

Pairwise sequence comparison

Dotplots and dynamic programming

Adapted from the courses of the Bonsai team,

CRISTAL UMR 9189

Sylvain.legrand@univ-lille.fr

Introduction

Why compare two sequences?

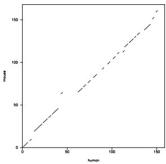
- Assemble a set of sequenced DNA fragments (fragment assembly)
- Search for homology (between genes, mRNA, proteins...)
- Find similar regions (protein domains)
- Identify intron/exon positions (comparison of a gene and its mRNA(s))



How to compare two sequences?

- 2 approaches
 - Dotplots (Dot-matrix plots)

A **graphical method** for the comparison of two sequences or a sequence against itself



- Alignment

A text comparison method

- → Using **dynamic programming** → optimal alignment
- → Using heuristic methods (Blast) → Fast, useful when classic methods are too slow. But favours speed at the expense of optimality or precision.



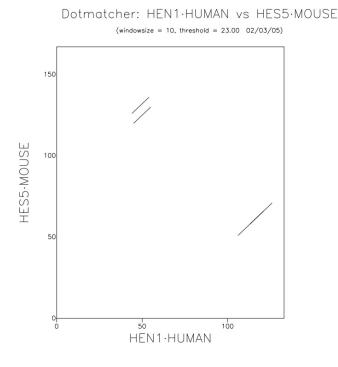
Dotplots

Dotplots

A graphical tool for the comparison of two sequences

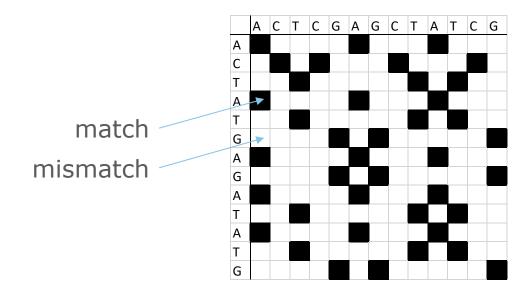
Method

- Put the two sequences along the axes of a matrix (x, y)
- Draw a point where there is a match between the two sequences
- A diagonal (a series of points) represents a similar region



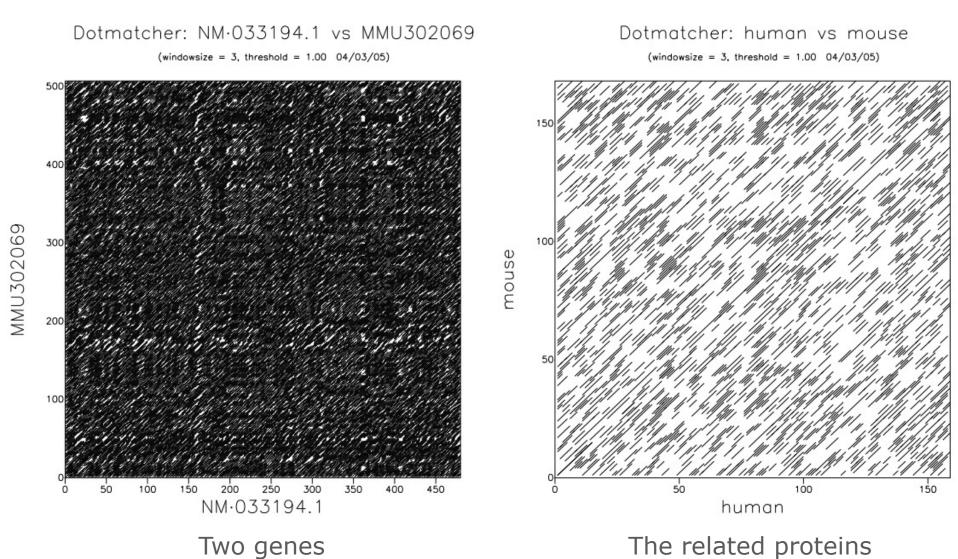


Dotplot example





Dotplot example



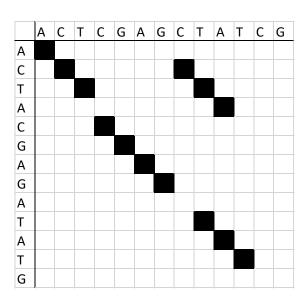
→ Noise problem (too low filtering)



Dotplot filtering

- Using a word of size k
 - a sliding window of a defined length (\Bbbk) that moves through the matrix comparing the two sequences
 - only represent exact windows: high selectivity/low sensitivity
 - Example of software: **dottup**
 - Example with k=3

