

# Class 17: Pertussis Mini-Project

AUTHOR

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Pertussis, or whooping cough, is a highly contagious lung infection caused by a bacteria *B. pertussis*.

The CDC tracks reported cases in the U.S. since the 1920s.

```
install.packages("datapasta", repos = "http://cran.us.r-project.org")
```

Installing package into 'C:/Users/charl/AppData/Local/R/win-library/4.2'  
(as 'lib' is unspecified)

package 'datapasta' successfully unpacked and MD5 sums checked

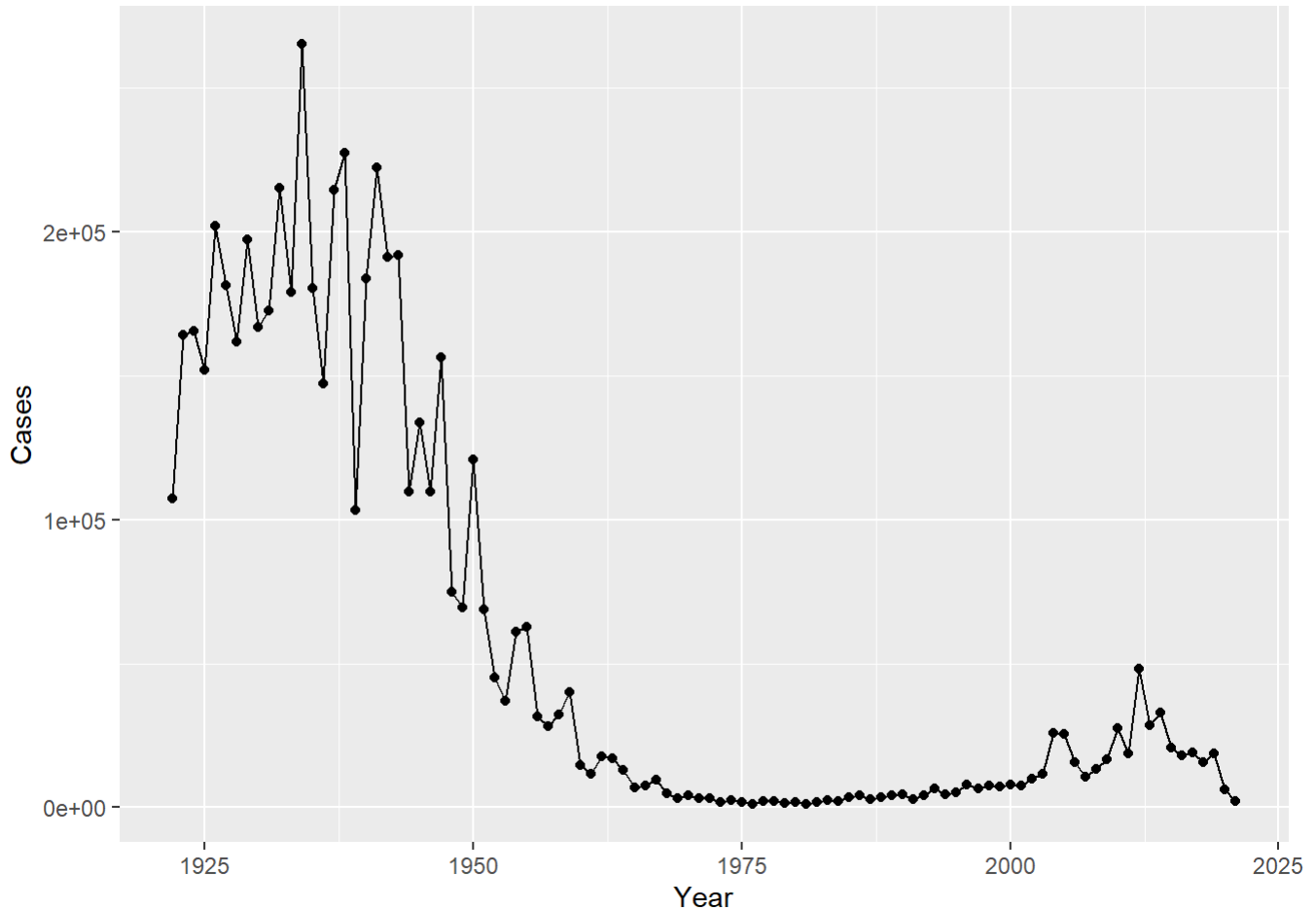
The downloaded binary packages are in  
C:\Users\charl\AppData\Local\Temp\Rtmp0YrKxT\downloaded\_packages

```
cdc <- data.frame(
  Year = c(1922L, 1923L, 1924L, 1925L,
           1926L, 1927L, 1928L, 1929L, 1930L, 1931L,
           1932L, 1933L, 1934L, 1935L, 1936L,
           1937L, 1938L, 1939L, 1940L, 1941L, 1942L,
           1943L, 1944L, 1945L, 1946L, 1947L,
           1948L, 1949L, 1950L, 1951L, 1952L,
           1953L, 1954L, 1955L, 1956L, 1957L, 1958L,
           1959L, 1960L, 1961L, 1962L, 1963L,
           1964L, 1965L, 1966L, 1967L, 1968L, 1969L,
           1970L, 1971L, 1972L, 1973L, 1974L,
           1975L, 1976L, 1977L, 1978L, 1979L, 1980L,
           1981L, 1982L, 1983L, 1984L, 1985L,
           1986L, 1987L, 1988L, 1989L, 1990L,
           1991L, 1992L, 1993L, 1994L, 1995L, 1996L,
           1997L, 1998L, 1999L, 2000L, 2001L,
           2002L, 2003L, 2004L, 2005L, 2006L, 2007L,
           2008L, 2009L, 2010L, 2011L, 2012L,
           2013L, 2014L, 2015L, 2016L, 2017L, 2018L,
           2019L, 2020L, 2021L),
  Cases = c(107473, 164191, 165418, 152003,
            202210, 181411, 161799, 197371,
            166914, 172559, 215343, 179135, 265269,
            180518, 147237, 214652, 227319, 103188,
            183866, 222202, 191383, 191890, 109873,
            133792, 109860, 156517, 74715, 69479,
            120718, 68687, 45030, 37129, 60886,
            62786, 31732, 28295, 32148, 40005,
            14809, 11468, 17749, 17135, 13005, 6799,
            7717, 9718, 4810, 3285, 4249, 3036,
```

```
3287, 1759, 2402, 1738, 1010, 2177, 2063,
1623, 1730, 1248, 1895, 2463, 2276,
3589, 4195, 2823, 3450, 4157, 4570,
2719, 4083, 6586, 4617, 5137, 7796, 6564,
7405, 7298, 7867, 7580, 9771, 11647,
25827, 25616, 15632, 10454, 13278,
16858, 27550, 18719, 48277, 28639, 32971,
20762, 17972, 18975, 15609, 18617,
6124, 2116)
)
```

