Mandatory 3

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```
rm(list = ls())
library(coda)
library(mvtnorm)
```

Note: For most of the simulations regarding random walk, burn-in has been used even though it is not necessary for all types of random walk. As long as the effective sample size is sufficient, burn-in is OK to use, which is why it has been used in most of the simulations. The reason burn-in is used in some random walks is because the first part of the MCMC simulation is spent "locating" the high density region. The process of "locating" the high density is not of interest, and is therefore removed by burn-in. Independent random walk does not need burn-in, as the samples are independent and therefore not used to "locating".

Problem 1: Random number generation and Monte Carlo integration Problem 1A

Generate n independent random variables from the distribution-density p(x).

$$p(x) = \frac{2^{\frac{1}{4}}\Gamma\left(\frac{3}{4}\right)}{\pi} \exp\left(-\frac{x^4}{2}\right), \quad -\infty < x < \infty$$

```
# The probability density p(x)
p <- function(x) {</pre>
  2^{(1/4)}*gamma(3/4)/pi*exp(-(x^4)/2)
oneD.IRWMH <- function(prob, sigma, theta, Nsim = 10000){
  res <- vector(length = Nsim)</pre>
  # allocate memory
  res[1] <- theta
  # old importance weight
  wt.old <- prob(res[1])/dnorm(res[1], sd = sigma)
  Nacc <- 0
    for(i in 2:Nsim){
      # proposal (note, independent of past)
      thetaStar <- rnorm(1, sd = sigma)</pre>
      # new importance weight
      wt.star <- prob(thetaStar)/dnorm(thetaStar, sd = sigma)
      # accept probability
      alpha <- min(1.0, wt.star/wt.old)</pre>
      # accept/reject
      if(runif(1) < alpha){</pre>
        res[i] <- thetaStar
        wt.old <- wt.star
        Nacc <- Nacc + 1
```

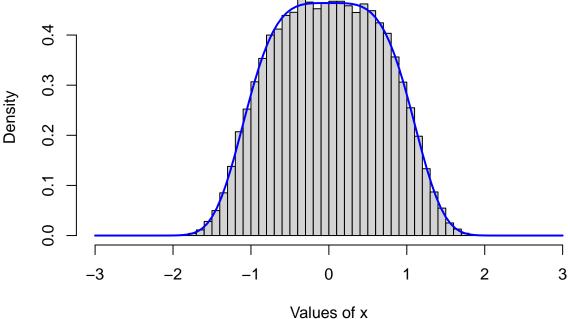
```
} else {
    res[i] <- res[i-1]
}

res
}

Nsim <- 100000
IndRandVariables <- oneD.IRWMH(p, theta = 0.0, sigma = 0.8, Nsim = Nsim)
IndRandVariables.burned <- IndRandVariables[201:length(IndRandVariables)]

# Plotting the random variables together with the probability density.
hist(IndRandVariables.burned, probability = TRUE, breaks = 50, xlim = c(-3, 3),
    main = "Independent random variables", xlab = "Values of x")
curve(p, add = TRUE, -3, 3, col = "blue", lwd = 2)</pre>
```

Independent random variables



n random

independent variables has to be generated. This is achieved by using the independent MH sampler, as finding the CDF and using the inverse transform method proves to be difficult for the given density distribution. The main difference between an independent MH sampler and a regular MH random walk is that the independent MH sampler preserves the independence of each random variable.

Although a regular MH random walk would return random variable values following the density in the long run, the variables would loose their independence as the regular MH random walk generates new random values based on the previous random value, making them autocorrelated.

Problem 1B

Generate n independent random variables from the distribution-density p(x).

$$p(x) = 2x \exp(-x^2), \qquad 0 < x < \infty$$

To generate independent random variables the most effective way, inverse transform sampling should be used. This is done by first finding the CDF of p(x):

$$F(x) = \int_0^x p(x)dx = \int_0^x 2x \exp(-x^2)dx = 1 - e^{-x^2}$$

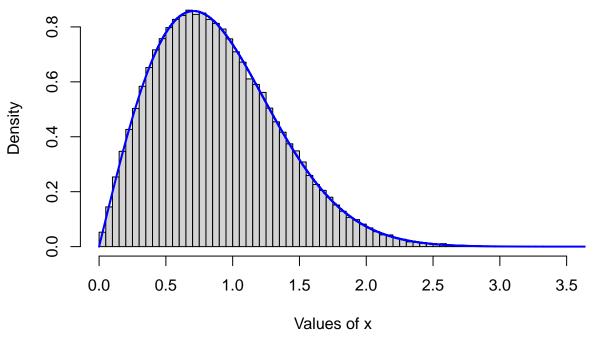
Because $0 \le F(x) \le 1$ and F(x) is uniformly distributed, we set F(X) = U and solve for x:

$$X = F^{-1}(U) = \pm \sqrt{-\log(1-U)}$$

Calculating the square root results in a positive and a negative solution. Because the problem states that $0 \le x \le \infty$ and the square root always gives a positive value, the negative solution is discarded.

$$X = \sqrt{-\log(1 - U)}$$

Independent random variables



Because the x-values are generated directly from p(x) through the inverse transform method, the random variables plotted into a histogram corresponds with the curve of the distribution density.

Problem 1C

Perform importance sampling with the integral:

$$\int_0^\infty \exp(\sqrt{x}) \exp(-20(x-4)^2) dx = \int_A g(x) dx$$

To perform importance sampling, g(x) has to be decomposed into g(x) = h(x)f(x) where f(x) is a probability density on the set A.

$$\exp(\sqrt{x}) \exp(-20(x-4)^2) = \sqrt{2\pi}\sigma \exp(\sqrt{x}) \frac{1}{\sqrt{2\pi}\sigma} \exp\left(-\frac{1}{2} \frac{(x-4)^2}{\frac{1}{40}}\right)$$

Thus resulting in:

$$h(x) = \sqrt{2\pi}\sigma \exp(\sqrt{x})$$

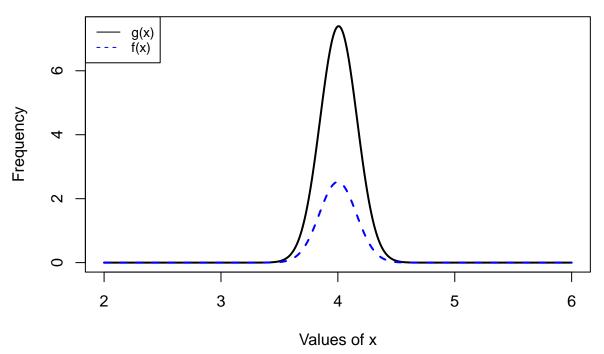
$$f(x) = \frac{1}{\sqrt{2\pi}\sigma} \exp\left(-\frac{1}{2} \frac{(x-4)^2}{\frac{1}{40}}\right)$$

f(x) is therefore normally distributed with $f(x) = N(4, \frac{1}{40})$, which can be used to generate x-values. Further, we have that:

$$\int_{A} g(x)dx = \int_{A} \frac{g(x)}{f(x)} f(x)dx = E\left(\frac{g(X)}{f(X)}\right)$$

