Plasma Proteomics with HDAnalyzeR:: CHEAT SHEET

Basics

With **HDAnalyzeR** you can perform complex plasma proteomics analysis with simple steps. Starting from Olink data and metadata via simple functions to biomarker discovery!









Remember that the Olink data should contain the columns: DAid, Assay, and NPX

The metadata should contain the columns: DAid, Disease (column with case-control groups), and Sex (M or F)

Utilities

create_dir(dir_name, date = FALSE)

Creates a directory with a specified name. The user can choose to create another inner directory with the current date as its name.

create_dir("my_directory", date = FALSE)

save_df(df, file_name, dir_name, date = FALSE, file_type = c("csv", "tsv", "rda"))

Saves a dataset in the specified format (CSV, TSV, or RDA) in a specified directory. The recommended file type is RDA.

save_df(example_metadata, "metadata", "my_data", file_type = "rda")

import df(file path)

Imports a dataset from a file. The file format can be CSV. TSV. TXT. RDA, RDS, XLSX, or Parquet format. It returns the dataset as tibble.

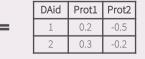
import_df("my_data/metadata.rda")

widen_data(olink_data)

Transforms Olink data from long to wide format.

widen_data(example_data)

DAid	Assay	NPX
1	Prot1	0.2
1	Prot2	-0.5
2	Prot1	0.3
2	Prot2	-0.2



* To improve performance, it is recommended to use tidied data throughout the analysis. Initially, widen the data in your script.

Preprocessing Data

PREPROCESSING

clean data(df in, keep cols = c("DAid", "Assay", "NPX"), cohort = NULL, filter plates = NULL, filter assays = NULL, filter assay warning = FALSE, remove_na_cols = c("DAid", "NPX"), replace_w_na = c(0, "0", "", "Unknown", "unknown", "none", NA, "na"))

clean_metadata(df_in, keep_cols = c("DAid", "Assay", "Sex", "Age", "BMI"), remove_na_cols = c("DAid", "Disease"), replace_w_na = c(0, "0", "", "Unknown", "unknown", "none", NA, "na"))

Select columns and filter rows based on the specified criteria.

clean data(example data, filter plates = c("Plate1", "Plate2"))

generate df(long_data, metadata = NULL, join = TRUE, metadata_cols = c("DAid", "Disease", "Sex", "Age", "BMI"), save = TRUE)

Creates wide and join dataframes from long Olink data and metadata.

generate_df(example_data, example_metadata, save = FALSE)

DATA NORMALIZATION & IMPUTATION

normalize_data(olink_data, metadata = NULL, wide = TRUE, center = TRUE, scale = TRUE, batch = NULL, batch2 = NULL, return_long = FALSE, save = FALSE, file_name = "normalized_data")

normalizes the data by scaling them and removing their batch effects.

normalize_data(example_data, example_metadata, wide = FALSE)

impute_median(olink_data, wide = TRUE, exclude_cols = c("DAid", "Disease"), show_na_percentage = TRUE)

impute knn(olink data, wide = TRUE, k = 5, exclude cols = c("DAid", "Disease"), show_na_percentage = TRUE)

impute_missForest(olink_data, wide = TRUE, maxiter = 10, ntree = 100, parallelize = "variables", ncores = 4, exclude_cols = c("DAid", "Disease"), show_na_percentage = TRUE)

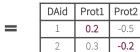
impute mice(olink data, wide = TRUE, m = 5, maxit = 5, method = "pmm",exclude_cols = c("DAid", "Disease"), show_na_percentage = TRUE)

0.2

Impute missing values in a dataset using different techniques.

impute_knn(example_data, wide = FALSE, k = 3)

DAid	Prot1	Prot2	
1	NA	-0.5	
2	0.3	NA	



Data Quality Control

QUALITY CONTROL

qc summary data(df, wide = TRUE, threshold = 0.8, report = TRUE)

HDAnalyze

qc_summary_metadata(metadata, categorical = "Sex", numeric = "Age", disease_palette = NULL, categ_palette = "sex_hpa", report = T)

Summarize the quality control results of data and metadata. Plot distributions of metadata variables.

qc_summary_data(example_data, wide = FALSE, threshold = 0.7)

CORRELATION & CLUSTERING

create_corr_heatmap(x, y = NULL, use = "pairwise.complete.obs", method = "pearson", threshold = 0.8, cluster_rows = TRUE, cluster cols = TRUE)

Calculates correlation matrix and creates heatmap. Returns list of protein pairs that correlate above the defined threshold.

