# **Bio-Data Science Task 1 Report**

**Summary:** In this week's task, I carried out two tasks to strengthen my skills inR-programming for data science.

First, I performed simple visualization on the data set "astrocyte\_data". I created barplots, histograms, boxplots, added legends and labels. Next, I installed all the packages that would enable me perform ggplots on the data set "aflatoxin\_data". I continued to melt data, stack barplots, create pie-chart, histogram and scatter plot while applying colors. I then created heatmaps which I manipulated severally to create what suits my taste.

In conclusion, this task has strengthened my skills in R-programming for statistical analysis/visualization.

Explanations of codes and visualizations are provided below:

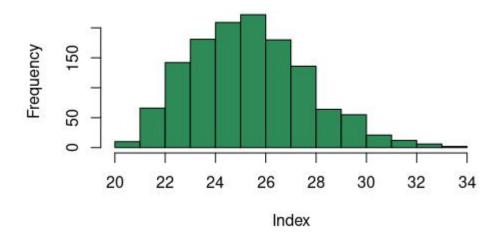
1. Mitochondria-endoplasmic reticulum contacts in reactive astrocytes promote vascular remodelling. Goebel et al. I want to perform the comparison of FACS-enriched astrocytes from uninjured and injured wild-type mice at different time points. I have already downloaded my data as .csv, next I will import my file in R

astrocyte\_data <- read.csv(file.choose())
astrocyte data</pre>

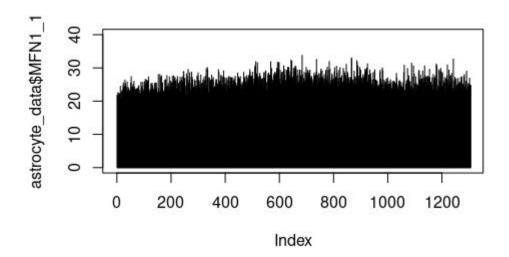
2. I want to perform Data Visualization in R. Let me construct a histogram for the distribution of MFN1 1 Gene

hist(astrocyte\_data\$MFN1\_1, col = "Sea green", main = "Distribution of MFN1\_1 Gene", xlab = "Index")

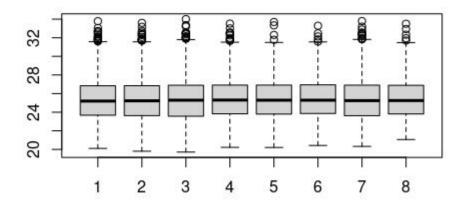
# Distribution of MFN1\_1 Gene



3. I want to plot lineplots for the distribution of MFN1\_1 Gene plot(astrocyte dataMFN1 1, type = "h", xlim = c(0,1300), ylim = c(0, 40))

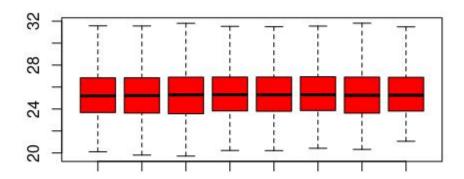


# 4. I want to create black and white boxplots for my genes boxplot(astrocyte\_data\$MFN1\_1, astrocyte\_data\$MFN1\_2, astrocyte\_data\$MFN1\_4, astrocyte\_data\$WT\_1, astrocyte\_data\$WT\_3, astrocyte\_data\$WT\_4, astrocyte\_data\$WT], astrocyte\_data\$WT



5. Let me add red color and title to my boxplot boxplot(astrocyte\_data\$MFN1\_1, astrocyte\_data\$MFN1\_2, astrocyte\_data\$MFN1\_4, astrocyte\_data\$WT\_1, astrocyte\_data\$WT\_3, astrocyte\_data\$WT\_4, astrocyte\_data\$WT, col = 'red', outline = F, main = 'BOXPLOT')