```
# The probability density q(x)
g \leftarrow function(x) \{ exp(sqrt(x))*exp(-20*(x-4)^2) \}
Nsim <- 10000
# Generating random variables based on f(x)
x \leftarrow rnorm(Nsim, 4, sqrt(1/40))
# Storing all values larger than O, as specified in the problem.
x \leftarrow x[x > 0]
intiReal <- integrate(g, 0, Inf)$value</pre>
# Solving the integral by importance sampling
intiMC \leftarrow mean(g(x) / dnorm(x, 4, sqrt(1/40)))
c("Acutal integral" = intiReal, "Sampled integral" = intiMC)
##
    Acutal integral Sampled integral
           2.929669
plot(g, xlim = c(2, 6), n= 1000, lwd = 2,
     xlab = "Values of x", ylab = "Frequency",
     main = "PDF of g(x) and f(x)")
curve(dnorm(x, 4, sqrt(1/40)), 2, 6, n = 1000, add = TRUE,
      col = "blue ", lty = 2, lwd = 2)
legend("topleft", legend=c("g(x)", "f(x)"),
       col=c("black", "blue"), lty = c(1, 2) , cex=0.8)
```

PDF of g(x) and f(x)



The plot illustrates that f(x) is proportional to g(x), meaning that f(x) can be used to generate x-values. The result from the definite integral of g(x) and the value from the importance sampling are approximately the same, showing that the importance sampling was done properly.

Problem 2: Smile-shaped target Problem 2A

Use a MCMC algorithm to obtain MCMC samples.

As the problem states, a MCMC algorithm has to be used as direct sampling from the distribution is too complicated to derive.

By trial and error, a MALA algorithm is shown to produce results with a very fluctuating effective sample size. Because of this, a two-dimensional random walk Metropolis Hastings algorithm (RWMH) is assumed to be most suitable for obtaining MCMC samples.

```
lp.log <- function(theta){</pre>
  -(theta[1]^2/2)-(theta[2] - theta[1]^2)^2/2 # log(g(theta))
}
Nsim <- 50000
twoDRWMH <- function(logProb, theta, sigma) {</pre>
  resMatrix <- matrix(0.0, Nsim, 2)</pre>
  resMatrix[1, ] <- theta</pre>
  # store old logProb
  lp.old <- logProb(theta)</pre>
  # accept counter
  acceptCount <- 0
  # main iteration loop
  for(i in 2:Nsim){
    # proposal
    # using rmunorm and a matrix as sigma to operate at both theta values
    thetaStar <- resMatrix[(i-1), ] + rmvnorm(1, theta, diag(sigma, 2, 2))
    # evaluate
    lp.star <- logProb(thetaStar)</pre>
    # accept prob
    alpha \leftarrow exp(min(0.0, lp.star - lp.old))
    # accept/reject
    if(runif(1) < alpha && is.finite(alpha)){</pre>
      resMatrix[i, ] <- thetaStar</pre>
      lp.old <- lp.star</pre>
      acceptCount <- acceptCount + 1</pre>
    } else {
      resMatrix[i, ] <- resMatrix[(i-1), ]</pre>
    }
  }
  print(c("Acceptance rate" = round(acceptCount/(Nsim - 1), 4)))
  return(resMatrix)
twoDRWMH.result <- twoDRWMH(logProb = lp.log , theta = c(0.0,0.0), sigma = 4.6)
```

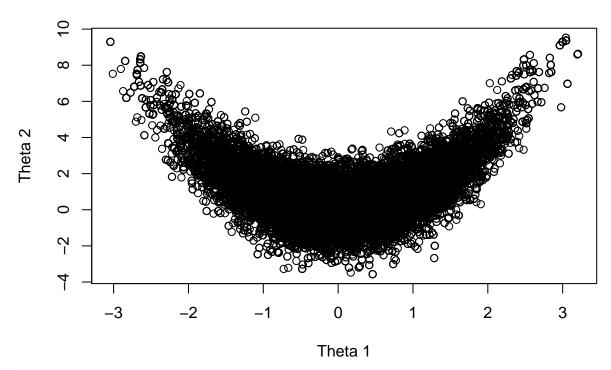
Acceptance rate

```
## 0.2323
```

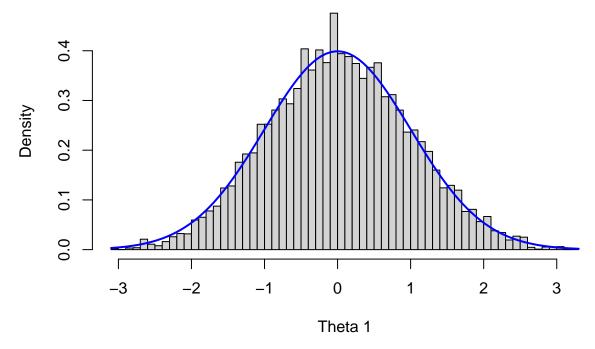
```
twoDRWMH.result.burned <- twoDRWMH.result[501:length(twoDRWMH.result[ ,1]), ]
colnames(twoDRWMH.result.burned) <- c("Theta 1", "Theta 2")

plot(twoDRWMH.result.burned, main = "MCMC samples")</pre>
```

MCMC samples



The marginal distribution of Theta 1



Tuning the algorithm to make it as efficient as possible is done by tweaking the sigma used, which defines how large the jumps are "too large" the chain sticks, and if the jumps are "too small" the chain explores the parameter space very slowly.

According to a paper of Sherlock and Roberts regarding the optimal scaling of the random walk Metropolis, the acceptance rate should be close to 0.234, with the reason being that this acceptance rate minimizes the expected square jump distance, which can be seen as minimizing the autocorrelation.

By trial and error, sigma = 4.6 is used to make the algorithm the most efficient, and the acceptance rate close to 0.234.

The histogram of theta 1 clearly illustrates that the theta1-marginal distribution of is still normally distributed with parameters $\mu = 0, \sigma^2 = 1$.

After the burn-in, the output shows that the effective sample size, both for alpha and beta, are over 1000.

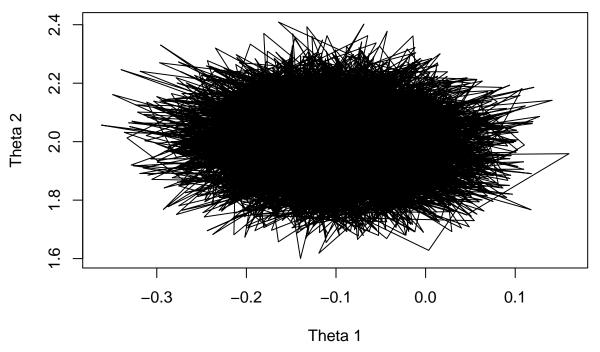
Problem 3: IMH for simple logistic regression problem Problem 3A

Use an independent MH sampler.

