

## METHODOLOGY

# Tissue Enrichment Analysis for *C. elegans* Genomics

David Angeles-Albores, Raymond YN Lee, Juancarlos Chan and Paul W Sternberg\*

\*Correspondence: [pws@caltech.edu](mailto:pws@caltech.edu)

HHMI and California Institute of Technology, Division of Biology and Biological Engineering, 1200 E California Blvd, 91125, Pasadena, USA

Full list of author information is available at the end of the article

### Abstract

**Background:** Over the last ten years, there has been explosive development in methods for measuring gene expression. These methods can identify thousands of altered genes between conditions, but understanding these datasets and forming hypotheses based on them remains challenging. One way to analyze these datasets is to associate ontologies (hierarchical, descriptive vocabularies with controlled relations between terms) with genes and to look for enrichment of specific terms. Although gene ontology (GO) is available for *Caenorhabditis elegans*, it does not include anatomical information.

**Results:** We have developed a tool for identifying enrichment of *C. elegans* tissues among gene sets and generated a website GUI where users can access this tool. Since a common drawback to ontology enrichment analyses is result verbosity, we developed a very simple trimming algorithm to reduce the ontology size by an order of magnitude. We adjusted these filters and validated our tool using a set of 30 gold standards from Expression Cluster data in WormBase and we show our tool can even discriminate between embryonic and larval tissues and can even identify tissues down to the single-cell level. We used our tool to identify multiple neuronal tissues that are down-regulated due to pathogen infection in *C. elegans*.

**Conclusions:** Our Tissue Enrichment Analysis (TEA) can be found within WormBase, and can be downloaded using Python's standard pip installer. It tests a slimmed-down *C. elegans* tissue ontology for enrichment of specific terms and provides users with a text and graphic representation of the results.

**Keywords:** Gene Ontology; Anatomy Ontology; WormBase; RNA-seq; High-throughput biology

### Background

RNA-seq and other high-throughput methods in biology have the ability to identify thousands of genes that are altered between conditions. These genes are often correlated in their biological characteristics or functions, but identifying these functions remains challenging. To interpret these long lists of genes, biologists need to abstract genes into concepts that are biologically relevant to form hypotheses about what is happening in the system. One such abstraction method relies on Gene Ontology (GO). GO provides a controlled set of hierarchically ordered terms [1, 2] that provide detailed descriptions about the molecular, cellular or biochemical functions of any gene. For a given gene list, existing software programs can query whether a particular term is enriched [3–5]. One area of biological significance that GO does not include is anatomy. One way to address this shortcoming is to use a ‘tissue

ontology’ that provides a complete anatomical description for an organism (e.g ‘tissue’, ‘organ’ or ‘specific cell’), in this case for *C. elegans* [6]. Cells and tissues are physiologically relevant units with broad, relatively well-understood functionalities amenable to hypothesis formation. The *C. elegans* database, WormBase [7], maintains a curated list of gene expression data from the literature. Here we provide a new framework that analyzes a user-input list for enrichment of specific cells and tissues.

Another problem frequently associated with GO enrichment analysis is that it is often difficult to interpret due to the large number of terms associated with a given gene. DAVID, a common tool for GO enrichment analysis, clusters enriched terms into broad categories [8], whereas PANTHER [3, 9] attempts to solve this issue by employing a manually reduced ontology, GOSlim (pers. comm., H. Yu and P. Thomas). To reduce verbosity, we have filtered our ontology using a small set of well-defined criteria to remove terms that do not contribute additional information. To our knowledge, such filtering has not been performed in an algorithmic fashion for a biological ontology before—indeed, DAVID does not employ term trimming *a priori* of testing, but rather fuzzy clustering *post* testing to reduce the number of ontology terms. Other pruning methods do exist (see for example [10]), but the pruning is query-dependent. We believe our trimming methodology strikes a good balance between detailed tissue calling and conservative testing.

## Results

### Generating a Gene-Tissue Dictionary

#### *Reducing term redundancy through a similarity metric*

As a first step to generate our tissue enrichment software, we wished to select tissue terms that were reasonably well-annotated, yet specific enough to provide insight and not redundant with other terms. For example, nematodes have a number of neurons that are placed symmetrically along the left/right body axis, and are functionally similar. These left/right neuronal pairs (which are sisters in the ontology) have almost identical annotations, with at most one or two gene differences between them, and therefore we cannot have statistical confidence in differentiating between them. As a result, testing these sister tissues provides no additional information compared with testing only the parent node to these sisters. To identify redundancy, we defined a similarity metric (see *Methods* section and Figure 1a) that can be used to identify sisters that have very high similarity between them; alternatively, redundant sisters could be identified if a single sister had a very high similarity score. We referred to these two scoring criteria as ‘avg’ and ‘any’ respectively.

#### *Terminal branch terms and parent terms can be safely removed in an algorithmic fashion*

Another problem arises from the ontology being scarcely populated. Many nodes have 0-10 annotations, which we consider too few to accurately test. To solve this issue, we implemented a straightforward trimming strategy. For a given terminal node, we test whether the node has more than a threshold number of annotations. If it does not, the node is removed. The next higher node in the branch is tested and removed recursively until a node that satisfies the condition is found. At that point,

no more nodes can be removed from that branch. This completion is guaranteed by the structure of the ontology: parent nodes inherit all of the annotations of all of their descendants, so the number of annotated terms monotonically increases with increasing term hierarchy (see Figure 1b). In this way, we ensure that our term dictionary includes only those tissues that are considered sufficiently well annotated for statistical purposes.

