Classifying_Mushrooms

March 26, 2022

This coursework aims to implement the universal workflow from Francois Chollet's Deep Learning with Python and apply it to classify mushrooms as either edible or not edible.

We are using the Mushroom Dataset from UCI's Machine Learning Repository to implement our model.

The overall workflow will reflect the literature.

The Universial Machine Learning Workflow

- 1. Define the problem and assemble a dataset
- 2. Choose a measure of success
- 3. Decide on an evaluation protocol
- 4. Prepare the data
- 5. Devlop a model that does better than a baseline
- 6. Devlop a model that overfits 7. Regularize the model and tune its hyperparameters

1 Define the problem and assemble a dataset

Our hypothetical problem is the classification of mushrooms as edible or non-edible, with the purpose of selecting mushrooms for use in human food production. Since mushrooms can be poisonous, the goal is to prevent humans from getting sick by eating poisonous mushrooms.

Let's frame the problem in more detail. Our input data is the aforementioned dataset, which is a multivariate set of data about mushrooms and 22 features about the sample mushrooms. Based on these features, we are trying to predict if the mushrooms are suitable for human consumption.

The task at hand is binary classification - we determine if a mushroom edible or not.

Using the data information file, we can load up a list of features for our data.

Feature Information:

- classes:
 - edible=e, poisonous=p
- cap-shape:
 - bell=b ,conical=c ,convex=x, flat=f, knobbed=k, sunken=s
- cap-surface:
 - fibrous=f, grooves=g, scaly=y, smooth=s
- cap-color:
 - brown=n, buff=b, cinnamon=c, gray=g, green=r, pink=p, purple=u, red=e, white=w, yellow=y

- bruises?:
 - bruises=t, no=f
- odor:
 - almond=a, anise=l, creosote=c, fishy=y, foul=f, musty=m, none=n, pungent=p, spicy=s
- gill-attachment:
 - attached=a, descending=d, free=f, notched=n
- gill-spacing:
 - close=c, crowded=w, distant=d
- gill-size:
 - broad=b, narrow=n
- gill-color:
 - black=k, brown=n, buff=b, chocolate=h, gray=g, green=r, orange=o, pink=p, purple=u, red=e, white=w, yellow=y
- stalk-shape:
 - enlarging=e, tapering=t
- stalk-root:
 - bulbous=b, club=c, cup=u, equal=e, rhizomorphs=z, rooted=r, missing=?
- stalk-surface-above-ring:
 - fibrous=f, scaly=y, silky=k, smooth=s
- stalk-surface-below-ring:
 - fibrous=f, scaly=y, silky=k, smooth=s
- stalk-color-above-ring:
 - brown=n, buff=b, cinnamon=c, gray=g, orange=o, pink=p, red=e, white=w, yellow=y
- stalk-color-below-ring:
 - brown=n, buff=b, cinnamon=c, gray=g, orange=o, pink=p, red=e, white=w, yellow=y
- veil-type:
 - partial=p, universal=u
- veil-color:
 - brown=n, orange=o, white=w, yellow=y
- ring-number:
 - none=n, one=o, two=t
- ring-type:
 - cobwebby=c, evanescent=e, flaring=f, large=l, none=n, pendant=p, sheathing=s, zone=z
- spore-print-color:
 - black=k, brown=n, buff=b, chocolate=h, green=r, orange=o, purple=u, white=w, yellow=y
- population:
 - abundant=a, clustered=c, numerous=n, scattered=s, several=v, solitary=y
- habitat:
 - grasses=g, leaves=l, meadows=m, paths=p, urban=u, waste=w, woods=d

From these features we can create a list of column names for our dataset

```
'cap-color',
'bruises?',
'odor',
'gill-attachment',
'gill-spacing',
'gill-size',
'gill-color',
'stalk-shape',
'stalk-root',
'stalk-surface-above-ring',
'stalk-surface-below-ring',
'stalk-color-above-ring',
'stalk-color-below-ring',
'veil-type',
'veil-color',
'ring-number',
'ring-type',
'spore-print-color',
'population',
'habitat']
```