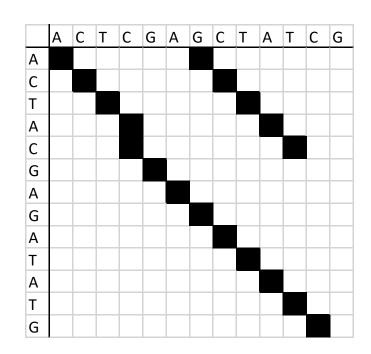
	Α	С	Т	С	G	Α	G	С	Т	Α	Т	С	G
Α													
С													
Т													
A C G A T A T G													
С													
G													
Α													
G													
Α													
Т													
Α													
Т													
G													





Dotplot filtering

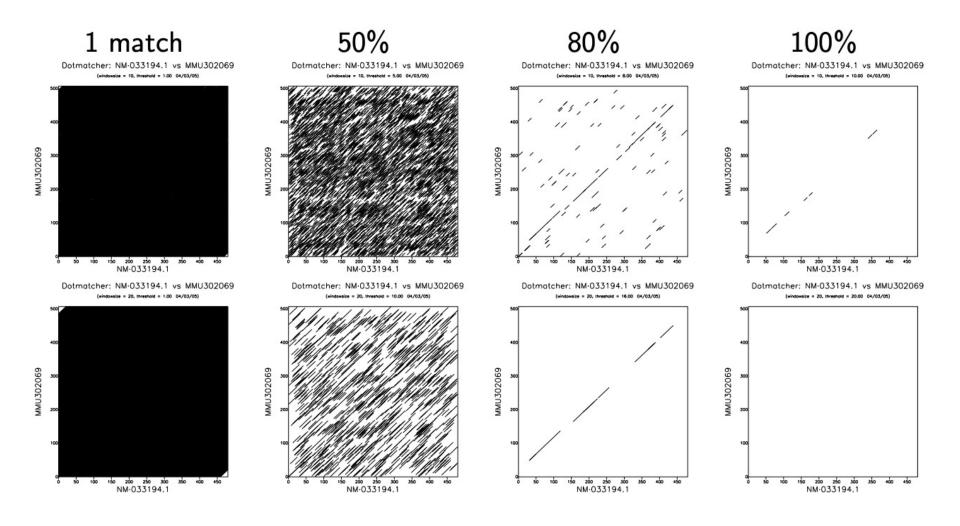
- Using a sliding window and score threshold (Maizel & Lenk - 1981)
 - a sliding window of a defined length (\Bbbk) that moves through the matrix comparing the two sequences
 - Represent windows with a score ≥ s: high selectivity/high sensitivity
 - Software example: dotmatcher
 - Example with k=4 and 75% identity





Dotmatcher, examples

• K= 10 and 20, threshold from 1% to 100%





Dotplot filtering

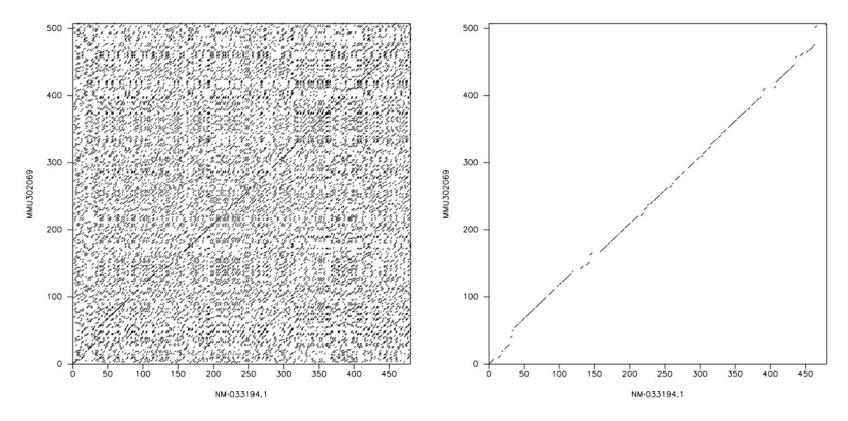
- By eliminating overlapping blocks
 - Observe the overall resemblance
 - Software example: dotpath
 - Example with k=2

Eliminated blocks

	Α	С	Т	С	G	Α	G	С	Т	Т	Т	С	G
Α													
A C T													
Т													
Α													
A C G A T A													
G													
Α													
G													
Α													
Т													
Α													
Т													
G													