Additionally, we reasoned that, for any parent node, if all its daughters were selected for testing, there was no additional benefit to test the parent. We removed parent nodes from the analysis if all their daughter nodes passed the annotation threshold (see Figure 1c). We called this a ceiling filter. Applying these three filters reduced the number of ontology terms by an order of magnitude.

#### *Filtering greatly reduces the number of nodes used for analysis*

By itself, each of these filters can reduce the number of nodes employed for analysis, but applying the filters in different orders removes different numbers of nodes (not all the filters are commutative). We chose to always execute annotation and similarity thresholding first, followed by the ceiling filter. For validation (see below) we made a number of different dictionaries. The original ontology has almost 6,000 terms of which 1675 have at least 5 gene annotations. After filtering, dictionary sizes ranged from 21 to a maximum of 460 terms, which shows the number of terms in a scarcely annotated ontology can be reduced by an order of magnitude through the application of a few simple filters (see Table 1). These filters were used to compile a static dictionary that we employ for all analyses (see *Validation of the algorithm and parameter selection* section for details).

#### **Tissue enrichment testing via a hypergeometric model**

Having built a static dictionary, we generated a Python script that implements a significance testing algorithm based on the hypergeometric model. Briefly, the hypergeometric model tests the probability of observing  $n_i$  occurrences of a tissue  $i$  in a list of size  $M$  if there are  $m_i$  labels for that tissue in a dictionary of total size  $N$  that are drawn without replacement. Mathematically, this is expressed as:

$$P(n_i|N, m_i, M) = \frac{\binom{m_i}{n_i} \binom{M - m_i}{N - n_i}}{\binom{N}{n_i}}. \quad (1)$$

Although a user will input gene IDs, we test the number of occurrences of a term within the gene list, so a single gene can contribute to multiple terms. Due to the discrete nature of the hypergeometric distribution, this algorithm can generate artifacts when the list is small. To avoid spurious results, a tissue is never considered significant if there are no annotations for it in the user-provided list.

Once the p-values for each term have been calculated, we apply a standard FDR correction using a Benjamini-Hochberg step-up algorithm [11]. FDR corrected p-values are called q-values. Genes that have a q-value less than a given alpha are considered significant. Our default setting is an alpha of 0.1.

Users input a gene list using any valid gene name for *C. elegans*. These names are processed into standard WormBase gene IDs (WBGene IDs). The program returns a table containing all the enriched terms and associated information such as number of terms in gene list and expected number of terms. Finally, the program can also return a bar chart of the enrichment fold change for the fifteen tissues with the lowest measured q-values. The bars in the graph are sorted in ascending order of q-value and then in descending order of fold-change. Bars are colored for ease of viewing, and color does not convey information. Our software is implemented in an easy to use GUI (see Figure 2). Anatomy terms are displayed in human-readable format followed by their unique ontology ID (WBbt ID). In summary, each time the ontology annotations are updated, a new trimmed ontology is generated using our filters; in parallel, users can submit their gene lists through WormBase for testing, with results output in a number of formats (see Figure 3).

### Validation of the algorithm and optimizing parameter selection

We wanted to select a dictionary that included enough terms to be specific beyond the most basic *C. elegans* tissues, yet would minimize the number of spurious results and which had a good dynamic range in terms of enrichment fold-change. Larger tissues are correlated with better annotation, so increasing term specificity is associated with losses in statistical power. To help us select an appropriate dictionary and validate our tool, we used a set of 30 gold standards based on microarray and RNA-seq literature which are believed to be enriched in specific tissues [12–19]. These data sets are annotated gene lists derived from the corresponding Expression Cluster data in WormBase. Some of these studies have been used to annotate gene expression, and so they did not constitute an independent testing set. To correct this flaw, we built a clean dictionary that specifically excluded all annotation evidence that came from these studies.

As a first attempt to select a dictionary, we generated all possible combinations of dictionaries with minimal annotations of 10, 25, 33, 50 and 100 genes and similarity cutoffs of 0.9, 0.95 and 1, using ‘avg’ or ‘any’ similarity thresholding methods (see Table 1). The number of remaining ontology terms was inversely correlated to the minimum annotation cutoff, and was largely insensitive to the similarity threshold in the range we explored. Next, we analyzed all 30 datasets using each dictionary. Because of the large number of results, instead of analyzing each set of terms individually, we measured the average q-value for significantly enriched terms in each dataset without regard for the perceived accuracy of the terms that tested significant. We found that the similarity threshold mattered relatively little for any dictionary. We also noticed that the ‘any’ thresholding method resulted in tighter histograms with a mode closer to 0. For this reason, we chose the ‘any’ method for dictionary generation. The average q-value increased with decreasing annotation cut-off (see Figure 4), which reflects the decreasing statistical power associated with fewer annotations per term, but we remained agnostic as to how significant is the trade-off between power and term specificity. Based on these observations, we ruled out the dictionary with the 100 gene annotation cut-off: it had the fewest terms and its q-values were not low enough in our opinion to compensate for the trade-off in specificity.

To select between dictionaries generated between 50, 33 and 25 annotation cutoffs, and also to ensure the terms that are selected as enriched by our algorithm are reasonable, we looked in detail at the enrichment analysis results. Most results were comparable and expected. For some sets, all dictionaries performed well. For example, in our ‘all neuron enriched sets’ [13, 15] all terms were neuron-related regardless of the dictionary used (see Table ). On the other hand, for a set enriched for germline precursor expression in the embryo [13], the 50 cutoff dictionary was only able to identify ‘oocyte WBbt:006797’ which is not a germline precursor, although it is germline related; whereas the two smaller dictionaries singled out actual germline precursor cells—at the 33 cutoff, our tool identified the larval germline precursor cells ‘Z2’ and ‘Z3’ as enriched, and at the 25 gene cutoff the embryonic germline precursor terms ‘P<sub>4</sub>’, ‘P<sub>3</sub>’ and ‘P<sub>2</sub>’ were identified in addition to ‘Z2’ and ‘Z3’. We also queried an intestine precursor set [13]. Notably, this gene set yielded no enrichment when using the 25 cutoff dictionary, nor when using the 50 cutoff dictionary. However, the 33 cutoff dictionary identified the E lineage, which is the intestinal precursor lineage in *C. elegans*, as enriched. Both of these results capture specific aspects of *C. elegans* that are well known to developmental biologists.