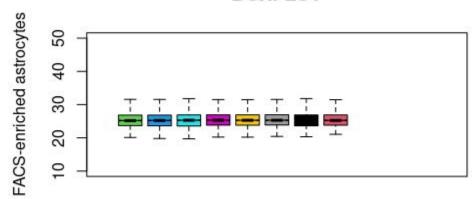
# BOXPLOT



#### 6. I want to color by gene types

boxplot(astrocyte\_data\$MFN1\_1, astrocyte\_data\$MFN1\_2, astrocyte\_data\$MFN1\_4, astrocyte\_data\$WT\_1, astrocyte\_data\$WT\_3, astrocyte\_data\$WT\_4, astrocyte\_data\$MFN1, astrocyte\_data\$WT, col = 3:10, notch = T, outline = F, main = 'BOXPLOT', xaxt = 'n', xlab = 'Wild-type mice at different time points', ylab = 'FACS-enriched astrocytes', ylim = c(10,50), xlim = c(0,12))

## BOXPLOT

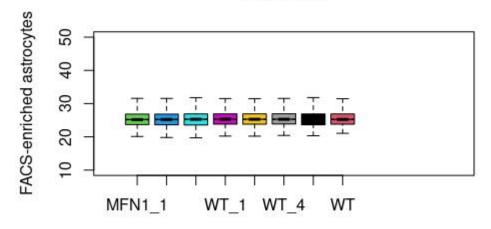


## Wild-type mice at different time points

#### 7. I want to add axis

axis(side = 1, at = c(1,2,3,4,5,6,7,8), labels = c('MFN1\_1', 'MFN1\_2', 'MFN1\_4', 'WT\_1', 'WT\_3', 'WT\_4', 'MFN1', 'WT'))

# **BOXPLOT**

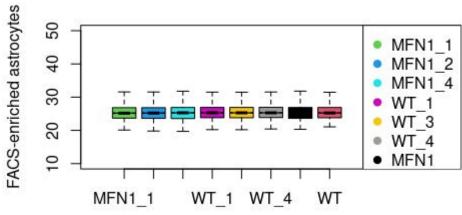


Wild-type mice at different time points

#### 8. Finally I will add legend

legend('topright', legend = c('MFN1\_1', 'MFN1\_2', 'MFN1\_4', 'WT\_1', 'WT\_3', 'WT\_4', 'MFN1', 'WT'), col = 3:10, pch = 19)



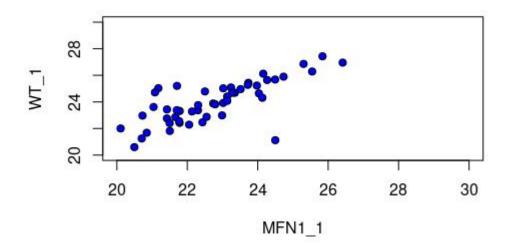


Wild-type mice at different time points

9. I want to create a scatterplot of MFN1\_1 vs WT\_1. I will color it black with a background of blue.

plot(x = astrocyte\_data\$MFN1\_1[1:50], y = astrocyte\_data\$WT\_1[1:50], col = 'black', pch = 21, bg = 'blue', main = 'MFN1\_1 vs WT\_1', xlab = 'MFN1\_1', ylab = 'WT\_1', xlim = c(20,30), ylim = c(20,30))

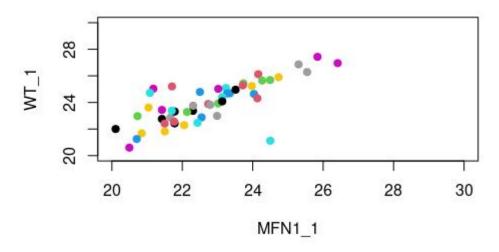
# MFN1\_1 vs WT\_1



10. I want to color by gene type.

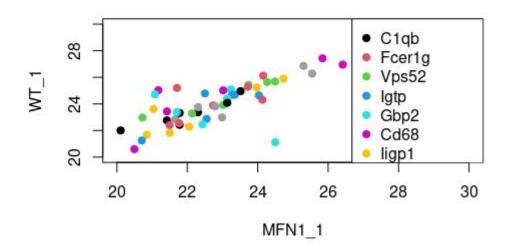
plot(astrocyte\_data\$MFN1\_1[1:50], astrocyte\_data\$WT\_1[1:50], col = 1:50, pch = 19, main
= 'MFN1\_1 vs WT\_1', xlab = 'MFN1\_1', ylab = 'WT\_1', xlim = c(20,30), ylim = c(20,30))