```
df <- data.frame(read.table("logistic_regression_data.txt"))</pre>
x \leftarrow df$x
y \leftarrow df$y
logistic.lp <- function(theta){</pre>
    alpha <- theta[1]</pre>
    beta <- theta[2]
    # log-likelihood
    Eeta <- exp(alpha + beta * x)</pre>
    p \leftarrow Eeta/(1.0 + Eeta)
    log.like <- sum(dbinom(y, size = 1, prob = p, log = TRUE))</pre>
    # priors
    log.prior <- dnorm(alpha, sd =10, log=TRUE) + dnorm(beta, sd = 10, log = TRUE)
    # log-posterior kernel
    return(log.like + log.prior)
}
Nsim <- 10000
iMH <- function(target_prob, theta, sigma, tuning) {</pre>
  # Note: Taking exp(log-expression) to work with "normal" values.
  res <- matrix(0, Nsim, 2)</pre>
  # allocate memory
  res[1, ] <- theta
  # old importance weight
  wt.old <- exp(target_prob(res[1, ]))/dmvnorm(res[1, ], mean = theta, sigma = sigma)
  Nacc <- 0
  for(i in 2:Nsim){
    # proposal (note, independent of past)
    thetaStar <- rmvnorm(1, mean = theta, sigma = sigma * tuning)</pre>
    # new importance weight
    wt.star <- exp(target_prob(thetaStar)) /</pre>
      dmvnorm(thetaStar, mean = theta, sigma = sigma)
    # accept probability
    alpha <- min(1.0, wt.star / wt.old)
    # accept/reject
    if(runif(1) < alpha){</pre>
      res[i, ] <- thetaStar</pre>
      wt.old <- wt.star
      Nacc <- Nacc + 1
    } else {
      res[i, ] <- res[i-1, ]
    }
  }
```

```
print(c("Acceptance rate" = round(Nacc/(Nsim - 1), 3)))
  return(res)
}
sigmas \leftarrow matrix(c(0.00653, -0.00058, -0.00058, 0.01689), 2, 2)
thetas <-c(-0.102, 1.993)
# Not necessarry to use burn-in, but a good rule of thumb
iMH.output <- iMH(logistic.lp, theta = thetas, sigma = sigmas, tuning = 0.8)
## Acceptance rate
             0.975
iMH.output.burned <- iMH.output[501:length(iMH.output[ ,1]), ]</pre>
print(
  c(
    "Theta 1" = round(colMeans(iMH.output.burned)[[1]], 3),
    "Theta 2" = round(colMeans(iMH.output.burned)[[2]], 3)
)
## Theta 1 Theta 2
## -0.103
             2.001
plot(iMH.output.burned, type = "l", xlab = "Theta 1", ylab = "Theta 2",
     main = "Independent MH samples")
```

Independent MH samples



```
ess <- ESS(iMH.output.burned)
c("ESS Alpha" = ess[[1]], "ESS Beta" = ess[[2]])</pre>
```

```
## ESS Alpha ESS Beta
## 9091.356 8084.730
```

As the function logistics.lp returns the log-posterior kernel, taking $e^{logistics.lp}$ allows the independent MH sampler to operate with "normal" values instead of log-values.

The mean from the independent MH sampler is approximately the same as the point estimate from the initial maximum likelihood-based classical analysis.

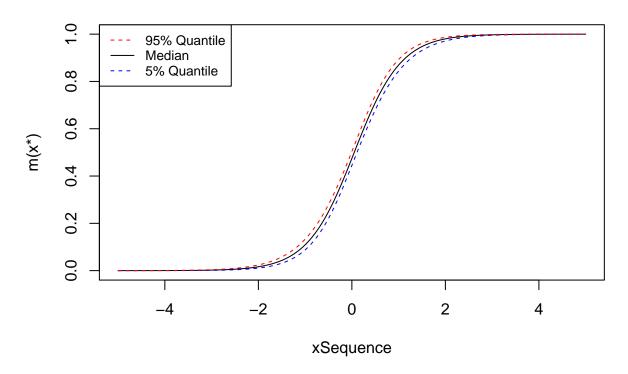
The effective sample size of both alpha and beta are well over 1000.

Problem 3B

Plot the median, 5% quantile, and the 95% quantile.

```
mFunc <- function(x_i) {</pre>
  exp(alpha + beta * x_i) / (1 + exp(alpha + beta * x_i))
stepInfo <- function(x i) {</pre>
  mstar <- exp(iMH.output.burned[ , 1] + iMH.output.burned[ , 2] * x_i) /</pre>
    (1 + exp(iMH.output.burned[ , 1] + iMH.output.burned[ , 2] * x_i))
  med <- median(mstar)</pre>
  # QuantO1 is used in problem 3c)
  quant01 <- quantile(mstar, 0.01)</pre>
  quant05 <- quantile(mstar, 0.05)</pre>
  quant95 <- quantile(mstar, 0.95)</pre>
  return(c(med, quant01, quant05, quant95))
}
xSequence <- seq(-5, 5, length.out = length(iMH.output.burned[ ,1]))</pre>
plotInfo <- matrix(0, length(xSequence), 4)</pre>
i <- 0
for (i in 1:length(xSequence)) {
  plotInfo[i,] <- stepInfo(xSequence[i])</pre>
}
plot(xSequence, plotInfo[, 1], type="l", col="black", ylab = "m(x*)",
     main = "95%, 5% and median of m(x*)")
lines(xSequence, plotInfo[, 3], type="1", col="blue", lty=2)
lines(xSequence, plotInfo[, 4], type="1", col="red", lty=2)
legend("topleft", legend=c("95% Quantile", "Median", "5% Quantile"),
       col=c("red", "black", "blue"), lty=c(2, 1, 2), cex=0.8)
```

95%, 5% and median of m(x*)

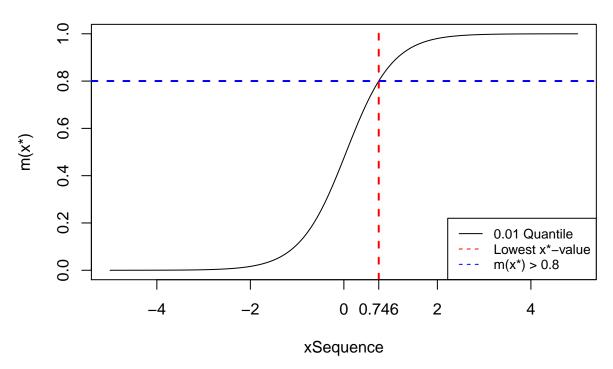


Problem 3C

Find x-values with 99% certainty that $m(x^*) > 0.8$.