Lets import our dataset and create a Pandas DataFrame from the .data file

```
[]: import pandas as pd
     url = 'https://archive.ics.uci.edu/ml/machine-learning-databases/mushroom/
      ⇔agaricus-lepiota.data'
     mushrooms = pd.read_csv(url, header=None, names=column_names)
     mushrooms.head()
[]:
       class cap-shape cap-surface cap-color bruises? odor gill-attachment \
           р
                    X
                                 s
                                           n
                                                    t
                                                         р
                                                                          f
     1
           е
                                                    t
                     X
                                           У
                                                         a
     2
                     b
                                                         1
                                                                          f
                                           W
                                                    t
                                 S
                                                                          f
     3
          р
                     X
                                 У
                                           W
                                                    t
                                                         р
     4
                                                    f
                                                                          f
                     X
                                           g
                                                         n
```

```
gill-spacing gill-size gill-color ... stalk-surface-below-ring \
0
             С
                        n
                                    k ...
                                    k ...
1
             С
                        b
                                                                   s
2
             С
                        b
                                    n ...
                                                                   s
3
              С
                        n
                                    n ...
                                                                   s
                                    k ...
```

stalk-color-above-ring stalk-color-below-ring veil-type veil-color \

\wedge			_		~	
U		V	V	W	Р	W
1		V	Į.	W	p	W
2		V	ī	W	p	W
3		V	ī	W	р	W
4		V	7	W	p	W
	ring-number	ring-type	${\tt spore-print-color}$	population	habitat	
0	0	р	k	S	11	
		Р		ъ	u	
1	0	p p	n	n		
1 2	0	p p		_	g	
1 2 3	· ·	p p p	n	n	g	
1 2 3 4	0	p p p	n n	n n	g m u	

[5 rows x 23 columns]

2 Choose a measure of success

Since our problem involves the prevention of human sickness or death, we will use precision as a measure of success. Precision is the ability of the classifier not to label mushrooms which are poisonous as edible. This places a preference for correct predictions of poisonous mushrooms, as we prefer if edible mushrooms are misidentified as poisonous rather than poisonous mushrooms being misidentified as edible.

```
Precision = \frac{TruePositives}{TruePositives + FalsePositive}
[]: from sklearn.metrics import precision_score
```

3 Decide on an evaluation protocol

I'll apply 10-fold cross validation to evaluate our model. While a simple holdout validation set might be enough, our dataset of 8.000 samples might not be enough.

```
[]: from sklearn.model_selection import train_test_split, RandomizedSearchCV
```

First lets split our data into a feature matrix (X), and a target vector(y). We will use OneHotEncoder to encode our catagorical variables.

```
[]: import category_encoders as ce

X = mushrooms.drop(columns='class')
X = ce.OneHotEncoder(use_cat_names=True).fit_transform(X)
y = mushrooms['class'].replace({'p':0, 'e':1})

print('Feature matrix size:',X.shape)
print('Target vector size:',len(y))
```

Feature matrix size: (8124, 117) Target vector size: 8124

Next we will split our data into a training set and a test set.

4 Prepare the data

Before we begin training our model, we shall explore the data and note some important characteristics, and potentially fix any issues.

We could use .dtypes(), .columns, and .shape to examine our dataset, but Pandas provides a .info function that will allow us to view all this information in one place.

```
[]: print(mushrooms.info())
```

<class 'pandas.core.frame.DataFrame'>
RangeIndex: 8124 entries, 0 to 8123
Data columns (total 23 columns):

	• • • • • • • • • • • • • • • • • • • •		
#	Column	Non-Null Count	Dtype
0	class	8124 non-null	object
1	cap-shape	8124 non-null	object
2	cap-surface	8124 non-null	object
3	cap-color	8124 non-null	object
4	bruises?	8124 non-null	object
5	odor	8124 non-null	object
6	gill-attachment	8124 non-null	object
7	gill-spacing	8124 non-null	object
8	gill-size	8124 non-null	object
9	gill-color	8124 non-null	object
10	stalk-shape	8124 non-null	object
11	stalk-root	8124 non-null	object

```
12 stalk-surface-above-ring 8124 non-null
                                              object
   stalk-surface-below-ring
                              8124 non-null
                                              object
13
14
   stalk-color-above-ring
                              8124 non-null
                                              object
   stalk-color-below-ring
                              8124 non-null
                                              object
15
   veil-type
                                              object
16
                              8124 non-null
17
   veil-color
                              8124 non-null
                                              object
   ring-number
                              8124 non-null
                                              object
   ring-type
19
                              8124 non-null
                                              object
20
   spore-print-color
                              8124 non-null
                                              object
   population
                                              object
21
                              8124 non-null
22 habitat
                              8124 non-null
                                              object
```

dtypes: object(23) memory usage: 1.4+ MB

None

Another useful step is to check is the number of null values and where they are in the DataFrame.