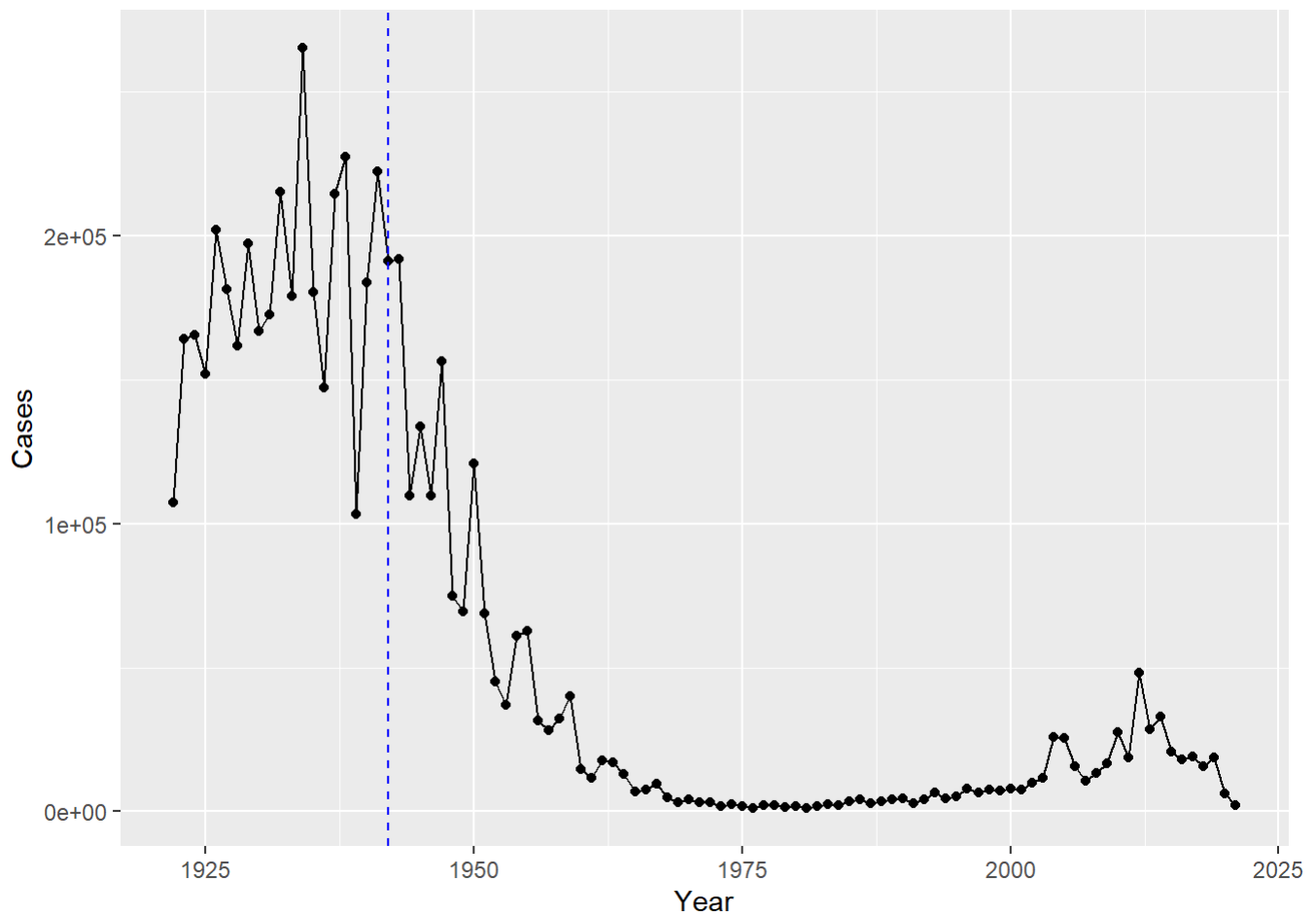
```
library(ggplot2)
```

```
ggplot(cdc,
  aes(Year, Cases)) +
  geom_point() + geom_line()
```