```
# First value larger than 0.8. 1% quantile
lowestValue <- plotInfo[, 1][plotInfo[, 1] > 0.8][1]
# First x-value of m-value with 99% probability of being larger than 0.8
firstOccurence <- xSequence[Position(function(x) x > 0.8, plotInfo[, 1])]
firstOccurence.rounded <- round(firstOccurence, 3)</pre>
print(c("Smallest x*-value" = firstOccurence))
## Smallest x*-value
##
            0.745868
plot(xSequence, plotInfo[, 1], type="1", col="black", ylab = "m(x*)",
     main = "1% Quantile of m(x*)")
abline(v = firstOccurence, col="red", lty = 2, lwd = 2)
abline(h = lowestValue, col="blue", lty = 2, lwd = 2)
axis(1, at=firstOccurence.rounded, labels=firstOccurence.rounded)
legend("bottomright", legend=c("0.01 Quantile", "Lowest x*-value", "m(x*) > 0.8"),
       col=c("black", "red", "blue"), lty = c(1, 2, 2), cex=0.8)
```

1% Quantile of m(x*)



As the plot from problem 3b illustrates, m(x*) is a strictly increasing function. This means that if a < b, m(a) < m(b). The 1% quantile provides x-values with 99% certainty that m(x*) > 0.8. This means that there is a lower x*, and all x*-values above satisfy the condition m(x*) > 0.8. This means that for any x*-values larger than or equal to 0.746, the condition m(x*) > 0.8 with a certainty of 99% is fulfilled.

Problem 4: Gibbs sampler for simple linear regression model Problem 4A

Find the distribution of $\alpha | \beta, \tau, y$ and $\beta | \alpha, \tau, y$

Given that the joint posterior of $\theta = (\alpha, \beta, \tau)$ has the log-density kernel:

$$\log g(\theta) = \frac{n}{2} \log (\tau) - \frac{\tau}{2} \sum_{i=1}^{n} (y_i - \alpha - \beta x_i)^2 - \frac{\alpha^2 + \beta^2}{200} - \tau$$

Finding the conditional α and β distributions is/are done by collecting all parts of the equation not containing the parameter in focus into a constant and (in this case) writing the expressions in the form $a + bx + cx^2 \Rightarrow x \sim N\left(-\frac{b}{2c}, -\frac{1}{2c}\right)$.

By first expanding the squared sum and factorizing each part of the expression, the expression becomes:

For α :

$$\alpha | \beta, \tau, \mathbf{y} = \alpha \sum_{i=1}^{n} (y_i \tau) + \alpha^2 \sum_{i=1}^{n} (-\frac{\tau}{2}) + \alpha \sum_{i=1}^{n} (-\beta x_i \tau) + \alpha^2 (-\frac{1}{200}) + C$$

$$\alpha | \beta, \tau, \mathbf{y} = C + \alpha (\tau \sum_{i=1}^{n} (y_i - \beta x_i)) + \alpha^2 (-\frac{\tau n}{2} - \frac{1}{200})$$

$$\alpha | \beta, \tau, \mathbf{y} \sim N \left(\frac{\tau \sum_{i=1}^{n} (y_i - \beta x_i)}{\tau n + 1/100}, \frac{1}{\tau n + 1/100} \right)$$

For β :

$$\beta | \alpha, \tau, \mathbf{y} = \beta \sum_{i=1}^{n} (y_i x_i \tau) + \beta \sum_{i=1}^{n} (-\alpha x_i \tau) + \beta^2 \sum_{i=1}^{n} (-\frac{\tau}{2} x_i^2) + \beta^2 (-\frac{1}{200}) + C$$

$$\beta | \alpha, \tau, \mathbf{y} = C + \beta (\tau \sum_{i=1}^{n} (y_i x_i - \alpha x_i)) + \beta^2 (-\frac{\tau}{2} \sum_{i=1}^{n} (x_i^2) - \frac{1}{200})$$

$$\beta | \alpha, \tau, \mathbf{y} \sim N \left(\frac{\tau \sum_{i=1}^{n} (y_i x_i - \alpha x_i)}{\tau \sum_{i=1}^{n} (x_i^2) + \frac{1}{100}}, \frac{1}{\tau \sum_{i=1}^{n} (x_i^2) + \frac{1}{100}} \right)$$

Also, $\tau | \alpha, \beta, \boldsymbol{y}$ is given as:

$$\tau | \alpha, \beta, \boldsymbol{y} \sim Gamma\left(\frac{n}{2} + 1, \frac{1}{\frac{1}{2} \sum_{i=1}^{n} (y_i - \alpha - \beta x_i)^2 + 1}\right)$$

Problem 4B + 4C

Gibbs sampler with 3 blocks targeting $p(\theta|y)$.

```
df <- data.frame(read.table(file="linear_regression_data.txt"))</pre>
x \leftarrow df$x
y <- df$y
gibbs3b <- function() {</pre>
  n <- 10000 # number of Gibbs iterations
  theta <- c(1, 1, 1) # variable used as state throughout
  res <- matrix(0.0, 3*n + 1, 3) # results, note stores twice per iteration
  res[1, ] <- theta
  k <- 2 # store counter
  for(i in 1:n){
      # update first block (alpha)
      theta[1] \leftarrow rnorm(1, theta[3]*sum(y-theta[2]*x) / (theta[3]*length(y) + 1/100),
                         1 / (theta[3]*length(y) + 1/100))
      # store state
      res[k, ] <- theta
      k < - k+1
      # update second block (beta)
      theta[2] <- rnorm(1, theta[3]*sum(x*y-x*theta[1]) / (theta[3]*sum(x^2) + 1/100),
                         1 / (theta[3]*sum(x^2) + 1/100)
      # store state
      res[k, ] <- theta
      k < - k+1
       # update third block (tau)
      theta[3] <- rgamma(1, shape = length(y)/2 + 1,
                          scale = 1/(1/2 * sum((y-theta[1]-theta[2]*x)^2) + 1))
      # store state
      res[k, ] <- theta
      k < - k+1
  }
  res
}
```

Problem 4C

Use the Gibbs sampler with 3 blocks.