# MFN1\_1 vs WT\_1



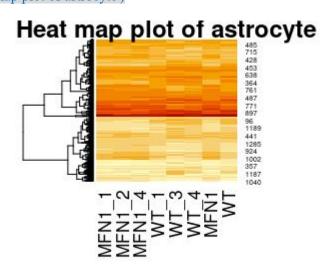
11. Let me add legend to my plot legend('topright', legend = c(astrocyte data\$T..Gene.names[1:50]), pch = 19, col = 1:50)

# MFN1\_1 vs WT\_1



## 12. I plot the heatmap next.

heatmap(as.matrix(astrocyte\_data[2:9]), Colv = NA, scale = 'col', margins = c(10,10), main = 'Heat map plot of astrocyte')



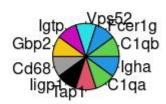
#### 13. Let me now plot Pie Charts

table(astrocyte\_data\$T..Gene.names[1:10])
item <- unique(astrocyte\_data\$T..Gene.names[1:10])
itemCount <- as.vector(table(astrocyte\_data\$T..Gene.names[1:10]))

#### 14. I will start by plotting a simple pie chart for astrocyte species

pie (x = itemCount, labels = item, radius = 0.5, col = 11:20, main = 'Pie chart for astrocyte species')

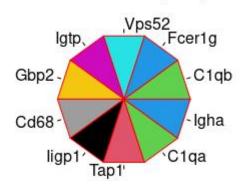
# Pie chart for astrocyte species



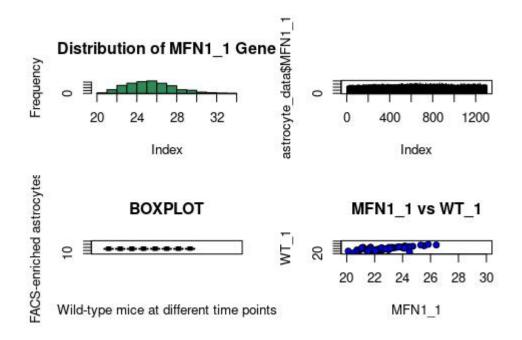
# 15. Next, let me add edges and color my borders pie (x = itemCount, labels = item, radius = 1.0, edge = T, border = 'red', col = 11:20, main =

'Pie chart for astrocyte species')

