# The twenty minute guide to mzTab

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## Introduction

The purpose of this guide is to give a quick introduction on how to use mzTab efficiently. It is targeted at both, developers and end-users alike. This guide is not intended to give a complete and detailed overview of mzTab but should only be a quick and easy to understand introduction. The complete format specification as well as example files can be found at <http://mztab.googlecode.com>.

## Basic structure

mzTab files can have five sections: The metadata section, the protein section, the peptide section, the peptide-spectrum match (PSM) section and the small molecule section (see Figure 1). All of these sections, apart from the metadata section are optional and may not be present in every file.

**Sections in an mzTab file**

Figure 1: Basic structure of an mzTab file.

All lines in an mzTab file start with a three letter code to identify the information held by the line:

**MTD** for metadata

**PRH** for the protein table header line (the column labels)

**PRT** for rows of the protein table

**PEH** for the peptide table header line (the column labels)

**PEP** for rows of the peptide table

**PSH** for the PSM table header line (the column labels)

**PSM** for rows in the PSM table

**SMH** for small molecule table header line (the column labels)

**SML** for rows of the small molecule table

**COM** for comment lines

The header lines of the table based sections (protein, peptide, PSM, small molecule) must be at the top of these sections and must only occur once in the file (since every section must only occur once).

*For developers:*

mzTab is a tab separated file format. The three letter codes must be separated by a tab from the next field. Also, field names and values in the metadata section are separated by tabs as are the columns in the table based sections.

## Modelling an experimental design in mzTab

mzTab supports the reporting of technical/biological replicates within experimental designs using an adaptation of the system originally developed for mzQuantML. This is made up of four components:



Figure 2: Diagram summarizing the relation between Study Variables (SVs), MS runs, assays and samples.

**- Study variable:** Study variables represent the core final results of the study (ie. ‘inflammatory response’ vs. ‘control’). Often, these will have been derived from averaging the results of a group of replicate measurements (assays). In files where such assays are reported, study variables reference and thereby group assays. The same concept has been defined as “experimental factor”.

**- MS run:** An MS run is effectively one run (or set of runs on pre-fractionated samples) on an MS instrument, and is referenced from assay in different contexts.

**- Assay:** Any quantitative measurement about the sample (in this case through MS) is reported as an assay. In label-free MS analysis one assay is usually mapped to one MS run. If multiplexed techniques are used multiple assays are mapped to one MS run (e.g. iTRAQ). In these cases additional information about the used tag (as a property of the assay) can be reported in the metadata section.

**- Sample:** A sample represents any analysed biological material to which descriptors of species such as cell/tissue type can be applied. In all mzTab files, these can be reported in the metadata section as “sample[1-n]-description”. Samples are not mandatory in mzTab, since many software packages cannot determine what type of sample was analysed (e.g. whether biological or technical replication was performed).

See below an example corresponding to one SILAC experiment:

## COM Report of a minimal "Complete Quantification report" SILAC experiment, quantification on 2 study variables (control/treatment), 3+3 assays (replicates) reported, no identifications reported.

## COM Internally 3 replicates/assays have been used to obtain quantification values, stdev and stderror

## MTD mzTab-version 1.0.0

## MTD mzTab-mode Complete

## MTD mzTab-type Quantification

## MTD description mzTab example file for reporting a summary report of quantification data quantified on the protein level