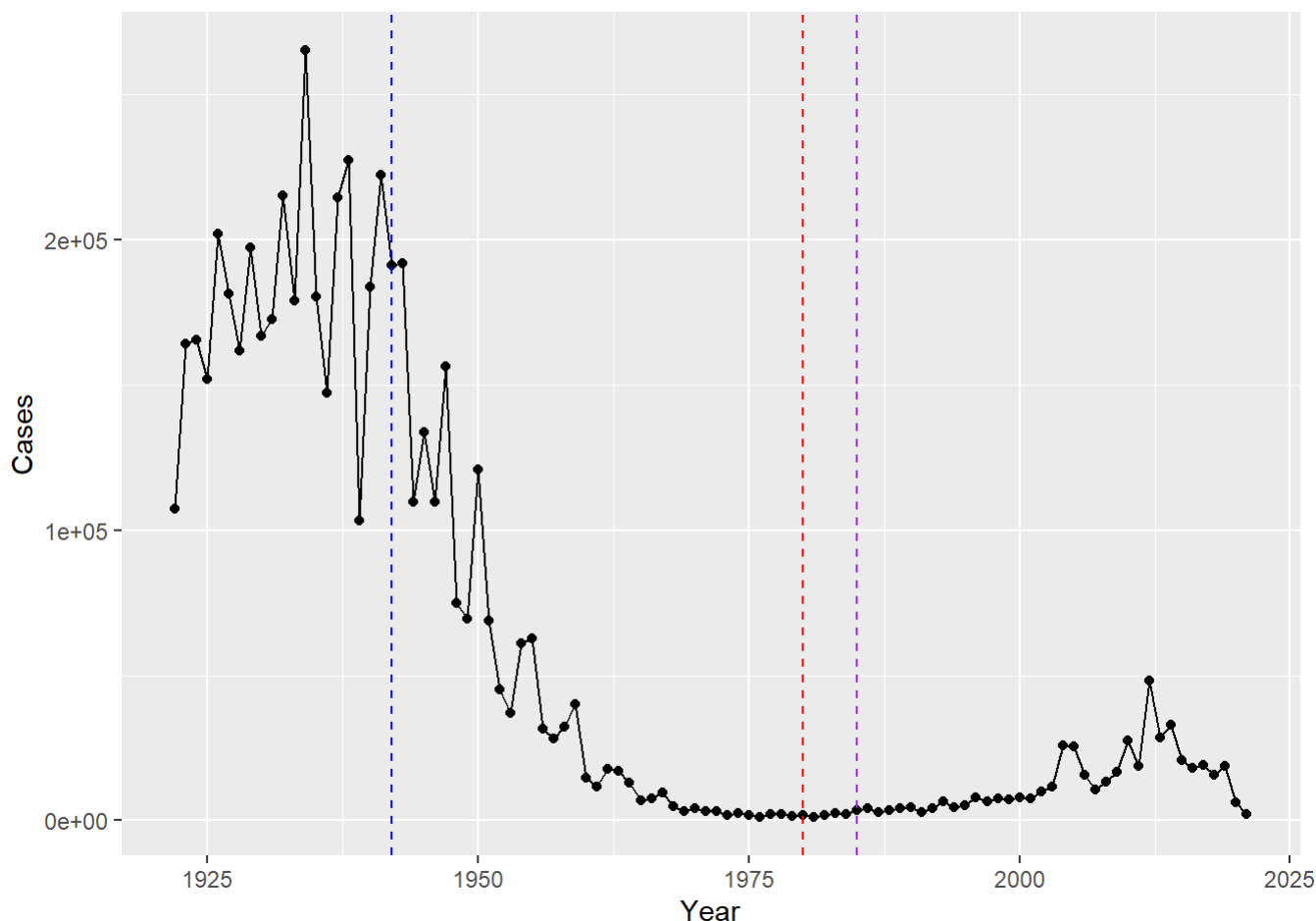


The first big “whole-cell” vaccine program started in 1942.

```
ggplot(cdc,
  aes(Year, Cases)) +
  geom_point() + geom_line() +
  geom_vline(xintercept=1942, linetype="dashed", col="blue")
```



```
ggplot(cdc,  
  aes(Year, Cases)) +  
  geom_point() + geom_line() +  
  geom_vline(xintercept=1942, linetype="dashed", col="blue") +  
  geom_vline(xintercept=1980, linetype="dashed", col="red ") +  
  geom_vline(xintercept=1985, linetype="dashed", col="purple")
```



Something big is happening with pertussis cases and big outbreaks are once again a major public health concern!

One of the main hypotheses for the increasing case numbers is waning vaccine efficiency with a newer aP vaccine.

Enter the CMI-PB project, which is studying this problem on a large scale. Let's see what data they have.

The data is available in a JSON format ("key.value" pair style). We will use the "jsonlite" package to read their data.

```
library(jsonlite)
subject <- read_json("https://www.cmi-pb.org/api/subject", simplifyVector = TRUE)

head(subject)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female	Not Hispanic or Latino	White
2	2	wP	Female	Not Hispanic or Latino	White
3	3	wP	Female	Unknown	White
4	4	wP	Male	Not Hispanic or Latino	Asian
5	5	wP	Male	Not Hispanic or Latino	Asian
6	6	wP	Female	Not Hispanic or Latino	White

	year_of_birth	date_of_boost	dataset
--	---------------	---------------	---------

1	1986-01-01	2016-09-12	2020_dataset
2	1968-01-01	2019-01-28	2020_dataset
3	1983-01-01	2016-10-10	2020_dataset
4	1988-01-01	2016-08-29	2020_dataset
5	1991-01-01	2016-08-29	2020_dataset
6	1988-01-01	2016-10-10	2020_dataset

Q4. How many aP and wP infancy vaccinated subjects are in the dataset?

```
table(subject$infancy_vac)
```

```
aP wP
47 49
```

Q5. How many Male and Female subjects/patients are in the dataset?

```
table(subject$biological_sex)
```