Not all queries worked equally well. For example, a number of intestinal sets [13, 16] were not enriched in intestine-related terms in any dictionary, but were enriched for pharynx and hypodermis. We were surprised that intestinal gene sets performed poorly, since the intestine is a relatively well-annotated tissue.

We assessed the internal agreement of our tool by using independent gene sets that we expected to be enriched in the same tissues. We used two pan-neuronal sets [13, 15]; two PVD sets [13, 19]; and two GABAergic sets [13, 14]. Overall, the tool has good internal agreement. On most sets, the same terms were enriched, although order was somewhat variable (see Table ), and most high-scoring terms were preserved between sets. All comparisons can be found online in our Github repository (see Availability of data and materials). Overall, the dictionary generated by a 33 gene annotation cutoff with 0.95 redundancy threshold using the ‘any’ criterion performed best, with a good balance between specificity, verbosity and accuracy, so we selected this parameter set to generate our static dictionary. As of this publication, the testable dictionary contains 261 terms.

### Applying the tool

We applied our tool to the RNA-seq datasets developed by Engelmann *et al.* [20] to gain further understanding of their underlying biology. Engelmann *et al.* exposed young adult worms to 5 different pathogenic bacteria or fungi for 24 hours, after which mRNA was extracted from the worms for sequencing. We ran TEA on the genes Engelmann *et al.* identified as up- or down-regulated. Initially we noticed that genes that are down-regulated tend to be twice better annotated on average than genes that were up-regulated, suggesting that our understanding of the worm immune system is scarce, in spite of important advances made over the last decade. Up-regulated tissues, when detected, almost always included the hypodermis and excretory duct. Three out of the five samples showed enrichment of neuronal tissues or neuronal precursor tissues amongst the down-regulated genes. As an independent verification, we also performed GO analysis using PANTHER on the down-regulated

genes for *D. coniospora*. These results also showed enrichment in terms associated with neurons (see Figure 6). A possible explanation for this neuronal association might be that the infected worms are sick and the neurons are beginning to shut down; an alternative hypothesis would be that the worm is down-regulating specific neuronal pathways as a behavioral response against the pathogen. Indeed, several studies [21, 22] have provided evidence that *C. elegans* uses chemosensory neurons to identify pathogens. Our results highlight the involvement of various *C. elegans* neuronal tissues in pathogen defense.

## Discussion

We have presented a tissue enrichment analysis tool that employs a standard hypergeometric model to test the *C. elegans* tissue ontology. We use a hypergeometric function to test a user-provided gene list for enrichment of anatomical terms in *C. elegans*. Our hope is that the physiological relevance of anatomical terms will enable researchers to make hypotheses about high-dimensionality data. Specifically, we believe an enriched term may broadly suggest one of two hypotheses: if a list is enriched in a particular anatomical region, that anatomical region is affected by the experimental treatment; alternatively, the anatomical regions that are enriched reflect biologically relevant interactions between tissues. We believe the first hypothesis is a reasonable hypothesis to make in the case of whole-worm RNA-seq data for example, whereas the second hypothesis may be more plausible in cases where a researcher already knows what tissues a particular gene list came from, as may be the case in single-cell RNA-seq.

Our tool relies on an annotation dictionary that is continuously updated primarily with data from single gene qualitative analyses, does not require retraining and does not require ranked genes. To our knowledge, this is the first tool that tests tissue enrichment in *C. elegans* via the hypergeometric method, but similar projects exist for humans and zebrafish [23, 24], highlighting the relevance of our tool for high-dimensionality biology. Chikina *et al* [25] have previously reported a tissue enrichment model for *C. elegans* based on a Support Vector Machine classifier that has been trained on microarray studies. SVMs are powerful tools, but they require continuous retraining as more tissue expression data becomes available. Moreover, classifiers require that data be rank-ordered by some metric, something which is not possible for certain studies. Furthermore, this tissue enrichment tool provides users with enrichment results for only 6 large tissues. In contrast, our tool routinely tests a much larger number of terms, and we have shown it can even accurately identify enrichment of embryonic precursor lineages for select data sets.

We have also presented the first, to our knowledge, ontology trimming algorithm for biomedical ontologies. This algorithm, which is very easy to execute, places strong limits on the number of terms selected for testing. Due to the nature of all ontologies as hierarchical, acyclical graphs with term inheritance, term annotations are correlated along any given branch. This correlation reduces the benefits of including all terms for statistical analysis: for any given term along a branch, if that term passes significance, there is a high probability that many other terms along that branch will also pass significance. If the branch is enriched by random chance, error propagation along a branch means that many more false positives will follow.

Thus, a researcher might be misled by the number of terms of correlated function and assign importance to this finding; the fact that the branching structure of GO amplifies false positive signals is a powerful argument for either reducing branch length or branch intracorrelation, or both. On the other hand, if a term is actually enriched, we argue that there is little benefit to presenting the user with additional terms along that branch. Instead, a user will benefit most from testing sparsely along the tree at a suitable specificity for hypothesis formation. Related terms of the same level should only be tested when there is sufficient annotation to differentiate, with statistical confidence, whether one term is enriched above the other. Our algorithm reduces branch length by identifying and removing nodes that are insufficiently annotated and parents that are likely to include sparse information.