## MTD ms\_run[1]-location file://C:\path\to\my\file1.mzML

## MTD ms\_run[2]-location file://C:\path\to\my\file2.mzML

## MTD ms\_run[3]-location file://C:\path\to\my\file3.mzML

## MTD ms\_run[4]-location file://C:\path\to\my\file4.mzML

## MTD protein-quantification\_unit [PRIDE, PRIDE:0000393, Relative quantification unit,]

## MTD software[1] [MS, MS:1001583, MaxQuant,] MTD psm\_search\_engine\_score[1] [MS, MS:1001979, MaxQuant:PTM score,]

## MTD fixed\_mod[1] [UNIMOD, UNIMOD:4, Carbamidomethyl, ]

## MTD fixed\_mod[2] [UNIMOD, UNIMOD:188, Label:13C(6), ]

## MTD variable\_mod[1] [UNIMOD, UNIMOD:35, Oxidation, ]

## MTD quantification\_method [MS, MS:1001835, SILAC, ]

## MTD assay[1]-quantification\_reagent [PRIDE, PRIDE:0000326, SILAC light, ]

## MTD assay[2]-quantification\_reagent [PRIDE, PRIDE:0000325, SILAC heavy, ]

## MTD assay[3]-quantification\_reagent [PRIDE, PRIDE:0000326, SILAC light, ]

## MTD assay[4]-quantification\_reagent [PRIDE, PRIDE:0000325, SILAC heavy, ]

## MTD assay[1]-ms\_run\_ref ms\_run[1]

## MTD assay[2]-ms\_run\_ref ms\_run[1]

## MTD assay[3]-ms\_run\_ref ms\_run[2]

## MTD assay[4]-ms\_run\_ref ms\_run[2]

## MTD study\_variable[1]-assay\_refs assay[1],assay[3]

## MTD study\_variable[2]-assay\_refs assay[2],assay[4]

## MTD study\_variable[1]-description heat shock response of control

## MTD study\_variable[2]-description heat shock response of treatment

## Metadata section in mzTab

The metadata section in mzTab files contains information about the units and consists of key - value pairs separated by a tab. A complete list of available fields can be found in the specification document.

COM Example of the metadata section for an identification file.

MTD mzTab-version 1.0 rc5

MTD mzTab-mode Complete

MTD mzTab-type Identification

MTD mzTab-ID PRIDE assay metadata example

MTD title COFRADIC N-terminal proteome of unstimulated human blood platelets, identified and unidentified spectra

MTD instrument[1]-name [PRIDE, PRIDE:0000131, Instrument model, Micromass Q-TOF I]

MTD instrument[1]-source [MS, MS:1000008, Ionization Type, ESI]

MTD instrument[1]-analyzer [1][MS, MS:1000010, Analyzer Type, Quadrupole-TOF]

MTD instrument[1]-detector [MS, MS:1000026, Detector Type, MultiChannelPlate]

MTD software[1] [MS, MS:1001456, analysis software, MassLynx v3.5]

MTD protein\_search\_engine\_score[1][MS, MS:1002367, probability for proteins,]

MTD publication[1] pubmed:16038019|pubmed:12665801|pubmed:16518876

MTD contact[1]-name Kristian Flikka

MTD contact[1]-affiliation Computational Biology Unit, Bergen Center for Computational Science, University of Bergen

MTD contact[1]-email flikka@ii.uib.no

MTD ms\_run[1]-format [MS, MS:1000564, PSI mzData file, ]

MTD ms\_run[1]-location ftp://ftp.ebi.ac.uk/pub/databases/pride/PRIDE\_Exp\_Complete\_Ac\_1643.xml

MTD ms\_run[1]-id\_format [MS, MS:1000777, spectrum identifier nativeID format, ]

MTD sample[1]-species[1] [NEWT, 9606, Homo sapiens (Human), ]

MTD sample[1]-cell\_type[1] [CL, CL:0000233, platelet, ]

MTD sample[1]-custom[1] [MeSH, D001792, blood\_platelets, ]

MTD assay[1]-sample\_ref sample[1]

MTD assay[1]-ms\_run\_ref ms\_run[1]

The number of required columns in the protein table depends on the type of mzTab file (‘Identification’ / ‘Quantification’) and the used mode (‘Complete’ / ‘Summary’):

