Package 'RoMA'

December 5, 2017

Type Package					
Title RNA-seq analyses of Molecular Abundance (RoMA)					
Version 1.0 Date 2019-09-17					
Description An efficient method to detect RNA-seq differential expression, by modeling mRNA abundance with precision weights from raw counts, provides an accurate quantification of mRNA abundance and a data adjustment-tolerant analysis and therefore integrative with various well-established methods of mRNA abundance.					
License GPL (>=2)					
Depends edgeR, limma					
R topics documented:					
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RoMA-package RNA-seq analyses of Molecular Abundance (RoMA)					
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Description

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Details

The DESCRIPTION file: This package was not yet installed at build time.

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Author(s)

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Examples

```
data(x)
e<-DEGList(counts=x)
e <- calcNormFactors(e, method="TMM")
e<-calcLibSizes(e)
e<-calcNormRPKMs(e)

des<-factor(c(rep("BR",7),rep("UHR",7)))

des<-as.matrix(cbind(1,as.numeric(des)))

fit<-roma(e, design = des, lib.size = NULL, normalize.method = "none", span = 0.5, plot = plotMA(fit,coef=2,h=1,n=3000,sort="p",p=0.05)

res<-topTable(fit,coef=2,n=Inf,sort = "p", p = 0.05)

res<-topTable(fit,coef=2,n=Inf)</pre>
```

calcLibSizes

Calculation of Library Sizes

Description

The effective library size is estimated by norm factor * total counts, where norm factor can be estimated by TMM.

Usage

```
calcLibSizes(e)
```

Arguments

e The DGEList object with estimated normalization factors.

calcNormRPKMs 3

Value

The DGEList object with estimated effective library sizes in e\$lib.size.

NormRPKMs Calculation of Normalized RPKMs

Description

Although RPKM/FPKM normalizes the RNA-seq data, it does not account for composition bias. Composition bias can be accounted for using TMM normalization by equaling the median of logratios of read counts between samples. RoMA use this function to normalize RPKMs by the TMM method.

Usage

```
calcNormRPKMs(e)
```

Arguments

e The DGEList object with estimated library sizes and transcript lengths.

Value

The DGEList object with calculated RPKMs in e\$rpkms.

|--|

Description

Creates a DGEList object with various information including library sizes, gene length and others.

Usage

```
DEGList(counts = matrix(0, 0, 0), lib.size = colSums(counts, na.rm = TRUE), norm
```

Arguments

counts	numeric matrix of read counts.
lib.size	numeric vector giving sequence depth for each library, which is the total count by default.
norm.factors	numeric vector of normalization factors that modify the library sizes, which is 1 by default.
group	vector or factor giving the experimental group/condition for each sample/library.
length	numeric matrix of gene length, which could be gene expression effective length estimated by RSEM.
length.gene	data frame containing annotation information for each gene, which is from the hg19 human assembly UCSC annotation by default.

4 length.isoform.hg19

length.isoform

frame containing annotation of gene length for each gene isoform, which is from

the hg19 human assembly UCSC annotation by default.

rpkms numeric matrix of rpkms, which is NULL by default.

genes data frame containing annotation information for each gene.

remove.zeros logical, whether to remove rows that have 0 total count.

level indicate the data is on gene level or isoform level.

Value

DGEList object

length.gene.hg19 Collapsed Gene Length from UCSC HG19 annotation

Description

Collapsed Gene Length from UCSC HG19 annotation

Usage

```
data("length.gene.hg19")
```

Format

The format is: int [1:47352, 1] 230 320 1774 2134 9544 2907 4689 2190 5400 454 ... - attr(*, "dimnames")=List of 2 ...\$: chr [1:47352] "5S_rRNA" "7SK" "A1BG" "A1BG-AS1"\$: chr "Length"

```
length.isoform.hg19
```

Transcript Length from UCSC HG19 annotation

Description

Transcript Length from UCSC HG19 annotation

Usage

```
data("length.isoform.hg19")
```

Format

```
The format is: int [1:82960, 1] 1652 1746 2412 1769 1577 1130 918 32 62 4370 ... - attr(*, "dimnames")=List of 2 ..$ : chr [1:82960] "uc001aaa.3" "uc001aac.4" "uc001aae.4" "uc001aah.4" ... ... $: chr "Length"
```

plotMA 5

plotMA MA-plot Maker

Description

Visualize the difference between two measurements by log ratio and mean average of data.

Usage

```
plotMA(fit, coef = 2, h = 1, n = 3000, sort = "p", p = 0.05, colors = NULL, legellation = NULL, legellat
```

Arguments

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	fit	the fit object from RoMA.
	coef	column number or column name of coefficient matrix which is of interest.
	h	threshold line on log ratio
	n	maximum number of significant genes will be shown
	sort	values which will be ranked to select signficiant genes.
	р	shreshold value
	colors	colors will be display
	legend	legend will be display

roma	The Main Function of RoMA	

Description

RoMA incorporates information from both mRNA abundance and raw counts by modeling RPKM (reads per kilobase per million), which represents the relative abundance of mRNA transcripts, and borrowing mean-variance dependency from CPM (counts per million) as a precision weight accounting for the variability in sequencing depth. LOWESS curve is fitted to capture the CPM mean-variance dependency and the inverse variance is taken into the modeling as the precision weight. Moderate-t statistics is performed by the eBayes function in limma.

Usage

```
roma(counts, rpkms = NULL, design = NULL, lib.size = NULL, normalize.method = "r
```

Arguments

counts	numeric matrix containing raw counts or a DGEList object.
rpkms	numeric vector containing RPKMs for each sample. If NULL and counts is a DGEList then it can be obtained from counts\$rpkms.
design	design matrix with rows corresponding to samples and columns to coefficients of study condition and covariates to be estimated.

6 x

lib.size numeric vector containing total library sizes for each sample. If NULL and

counts is a DGEList then library sizes are calculated from the columnwise counts $% \left(1\right) =\left(1\right) \left(1\right)$

totals.

normalize.method

normalization method to be applied to the logCPM values. Choices are as for the method argument of normalizeBetweenArrays of limma. As normalization is performed ahead in RoMA standard analysis, this argument is suggested to

set as "none".

span width of the lowess smoothing window as a proportion.

plot logical, should a plot of the mean-variance trend be displayed?

save.plot logical, should the coordinates and line of the plot be saved in the output?

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Counts Matrix

Description

Counts Matrix

Usage

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data("x")

Format

A data frame with 15998 observations on the following 14 variables.

X127_SRR037452 a numeric vector

X127_SRR037453 a numeric vector

X127_SRR037454 a numeric vector

X127_SRR037455 a numeric vector

 $X127_SRR037456$ a numeric vector

X127_SRR037457 a numeric vector

X127_SRR037458 a numeric vector

X128_SRR037459 a numeric vector

X128_SRR037460 a numeric vector

X128_SRR037461 a numeric vector

X128_SRR037462 a numeric vector

 $X128_SRR037463$ a numeric vector

X128_SRR037464 a numeric vector

X128_SRR037465 a numeric vector

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