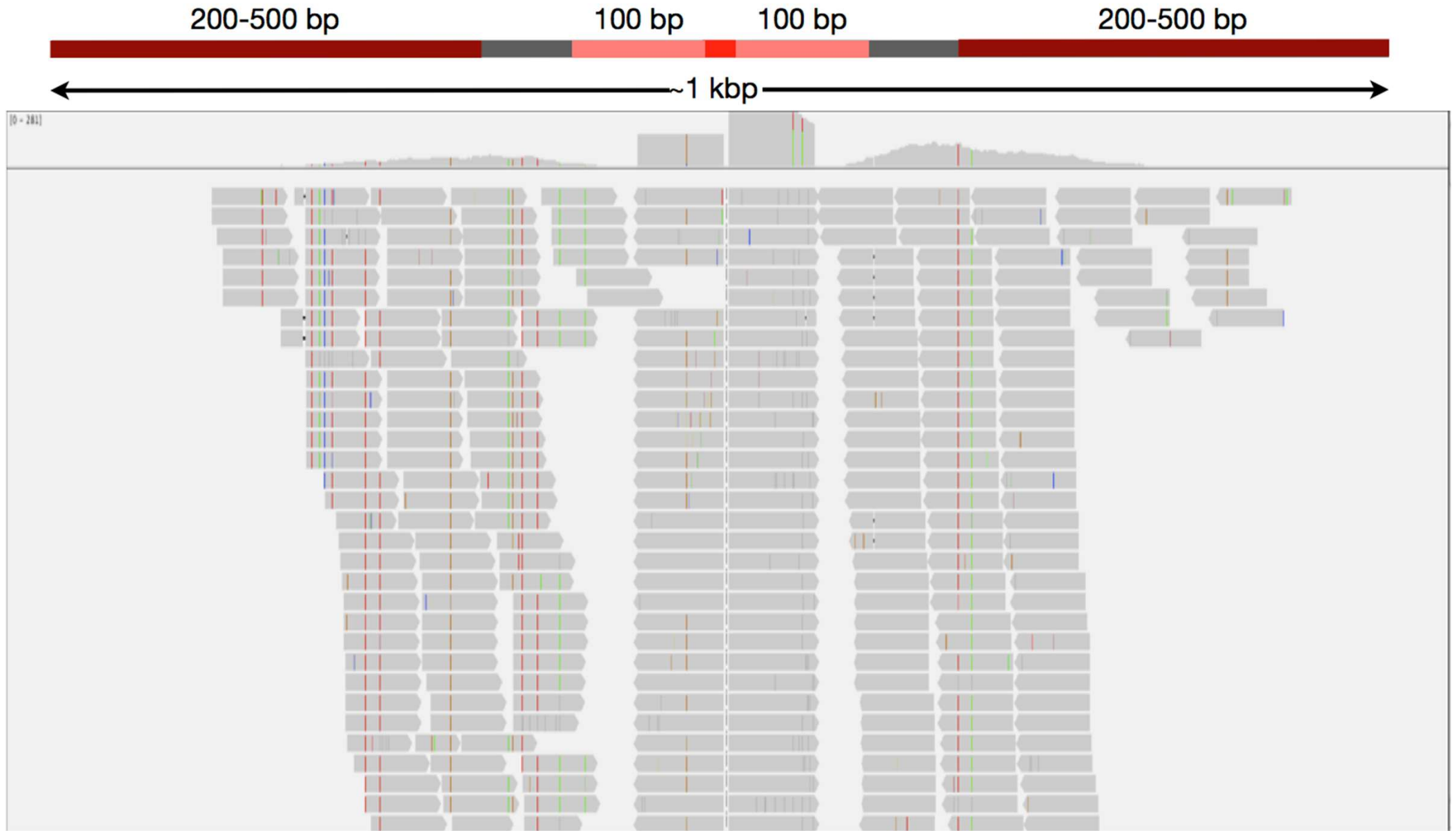


**The coverage** in DNA sequencing is the number of unique reads that include a given nucleotide in the reconstructed sequence.