```
Female  Male
66      30
```

Q6. What is the breakdown of race and biological sex (e.g. number of Asian females, White males etc...)?

```
table(subject$race, subject$biological_sex)
```

	Female	Male
American Indian/Alaska Native	0	1
Asian	18	9
Black or African American	2	0
More Than One Race	8	2
Native Hawaiian or Other Pacific Islander	1	1
Unknown or Not Reported	10	4
White	27	13

Now let's read in some more database tables from CMI-PB:

```
specimen <- read_json("https://www.cmi-pb.org/api/specimen", simplifyVector=TRUE)
head(specimen)
```

	specimen_id	subject_id	actual_day_relative_to_boost
1	1	1	-3
2	2	1	736

3	3	1	1
4	4	1	3
5	5	1	7
6	6	1	11

	planned_day_relative_to_boost	specimen_type	visit
1	0	Blood	1
2	736	Blood	10
3	1	Blood	2
4	3	Blood	3
5	7	Blood	4
6	14	Blood	5

I want to join (a.k.a. merge) the `subject` and `specimen` tables together. I will use the **dplyr** package to do this.

```
library(dplyr)
```

Attaching package: 'dplyr'

The following objects are masked from 'package:stats':

`filter`, `lag`

The following objects are masked from 'package:base':

`intersect`, `setdiff`, `setequal`, `union`

```
meta <- inner_join(subject, specimen)
```

Joining with `by = join\_by(subject\_id)`

```
head(meta)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female	Not Hispanic or Latino	White
2	1	wP	Female	Not Hispanic or Latino	White
3	1	wP	Female	Not Hispanic or Latino	White
4	1	wP	Female	Not Hispanic or Latino	White
5	1	wP	Female	Not Hispanic or Latino	White
6	1	wP	Female	Not Hispanic or Latino	White

	year_of_birth	date_of_boost	dataset	specimen_id
1	1986-01-01	2016-09-12	2020_dataset	1
2	1986-01-01	2016-09-12	2020_dataset	2
3	1986-01-01	2016-09-12	2020_dataset	3
4	1986-01-01	2016-09-12	2020_dataset	4
5	1986-01-01	2016-09-12	2020_dataset	5
6	1986-01-01	2016-09-12	2020_dataset	6

	actual_day_relative_to_boost	planned_day_relative_to_boost	specimen_type
1	-3	0	Blood