[]: print(mushrooms.isna().sum())

class	0
cap-shape	0
cap-surface	0
cap-color	0
bruises?	0
odor	0
gill-attachment	0
gill-spacing	0
gill-size	0
gill-color	0
stalk-shape	0
stalk-root	0
stalk-surface-above-ring	0
stalk-surface-below-ring	0
stalk-color-above-ring	0
stalk-color-below-ring	0
veil-type	0
veil-color	0
ring-number	0
ring-type	0
spore-print-color	0
population	0
habitat	0
dtype: int64	

It is unlikely that this dataset actually has no null values at all. We should check if there are placeholder values, often denoted with question marks.

```
[]: import numpy as np
     mushrooms = mushrooms.replace({'?':np.NaN})
```

```
[]: print(mushrooms.isna().sum())
                                     0
    class
                                     0
    cap-shape
    cap-surface
                                     0
    cap-color
                                     0
    bruises?
                                     0
    odor
                                     0
    gill-attachment
                                     0
    gill-spacing
                                     0
    gill-size
                                     0
    gill-color
                                     0
    stalk-shape
                                     0
    stalk-root
                                  2480
    stalk-surface-above-ring
                                     0
    stalk-surface-below-ring
                                     0
    stalk-color-above-ring
                                     0
    stalk-color-below-ring
                                     0
    veil-type
    veil-color
    ring-number
    ring-type
                                     0
    spore-print-color
                                     0
    population
                                     0
    habitat
                                     0
    dtype: int64
```

As we can see, stalk_root has 2480 blank features. We can replace these with n for none.

```
[]: mushrooms['stalk-root'] = mushrooms['stalk-root'].replace(np.NaN,'n')

[]: print(mushrooms['stalk-root'].value_counts())

b     3776
     n     2480
     e     1120
     c     556
     r     192
Name: stalk-root, dtype: int64
```

5 Devlop a model that does better than a baseline

5.1 Baseline Model

Using the most common label from our dataset we will create a baseline model that we hope to beat. First lets look at how class is distributed.

```
[]: mushrooms['class'].value_counts(normalize=True)
```

[]: e 0.517971 p 0.482029 Name: class, dtype: float64

We will use the mode of the class attribute to create our baseline prediction.

```
[]: majority_class = y_train.mode()[0]
baseline_predictions = [majority_class] * len(y_train)
```

Let's see how accurate our baseline model is.

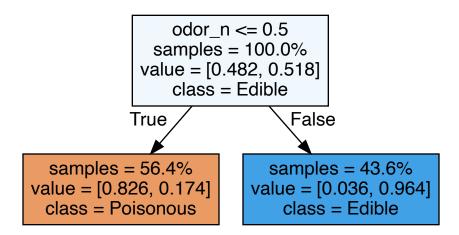
[]: 0.5179258347438067

The accuracy is around 52%, which is expected given the nature of our dataset.

5.2 Decision Tree

We will attempt to fit a decision stump to our training data and produce an accuracy score greater than 52%.

8



Let's look at the prediction probability distribution for our model. In simple terms this represents how sure the model is about its classification label. sklearn provides us with a simple way to see many of the ralevant scores for classification models with classification_report. We can also generate a confusion matrix using sklearn's confusion_matrix.