We endeavoured to benchmark our tool well, but our analysis cannot address problems related to spurious term enrichment. Although we were unable to determine false-positive and false-negative rates, we do not believe this should deter scientists from using our tool. Rather, we encourage researchers to use our tool as a guide, integrating evidence from multiple sources to inform the most likely hypotheses. As with any other tool based on statistical sampling, our analysis is most vulnerable to bias in the data set. For example, expression reports are negatively biased against germline expression because of the difficulties associated with expressing DNA in this tissue [26]. As time passes, we are certain the accuracy and power of this tool will improve thanks to the continuing efforts of the worm research community; indeed, without the community reports of tissue expression in the first place, this tool would not be possible.

## Methods

### Fetching annotation terms

We used WormBase-curated gene expression data, which includes annotated descriptions of spatial-temporal expression patterns of genes, to build our dictionary. Gene lists per anatomy term were extracted from a Solr document store of gene expression data from the WS252 database provided by WormBase [7]. We used the Solr document store because it provided a convenient access to expression data that included inferred annotations. That is, for each anatomy term, the expression gene list includes genes that were directly annotated to the term, as well as those that were annotated to the term's descendant terms (if there were any). Descendant terms were those connected with the focus term by *is\_a/part\_of* relationship chains defined in the anatomy term ontology hierarchy.

### Filtering nodes

#### *Defining a Similarity Metric*

To identify redundant sisters, we defined the following similarity metric:

$$s_i = \frac{|g_i|}{|\bigcup_{i=0}^k g_i|} \quad (2)$$

Where  $s_i$  is the similarity for a tissue  $i$  with  $k$  sisters;  $g_i$  refers to the set of tissues associated with tissue  $i$  and  $|g|$  refers to the cardinality of set  $g$ . For a given set



of sisters, we called them redundant if they exceeded a given similarity threshold. We envisioned two possible criteria and built different dictionaries using each one. Under a threshold criterion ‘any’ with parameter  $S$  between  $(0, 1)$ , a given set of sisters  $j$  was considered redundant if the condition

$$s_{i,j} > S \quad (3)$$

was true for any sister  $i$  in set  $j$ . Under a threshold criterion ‘avg’ with parameter  $S$ , a given set of sisters  $j$  was considered redundant if the condition

$$E[s_i]_j > S \quad (4)$$

was true for the set of sisters  $j$  (see Figure 1a).

### Implementation

All scripts were written in Python 3.5. Our software relies on the pandas, NumPy, Seaborn and SciPy modules to perform all statistical testing and data handling [27–29].

## Availability of data and materials

Our web implementation is available at <http://www.wormbase.org/tools/enrichment/tea/tea.cgi>. Our software can also be downloaded using Python’s pip installer via the command

```
pip install tissue_enrichment_analysis
```

Alternatively, our software is available for download at: <http://dangeles.github.io/TissueEnrichmentAnalysis>

All benchmark gene sets, benchmarking code and Figures can also be found at the same address, under the ‘tests’ folder.

### Abbreviations

- TEA — Tissue Enrichment Analysis
- GO — Gene Ontology
- WBbt ID — A unique ID assigned to reference ontology terms
- WBgene ID — A unique ID assigned to reference nematode genes

### Ethics Approval and Consent to Participate

Not applicable

### Consent for Publication

Not applicable

### Competing interests

The authors declare that they have no competing interests.

### Author’s contributions

DA and PWS conceived of the project; DA developed algorithm; RYL made intellectual contributions to the project; RYL and JC developed the web GUI.

### Acknowledgements

We thank Justin Bois for his help and support. We would like to acknowledge all members of the Sternberg lab for helpful discussion.

### Additional Files

Additional file 1 — TEA Tutorial

Tutorial for users interested in using our software within a python script



Additional file 2 — Folder Structure for SI files 3 and 4  
A file detailing the folder structure of the zipped folders 3 and 4.

Additional file 3 — Golden Gene Sets  
A list of all the genes used for our benchmarking process.

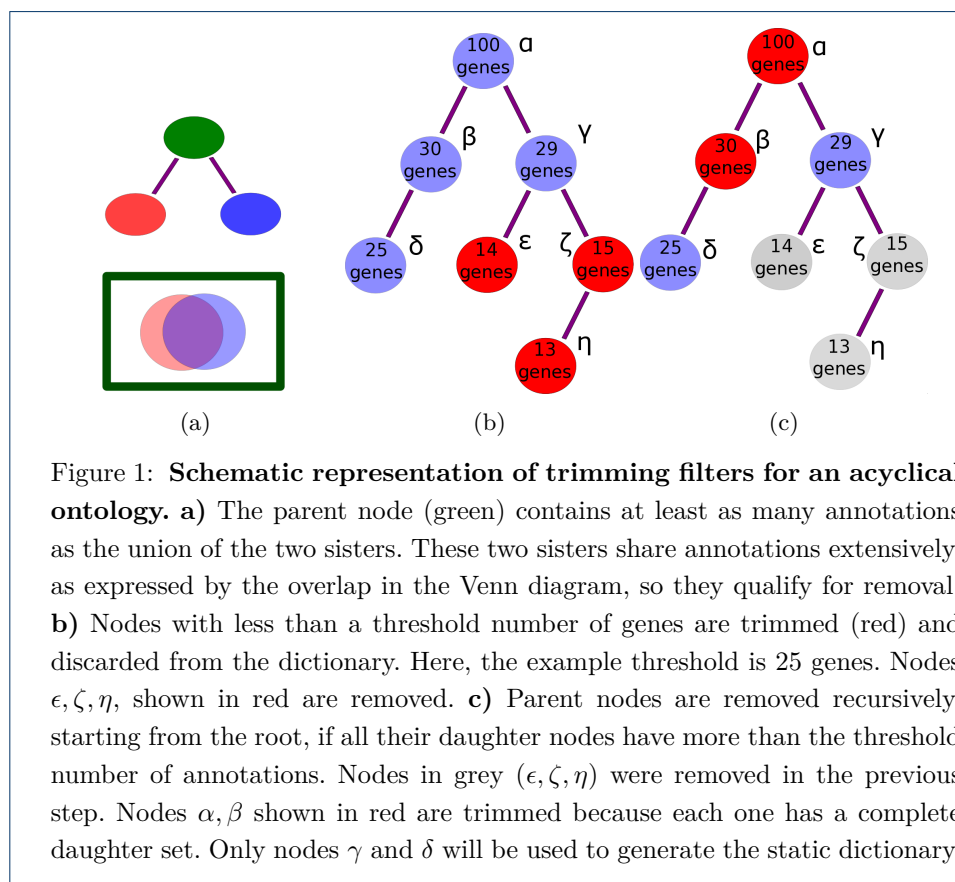
Additional file 4 — Results  
A folder containing a complete version of the results we generated for this paper.