**Metadata Section**

|  |  |  |
| --- | --- | --- |
| **Field Name** | **Identification** | **Quantification** |
| mzTab-version | **SC** | **SC** |
| mzTab-mode | **SC** | **SC** |
| mzTab-type | **SC** | **SC** |
| description | **SC** | **SC** |
| ms\_run[1-n]-location | **SC** | **SC** |
| protein\_search\_engine\_score[1-n] | **SC** | **SC** |
| peptide\_search\_engine\_score[1-n] | **SC** | **SC** |
| psm\_search\_engine\_score[1-n] | **SC** | **SC** |
| smallmolecule\_search\_engine\_score[1-n] | **SC** | **SC** |
| fixed\_mod[1-n] | **SC** | **SC** |
| variable\_mod[1-n] | **SC** | **SC** |
| protein-quantification-unit | (not used) | **SC** (if protein section present) |
| peptide-quantification-unit | (not used) | **SC** (if peptide section present) |
| smallmolecule- quantification -unit | (not used) | **SC** (if small molecule section present) |
| study\_variable[1-n]-description | (not used) | **SC** |
| software[1-n] | *s***C** | *s***C** |
| quantification\_method | (not used) | *s***C** |
| assay[1-n]-ms\_run\_ref | *sc* (required if assays reported) | *s***C** (required if assays reported) |
| assay[1-n]-quantification\_reagent | (not used) | *s***C** |
| mzTab-ID | *sc* | *sc* |
| title | *sc* | *sc* |
| sample\_processing[1-n] | *sc* | *sc* |
| instrument[1-n]-name | *sc* | *sc* |
| instrument[1-n]-source | *sc* | *sc* |
| instrument[1-n]-analyzer[1-n] | *sc* | *sc* |
| instrument[1-n]-detector | *sc* | *sc* |
| software[1-n]-setting[1-n] | *sc* | *sc* |
| false\_discovery\_rate | *sc* | *sc* |
| publication[1-n] | *sc* | *sc* |
| contact[1-n]-name | *sc* | *sc* |
| contact[1-n]-affiliation | *sc* | *sc* |
| contact[1-n]-email | *sc* | *sc* |
| uri[1-n] | *sc* | *sc* |
| fixed\_mod[1-n]-site | *sc* | *sc* |
| fixed\_mod[1-n]-position | *sc* | *c* |
| variable\_mod[1-n]-site | *sc* | *sc* |
| variable\_mod[1-n]-position | *sc* | *sc* |
| ms\_run[1-n]-format | *sc* | *sc* |
| ms\_run[1-n]-id\_format | *sc* | *sc* |
| ms\_run[1-n]-fragmentation\_method | *sc* | *sc* |
| ms\_run[1-n]-hash | ***sc*** | ***sc*** |
| ms\_run[1-n]-hash\_method | ***sc*** | ***sc*** |
| custom[1-n] | *sc* | *sc* |
| sample[1-n]-species[1-n] | *sc* | *sc* |
| sample[1-n]-tissue[1-n] | *sc* | *sc* |
| sample[1-n]-cell\_type[1-n] | *sc* | *sc* |
| sample[1-n]-disease[1-n] | *sc* | *sc* |
| sample[1-n]-description | *sc* | *sc* |
| sample[1-n]-custom[1-n] | *sc* | *sc* |
| study\_variable[1-n]-description | *sc* (required if SV reported) | *sc* (required if SV reported) |
| study\_variable[1-n]-sample\_refs | *sc* | *sc* |
| study\_variable[1-n]-assay\_ref | *sc* | *s***C** |
| assay[1-n]-quantification\_mod[1-n] | (not used) | *sc* |
| assay[1-n]-quantification\_mod[1-n]-position | (not used) | *sc* |
| assay[1-n]-quantification\_mod[1-n]-site | (not used) | *sc* |
| assay[1-n]-sample\_refs | (not used) | *sc* |
| cv[1-n]-label | *sc* | *sc* |
| cv[1-n]-full\_name | *sc* | *sc* |
| cv[1-n]-version | *sc* | *sc* |
| cv[1-n]-url | *sc* | *sc* |
| colunit\_protein | *sc* | *sc* |
| colunit\_peptide | *sc* | *sc* |
| colunit\_psm | *sc* | *sc* |
| colunit\_small\_molecule | *sc* | *sc* |

**S** … required in summary file *s* … optional in summary file  
**C** … required in complete file *c* … optional in complete file

## Proteins in mzTab

Protein identifications are reported in the protein section. The protein section is table based. The table header is identified by the prefix “PRH”, entries in the protein table are identified through “PRT”. The protein section must only be present once. Columns are separated by a tab.