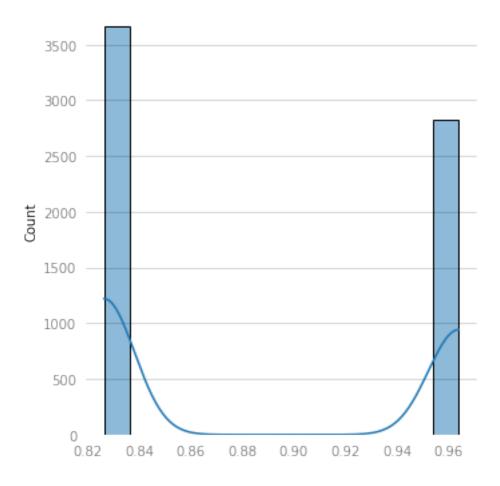
```
[]: import matplotlib.pyplot as plt
     import seaborn as sns
     from sklearn.metrics import classification_report, confusion_matrix
     def model_analysis(model, train_X, train_y, model_name):
       model_probabilities = model.predict_proba(train_X)
       Model_Prediction_Probability = []
       for _ in range(len(train_X)):
         x = max(model_probabilities[_])
         Model_Prediction_Probability.append(x)
      plt.figure(figsize=(15,10))
       sns.displot(Model_Prediction_Probability, kde=True)
      plt.title(model_name+'Prediction Probabilities')
       # Set x and y ticks
      plt.xticks(color='gray')
       \#plt.xlim(.5,1)
      plt.yticks(color='gray')
       # Create axes object with plt. get current axes
       ax = plt.gca()
       # Set grid lines
       ax.grid(visible=True, which='major', axis='y', color='black', alpha=.2)
```

```
# Set facecolor
ax.set_facecolor('white')
# Remove box
ax.spines['top'].set_visible(False)
ax.spines['right'].set_visible(False)
ax.spines['bottom'].set_visible(False)
ax.spines['left'].set_visible(False)
ax.tick_params(color='white')
plt.show();
model_predictions = model.predict(train_X)
print('\n\n', classification_report(train_y, model_predictions,
                           target_names=['0-Poisonous', '1-Edible']))
con_matrix = pd.DataFrame(confusion_matrix(train_y, model_predictions),
                                       columns=['Predicted Poison', 'Predicted⊔
⇔Edible'],
                                       index=['Actual Poison', 'Actual__
⇔Edible'])
plt.figure(figsize=(15,10))
sns.heatmap(data=con_matrix, cmap='cool');
plt.title(model_name + 'Confusion Matrix')
plt.show();
\#print(' \ n \ n', con_matrix)
return con_matrix
```

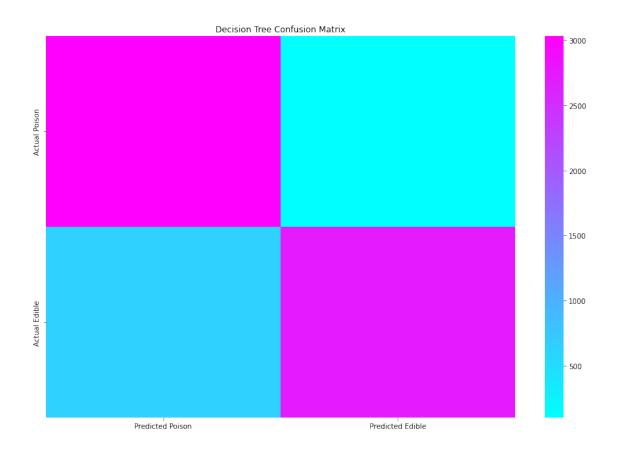
```
[]: model_analysis(tree, X_train, y_train, 'Decision Tree ')
```

<Figure size 1080x720 with 0 Axes>

Decision Tree Prediction Probabilities



	precision	recall	f1-score	support
0-Poisonous	0.83	0.97	0.89	3133
1-Edible	0.96	0.81	0.88	3366
accuracy			0.89	6499
macro avg	0.90	0.89	0.89	6499
weighted avg	0.90	0.89	0.89	6499



[]: Predicted Poison Predicted Edible
Actual Poison 3031 102
Actual Edible 637 2729

We will store our predictions as a tree_predictions variable for use in interpreting the models accuracy.

```
[]: tree_predictions = tree.predict(X_train)
[]: accuracy_score(y_train, tree_predictions)
```

[]: 0.8862901984920757

6 Devlop a model that overfits

We will use the RandomForestClassifier for our overfitting model.

```
[]: from sklearn.ensemble import RandomForestClassifier from sklearn.model_selection import cross_validate
```