## References

1. The Gene Ontology Consortium. Gene Ontology: tool for the unification of biology. *Nature Genetics*, 25(may):25–29, 2000.
2. The Gene Ontology Consortium. Gene Ontology Consortium: going forward. *Nucleic Acids Research*, 43(D1):D1049–D1056, 2015.
3. Huaiyu Mi, Qing Dong, Anushya Muruganujan, Pascale Gaudet, Suzanna Lewis, and Paul D. Thomas. PANTHER version 7: Improved phylogenetic trees, orthologs and collaboration with the Gene Ontology Consortium. *Nucleic Acids Research*, 38(SUPPL.1), 2009.
4. Cory Y McLean, Dave Bristor, Michael Hiller, Shoa L Clarke, Bruce T Schaar, Craig B Lowe, Aaron M Wenger, and Gill Bejerano. GREAT improves functional interpretation of cis-regulatory regions. *Nature biotechnology*, 28(5):495–501, 2010.
5. Da Wei Huang, Richard A Lempicki, and Brad T Sherman. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nature Protocols*, 4(1):44–57, 2009.
6. R. Y N Lee and Paul W. Sternberg. Building a cell and anatomy ontology of *Caenorhabditis elegans*, 2003.
7. Kevin L Howe, Bruce J Bolt, Scott Cain, Juan Carlos Chan, Wen J Chen, Paul Davis, James Done, Thomas Down, Sibyl Gao, Christian Grove, Todd W Harris, Ranjana Kishore, Raymond Lee, Jane Lomax, Yuling Li, Hans-Michael Muller, Cecilia Nakamura, Paulo Nuin, Michael Paulini, Daniela Raciti, Gary Schindelman, Eleanor Stanley, Mary Ann Tuli, Kimberly Van Aiken, Daniel Wang, Xiaodong Wang, Gary Williams, Adam Wright, Karen Yook, Matthew Berriman, Paul Kersey, Tim Schedl, Lincoln Stein, and Paul W Sternberg. WormBase 2016: expanding to enable helminth genomic research. *Nucleic acids research*, 44(November 2015):D774–D780, 2016.
8. Da Wei Huang, Brad T. Sherman, Qina Tan, Joseph Kir, David Liu, David Bryant, Yongjian Guo, Robert Stephens, Michael W. Baseler, H. Clifford Lane, and Richard A. Lempicki. DAVID Bioinformatics Resources: Expanded annotation database and novel algorithms to better extract biology from large gene lists. *Nucleic Acids Research*, 35(SUPPL.2), 2007.
9. Huaiyu Mi, Anushya Muruganujan, and Paul D. Thomas. PANTHER in 2013: Modeling the evolution of gene function, and other gene attributes, in the context of phylogenetic trees. *Nucleic Acids Research*, 41(D1), 2013.
10. Jong Woo Kim, Jordi Conesa Caralt, and Julia K. Hilliard. Pruning bio-ontologies. *Proceedings of the Annual Hawaii International Conference on System Sciences*, pages 1–10, 2007.
11. Yoav Benjamini and Yoel Hochberg. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing, 1995.
12. Jeb Gaudet, Srikanth Muttumu, Michael Horner, and Susan E. Mango. Whole-genome analysis of temporal gene expression during foregut development. *PLoS Biology*, 2(11), 2004.
13. W. Clay Spencer, Georg Zeller, Joseph D. Watson, Stefan R. Henz, Kathie L. Watkins, Rebecca D. McWhirter, Sarah Petersen, Vipin T. Sreedharan, Christian Widmer, Jeanyoung Jo, Valerie Reinke, Lisa Petrella, Susan Strome, Stephen E. Von Stetina, Menachem Katz, Shai Shaham, Gunnar Rätsch, and David M. Miller. A spatial and temporal map of *C. elegans* gene expression. *Genome Research*, 21(2):325–341, 2011.
14. Hulusi Cinar, Sunduz Keles, and Yishi Jin. Expression profiling of GABAergic motor neurons in *Caenorhabditis elegans*. *Current Biology*, 15(4):340–346, 2005.
15. Joseph D Watson, Shenglong Wang, Stephen E Von Stetina, W Clay Spencer, Shawn Levy, Phillip J Dexheimer, Nurith Kurn, Joe Don Heath, D M Miller 3rd, and David M Miller. Complementary RNA amplification methods enhance microarray identification of transcripts expressed in the *C. elegans* nervous system. *BMC Genomics*, 9:84, 2008.
16. Florencia Pauli, Yueyi Liu, Yoona A Kim, Pei-Jiun Chen, and Stuart K Kim. Chromosomal clustering and GATA transcriptional regulation of intestine-expressed genes in *C. elegans*. *Development (Cambridge, England)*, 133(2):287–295, 2006.
17. Douglas S. Portman and Scott W. Emmons. Identification of *C. elegans* sensory ray genes using whole-genome expression profiling. *Developmental Biology*, 270(2):499–512, 2004.
18. Rebecca M Fox, Joseph D Watson, Stephen E Von Stetina, Joan McDermott, Thomas M Brodigan, Tetsunari Fukushima, Michael Krause, D M Miller 3rd, and David M Miller. The embryonic muscle transcriptome of *Caenorhabditis elegans*. *Genome Biol*, 8(9):R188, 2007.
19. Cody J. Smith, Joseph D. Watson, W. Clay Spencer, Tim O’brien, Byeong Cha, Adi Albeg, Millet Treinin, and David M. Miller. Time-lapse imaging and cell-specific expression profiling reveal dynamic branching and molecular determinants of a multi-dendritic nociceptor in *C. elegans*. *Developmental Biology*, 345(1):18–33, 2010.
20. Ilka Engelmann, Aurélien Griffon, Laurent Tichit, Frédéric Montaña-Sanchis, Guilin Wang, Valerie Reinke, Robert H. Waterston, LaDeana W. Hillier, and Jonathan J. Ewbank. A comprehensive analysis of gene expression changes provoked by bacterial and fungal infection in *C. elegans*. *PLoS ONE*, 6(5), 2011.
21. Joshua D. Meisel and Dennis H. Kim. Behavioral avoidance of pathogenic bacteria by *Caenorhabditis elegans*. *Trends in Immunology*, 35(10):465–470, 2014.
22. Y Zhang, H Lu, and C I Bargmann. Pathogenic bacteria induce aversive olfactory learning in *Caenorhabditis elegans*. *Nature*, 438(7065):179–184, 2005.