COM Example of the protein section. Other sections are omitted.  
PRH accession description taxid species database database\_version …  
PRT P12345 mAspAT 9986 Rabbit UniProtKB 2013\_08 …  
PRT P02042 Hemoglobin 9606 Human UniProtKB 2013\_08 …

The number of required columns in the protein table depends on the type of mzTab file (‘Identification’ / ‘Quantification’) and the used mode (‘complete’ / ‘summary’):

|  |  |  |
| --- | --- | --- |
| **Field Name** | **Identification** | **Quantification** |
| accession | **SC** | **SC** |
| description | **SC** | **SC** |
| taxid | **SC** | **SC** |
| species | **SC** | **SC** |
| database | **SC** | **SC** |
| database\_version | **SC** | **SC** |
| search\_engine | **SC** | **SC** |
| best\_search\_engine\_score[1-n] | **SC** | **SC** |
| ambiguity\_members | **SC** | **SC** |
| modifications | **SC** | **SC** |
| protein\_coverage | ***s*C** | ***s*C** |
| protein\_abundance\_study\_variable[1-n] | (not used) | **SC** |
| protein\_abundance\_stdev\_study\_variable[1-n] | (not used) | **SC** |
| protein\_abundance\_std\_error\_study\_variable[1-n] | (not used) | **SC** |
| search\_engine\_score[1-n]\_ms\_run[1-n] | *s***C** | *s***C** |
| num\_psms\_ms\_run[1-n] | *s***C** | *sc* |
| num\_peptides\_distinct\_ms\_run[1-n] | *s***C** | *sc* |
| num\_peptide\_unique\_ms\_run[1-n] | *s***C** | *sc* |
| protein\_abundance\_assay[1-n] | (not used) | *s***C** |
| opt\_{identifier}\_\* | *sc* | *sc* |
| go\_terms | *sc* | *sc* |
| reliability | *sc* | *sc* |
| uri | *sc* | *sc* |

**S** … required in summary file *s* … optional in summary file  
**C** … required in complete file *c* … optional in complete file

## Peptides in mzTab

The peptide section is similar to the PSM section but used to report quantitative results aggregated on the peptide level. It should therefore not be used in ‘Identification’ files. It is table based and columns are separated by a tab. The header of the peptide table is indicated by “PEH”, and entries in the table by “PEP”. The peptide section must also be present only once.

PEH sequence accession unique database database\_version search\_engine

PEP KLVILEGELER IPI00010779 0 UniProtKB 2013\_08 [MS,MS:1001207,Mascot,

PEP KQAEDRCK IPI00513698 0 UniProtKB 2013\_08 [MS,MS:1001207,Mascot,

PEP LATALQK IPI00218319 1 UniProtKB 2013\_08 [MS,MS:1001207,Mascot,]

PEP LATALQKLEEAEK IPI00218319 1 UniProtKB 2013\_08 [MS,MS:1001207,Mascot,]

PEP RIQLVMEEELDRAQER IPI00212519 0 UniProtKB 2013\_08 [MS,MS:1001207,Mascot,]

The number of required columns depends on the mzTab file’s type and mode:

|  |  |  |
| --- | --- | --- |
| **Field Name** | **Identification** | **Quantification** |
| sequence | (not used) | **SC** |
| accession | (not used) | **SC** |
| unique | (not used) | **SC** |
| database | (not used) | **SC** |
| database\_version | (not used) | **SC** |
| search\_engine | (not used) | **SC** |
| best\_search\_engine\_score[1-n] | (not used) | **SC** |
| modifications | (not used) | **SC** |
| retention\_time | (not used) | **SC** |
| retention\_time\_window | (not used) | **SC** |
| charge | (not used) | **SC** |
| mass\_to\_charge | (not used) | **SC** |
| peptide\_abundance\_study\_variable[1-n] | (not used) | **SC** |
| peptide\_abundance\_stdev\_study\_variable[1-n] | (not used) | **SC** |
| peptide\_abundance\_std\_error\_study\_variable[1-n] | (not used) | **SC** |
| search\_engine\_score[1-n]\_ms\_run[1-n] | (not used) | *s***C** |
| peptide\_abundance\_assay[1-n] | (not used) | *s***C** |
| spectra\_ref | (not used) | *s***C** (if MS2 based quantification is used) |
| opt\_{identifier}\_\* | (not used) | *sc* |
| reliability | (not used) | *sc* |
| uri | (not used) | *sc* |

**S** … required in summary file *s* … optional in summary file  
**C** … required in complete file *c* … optional in complete file

## PSMs in mzTab

The PSM section is used to report peptide identifications on a per spectrum level and is the recommended way to report peptides in ‘Identification’ files. It is similar to the protein section and also table based with the columns separated by a tab. If a peptide can be assigned to multiple proteins, this PSM MUST be reported multiple times (see PSM\_ID 4 in the example below). The PSM section must also be present only once.