# Pie chart for astrocyte species



16. Let's practice how to arrange some of these plots as we usually see in publications par(mfrow = c(2,2))



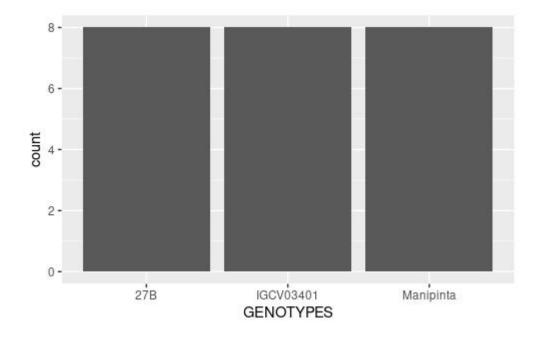
# # LET US NOW GO FULLY INTO GGPLOTS

17. We are going to work with a new dataset

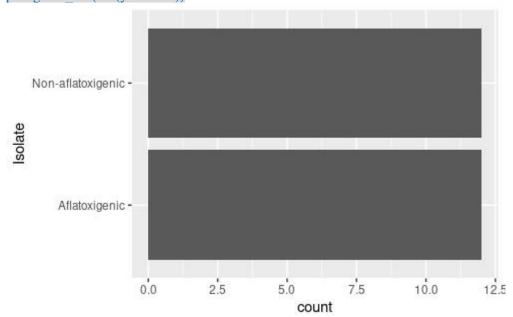
About data set: Growth and Toxigenicity of A. flavus on Resistant and Susceptible Peanut Genotypes. This study seeks to determine the reaction of peanut genotypes to Aflatoxigenic and non-aflatoxigenic A. flavus inoculation and also determine the mechanisms of their resistance. It was established that non-aflatoxigenic A. flavus grows faster than aflatoxigenic A. flavus. There was no significant difference in the incidence and severity of the A. flavus resistant genotypes (L027B and ICGV-03401) however, there were significant differences between resistant genotypes and the susceptible check Manipinta. This study also confirmed that non-aflatoxigenic A. flavus inoculation did not lead to aflatoxin production. Non-aflatoxigenic A. flavus identified could serve as a good biocontrol against aflatoxin contamination under field conditions. Additionally, peanut genotypes with resistance to post-harvest aflatoxin accumulation will resist the growth of A. flavus and subsequent aflatoxin accumulation.

# 18. Install ggplot2 install.packages("ggplot2") library(ggplot2)

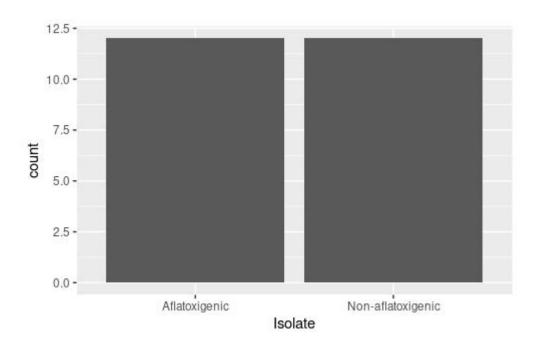
- 19. Import .csv file in R
  aflatoxin\_data <- read.csv(file.choose())
  aflatoxin\_data
- 20. Start with defining the base data for ggplot pl <- ggplot(data = aflatoxin data)
- 21. To the base, I will add what I want to plot and color pl + geom\_bar(aes(x=GENOTYPES))



# 22. I will plot frequency of the data from a single column pl + geom bar(aes(y=Isolate))



# 23. Let me flip the coordinates pl + geom\_bar(aes(y=Isolate))+coord\_flip()

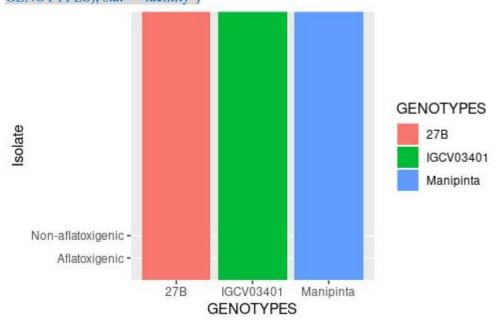


24. See the frequency of each variable within the species. Install.package(reshape2). Melt the data to have something understandable by ggplot better.

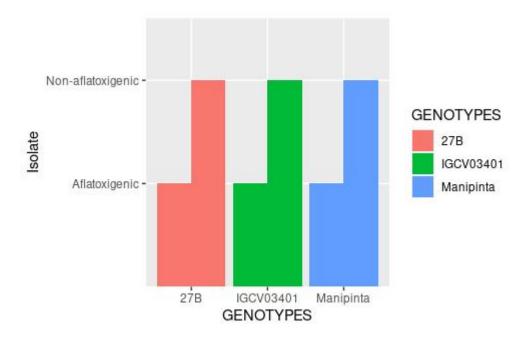
install.packages("reshape2")
install.packages("Rcpp")
library(reshape2)
melted aflatoxin data <- melt(aflatoxin data)</pre>