23. Young Suk Lee, Arjun Krishnan, Qian Zhu, and Olga G. Troyanskaya. Ontology-aware classification of tissue and cell-type signals in gene expression profiles across platforms and technologies. *Bioinformatics*, 29(23):3036–3044, 2013.
24. Sergey V Prykhodzhiy, Annalisa Marsico, and Sebastiaan H Meijnsing. Zebrafish Expression Ontology of Gene Sets (ZEOGS): a tool to analyze enrichment of zebrafish anatomical terms in large gene sets. *Zebrafish*, 10(3):303–15, 2013.
25. Maria D. Chikina, Curtis Huttenhower, Coleen T. Murphy, and Olga G. Troyanskaya. Global prediction of tissue-specific gene expression and context-dependent gene networks in *Caenorhabditis elegans*. *PLoS Computational Biology*, 5(6), 2009.
26. William G. Kelly, SiQun Xu, Mary K. Montgomery, and Andrew Fire. Distinct requirements for somatic and germline expression of a generally expressed *Caenorhabditis elegans* gene. *Genetics*, 146(1):227–238, 1997.
27. Wes McKinney. pandas: a Foundational Python Library for Data Analysis and Statistics. *Python for High Performance and Scientific Computing*, pages 1–9, 2011.
28. Stéfan Van Der Walt, S. Chris Colbert, and Gael Varoquaux. The NumPy array: A structure for efficient numerical computation. *Computing in Science and Engineering*, 13(2):22–30, 2011.
29. Travis E Oliphant. SciPy: Open source scientific tools for Python. *Computing in Science and Engineering*, 9:10–20, 2007.