COM Example of the PSM section. Other sections and several columns are omitted.

PSH sequence PSM\_ID accession unique database database\_version …

PSM QTQTFTTYSDNQPGVL 1 P63017 1 UniProtKB 2013\_08 …

PSM AVVNGYSASDTVGAGFAQAK 2 Q8K0U4 1 UniProtKB 2013\_08 …

PSM ALLRLHQECEKLK 3 Q61699 1 UniProtKB 2013\_08 …

PSM DWYPAHSR 4 P14602 0 UniProtKB 2013\_08 …

PSM DWYPAHSR 4 Q340U4 0 UniProtKB 2013\_08 …

PSM DWYPAHSR 4 P16627 0 UniProtKB 2013\_08 …

PSM MNQSNASPTLDGLFR 5 P14602 1 UniProtKB 2013\_08 …

The required columns depend on the mzTab file’s type and mode:

|  |  |  |
| --- | --- | --- |
| **Field Name** | **Identification** | **Quantification** |
| sequence | **SC** | **SC** |
| PSM\_ID | **SC** | **SC** |
| accession | **SC** | **SC** |
| unique | **SC** | **SC** |
| database | **SC** | **SC** |
| database\_version | **SC** | **SC** |
| search\_engine | **SC** | **SC** |
| search\_engine\_score[1-n] | **SC** | **SC** |
| modifications | **SC** | **SC** |
| spectra\_ref | **SC** | **SC** |
| retention\_time | **SC** | **SC** |
| charge | **SC** | **SC** |
| exp\_mass\_to\_charge | **SC** | **SC** |
| calc\_mass\_to\_charge | **SC** | **SC** |
| pre | **SC** | **SC** |
| post | **SC** | **SC** |
| start | **SC** | **SC** |
| end | **SC** | **SC** |
| opt\_{identifier}\_\* | *sc* | *sc* |
| reliability | *sc* | *sc* |
| uri | *sc* | *sc* |

**S** … required in summary file *s* … optional in summary file  
**C** … required in complete file *c* … optional in complete file

## Small Molecules in mzTab

The small molecule section is also a table based section (same rules apply). Small molecules are identified through an “identifier” in mzTab. This identifier can be any text that sensibly identifies the given small molecule in the given field of research. These identifiers should generally be entries in compound databases used in the respective field (for example, Human Metabolome Database entries, ChEBI identifiers, PubChem IDs or LIPID MAPS IDs). Apart from this identifier, small molecules can be assigned a chemical formula, SMILES and/or InChi identifier, a human readable description, a *m/z* value, a charge state, retention time(s), a species, source database and search engine including score. We are aware, that these fields are not applicable to all fields of metabolomics, but we believe that they represent a sensible selection.

COM Example of the small molecule section. Other sections are omitted. ‘smiles’ and

COM ‘inchi\_key’ are not complete.

identifier chemical\_formula smiles inchi\_key description exp\_mass\_to\_charge

CHEBI:17562 C9H13N3O5 Nc1ccn([C@@H] UHDGCWIWMR… Cytidine 244.0928 2O[C@H](CO)…

The required columns depend on the mzTab file’s type and mode:

|  |  |  |
| --- | --- | --- |
| **Field Name** | **Identification** | **Quantification** |
| identifier | **SC** | **SC** |
| chemical\_formula | **SC** | **SC** |
| smiles | **SC** | **SC** |
| inchi\_key | **SC** | **SC** |
| description | **SC** | **SC** |
| exp\_mass\_to\_charge | **SC** | **SC** |
| calc\_mass\_to\_charge | **SC** | **SC** |
| charge | **SC** | **SC** |
| retention time | **SC** | **SC** |
| taxid | **SC** | **SC** |
| species | **SC** | **SC** |
| database | **SC** | **SC** |
| database\_version | **SC** | **SC** |
| spectra\_ref | **SC** | **SC** |
| search\_engine | **SC** | **SC** |
| best\_search\_engine\_score[1-n] | **SC** | **SC** |
| modifications | **SC** | **SC** |
| smallmolecule\_abundance\_assay[1-n] | (not used) | **SC** (if assays reported) |
| smallmolecule\_abundance\_study\_variable[1-n] | (not used) | **SC** (if study vars. reported) |
| smallmolecule\_stdev\_study\_variable[1-n] | (not used) | **SC** (if study vars. reported) |
| smallmolecule\_std\_error\_study\_variable[1-n] | (not used) | **SC** (if study vars. reported) |
| search\_engine\_score[1-n]\_ms\_run[1-n] | (not used) | s**C** |
| opt\_{identifier}\_\* | *sc* | *sc* |
| reliability | *sc* | *sc* |
| uri | *sc* | *sc* |

**S** … required in summary file *s* … optional in summary file  
**C** … required in complete file *c* … optional in complete file

## Missing values

In the table-based sections (protein, peptide, and small molecule) there MUST NOT be any empty cells. In case a given property is not available “null” MUST be used.

This is, for example, the case when a URI is not available for a given protein (*i.e.* the table cell MUST NOT be empty but “null” has to be reported). If ratios are included and the denominator is zero, the “INF” value MUST be used. If the result leads to calculation errors (for example 0/0), this MUST be reported as “not a number” (“NaN”). In some cases, there is ambiguity with respect to these cases: e.g. in spectral counting if no peptide spectrum matches are observed for a given protein, it is open for debate as to whether its abundance is zero or missing (“null”).

## Reliability score

All protein, peptide, psm and small molecule identifications reported in an mzTab file can be assigned a reliability score (optional column “reliability” in all tables). The idea is to provide a way for researcher and/or MS proteomics or metabolomics repositories or data producers to score the reported identifications based on their own criteria. This score is completely resource-dependent and must not be seen as a comparable score between mzTab files generated from different resources. The criteria used to generate this score should be documented by the data providers.

The reliability is reported as an integer between 1-3 in all but the *small molecule* section (see below) and should be interpreted as follows:

1: high reliability

2: medium reliability

3: poor reliability

For metabolomics (*small molecule* section), according to current MSI agreement, it should be reported as an integer between 1-4 and should be interpreted as follows:

1: identified metabolites

2: putatively annotated compounds

3: putatively characterized compound classes

4: unknown compounds

The idea behind this score is to mimic the general concept of “resource based trust”. For example, if one resource reports identifications with a given reliability this would be interpreted differently as an identification reported from another resource. If resources now report their reliabilities using this metric and document how their metric is generated, a user can base his own interpretation of the results based on his trust in the resource. Furthermore, approaches to make various, for example search engine scores comparable have failed so far. To prevent the notion that the reported scores represent comparable probabilities this very abstract metric was chosen.

## Quantitative Data

There are multiple quantification techniques available for MS-based experiments that often result in slightly different types of data. mzTab was not designed to capture any of these specific differences. The goal for mzTab was to provide a generic view on quantitative MS-based identification data that is applicable to as many different quantitation methods as possible. The method used in mzTab to model quantitative data is similar to the one used in mzQuantML and relies on “assays” and “study variables”. “Assays” are used to report the actual measured values (*ie.* tag intensities) while “study variables” correspond to the final results from the study. A description of these items can be found above in “Modelling an experimental design in mzTab”. Extensive example files on how to report different types of quantitation techniques can be found at <https://code.google.com/p/mztab/wiki/ExampleFiles>. See below an example corresponding to one SILAC experiment:

## COM Report of a minimal "Complete Quantification report" SILAC experiment, quantification on 2 study variables (control/treatment), 3+3 assays (replicates) reported, no identifications reported.