25. I want to produce a ttacked/continuous bar plot for genotypes.

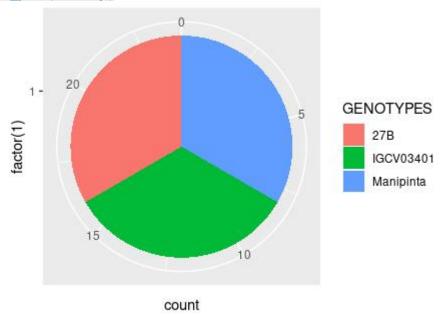
ggplot(data = aflatoxin\_data) + geom\_bar(aes(x=GENOTYPES, y=Isolate, fill = GENOTYPES), stat = "identity")



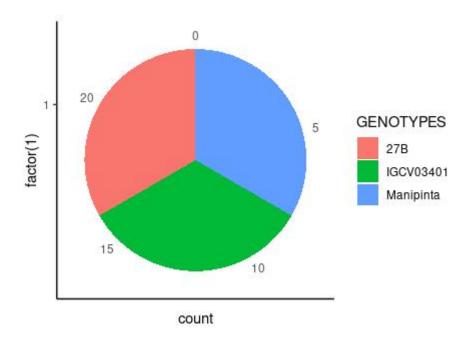
26. Let me produce a multiple bar plot ggplot(data = aflatoxin\_data) + geom\_bar(aes(x=GENOTYPES, y=Isolate, fill = GENOTYPES), stat = "identity", position = 'dodge')



# 27. Let me plot a simple pie chart and label by genotypes ggplot(data = aflatoxin\_data) + geom\_bar(aes(x=factor(1), fill = GENOTYPES), width = 1) + coord\_polar(theta = 'y')



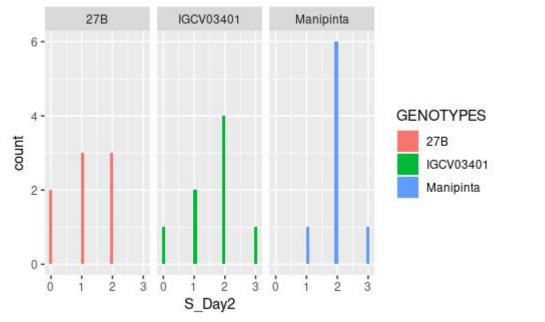
28. I will add a theme of my choice ggplot(data = aflatoxin\_data) + geom\_bar(aes(x=factor(1), fill = GENOTYPES), width = 1) + coord\_polar(theta = 'y') + theme\_classic()



# 29. Let me plot a simple histogram for aflatoxin\_data

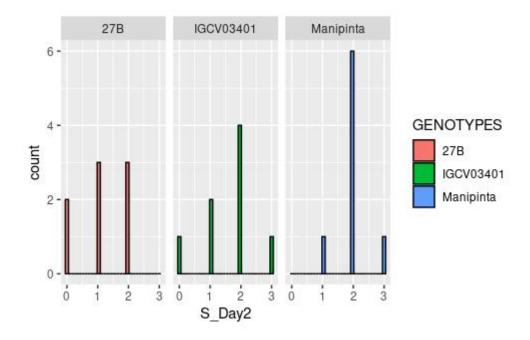
pl <- ggplot(data = aflatoxin data)

pl + geom histogram(aes(x=S Day2, fill = GENOTYPES)) + facet grid(. ~GENOTYPES)

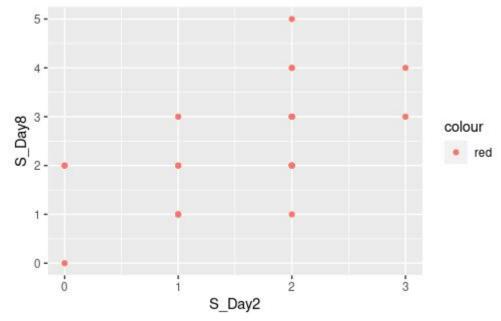


30. I will add a color parameter

pl + geom\_histogram(aes(x=S\_Day2, fill = GENOTYPES), color = "black") + facet\_grid(. ~GENOTYPES)