```
gibbs3 <- gibbs3b()

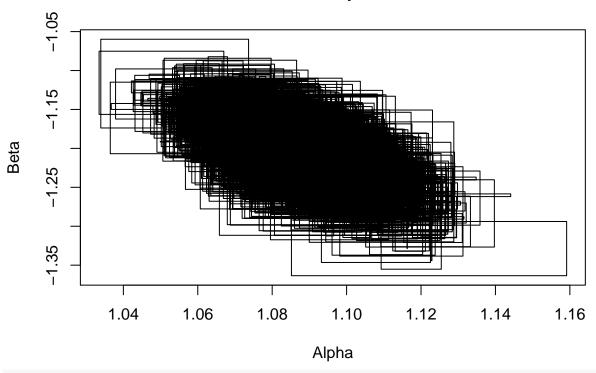
gibbs3.burned <- gibbs3[501:length(gibbs3[ ,1]), ]
colnames(gibbs3.burned) <- c("Alpha", "Beta", "Tau")

round(colMeans(gibbs3.burned), 4)

## Alpha Beta Tau
## 1.0884 -1.2067 1.0992

plot(gibbs3.burned[,1], gibbs3.burned[,2], type="l",
    main = "Correlation of alpha and beta",
    xlab = "Alpha", ylab = "Beta")</pre>
```

Correlation of alpha and beta



```
ess <- ESS(gibbs3.burned)
c("ESS Alpha" = ess[[1]], "ESS Beta" = ess[[2]], "ESS Tau" = ess[[3]])

## ESS Alpha ESS Beta ESS Tau
## 5234.634 5438.059 9886.550

# Classical linear regression
lm.out <- lm(y~x,data=df)
c("Alpha" = lm.out$coefficients[[1]], "Beta" = lm.out$coefficients[[2]])</pre>
```

Alpha Beta ## 1.08871 -1.20703

After burn-in, the effective sample size of all α , β and τ are above the required size of 1000.

It can be seen from the plots that α and β has a correlation of -0.5047241, while the other plots show a correlation close to zero. Thus, α and β are dependent on each other, and the Gibbs sampler would be improved by combining α and β , and reducing the number of blocks from three to two.

The last output can be used to compare the posterior means with a classical linear regression, which gives approximately the same α - and β -values.

Problem 4D

Gibbs sampler with 2 blocks.

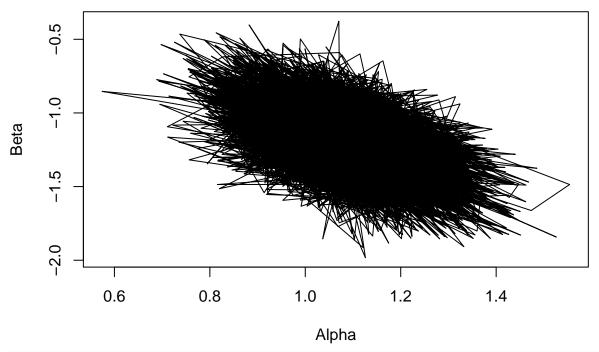
From the text, we have that $(\alpha, \beta)|\tau, y \sim N(-c^{-1}b, -c^{-1})$, where:

$$\boldsymbol{c} = \begin{pmatrix} -n\tau & 0.01 \\ -\tau \sum_{i=1}^{n} x_i & -\tau \sum_{i=1}^{n} x_i^2 - 0.01 \end{pmatrix}$$
$$\boldsymbol{b} = \begin{pmatrix} \tau \sum_{i=1}^{n} y_i \\ \tau \sum_{i=1}^{n} y_i x_i \end{pmatrix}$$

Using $(\alpha, \beta)|\tau, y$ and $\tau|\alpha, \beta, y$ gives the following Gibbs sampler with two blocks:

```
gibbs2b <- function() {</pre>
  n <- 10000 # number of Gibbs iterations
 theta <- c(1, -1.2, 1.0) # variable used as state throughout
 res <- matrix(0.0, 2*n + 1, 3) # results, note stores twice per iteration
 res[1, ] <- theta
  k <- 2 # store counter
  # Calculating some constants for performance:
  sumx \leftarrow sum(x)
  # Sum of x^2
  sumx2 <- sum(x^2)
  # Sum of y
  sumy <- sum(y)</pre>
  # Sum of xy
  sumyx <- sum(y*x)</pre>
  # Length of both y (or x)
  dataLength <- length(y)</pre>
  for(i in 1:n){
    cVal = matrix(c(-dataLength*theta[3]-0.01, -theta[3]*sumx,
                     -theta[3]*sumx, -theta[3]*sumx2-0.01), 2, 2)
    bVal = c(theta[3]*sumy, theta[3]*sumyx)
    # update first block
    theta[1:2] <- rmvnorm(1, -solve(cVal) %*% bVal, -solve(cVal))
    # store state
    res[k,] <- theta
    k \leftarrow k+1
    # update the second block
    theta[3] <- rgamma(1, shape = dataLength/2 + 1,
                        scale = 1/(1/2 * sum((y-theta[1] - theta[2]*x)^2) + 1))
    #store state
    res[k,] <- theta
    k < - k+1
 }
 res
}
```

Correlation of alpha and beta



```
ess <- ESS(gibbs2.burned)
c("ESS Alpha" = ess[[1]], "ESS Beta" = ess[[2]], "ESS Tau" = ess[[3]])</pre>
```

```
## ESS Alpha ESS Beta ESS Tau
## 9264.174 9858.245 8135.674
```

The new effective sample sizes clearly shows that fewer simulations are needed to get the same ESS as in problem 3C, meaning that the performance of the Gibbs sampling procedure has been improved.

Having fewer blocks and improving independence between the blocks gives better samples, which is clearly showed in the solution of the problem.

As in problem 3C, the mean of α , β and τ are still the same, meaning that the sampling still works as intended. In addition, the plot shows that the dependence of α and β are still intact.

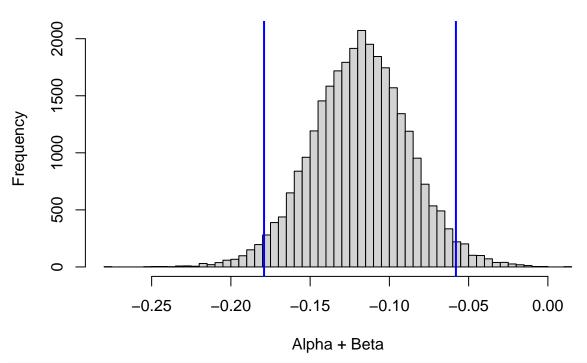
Problem 4E

Verify that $\alpha + \beta = 0$.

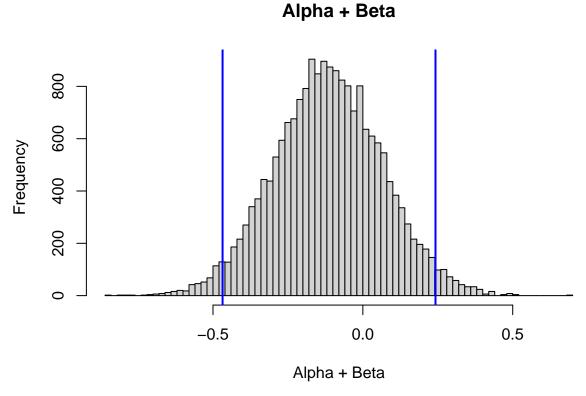
This can be seen as a hypothesis test, using the 15- and 75-percentile on both Gibbs samplers.