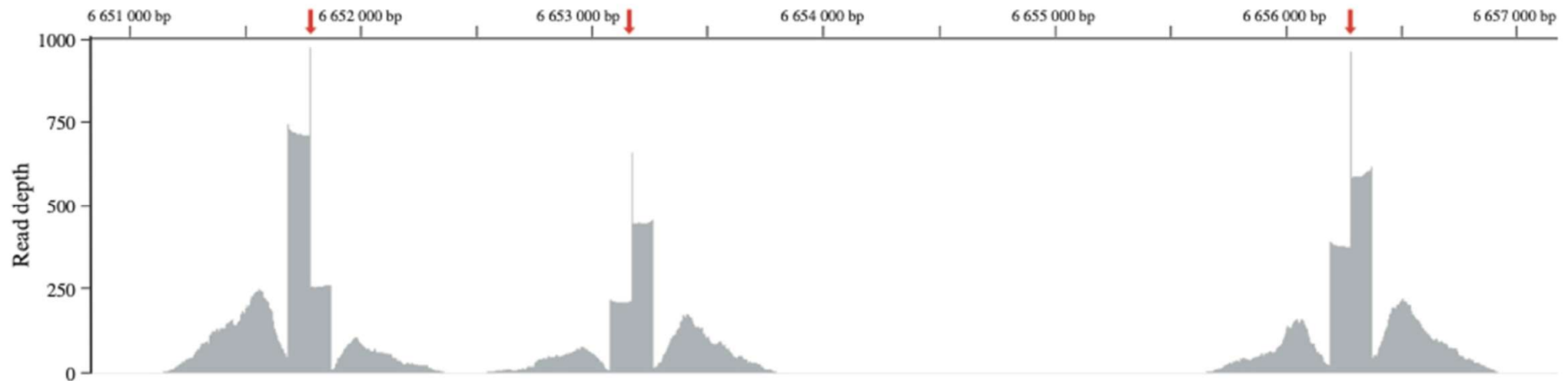
**The sequencing depth (also known as read depth)** is the ratio of the total number of bases obtained by sequencing to the size of the genome or the average number of times each base is measured in the genome

Coverage and sequencing depth are basically the same thing

# RAD-Seq bioinformatics



# RAD-Seq bioinformatics



**Fig. 1** Characteristic pileup of RAD-Seq data. Three *Pst*I restriction sites (red arrows) in *Caenorhabditis elegans* chromosome I (6.651 Mb–6.657 Mb) are covered both upstream and downstream by RAD-Seq raw reads (dark grey). Read 1 sequences are piled up in stacks either side of each restriction site; read 2 sequences are spread out in heaps up to 700 bp beyond the restriction site. The restriction site overhang TGCA is covered by reads belonging to both upstream and downstream RAD loci, producing narrow peaks of read coverage at the restriction sites. RAD loci on either side of a restriction site have different read depths; however, loci from the same restriction fragment have similar read depths. Bases in the read 2 regions are covered at much lower depth overall; read 2 sequences also partially cover the read 1 regions, as seen by the increase in read depth at bases towards the end of read 1 regions.