31. Let me construct a Scatter plot of S\_Day8 against S\_Day2 with red color pl + geom\_point(aes(x = S\_Day2, y = S\_Day8, color = 'red'))



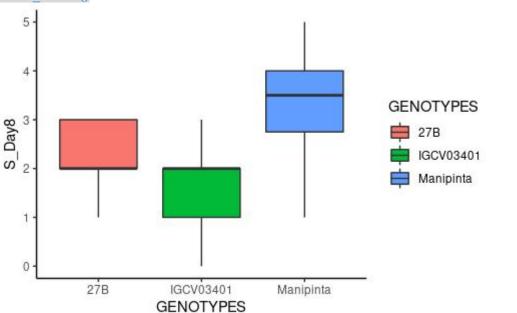
32. I will Color by Genotype (quite automated)

pl + geom\_point(aes(x = S\_Day2, y = S\_Day8, color = GENOTYPES)) + theme\_bw() + ggtitle(label = 'Scatterplot', subtitle = 'Days')

# Scatterplot Days GENOTYPES 27B IGCV03401 Manipinta

#### 33. I will construct a boxplot plus a theme

pl + geom\_boxplot(notch = F, aes(x= GENOTYPES, y = S\_Day8, fill = GENOTYPES)) + theme classic()

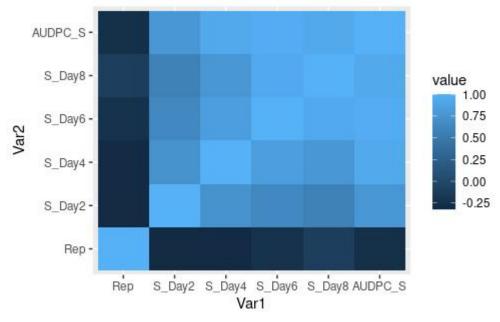


34. Heat map plotting. I will use melting methods to create heat map with ggtiles. meltCorData <- melt(cor(aflatoxin\_data[3:8]))

# 35. I start by setting my new ggplot

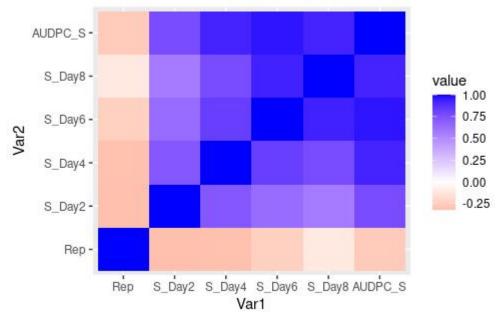
hm <- ggplot(data = meltCorData)

hm + geom tile(aes(x = Var1, y = Var2, fill = value))



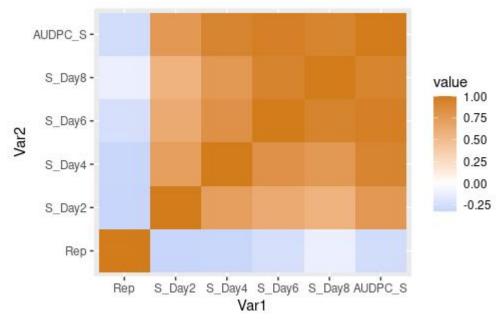
## 36. Let me start changing colors in heatmaps

hm + geom\_tile(aes(x = Var1, y = Var2, fill = value)) + scale\_fill\_gradient2(low = 'red', high = 'blue')



#### 37. I will also use hex code

hm + geom\_tile(aes(x = Var1, y = Var2, fill = value)) + scale\_fill\_gradient2(low = '#1687ee', high = '#d27C1e')



38. Let me change my x and y labels

hm + geom\_tile(aes(x = Var1, y = Var2, fill = value)) + scale\_fill\_gradient2() + xlab('First Variable') + ylab('Second Variable')