2	736	736	Blood
3	1	1	Blood
4	3	3	Blood
5	7	7	Blood
6	11	14	Blood

visit

1	1
2	10
3	2
4	3
5	4
6	5

```
ncol(subject)
```

```
[1] 8
```

```
ncol(specimen)
```

```
[1] 6
```

```
ncol(meta)
```

```
[1] 13
```

```
sum(ncol(subject), ncol(specimen))
```

```
[1] 14
```

```
ab <- read_json("http://cmi-pb.org/api/ab_titer", simplifyVector=TRUE)
head(ab)
```

	specimen_id	isotype	is_antigen_specific	antigen	MFI	MFI_normalised
1	1	IgE	FALSE	Total	1110.21154	2.493425
2	1	IgE	FALSE	Total	2708.91616	2.493425
3	1	IgG	TRUE	PT	68.56614	3.736992
4	1	IgG	TRUE	PRN	332.12718	2.602350
5	1	IgG	TRUE	FHA	1887.12263	34.050956
6	1	IgE	TRUE	ACT	0.10000	1.000000

	unit	lower_limit_of_detection
1	UG/ML	2.096133
2	IU/ML	29.170000
3	IU/ML	0.530000
4	IU/ML	6.205949
5	IU/ML	4.679535
6	IU/ML	2.816431

Now I can join the “meta” table that we made above and contains all the data about the subjects and specimens with this `ab` data.

```
abdata <- inner_join(meta, ab)
```

Joining with ``by = join_by(specimen_id)``

```
head(abdata)
```

	subject_id	infancy_vac	biological_sex	ethnicity	race
1	1	wP	Female Not Hispanic or Latino	White	
2	1	wP	Female Not Hispanic or Latino	White	
3	1	wP	Female Not Hispanic or Latino	White	
4	1	wP	Female Not Hispanic or Latino	White	
5	1	wP	Female Not Hispanic or Latino	White	
6	1	wP	Female Not Hispanic or Latino	White	

	year_of_birth	date_of_boost	dataset	specimen_id
1	1986-01-01	2016-09-12	2020_dataset	1
2	1986-01-01	2016-09-12	2020_dataset	1
3	1986-01-01	2016-09-12	2020_dataset	1
4	1986-01-01	2016-09-12	2020_dataset	1
5	1986-01-01	2016-09-12	2020_dataset	1
6	1986-01-01	2016-09-12	2020_dataset	1

	actual_day_relative_to_boost	planned_day_relative_to_boost	specimen_type
1	-3	0	Blood
2	-3	0	Blood
3	-3	0	Blood
4	-3	0	Blood
5	-3	0	Blood
6	-3	0	Blood

	visit	isotype	is_antigen_specific	antigen	MFI	MFI_normalised	unit
1	1	IgE	FALSE	Total	1110.21154	2.493425	UG/ML
2	1	IgE	FALSE	Total	2708.91616	2.493425	IU/ML
3	1	IgG	TRUE	PT	68.56614	3.736992	IU/ML
4	1	IgG	TRUE	PRN	332.12718	2.602350	IU/ML
5	1	IgG	TRUE	FHA	1887.12263	34.050956	IU/ML
6	1	IgE	TRUE	ACT	0.10000	1.000000	IU/ML

	lower_limit_of_detection
1	2.096133
2	29.170000
3	0.530000
4	6.205949
5	4.679535
6	2.816431

```
dim(abdata)
```

```
[1] 32675    20
```



Q11. How many specimens (i.e. entries in abdata) do we have for each isotype?

```
table(abdata$isotype)
```

```

IgE  IgG  IgG1  IgG2  IgG3  IgG4
6698 1413 6141 6141 6141 6141

```

Q12. What do you notice about the number of visit 8 specimens compared to other visits?

```
table(abdata$visit)
```

```

 1    2    3    4    5    6    7    8
5795 4640 4640 4640 4640 4320 3920  80

```

visit 8 is significantly less than other visit counts. This is because this data is largely still being generated.