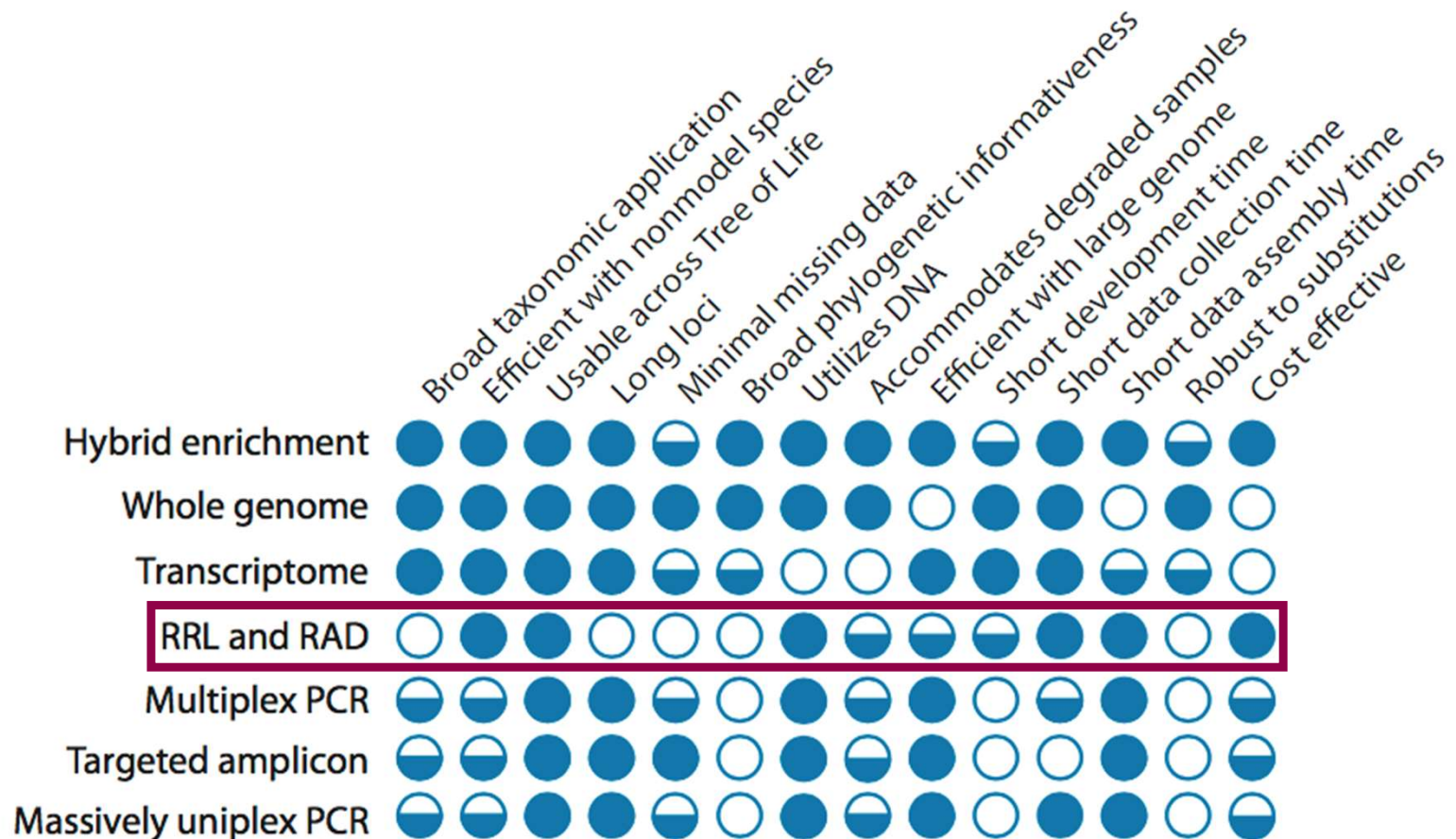
**The coverage** in DNA sequencing is the number of unique reads that include a given nucleotide in the reconstructed sequence.

**The sequencing depth (also known as read depth)** is the ratio of the total number of bases obtained by sequencing to the size of the genome or the average number of times each base is measured in the genome

Coverage and sequencing depth are basically the same thing

# Properties of RAD-Seq data

- wide genomic distribution
- relatively short loci
- allelic dropout/null alleles
- large proportion of missing data
- orthology/paralogy – bioinformatic assessment





# Applications des données de RAD-Seq

- shallow phylogenomics
- population structure, phylogeography
- population genomics
- evolution of recently radiated groups
- hybridization, introgression
- genetic mapping
- ...