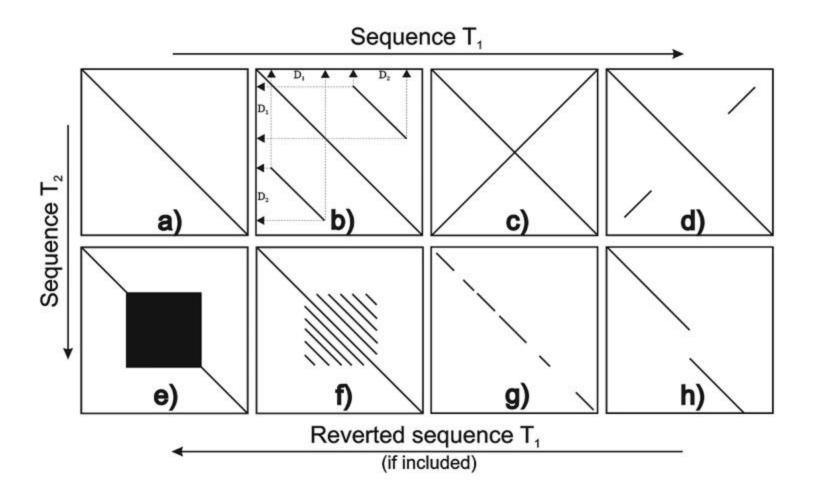
Dotpath, examples



dotpath finds all matches of size wordsize or greater between two sequences. It then reduces the matches found to the minimal set of long matches that do not overlap. This is a way of finding the (nearly) optimal path aligning two sequences. It is not the true optimal path as produced by the algorithms used in water or needle, but for very closely related sequences it will produce the same result and will work well with very long sequences



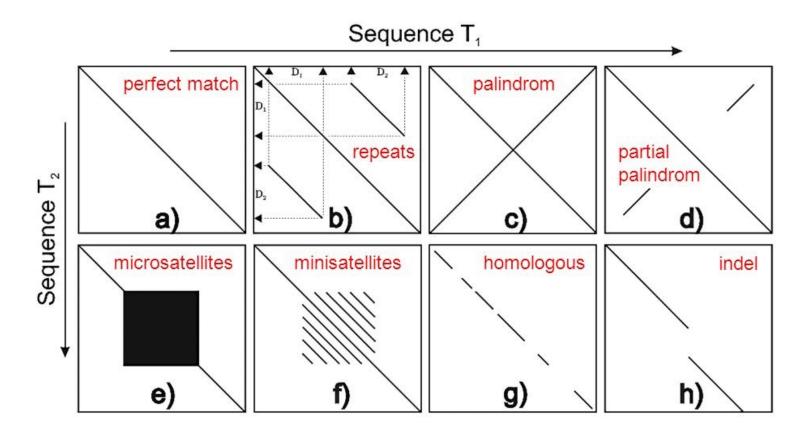
Interpretation of dotplots



http://www.code10.info/index.php?option=com_content&view=article&id=64:inroduction-to-dot-plots&catid=52:cat_coding_algorithms_dot-plots<emid=76



Interpretation of dotplots





Dotplots, summary

Advantages

- Simple
- Very informative

Cons

- Identification: no automatic detection method
- Interpretation: no objective measurement
- → Need for a quantitative measure of similarity



Pairwise alignment

Pairwise alignment

- 2+1 types of alignments
 - **global**: align **every residue** in the two sequences. When the two **sequences are similar** and of roughly equal size
 - local: match on sub-sequences
 For dissimilar sequences that are suspected to contain regions of similarity
 - semi-global: derived from global
 When you align a short sequence against a longer one or when the downstream part of one sequence overlaps with the upstream part of the other sequence