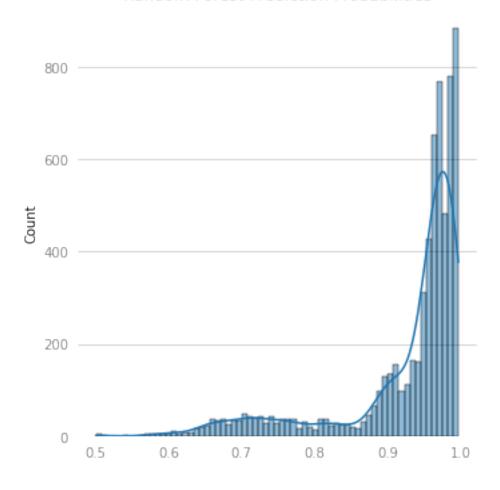
```
random_forest = RandomForestClassifier(n_estimators=100, max_depth=5)
[]: cv = cross_validate(estimator = random_forest,
                         X = X_{train}
                         y = y_train,
                         scoring='accuracy',
                         n_{jobs=-1},
                         cv=10,
                         verbose=5,
                         return_train_score=True)
    [Parallel(n_jobs=-1)]: Using backend LokyBackend with 10 concurrent workers.
    [CV] END ..., score=(train=0.990, test=0.986) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.993, test=0.992) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.991, test=0.995) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.990, test=0.994) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.990, test=0.986) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.990, test=0.991) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.989, test=0.997) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.990, test=0.992) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.991, test=0.985) total time=
                                                                0.2s
    [CV] END ..., score=(train=0.991, test=0.985) total time=
                                                                0.2s
    [Parallel(n_jobs=-1)]: Done
                                   3 out of 10 | elapsed:
                                                               0.8s remaining:
                                                                                  1.8s
    [Parallel(n_jobs=-1)]: Done
                                   6 out of 10 | elapsed:
                                                               0.8s remaining:
                                                                                  0.5s
    [Parallel(n_jobs=-1)]: Done 10 out of 10 | elapsed:
                                                               0.9s finished
[]: cv_df = pd.DataFrame(cv)
     cv_df
[]:
        fit_time score_time test_score train_score
     0 0.160542
                    0.011479
                                0.986154
                                              0.990426
     1 0.162347
                    0.006062
                                0.986154
                                              0.990426
     2 0.158364
                    0.006197
                                0.992308
                                              0.993161
     3 0.169151
                    0.016858
                                0.990769
                                              0.989913
     4 0.173562
                    0.006016
                                0.993846
                                              0.990255
     5 0.149473
                    0.006624
                                0.995385
                                              0.990939
     6 0.172313
                    0.006033
                                0.992308
                                              0.989742
     7 0.171101
                    0.006212
                                0.996923
                                              0.989229
     8 0.172027
                    0.006203
                                0.984615
                                              0.990597
     9 0.171147
                    0.005821
                                              0.990940
                                0.984592
[]: random_forest.fit(X_test, y_test)
     test_predictions = random_forest.predict(X_train)
     accuracy_score(y_train, test_predictions)
```

[]: 0.9890752423449761

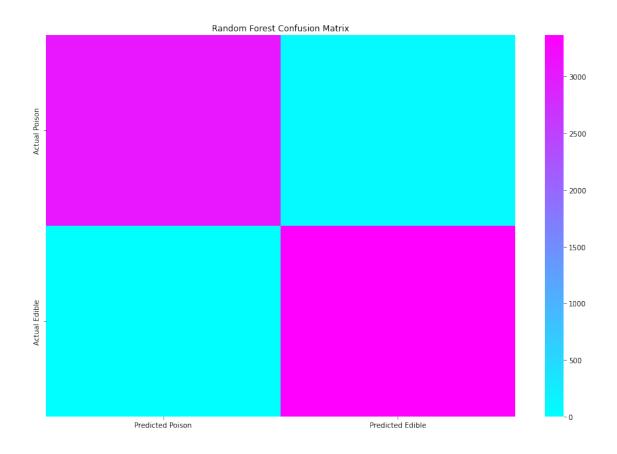
A very high accuracy score such as 99% is a sign of overfitting. We will write a function to preform all of our classification analysis from earlier.