## Examine IgG1 Ab titer levels

We will use the `filter()` function from dplyr to focus on just IgG1 isotype and visits 1-7.

```
ig1 <- abdata %>% filter(isotype == "IgG1", visit!=8)
```

```
head(ig1)
```

```

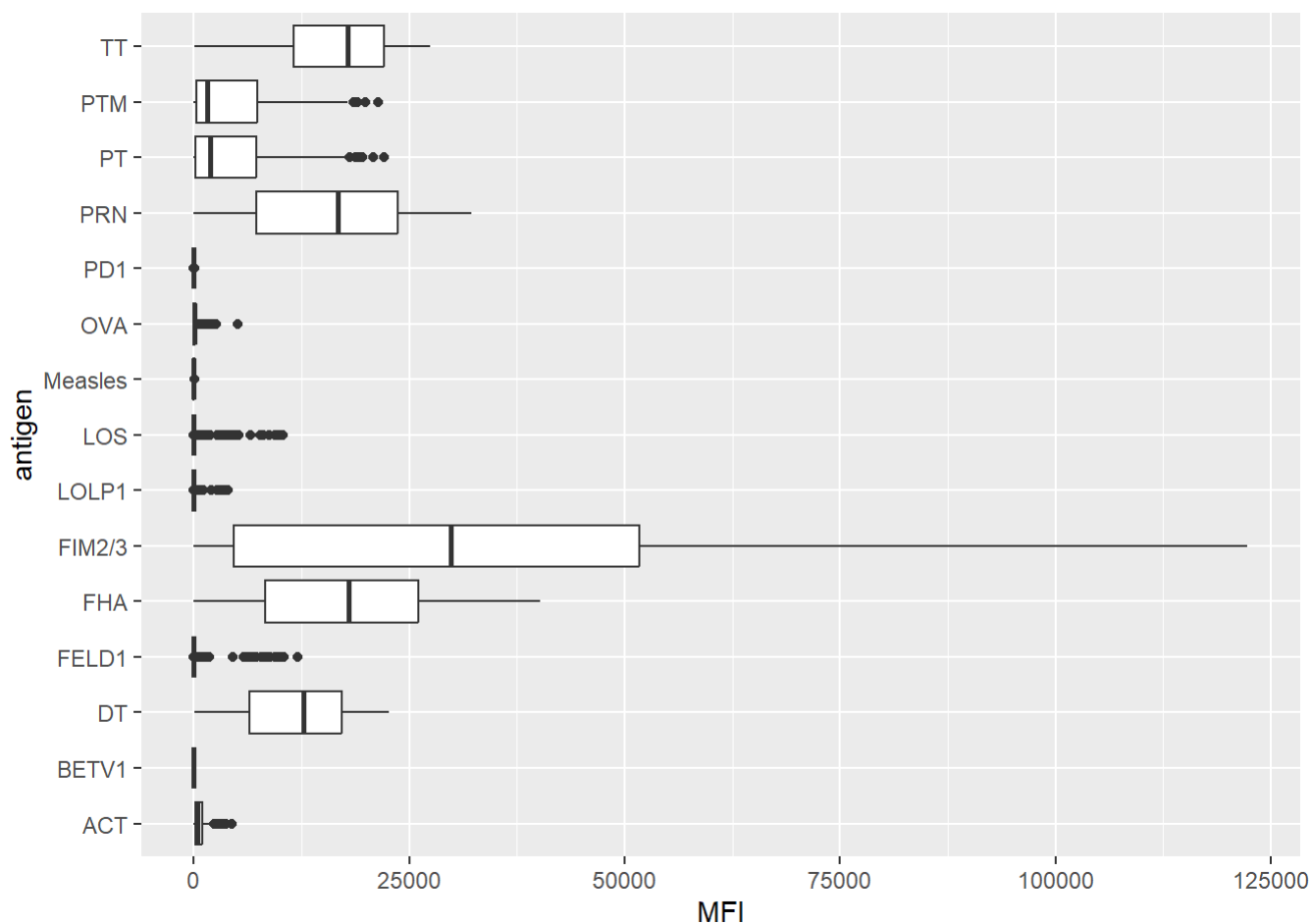
subject_id infancy_vac biological_sex ethnicity race
1          1          wP      Female Not Hispanic or Latino White
2          1          wP      Female Not Hispanic or Latino White
3          1          wP      Female Not Hispanic or Latino White
4          1          wP      Female Not Hispanic or Latino White
5          1          wP      Female Not Hispanic or Latino White
6          1          wP      Female Not Hispanic or Latino White
 year_of_birth date_of_boost   dataset specimen_id
1  1986-01-01   2016-09-12 2020_dataset          1
2  1986-01-01   2016-09-12 2020_dataset          1
3  1986-01-01   2016-09-12 2020_dataset          1
4  1986-01-01   2016-09-12 2020_dataset          1
5  1986-01-01   2016-09-12 2020_dataset          1
6  1986-01-01   2016-09-12 2020_dataset          1
actual_day_relative_to_boost planned_day_relative_to_boost specimen_type
1                -3                0          Blood
2                -3                0          Blood
3                -3                0          Blood
4                -3                0          Blood
5                -3                0          Blood