```
alpha_beta3 <- gibbs3.burned[, 1] + gibbs3.burned[, 2]
hist(alpha_beta3, breaks = 60, main = "Alpha + Beta", xlab = "Alpha + Beta")
abline(v = quantile(alpha_beta3, 0.975), lwd="2", col = "blue")
abline(v = quantile(alpha_beta3, 1-0.975), lwd="2", col = "blue")</pre>
```

Alpha + Beta



```
alpha_beta2 <- gibbs2.burned[, 1] + gibbs2.burned[, 2]
hist(alpha_beta2, breaks = 60, main = "Alpha + Beta", xlab = "Alpha + Beta")
abline(v = quantile(alpha_beta2, 0.975), lwd="2", col = "blue")
abline(v = quantile(alpha_beta2, 1-0.975), lwd="2", col = "blue")</pre>
```



From the Gibbs sampler with three blocks shows that $\alpha + \beta$ equals zero falls outside the quantile limits, and the hypothesis can be rejected.

On the other hand, the Gibbs sampler with two blocks shows that $\alpha + \beta$ equals zero falls inside the quantile limits. This means that $\alpha + \beta = 0$ might hold true. Thus, the hypothesis can not be rejected in this case.

Problem 5: Bootstrapping Problem 5A

Check if the claims given in the problem holds true.

```
Claim 1: \sigma = \frac{1}{\sqrt{\tau}} is strictly smaller than 1.0.
Claim 2: \alpha is equal to 0.
Claim 3: \alpha + \beta is equal to 0.
```

```
df <- data.frame(read.table(file = "linear_regression_data.txt"))
x <- df$x
y <- df$y

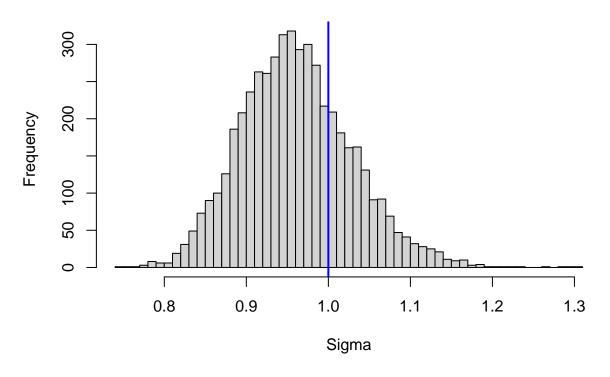
library(boot)
sdestfunc <- function(data, i) {
   lm(data[i, 1] ~ data[i, 2], data = df)$coefficients
}

boot.obj <- boot(data = df, statistic = sdestfunc, R = 5000)

sigma <- 1/sqrt(
   rgamma(5000, shape = length(y)/2 + 1,
   scale = 1/(1/2 * sum((y - mean(boot.obj$t[,1]) - mean(boot.obj$t[,2])*x)^2) + 1))

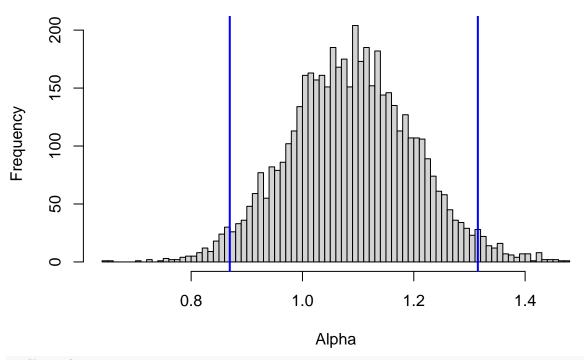
# Claim 1
hist(sigma, breaks = 60, xlab = "Sigma")
abline(v = 1, lwd = 2, col = "blue")</pre>
```

Histogram of sigma

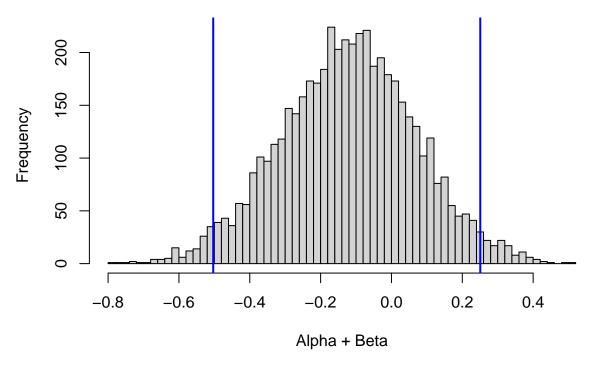


```
# Claim 2
hist(boot.obj$t[,1], breaks = 60, main = "Histogram of alpha", xlab = "Alpha")
abline(v = quantile(boot.obj$t[,1], 0.975), lwd = 2, col = "blue")
abline(v = quantile(boot.obj$t[,1], 1 - 0.975), lwd = 2, col = "blue")
```

Histogram of alpha



Histogram of alpha + beta



The first histogram clearly shows that σ is not strictly smaller than 1.0. Because of this, claim 1 is rejected. Claim 2 is also rejected as the second histogram shows that $\alpha=0$ falls outside the 95% confidence interval. As $\alpha+\beta=0$ falls inside the 95% confidence interval, claim 3 can not be rejected.

Problem 6: Bayesian inference for the SEIR model Problem 6A

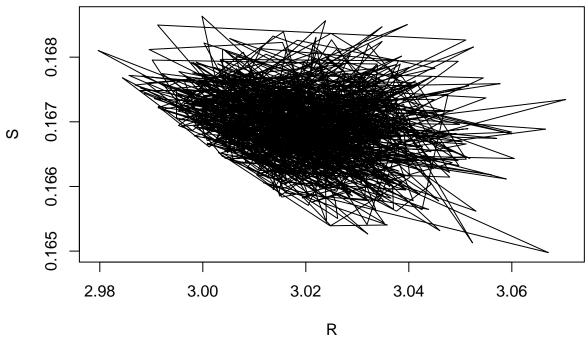
Obtain MCMC samples targeting $p(\theta|\mathbf{y})$

Using a bivariate RWMH does not yield effective sample sizes of R and S of 200 or more. To fix this, an independent RWMH is implemented. Because the independent random walk returns independent values of R and S, the effective sample size will increase.