# Figures



# Tables

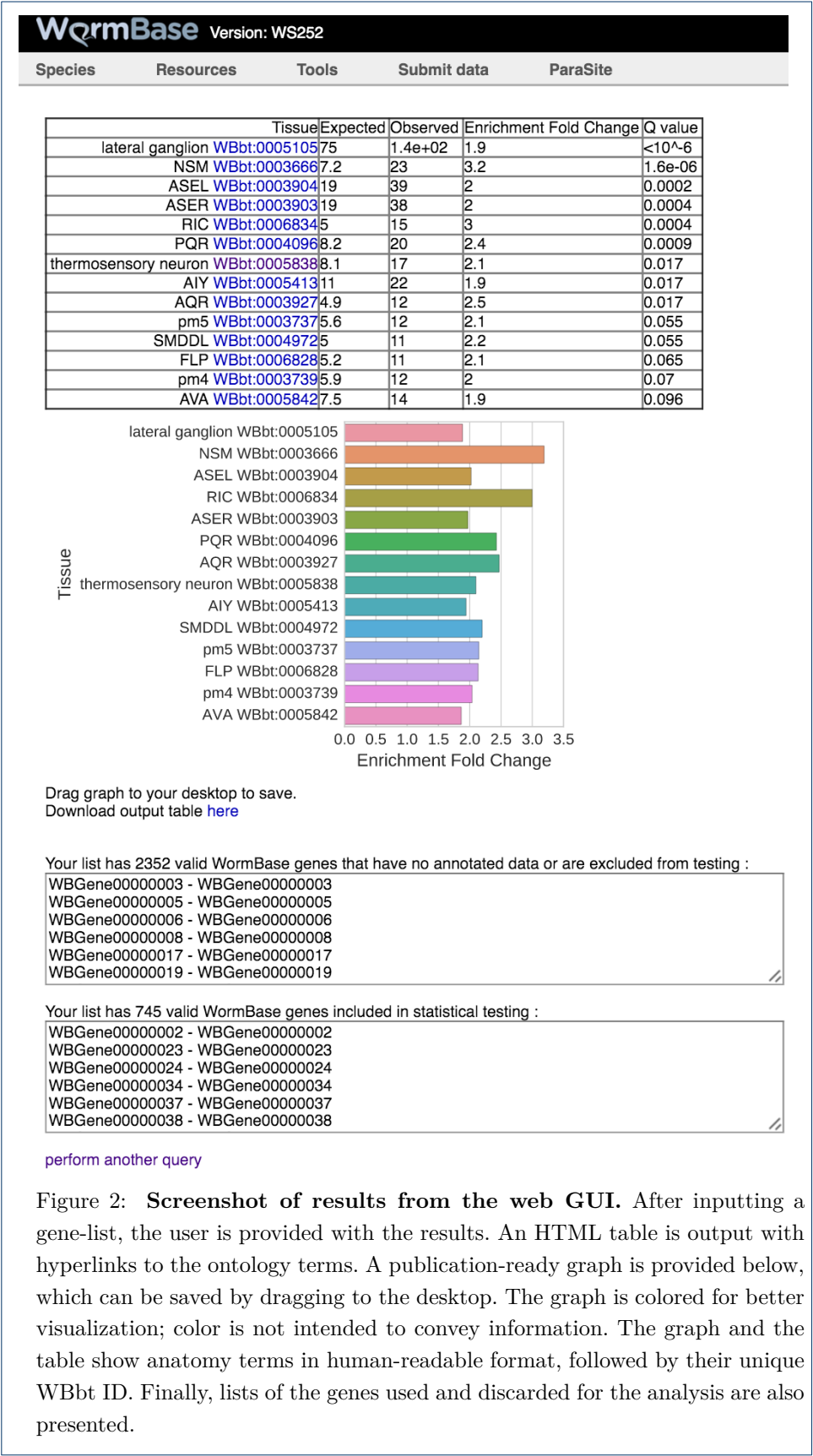


Figure 2: **Screenshot of results from the web GUI.** After inputting a gene-list, the user is provided with the results. An HTML table is output with hyperlinks to the ontology terms. A publication-ready graph is provided below, which can be saved by dragging to the desktop. The graph is colored for better visualization; color is not intended to convey information. The graph and the table show anatomy terms in human-readable format, followed by their unique WBbt ID. Finally, lists of the genes used and discarded for the analysis are also presented.

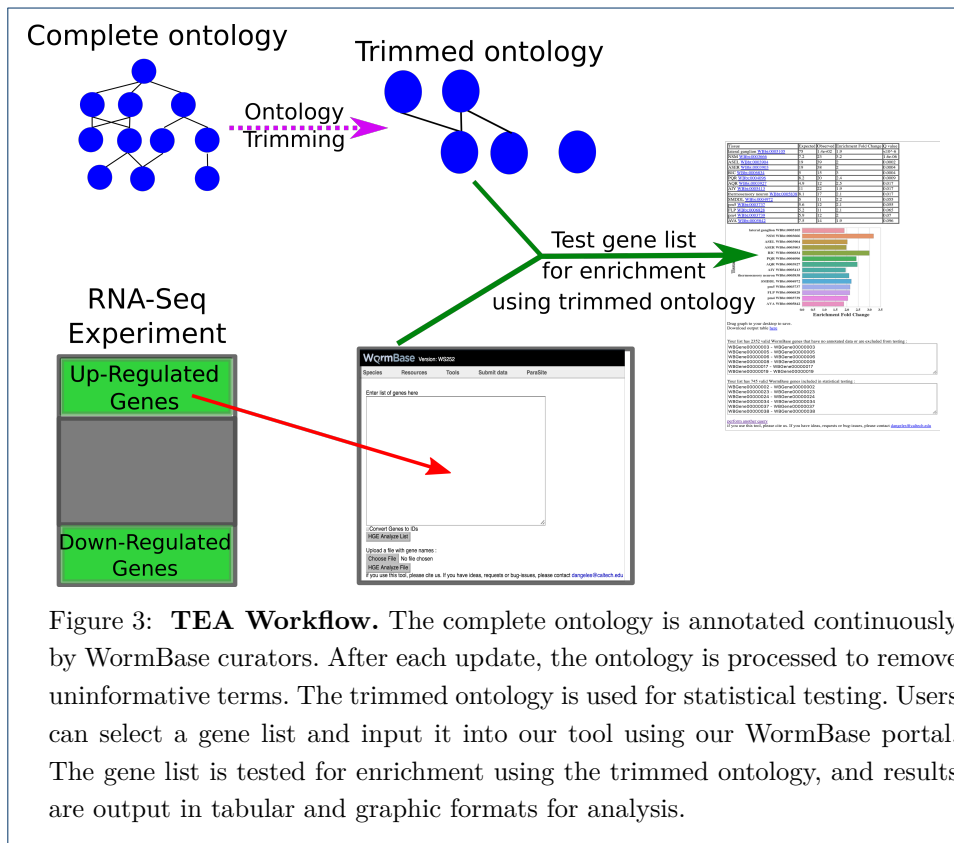


Figure 3: **TEA Workflow.** The complete ontology is annotated continuously by WormBase curators. After each update, the ontology is processed to remove uninformative terms. The trimmed ontology is used for statistical testing. Users can select a gene list and input it into our tool using our WormBase portal. The gene list is tested for enrichment using the trimmed ontology, and results are output in tabular and graphic formats for analysis.