```

6	-3				0	Blood	
visit	isotype	is_antigen_specific	antigen	MFI	MFI_normalised	unit	
1	1	IgG1	TRUE	ACT	274.355068	0.6928058	IU/ML
2	1	IgG1	TRUE	LOS	10.974026	2.1645083	IU/ML
3	1	IgG1	TRUE	FELD1	1.448796	0.8080941	IU/ML
4	1	IgG1	TRUE	BETV1	0.100000	1.0000000	IU/ML
5	1	IgG1	TRUE	LOLP1	0.100000	1.0000000	IU/ML
6	1	IgG1	TRUE	Measles	36.277417	1.6638332	IU/ML
lower_limit_of_detection							
1		3.848750					
2		4.357917					
3		2.699944					
4		1.734784					
5		2.550606					
6		4.438966					

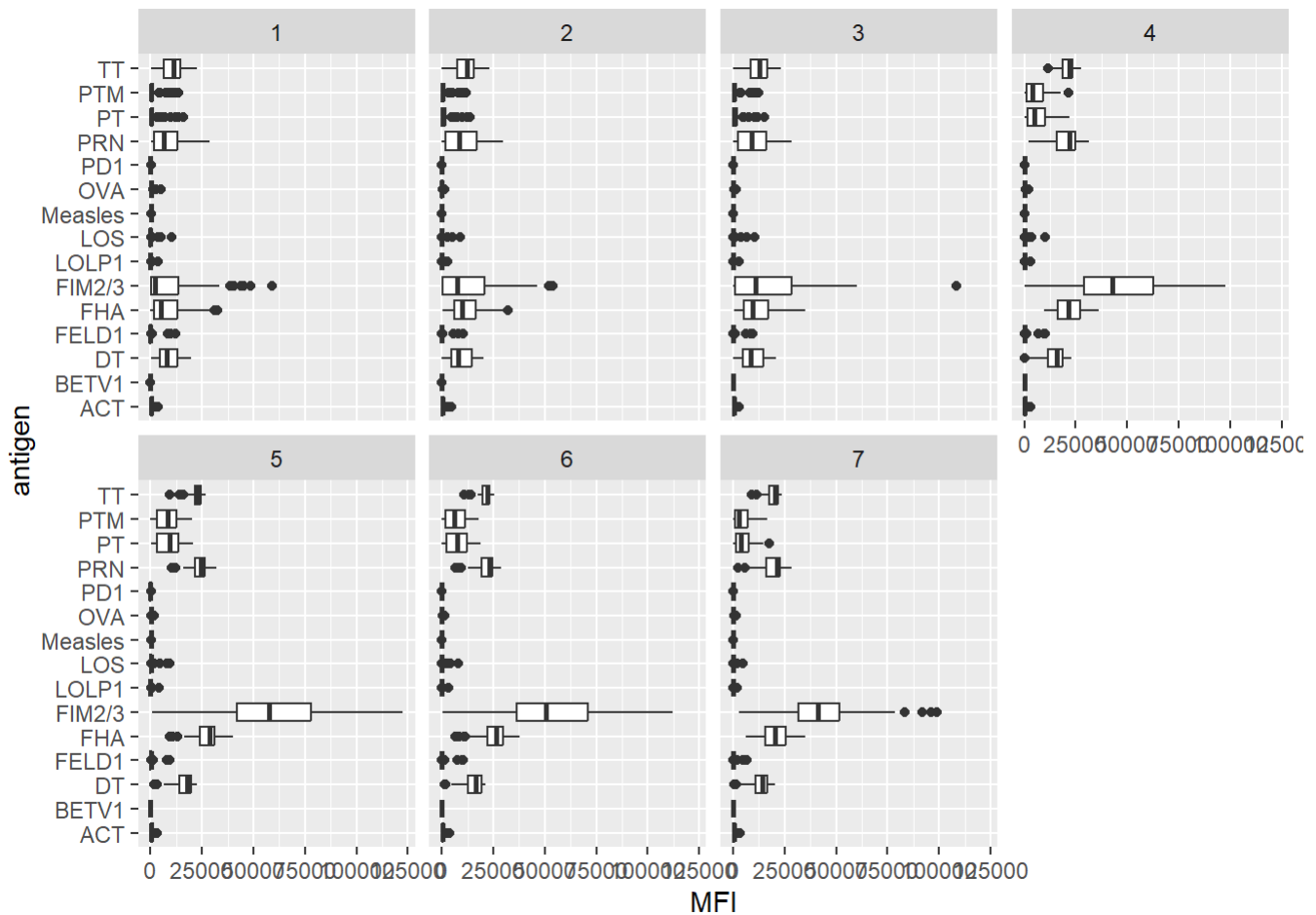
Boxplot of antigen levels over time:

```
ggplot(ig1) +
  aes(MFI, antigen) +
  geom_boxplot()
```



and faceted by visit:

```
ggplot(ig1) +
  aes(MFI, antigen) +
  geom_boxplot() +
  facet_wrap(vars(visit), nrow=2)
```