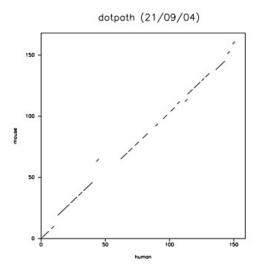


Example global alignment

Example: heat shock protein beta 9 from human and mouse

```
>human
MQRVGNTFSN ESRVASRCPS VGLAERNRVA TMPVRLLRDS PAAQEDNDHA RDGFQMKLDA
HGFAPEELVV QVDGQWLMVT GQQQLDVRDP ERVSYRMSQK VHRKMLPSNL SPTAMTCCLT
PSGQLWVRGQ CVALALPEAQ TGPSPRLGSL GSKASNLTR
>mouse
MQRVGSSFST GQREPGENRV ASRCPSVALA ERNQVATLPV RLLRDEVQGN GCEQPSFQIK
VDAQGFAPED LVVRIDGQNL TVTGQRQHES NDPSRGRYRM EQSVHRQMQL PPTLDPAAMT
CSLTPSGHLW LRGQNKCLPP PEAQTGQSQK PRRGGPKSSL QNESVKNP
```

```
1 MQRVGNTFS----NESRVASRCPSVGLAERNRVATMPVRLLRDSPAAQ
human
         111111::11
                       .1:111111111.11111:111:111111.
       1 MQRVGSSFSTGQREPGENRVASRCPSVALAERNQVATLPVRLLRDE---V
mouse
      45 EDNDHARDGFQMKLDAHGFAPEELVVQVDGQWLMVTGQQQLDVRDPERVS
         :.|...:..||:|:||.||||:|||::|||.|.|||:|.:..||.|...
      48 QGNGCEQPSFQIKVDAQGFAPEDLVVRIDGQNLTVTGQRQHESNDPSRGR
      95 YRMSQKVHRKM-LPSNLSPTAMTCCLTPSGQLWVRGQCVALALPEAQTG
         98 YRMEQSVHRQMQLPPTLDPAAMTCSLTPSGHLWLRGQNKCLPPPEAQTGQ
human 144 S--PRLGSLGSKASNLTR-----
                                    159
         1 11.1 1.1:1....
mouse 148 SQKPRRG--GPKSSLQNESVKNP
                                    168
```





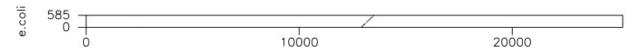
Example semi-global alignment

Example: alignment of a genomic region and a tRNA

AE008779.1 : Salmonella typhimurium LT2, section 83 of 220 of the complete genome. 25184 bp

e.coli : Escherichia coli peptidyl tRNA hydrolase. 585 bp

dotpath (21/09/04)



AE008779.1

AE008779.1 12901	TCAGGACAAAAAACGTGGCAATTAAATTGATTGTTCTGGCGAATCCC	13201	TGGATCTCCCTCCGGGCGTCGCGAAATTTAAACTTGGCGGCGGCCACGGC
e.coli 1	gtgacgattaaattgattgtcggcctggcgaacccc	287	${\tt tggatctgcctcctggcgtcgccaaatttaaattgggcggtggccatggt}$
AE008779.1 12951	GGTGCGGAATATGCCGCGACGCGACACAATGCAGGCGCATGGTACGTCGA	13251	GGCCACAATGGTCTGAAAGACATCATCAGCAAGCTGGGCAATAATCCCAA
e.coli 37	ggtgctgaatacgccgcaacgcgacataatgctggtgcctggttcgttga	337	ggtcacaatggactgaaagacatcatcagtaaattgggtaataaccctaa
AE008779.1 13001	TTTACTGGCGGAGCGCCTGCGCGCGCGTTGCGTGAAAATTCT	13301	CTTTCACCGATTACGCGTTGGAATTGGTCATCCAGGCGATAAAAATAAAG
e.coli 87	cttactggcagagcgtttgcgcgctccgctgcgcgaagaggctaaattct	387	ctttcaccgtttacgcatcgggaatcggtcatccgggcgataaaaataaag
AE008779.1 13051	TTGGCTATACCTCACGCATCACGCTGGAAGGGGAAGATGTTCGCCTGCTG	13351	TTGTTGGTTTCGTGCTGGGTAAACCCCCTGTTTCTGAACAAAATTAATT
e.coli 137	ttggttatacttcgcgagtcactcttggaggcgaagatgtccgcctgtta	437	ttgtcggttttgtgttaggcaaaccgcctgttagtgaacagaagttaatt
AE008779.1 13101	GTACCCACCACGTTCATGAACCTCAGTGGTAAAGCAGTTGGCGCAATGGC	13401	GATGAGGCCATTGACGAAGCGGCACGCTGTACGGAATTGTGGTTCAAAGA
e.coli 187	gtcccgactacatttatgaatctcagcggcaaagccgttgcggcgatggc	487	gatgaagccattgacgaagcggcgcgttgtactgaaatgtggtttacaga
AE008779.1 13151	CAGTTTTTACCGTATTCAGCCGGACGAAATTTTGGTCGCTCACGACGAGC	13451	GGGTCTGGCCAAAGCAACAAGCCGTTTGCATACCTTTAAGGCGCAATAAC
e.coli 237	cagttttttccgcattaatccggacgaaattctggtggcccacgacgaac	537	tggcttgaccaaagcaacgaaccgattgcacgcctttaaagcgcaataa