create corr heatmap(wide data, threshold = 0.7)

cluster_data(df, distance_method = "euclidean", clustering_method = "ward.D2", cluster_rows = TRUE, cluster_cols = TRUE, wide = TRUE)

Takes a dataset and returns the same dataset ordered according to the hierarchical clustering of the rows and columns

cluster_data(example_data, wide = FALSE)

DAid	Prot1	Prot2	Prot3
1	0.2	-0.5	1.2
2	0.3	-0.2	1.3
3	0.2	-0.4	1.1



DAid	Prot1	Prot3	Prot
2	0.3	1.3	-0.2
1	0.2	1.2	-0.5
3	0.2	1.1	-0.4

DIMENSIONALITY REDUCTION

do_pca(olink_data, metadata = NULL, pcs = 5, color = "Disease", palette = NULL, wide = TRUE, assay = FALSE, impute = TRUE, plots = TRUE, x = "PC1", y = "PC2", npcs = 4, nproteins = 8, loadings = FALSE, save = FALSE)

do_umap(olink_data, metadata = NULL, color = "Disease", palette = NULL, wide = TRUE, assay = FALSE, impute = TRUE, plots = TRUE, save =

Run a PCA or UMAP analysis on the the data. Visualize the data points on 2D planes.

do_pca(example_data, example_metadata, wide = FALSE, color = "Disease", palette = "cancers12")









Main Analysis

DIFFERENTIAL EXPRESSION

do_limma(olink_data, metadata, variable = "Disease", case, control, correct = c("Sex", "Age"), correct_type = c("factor", "numeric"), wide = TRUE, only_female = NULL, only_male = NULL, volcano = TRUE, pval_lim = 0.05, logfc_lim = 0, top_up_prot = 40, top_down_prot = 10, palette = "diff_exp", report_nproteins = TRUE, subtitle = NULL, save = FALSE)

Perform differential expression analysis. Ability to correct for cofactors.

do_limma(example_data, example_metadata, case = "AML", control =
c("CLL", "MYEL"), wide = FALSE)

do_ttest(olink_data, metadata, variable = "Disease", case, control,
wide = TRUE, only_female = NULL, only_male = NULL, volcano =
TRUE, pval_lim = 0.05, logfc_lim = 0, top_up_prot = 40,
top_down_prot = 10, palette = "diff_exp", report_nproteins = TRUE,
subtitle = NULL, save = FALSE)

Perform differential expression analysis by comparing the groups with t-test.

do_ttest(example_data, example_metadata, case = "AML", control =
c("CLL", "MYEL"), wide = FALSE)

do_limma_continuous(olink_data, metadata, variable, correct =
c("Sex"), correct_type = c("factor"), wide = TRUE, volcano = TRUE,
pval_lim = 0.05, logfc_lim = 0, top_up_prot = 40, top_down_prot = 10,
palette = "diff_exp", report_nproteins = TRUE, subtitle = NULL, save =
FALSE)

Perform differential expression analysis against continuous variable.

do_limma_continuous(example_data, example_metadata, "Age", wide = FALSE)

Assay	logFC	pval	Av. Expr
Prot1	0.01	0.01	1.2
Prot2	-0.2	0.02	1.3
Prot3	0.04	0.06	1.1



VISUALIZE RESULTS

Summarize the results from differential expression and classification models. Plot boxplots and correlation plots with regression on top.

plot_de_summary(de_results, disease_palette = NULL, diff_exp_palette = "diff_exp")

plot_features_summary(ml_results, importance = 50,
upset_top_features = FALSE, case_palette = NULL,
feature_type_palette = c(`all-features` = "pink", `top-features` =
"darkblue"))

plot_protein_boxplot(join_data, variable = "Disease", proteins,
case, points = TRUE, xaxis_names = FALSE, palette = NULL)

plot_scatter_wth_regression(plot_data, x, y, se = FALSE, line_color = "black", r_2 = TRUE)







do_rreg(olink_data, metadata, variable = "Disease", case, control,
wide = TRUE, strata = TRUE, balance_groups = TRUE, only_female =
NULL, only_male = NULL, exclude_cols = "Sex", ratio = 0.75, type =
"lasso", cor_threshold = 0.9, cv_sets = 5, grid_size = 10, ncores = 4,
hypopt_vis = TRUE, palette = NULL, vline = TRUE, subtitle =
c("accuracy", "sensitivity", "specificity", "auc", "features", "topfeatures", "mixture"), varimp_yaxis_names = FALSE, nfeatures = 9,
points = TRUE, boxplot_xaxis_names = FALSE, seed = 123)

do_rf(olink_data, metadata, variable = "Disease", case, control,
wide = TRUE, strata = TRUE, balance_groups = TRUE, only_female =
NULL, only_male = NULL, exclude_cols = "Sex", ratio = 0.75,
cor_threshold = 0.9, normalize = TRUE, cv_sets = 5, grid_size = 10,
ncores = 4, hypopt_vis = TRUE, palette = NULL, vline = TRUE, subtitle
= c("accuracy", "sensitivity", "specificity", "auc", "features", "top-features"), varimp_yaxis_names = FALSE, nfeatures = 9, points =
TRUE, boxplot_xaxis_names = FALSE, seed = 123)