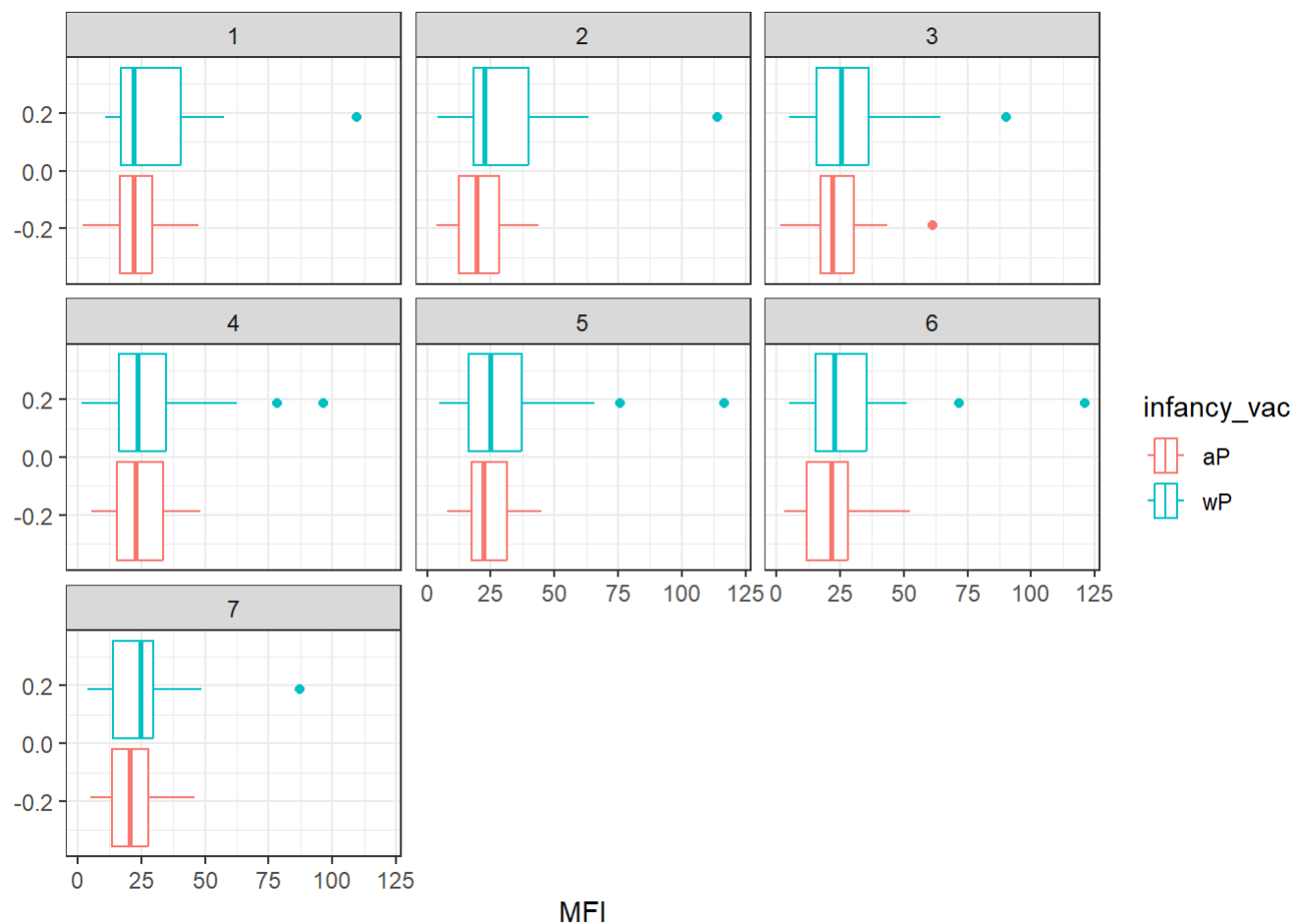
Clearly FIM2/3 changes, this is "Fimbrial Protein" that makes the bacterial pilus and is involved in cell adhesion.

PT (Pertussis toxin) changes

FHA (Filamentous hemagglutinin) also changes

Plot of measles antigen expression per visit.

```
filter(ig1, antigen=="Measles") %>%
  ggplot() +
  aes(MFI, col=infancy_vac) +
  geom_boxplot(show.legend = TRUE) +
  facet_wrap(vars(visit)) +
  theme_bw()
```



Plot of the FIM2/3 leves per visit:

```
filter(ig1, antigen=="FIM2/3") %>%
  ggplot() +
  aes(MFI, col=infancy_vac) +
  geom_boxplot(show.legend = TRUE) +
  facet_wrap(vars(visit)) +
  theme_bw()
```