Example local alignment

Example: 2 dissimilar sequences with a conserved domain

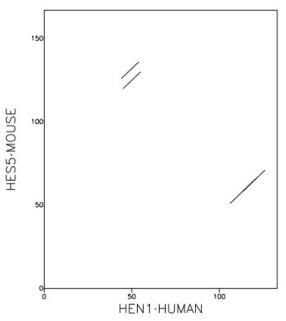
>HEN1_HUMAN

MMLNSDTMELDLPPTHSETESGFSDCGGGAGPDGAGPGGGGQARGPEPGEPGRKDLQHLSREERRRR RATAKYRTAHATRERIRVEAFNLAFAELRKLLPTLPPDKKLSKIEILRLAICYISYLNHVLDV > HES5_MOUSE

MAPSTVAVEMLSPKEKNRLRKPVVEKMRRDRINSSIEQLKLLLEQEFARHQPNSKLEKADILEMAVSYLK HSKAFAAAAGPKSLHQDYSEGYSWCLQEAVQFLTLHAASDTQMKLLYHFQRPPAPAAPAKEPPAPGAAPQ PARSSAKAAAAAVSTSRQPACGLWRPW

Dotmatcher: HEN1·HUMAN vs HES5·MOUSE

(windowsize = 10, threshold = 23.00 02/03/05)





Pairwise alignment

· Data:

- A pair of sequences (DNA / protein)
- A scoring system: how to count what is similar?

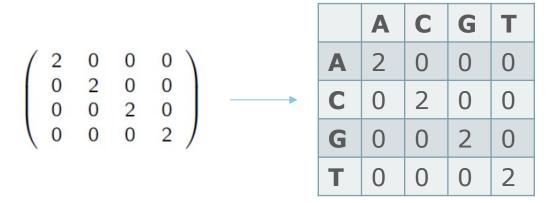
goal:

- To determine the degree of similarity (best score)
- Show similarity (better alignment)
- Describes the resemblance through 3 operations (point mutations)
 - Insertion
 - Deletion
 - Identity/substitution (match/mismatch)
- Measures similarity by giving weight to each operation
- Positive weight ("reward") to the good parts of the alignment (matching of two identical or close letters)
- Negative (or zero) weight ("penalty") associated with bad (matching of two unrelated letters, mismatch)



Scoring system

- Score (or weight) for an identity/substitution
 - **Substitution matrix** → See dedicated course
 - s(a, b) = alignment score of nucleotides a and b



More complicated for proteins!

 Score (or weight) of an **indel** (insertion/deletion): -2 for example per indel



Alignment score

- Alignment score = sum of the scores of the elementary events
- For **example**:



Why we need a smart algorithm

- 2 sequences of length n: max. length of the alignment 2n
- Example with the sequences TA and CA
- Naive algorithm: enumerate all the alignments
 (match=+1 mismatch=-1 indel = -1)
 i= number of overlapping nucleotides

	TA			
i=0	CA -4			
	TA-	TA-	TA-	T-A
i=1	-CA -3	-CA -3	C-A -3	-CA -1
	TA			
i=2	CA 0			



Why we need a smart algorithm

Maximum number of alignments (sequences of length n)

number of alignments (sequences of length n)
$$\frac{(2n)!}{(n)!^2} \Rightarrow \text{For 2 sequences of length 100: 2.10}^{57}$$
 alignments

- Using dynamic programming: matrix representation, complexity n²
 - → For 2 sequences of length 100: 10 000 calculations

	Т	A
С		
A		



Dynamic programming

- The concept was developed by Richard Bellman in the 1950s
- Aims to simplify a complicated problem by breaking it down into simpler sub-problems in a recursive way
- See "Pyramide de nombres": https://fr.wikipedia.org/wiki/Programmation_dynamique#opc
- Widely used in **bioinformatics**: sequence alignment, protein folding, protein binding, nucleic acid structures...



Global alignment

- Algorithm by Needleman & Wunsch, 1970
- For nucleic acids or proteins
- Dynamic programming
- Optimal alignment



Sequences

- Sequence A: ATT

- Sequence B: TTC

Define a scoring table:

- match: +1 \rightarrow s(a,b) = +1 if a = b

- mismatch: -1 \rightarrow s(a,b) = -1 if a \neq b

- indel: -1 \rightarrow s(a,-) = s(-,b) = -1

• Build scoring matrix by following these rules

- Matrix initialization: 0

-
$$S_{i,J}$$
 = MAX $\begin{bmatrix} S_{i-1}, J-1+s (a,b) \\ S_{i-1}, J+s (a,-) \\ S_{i}, J-1+s (-,b) \end{bmatrix}$ = $\begin{bmatrix} S(\nwarrow)+match \text{ or mismatch} \\ S(\leftarrow)+indel \\ S(\uparrow)+indel \end{bmatrix}$