```
source("seir mcmc funs.R")
Nsim <- 1500
# Independent sampler
ind.2DRW <- function(l_target_prob, theta, sigma, tuning){</pre>
  # allocate memory
 res <- matrix(0, Nsim, 2)
 res[1, ] <- theta
  # old importance weight
  wt.old <- 1_target_prob(theta) - dmvnorm(theta, theta, sigma, log = TRUE)
  Nacc <- 0
  for (i in 2:Nsim){
    # proposal (note, independent of past)
    thetaStar <- rmvnorm(1, theta, tuning*sigma)</pre>
    # new importance weight
    wt.star <- l_target_prob(thetaStar) - dmvnorm(thetaStar, theta, sigma, log = TRUE)
    # accept probability
    alpha <- exp(min(0.0, wt.star - wt.old))</pre>
    # accept/reject
    if(runif(1) < alpha && is.finite(alpha)){</pre>
      res[i, ] <- thetaStar</pre>
      wt.old <- wt.star
      Nacc <- Nacc + 1
    } else {
      res[i,] <- res[(i-1),]
    }
  print(paste0("Accept rate : ",Nacc/Nsim))
  return(res)
}
ind.result <- ind.2DRW(seir.lp,</pre>
             theta=c(3.019, 0.167),
             sigma=matrix(c(0.0003,0,0,0.0000005),2,2),
             tuning=0.8)
```

```
## [1] "Accept rate : 0.58333333333333333333"
plot(ind.result[,1], ind.result[,2], type="1", main="MCMC samples", xlab="R", ylab="S")
```

MCMC samples



```
ess <- ESS(ind.result)
c("ESS R" = ess[[1]], "ESS S" = ess[[2]])</pre>
```

ESS R ESS S ## 363.7848 259.8973

The effective sample size for R and S are both over the required 200.

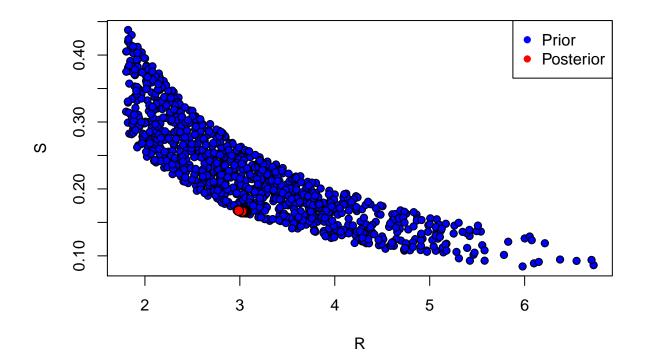
The plot shows that the sampler explores the distribution quite well, avoiding the chain to stick, while also avoiding exploring the parameter space too slowly.

Problem 6B

Compare the posterior distribution and the prior distribution.

```
Nsim <- 1000
# inverse transform method (from mandatory 2)
randNum <- function(1, mu, sigma) {</pre>
    qnorm(
      pnorm(1, mu, sigma) + runif(Nsim, 0, 1)*(1 - pnorm(1, mu, sigma)),
          mu, sigma
}
# generate random R- and S-variables (from mandatory 2)
prior <- function() {</pre>
  rNum <- randNum(1 = 1.8, mu = 2.2, sigma = 1.5)
  sNum <- runif(Nsim, 0.5/rNum, 0.8/rNum)</pre>
  cbind(rNum, sNum)
}
prior.values <- prior()</pre>
plot(prior.values[ ,1], prior.values[ ,2],
     xlab = "R", ylab = "S",
     pch = 21, bg = "blue",
     main = "Prior and Posterior distribution")
points(ind.result[ ,1], ind.result[ ,2], pch = 21, bg = "red", cex = 1.3)
legend("topright", legend = c("Prior", "Posterior"),
       pch = c(16, 16), col = c("blue", "red"))
```

Prior and Posterior distribution



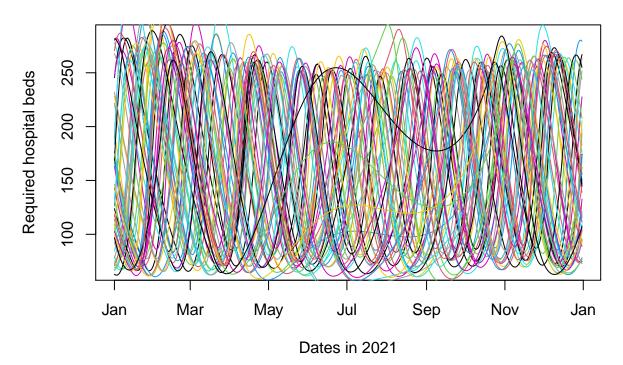
The hospital data provides a lot of information of the R- and S-values, which can clearly be seen by comparing the posterior distribution with the prior distribution through the plot.

Problem 6C

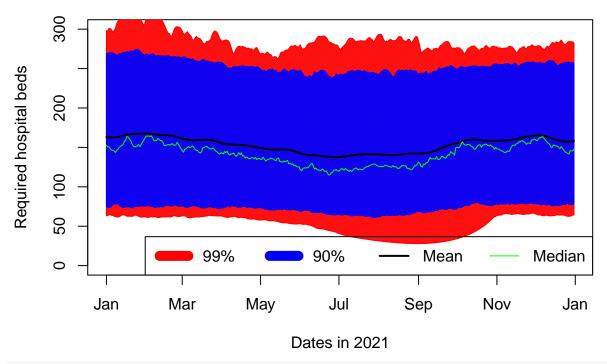
Re-do Problem 4F) of mandatory assignment 2.