<Figure size 1080x720 with 0 Axes>

Random Forest Prediction Probabilities



	precision	recall	f1-score	support
0-Poisonous	1.00	0.98	0.99	3133
1-Edible	0.98	1.00	0.99	3366
266117261			0.99	6499
accuracy				
macro avg	0.99	0.99	0.99	6499
weighted avg	0.99	0.99	0.99	6499



[]:			Predicted Poison	Predicted Edible
	Actual	Poison	3062	71
	Actual	Edible	0	3366

7 Regularize the model and tune its hyperparameters

Now we will tune the hyperparameters of our RandomForestClassifier and attempt to walk the line between underfitting and overfitting.

Fitting 10 folds for each of 30 candidates, totalling 300 fits

[]: RandomizedSearchCV(cv=10, estimator=RandomForestClassifier(), n_iter=30, n_jobs=-1, param_distributions={'max_depth': [1, 2, 3, 4, 5], 'n_estimators': [10, 25, 50, 100, 150, 200]}, return_train_score=True, scoring='precision', verbose=1)

We can use search.best_estimator_ to see which model has the highest precision score.

```
[ ]: best_model = search.best_estimator_
best_model
```

[]: RandomForestClassifier(max_depth=5, n_estimators=50)

From the model description we can see that a RandomForestClassifier with a max_depth of 5 and 25 estimators is our optimal model. Now we can run our analysis function.

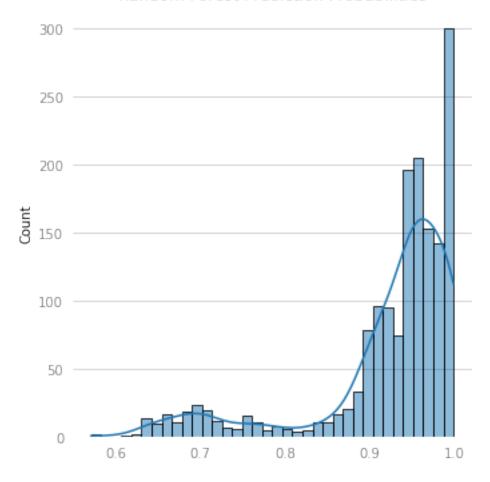
```
[]: search.best_score_
```

[]: 0.9836836764257519

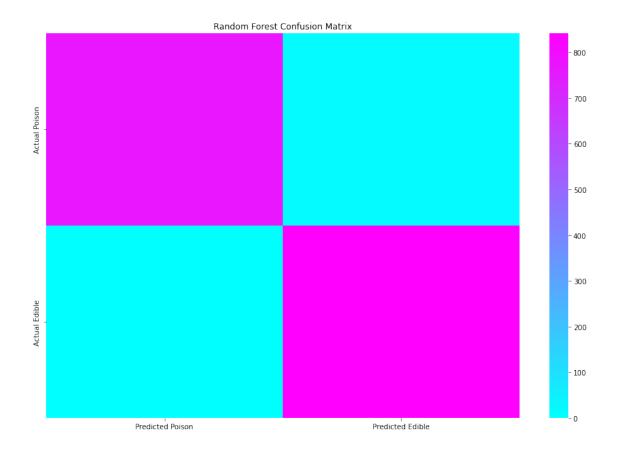
```
[]: model_analysis(best_model, X_test, y_test, 'Random Forest ')
```

<Figure size 1080x720 with 0 Axes>

Random Forest Prediction Probabilities



	precision	recall	f1-score	support
0-Poisonous	1.00	0.98	0.99	783
1-Edible	0.98	1.00	0.99	842
accuracy			0.99	1625
macro avg	0.99	0.99	0.99	1625
weighted avg	0.99	0.99	0.99	1625



[]:			Predicted Poison	Predicted Edible
	Actual	Poison	768	15
	Actual	Edible	0	842

The final model produced 6 false positives, which isn't perfect but quite close to a good model. In conclusion, we can state that we have found a model based on random forest generation that accurately predicts if mushrooms are edible or not 98% of the time.