	-	A	Т	Т
-				
Т				
Т				
С				

match: +1 mismatch: -1

$$S_{i,J} = MAX S(\)+match/mis.$$
 $S(\leftarrow)+indel$ $S(\uparrow)+indel$



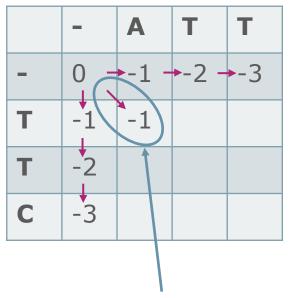
	-	A	Т	Т
-	0 -	-1 -	- -2 −	- 3
Т	-1			
Т	-2			
С	-3			

match: +1

mismatch: -1

$$S_{i,J} = MAX S(\)+match/mis.$$
 $S(\leftarrow)+indel$ $S(\uparrow)+indel$





match: +1

mismatch: -1

$$S_{i,J} = MAX S(\searrow) + match/mis.$$

 $S(\leftarrow) + indel$
 $S(\uparrow) + indel$

$$MAX (0-1;-1-1;-1-1) = MAX (-1;-2;-2) = -1$$



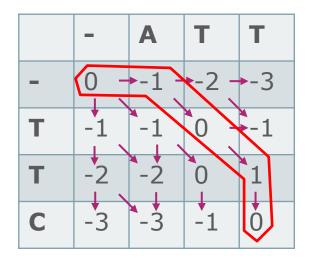
	-	A	Т	Т
-	0 -	-1 -	- -2 -	- 3
Т	-1	-1	0 -	-1
Т	-2	-2	0	1
С	-3	-3	-1	Ŏ

match: +1

mismatch: -1

$$S_{i,J} = MAX S(\)+match/mis.$$
 $S(\leftarrow)+indel$ $S(\uparrow)+indel$





- . Backtracking from the lower-right corner
- . Reconstructing alignment



	-	A	Т	С	G	G	A	G
-								
A								
Т								
G								
G								
С								
A								
A								

Other example from https://www.youtube.com/watch?v=BYdTqq8AGgc

Sequence A: ATCGGAG Sequence B: ATGGCAA

match: +1 mismatch: -1

$$S_{i,J} = MAX S(\)+match/mis.$$

 $S(\leftarrow)+indel$
 $S(\uparrow)+indel$



					G			
-					-4			
A					2 -			
Т	-2	Ŏ	2 -	+1 -	•0 -	-1 -	-2 -	→ -3
G					2 -			
G					2			
С	_	_		_	1			
A					Ŏ			
A	-7	-5	-3	-1	-1	0	2	2

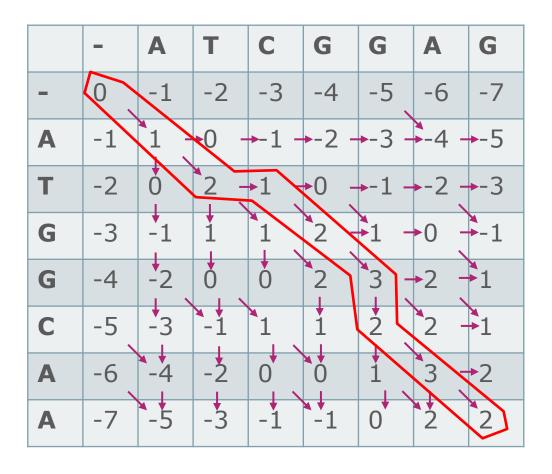
match: +1

mismatch: -1

$$S_{i,J} = MAX S(\)+match/mis.$$
 $S(\leftarrow)+indel$ $S(\uparrow)+indel$



Needleman Wunsch algorithm



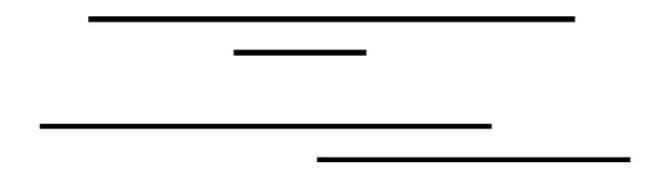
- . Backtracking from the lower-right corner
- . Reconstructing alignment

ATCGG-AG AT-GGCAA



Semi-global alignment

- A variant of global alignment that allows for gaps at the beginning and/or the end of one of the sequences
- Useful when you align a short against a long sequence or when the downstream part of one sequence overlaps with the upstream part of the other sequence