```
Nsim <- 500
source("seir model.R")
uncertainty <- function(dataInfo) {</pre>
  # run simulations
 M <- numeric(Nsim)</pre>
  H <- matrix(0.0,Nsim,365) # prepare for next point
  for(i in 1:Nsim){
    \#R[i] \rightarrow dataInfo[i, 1], S[i] \rightarrow dataInfo[i, 2]
    ff <- seir_sim(ROmax = dataInfo[i, 1],soc_dist_Rfac = dataInfo[i, 2])</pre>
    t1 <- which(ff$dates=="2021-01-01")
    t2 <- which(ff$dates=="2021-12-31")
    H[i,] <- rowSums(1000000*ff$ode[t1:t2,c("HH","HC","CC")])
    M[i] <- max(H[i,])
  # make a data frame to facilitate plotting with dates
  df <- data.frame(t(H),dates=ff$dates[t1:t2])</pre>
  plot1Name <- ifelse(plotNum == 1,</pre>
                        "Prior example trajetories",
                        "Posterior example trajetories")
  # in the upper plot, simply show some of the trajectories:
  plot(df$dates,df$X1,type="l",
       xlab="Dates in 2021",
       ylab="Required hospital beds",
       main = plot1Name)
  for(i in 2:50) lines(df$dates,df[,i],col=i)
  # compute daily statistics for plotting
  daily.stats <- matrix(0.0,365,6)</pre>
  for(t in 1:365){
    daily.stats[t,1] <- mean(H[,t])</pre>
    daily.stats[t,2] <- median(H[,t])</pre>
    daily.stats[t,3:6] \leftarrow quantile(H[,t],probs=c(0.005,0.1,0.9,0.995))
  }
  df2 <- data.frame(daily.stats,ff$dates[t1:t2])</pre>
  colnames(df2) <- c("mean", "median", "q05", "q10", "q90", "q995", "dates")</pre>
  plot2Name <- ifelse(plotNum == 1,</pre>
                        "Prior distribution representation",
                        "Posterior distribution representation")
  plot(df2$dates, df2$mean, ylim=c(0,300), col=0,
       xlab="Dates in 2021",
       ylab="Required hospital beds",
       main=plot2Name)
  polygon(c(df2$dates,rev(df2$dates)),c(df2$q05,rev(df2$q995)),
```

Prior example trajetories

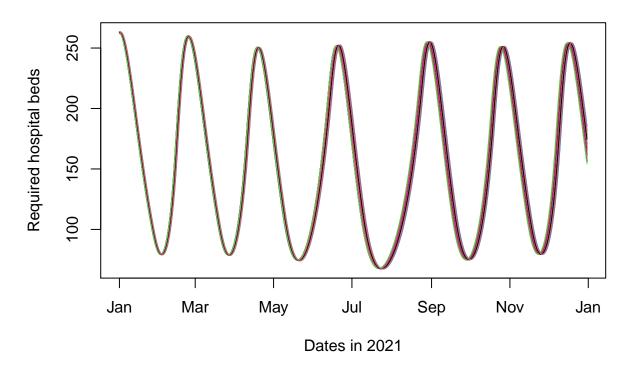


Prior distribution representation

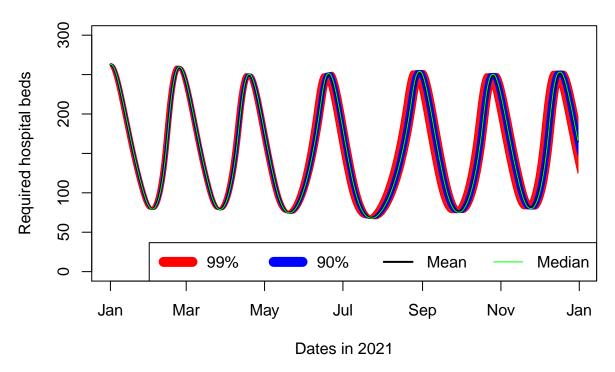


plotNum <- 2
uncertainty(ind.result)</pre>

Posterior example trajetories



Posterior distribution representation



The plots correspond to the data received from problem 6B. When observing the data, the variance of the R-and S-values decreases, which means that the posterior distribution will represent the required hospital beds more accurate than the prior distribution by itself.

The 90% confidence interval of the prior distribution ranges between 80 and 260 beds almost all the time, which from the prior example trajectories appears to be correct as the R- and S-values have a wide variety of trajectories to "choose from".

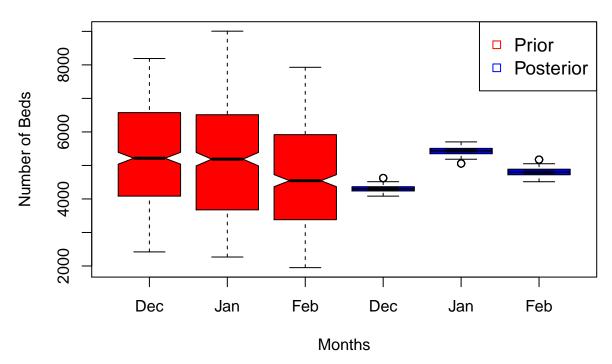
The 90% confidence interval of the posterior distribution has a width of around 10 required hospital beds, and follows the mean closely. Because all of the trajectories of the posterior distribution are close to equal, the result makes sense.

Problem 6D

Distribution of the total number of "hospital-bed-days" for December 2020, January 2021 and February 2021.

```
Nsim <- 500
hospBeds <- function(dataInfo) {</pre>
  december <- matrix(0.0,Nsim,31)</pre>
  january <- matrix(0.0,Nsim,31)</pre>
  february <- matrix(0.0,Nsim,28)</pre>
  for(i in 1:Nsim){
    f <- seir_sim(ROmax = dataInfo[i, 1], soc_dist_Rfac = dataInfo[i, 2])</pre>
    days <- which(grep1("2020-12", f$dates))</pre>
    december[i, ] <- rowSums(1000000*f$ode[days, c("HH","HC","CC")])</pre>
    days <- which(grep1("2021-01", f$dates))</pre>
    january[i, ] <- rowSums(1000000*f$ode[days, c("HH","HC","CC")])</pre>
    days <- which(grep1("2021-02", f$dates))</pre>
    february[i, ] <- rowSums(1000000*f$ode[days, c("HH","HC","CC")])
  }
  cbind("Dec" = rowSums(december), "Jan" = rowSums(january), "Feb" = rowSums(february))
}
priorList <- hospBeds(prior.values)</pre>
posteriorList <- hospBeds(ind.result)</pre>
boxplot(cbind(priorList, posteriorList),
        col = c("red", "red", "red", "blue", "blue", "blue"),
        notch = TRUE, varwidth = TRUE,
        xlab = "Months", ylab = "Number of Beds",
        main = "Hospital daybeds")
legend("topright", legend = c("Prior", "Posterior"), col = c("red", "blue"),
       pch = 22, cex = 1.2
```

Hospital daybeds



The plot explains that without the given dataset of required beds from December, January and February, the uncertainty of how many beds are required increases. In this case, the number of required hospital beds ranges between 4000 and 7000 per month.

By taking the observed data from 21th March to 18th July into consideration, the uncertainty drastically decreases. This means that if the data is representative for the given situation, the estimation of required hospital beds is much more exact. With a high confidence, it can be seen that the required number of hospital beds for December, January and February is 4300, 5500 and 4800, respectively, with a standard deviation of around 90 hospital beds.