Table 1: Parameter specifications and number of tissues for all dictionaries. The ‘Method’ column refers to the trimming criterion for the similarity metric. We used two such criteria, ‘any’ and ‘avg’: For a given sister set, if any sister had a similarity exceeding the corresponding threshold, all sisters were removed from the final dictionary. ‘avg’: For a given sister set, if the average similarity across all the sisters in the set was greater than the corresponding threshold, all sisters were removed from the final dictionary.

Annotation Cutoff	Similarity Threshold	Method	No. Of Terms in Dictionary
25	0.9	any	460
25	0.9	avg	461
25	0.95	any	466
25	0.95	avg	468
25	1.0	any	476
25	1.0	avg	476
33	0.9	any	261
33	0.9	avg	255
33	0.95	any	261
33	0.95	avg	262
33	1.0	any	247
33	1.0	avg	247
50	0.9	any	83
50	0.9	avg	77
50	0.95	any	82
50	0.95	avg	81
50	1.0	any	70
50	1.0	avg	70
100	0.9	any	45
100	0.9	avg	35
100	0.95	any	42
100	0.95	avg	36
100	1.0	any	21
100	1.0	avg	21

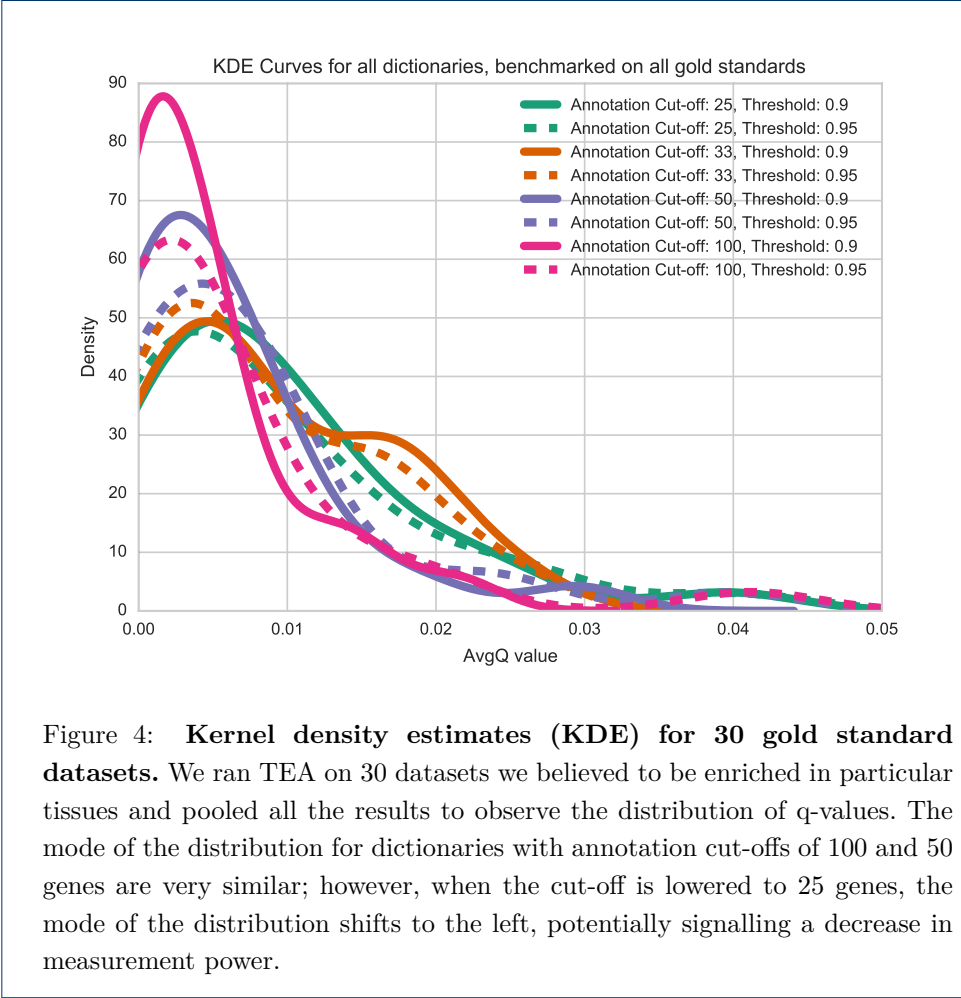


Table 2: Comparison of results for a neuronal-enriched gene set from Watson [15]. We ran the same genelist on a dictionary with a minimum annotation cutoff of 50, similarity threshold of 0.95 and similarity method ‘any’ versus another with a minimum annotation cutoff of 33, similarity threshold of 0.95 and similarity method ‘any’. In the table, columns are labeled with their significance value (Q-value) or enrichment fold change followed by a hyphen and a number which indicates which the cutoff for the dictionary that was used for testing. Not all tissues are present in either dictionary. Hyphens denote not-applicable values, which occurs when a particular tissue is not present in both dictionaries. Full table is available in Additional File 2.

Tissue	Q-value-33	Q-value-50	Enrichment-33	Enrichment-50
nerve ring WBbt:0006749	8.7e-11	2.7e-11	2.9	2.9
nervous system WBbt:0005735	1.1e-09	3.5e-10	1.4	1.4
retrovesicular ganglion WBbt:0005656	1.4e-08	4.2e-09	5.7	5.7
lateral ganglion WBbt:0005105	2e-08	6.1e-09	2.8	2.8
dorsal nerve cord WBbt:0006750	1.3e-05	4e-06	3.2	3.2
tail WBbt:0005741	3e-05	9.4e-06	1.6	1.6
SMDDL WBbt:0004972	7.4e-05	-	7.8	-
BAG WBbt:0006825	0.0002	-	6.8	-
ventral nerve cord WBbt:0005829	0.00032	0.00013	2.3	2.3
PVC WBbt:0005840	0.00034	-	6.1	-