Local alignment

- Algorithm by Smith & Waterman, 1981
- For nucleic acids or proteins
- Look for subsequence matches
- Modification of the Needleman Wunsch algorithm: negative scores are set to zero
- Dynamic programming
- **Optimal** alignment



Sequences

- Sequence A: ACCGTGA

- Sequence B: GTGAATA

Example from https://www.youtube.com/watch?v=BYdTqq8AGgc

Define a scoring table:

- match: +1
$$\rightarrow$$
 s(a,b) = +1 if a = b

- mismatch: -1
$$\rightarrow$$
 s(a,b) = -1 if a \neq b

- indel: -1
$$\rightarrow$$
 $s(a,-) = s(-,b) = -1$

Build scoring matrix by following these rules

- Matrix initialization: 0; No negative scores!

-
$$S_{i,J} = MAX$$
 $S_{i-1,J-1}+s(a,b)$ $S_{i-1,J}+s(a,-)$ $S_{i,J-1}+s(-,b)$ $S_{i,J-1}+s(-,b)$ $S_{i,J-1}+s(-,b)$ $S_{i,J-1}+s(-,b)$ $S_{i,J-1}+s(-,b)$ $S_{i,J-1}+s(-,b)$ $S_{i,J-1}+s(-,b)$ $S_{i,J-1}+s(-,b)$



	-	A	С	С	G	Т	G	A
-								
G								
Т								
G								
A								
A								
Т								
A								

match: +1 mismatch: -1

indel: -1

$$S_{i,J} = MAX$$
 $S(\)+match/mis.$ $S(\leftarrow)+indel$ $S(\uparrow)+indel$ 0



	-	A	С	С	G	Т	G	A
-	0	0	0	0	0	0	0	0
G	0	0	0	0	1	0	1	0
Т	0	0	0	0	0	2	1	0
G	0	0	0	0	1	1	3	2
A	0	1	0	0	0	0	2	4
A	0	1	0	0	0	0	1	3
Т	0	0	0	0	0	1	0	2
A	0	1	0	0	0	0	0	Ì

match: +1

mismatch: -1

indel: -1

$$S_{i,J} = MAX$$
 $S(\)+match/mis.$ $S(\leftarrow)+indel$ $S(\uparrow)+indel$ 0



	-	A	С	С	G	Т	G	A
-	0	0	0	0	0	0	0	0
G	0	0	0	0	1	0	1	0
Т	0	0	0	0	0	2	1	0
G	0	0	0	0	1	1	3	2
A	0	1	0	0	0	0	2	4
A	0	1	0	0	0	0	1	3
Т	0	0	0	0	0	1	0	2
A	0	1	0	0	0	0	0	İ

- . Backtracking from max score in the matrix until zero
- . Reconstructing local alignment

GTGA GTGA



Gaps penalties

- The different algorithms use different functions to calculate gap penalties
- The simplest function: linear function: g × 1
 - g: indel penalty
 - 1: length of the gap
- More realistic functions
 - Affine functions: o + e × 1
 - → o: gap opening penalty
 - → e: gap extension penalty
 - Logarithmic function



Influence of the scoring system, exercise

	Α	В	C
Match Cost	1	1	1
Mismatch Cost	-1	-1	-1
Gap Open Penalty (o)	0	1	4
Gap Extension Penalty (e)	0	0.1	0.1

AT-GCGGGACA-TG

| ||| | || A-GGCG---C-CTG

7 matches, 0 mismatches, 5 ogaps, 2 egaps

ATGCGGGACATG

|.||| |.|| AGGCG---CCTG 7 matches, 2 mismatches, 1 ogap, 2 egaps

ATGCGGGACATG

AL3: .||..|.||
---AGGCGCCTG

5 matches, 4 mismatches, 1 ogap, 2 egaps

For each scoring system (A, B or C), determine which alignment (AL1, AL2 or AL3) will be returned



Choose the right gap penalties

- Little *a priori* knowledge
- Data specificity
- Typical values for an affine gap function

$$0.5 < o < 5.0$$

 $0.05 < e < 1.0$

Always take (in absolute value) 0 > 1/2 substitution



Assess the quality of an alignment



Assess the quality of an alignment

Robustness of the score when changing the parameters

- Alignment is doubtful if small changes (about 10%) in penalties of insertion/deletion significantly change this alignment