Perform binary classification with regularized regression (elastic net, lasso, ridge) and random forest models. Ability to optimize model hyperparameters.

do_rf(example_data, example_metadata, case = "AML", control = c("CLL", "MYEL"), wide = FALSE, palette = "cancers12", cv_sets = 5, grid_size = 10)

do_rreg_multi(olink_data, metadata, variable = "Disease", wide =
TRUE, strata = TRUE, exclude_cols = "Sex", ratio = 0.75, type =
"lasso", cor_threshold = 0.9, cv_sets = 5, grid_size = 10, ncores = 4,
hypopt_vis = TRUE, palette = NULL, seed = 123)

do_rf_multi(olink_data, metadata, variable = "Disease", wide =
TRUE, strata = TRUE, exclude_cols = "Sex", ratio = 0.75,
cor_threshold = 0.9, normalize = TRUE, cv_sets = 5, grid_size = 10,
ncores = 4, hypopt_vis = TRUE, palette = NULL, seed = 123)

Perform multi classification with regularized regression (elastic net, lasso, ridge) and random forest models. Ability to optimize model hyperparameters.

do_rf_multi(example_data, example_metadata, wide = FALSE, palette = "cancers12", cv_sets = 5, grid_size = 5)

do_lreg(olink_data, metadata, variable = "Disease", case, control,
wide = TRUE, strata = TRUE, balance_groups = TRUE, only_female =
NULL, only_male = NULL, exclude_cols = "Sex", ratio = 0.75,
cor_threshold = 0.9, normalize = TRUE, cv_sets = 5, ncores = 4,
palette = NULL, points = TRUE, boxplot_xaxis_names = FALSE, seed =
123)

Perform binary classification with logistic regression model. Ideal for data with single predictor. If data contain multiple predictors prefer to use do rreg

do_lreg(single_predictor_data, example_metadata, case = "AML", control
= "CLL", wide = FALSE, ncores = 1, palette = "cancers12")







Post Analysis

LITERATURE SEARCH

literature_search(prot_dis_list, max_articles = 10)

searches for articles for protein-disease pairs in PubMed.

prot_dis_list <- list("acute myeloid leukemia" = c("FLT3", "EPO"))</pre>

lit_search_results <- literature_search(prot_dis_list, max_articles = 1))</pre>

ENRICHMENT ANALYSIS

do_ora(protein_list, database = c("GO", "Reactome"), background =
NULL, pval_lim = 0.05)

plot_ora(enrichment, protein_list, pval_lim = 0.05, ncateg = 10,
fontsize = 10)

Performs over-representation analysis (ORA) using the clusterProfiler package and plot results.

control = c("BRC", "CLL", "CRC", "CVX", "ENDC", "GLIOM", "LUNGC",
 "LYMPH", "MYEL", "OVC", "PRC")
de_res <- do_limma(example_data, example_metadata, case = "AML",
 control = control, wide = FALSE)</pre>

sig_up_proteins_aml <- de_res\$de_results |>
dplyr::filter(sig == "significant up") |>
dplyr::pull(Assay)

do_ora(sig_up_proteins_aml, database = "GO")

Enrichment <- plot_ora(enrichment, protein_list, pval_lim = 0.05, ncateg = 10, fontsize = 10)

do_gsea(de_results, database = c("GO", "Reactome"), pval_lim = 0.05)

plot_gsea(enrichment, de_results, pval_lim = 0.05, ncateg = 10, fontsize = 10)

Performs Gene Set Enrichment Analysis (GSEA) using the clusterProfiler package and plot results.

control = c("BRC", "CLL", "CRC", "CVX", "ENDC", "GLIOM", "LUNGC",
 "LYMPH", "MYEL", "OVC", "PRC")
de_res <- do_limma(example_data, example_metadata, case = "AML",
 control = control, wide = FALSE)</pre>

de_results <- de_res\$de_results
Enrichment <- do_gsea(de_results, database = "GO", pval_lim = 0.9)</pre>

plot gsea(enrichment, de results, pval lim = 0.9, ncateg = 7, fontsize = 7)