## COM Internally 3 replicates/assays have been used to obtain quantification values, stdev and stderror

## MTD mzTab-version 1.0.0

## MTD mzTab-mode Complete

## MTD mzTab-type Quantification

## MTD description mzTab example file for reporting a summary report of quantification data quantified on the protein level

## MTD ms\_run[1]-location file://C:\path\to\my\file1.mzML

## MTD ms\_run[2]-location file://C:\path\to\my\file2.mzML

## MTD ms\_run[3]-location file://C:\path\to\my\file3.mzML

## MTD ms\_run[4]-location file://C:\path\to\my\file4.mzML

## MTD protein-quantification\_unit [PRIDE, PRIDE:0000393, Relative quantification unit,]

## MTD software[1] [MS, MS:1001583, MaxQuant,]

## MTD psm\_search\_engine\_score[1] [MS, MS:1001979, MaxQuant:PTM score,]

## MTD fixed\_mod[1] [UNIMOD, UNIMOD:4, Carbamidomethyl, ]

## MTD fixed\_mod[2] [UNIMOD, UNIMOD:188, Label:13C(6), ]

## MTD variable\_mod[1] [UNIMOD, UNIMOD:35, Oxidation, ]

## MTD quantification\_method [MS, MS:1001835, SILAC, ]

## MTD assay[1]-quantification\_reagent [PRIDE, PRIDE:0000326, SILAC light, ]

## MTD assay[2]-quantification\_reagent [PRIDE, PRIDE:0000325, SILAC heavy, ]

## MTD assay[3]-quantification\_reagent [PRIDE, PRIDE:0000326, SILAC light, ]

## MTD assay[4]-quantification\_reagent [PRIDE, PRIDE:0000325, SILAC heavy, ]

## MTD assay[1]-ms\_run\_ref ms\_run[1]

## MTD assay[2]-ms\_run\_ref ms\_run[1]

## MTD assay[3]-ms\_run\_ref ms\_run[2]

## MTD assay[4]-ms\_run\_ref ms\_run[2]

## MTD study\_variable[1]-assay\_refs assay[1],assay[3]

## MTD study\_variable[2]-assay\_refs assay[2],assay[4]

## MTD study\_variable[1]-description heat shock response of control

## MTD study\_variable[2]-description heat shock response of treatment

## Protein Inference

There are multiple approaches to how protein inference can be reported. mzTab is designed to only hold experimental results, which in proteomics experiments can be very complex. At the same time, for downstream statistical analysis there is a need to simplify this problem. It is not possible to model detailed protein inference data without a significant level of complexity at the file format level. Therefore, it was decided to have only limited support for protein inference/grouping reporting in mzTab files. Protein entries in mzTab files contain the field ambiguity\_members. The protein accessions listed in this field should identify proteins that were also identified through the same set of peptides or spectra, or proteins supported by a largely overlapping set of evidence, and could also be a viable candidate for the “true” identification of the entity reported. It is RECOMMENDED that “subset proteins” that are unlikely to have been identified SHOULD NOT be reported here. The mapping of a single peptide-spectrum match (PSM) to multiple accessions is supported through the reporting of the same PSM on multiple rows of the PSM section, as exemplified below.

COM In the following example only one peptide was identified that can be attributed to  
COM multiple proteins. The choice which one to pick as primary accession depends on COM the resource generating the mzTab file.  
...  
PRH accession … ambiguity\_members …  
PRT P14602 … Q340U4, P16627 …  
...  
PSH sequence PSM\_ID accession unique …

PSM DWYPAHSR 4 P14602 0 …

PSM DWYPAHSR 4 Q340U4 0 …

PSM DWYPAHSR 4 P16627 0 …

## Advanced topics

There are several other features in mzTab that could not be introduced here. Detailed information about these features can be found in the specification document such as:

* Reporting post-translational modifications (PTMs) including modification position ambiguity.
* Reporting results from multiple search engines.
* Referencing external spectra.
* Referencing external resources such as mzIdentML or mzQuantML files.
* Adding optional columns.
* Specifying a column’s unit.

An up-to-date list of example files can be found at <http://code.google.com/p/mztab/wiki/ExampleFiles>. The specification document can be found at <http://code.google.com/p/mztab/>.