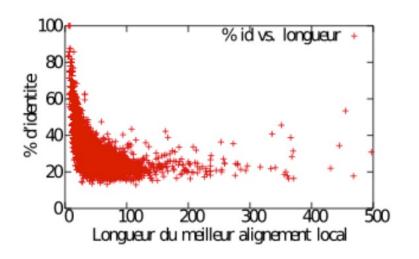
Frequency of gaps

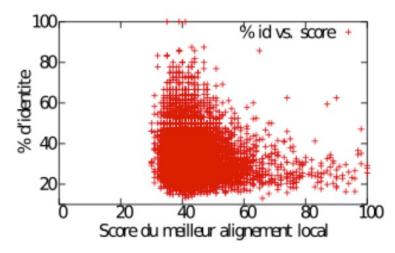
- Alignment is doubtful if it requires more than one insertion on average for 20 amino acids
- Two nucleic sequences of at least 100 bases and 50% identities do not necessarily have a biological relationship
- Protein sequences of 100 or more residues, with at least 25% identity have certainly a common ancestor (Doolittle, 1990 PDB).



Use % of identity?

- Depends on the composition of bases or amino acids
- Depends on the **length** of the sequences





Not a good idea!



Empirical approach

Score robustness test

- Two sequences : u and v
- s: score of the alignment between $\ensuremath{\mathtt{U}}$ and $\ensuremath{\mathtt{V}}$

Method:

- 1. Generation of 100 (or more) permutations of ∨ (same length, same composition)
- 2. Alignment with \cup \rightarrow score calculation
- 3. Distribution of alignment scores
- 4. Where does s fit into this distribution?



Statistical approach

- **E-value**: number of times one expects to find a score alignment greater than s by chance when a sequence of length n is aligned with a length sequence m
 - Describes the random noise that exists when aligning sequences increases proportionally with n and m
 - Decreases exponentially as a function of the score S
 - The closer the E-value is to 0, the more the similarity is significant



Example

Human alpha haemoglobin (141 aa) vs. Human myoglobin (153 aa) VLSPADKTNVKAAWGKVGAHAGEYGAEALERMFLSFPTTKTYFPHF-DLS-----HGSAQVKGHGKKVADALTNAVAHVDDMPNALSAL GLSDGEWQLVLNVWGKVEADIPGHGQEVLIRLFKGHPETLEKFDKFKHLKSEDEMKASEDLKKHGATVLTALGGILKKKGHHEAEIKPL SDLHAHKLRVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLTSKYR-----AQSHATKHKIPVKYLEFISECIIQVLQSKHPGDFGADAQGAMNKALELFRKDMASNYKELGFQG Chicken lysozyme (129 aa) vs. Bovine ribonuclease (124 aa) KVFGRCELAAAMKRHGLDNYRGYSLGNWVCAAKFESNFNTQATNRNTDGSTDYGILQINSRWWCNDGRTP--GSRNLCNIPCSALLSSD :.. : ..:.. .: KETA----AAKFERQHMDSSTSAASSSNYCNQMMKSRNLTKDRCKPVNTFVHESLADVQAV--CSQKNVACKNGQTNCYQSYSTMSITD ITASVNCAKKIVSDGDGMNAWVAWRNRCKGTDVQAWIRGCRL:: CRET-GSSKYPNCAYKTTQANKHIIVACEGNPYVPVHFDASV



Use of PRSS

PRSS : Probability of Random Shuffle Sequence

```
< 20
 22
                 one = represents 1 library sequences
 26
         0:
 30
         1:*
 32
         3:==*
 34
         7:=====*==
 36
        15:========
 40
    29
        34:==========
 42
    33
    51
        46
    41
        48
    32
        50
    51
        41:======*===*======
 52
        36:==========
 54
    24
        31:=========
 56
    30
        26:=================
        21:======= *
 60
        17:==========
 62
        14:=========
        11:====
 66
         9:======*=
 68
         7:==== *
 70
         5:====*
 72
         4:===*===
         3:==*
 76
         3:==*
 78
         2:=*=
         2:=*
 82
         1:*
         1:*
 86
         1:*
 88
         1:*
         0:
              unshuffled s-w score: 177
 92
         0:=
             For 500 sequences, a score >= 177 is expected 3.096e-06 times
```



Sylvain Legrand
Maître de Conférences
UMR CNRS 8198 EVO-ECO-PALEO
Evolution, Ecologie et Paléontologie
Université de Lille - Faculté des Sciences et Technologies
Bât SN2, bureau 208 - 59655 Villeneuve d'Ascq

sylvain.legrand@univ-lille.fr | http://eep.univ-lille.fr/ Tél. +33 (0)3 20 43 40 16