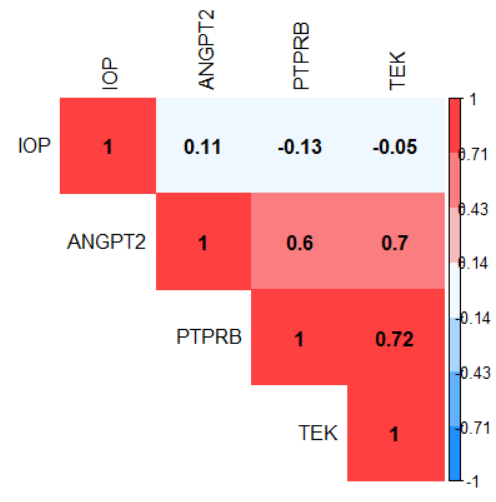
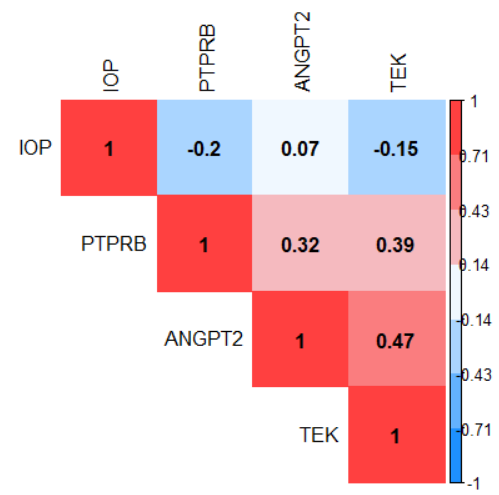
**Report date: March 22 Heatmap and Distribution No Normalization (Real Counts)**

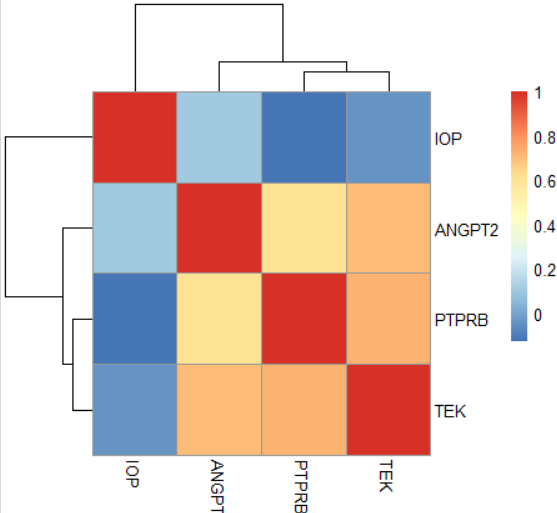
**Correlation method: Pearson**



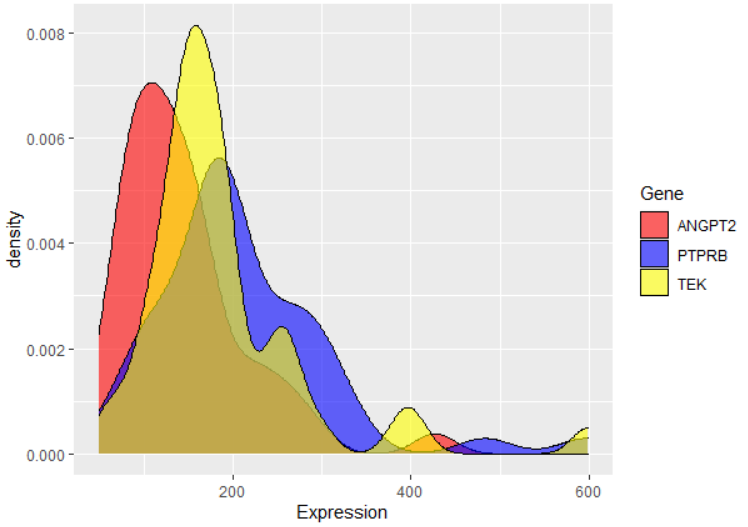
**Correlation method: Spearman**



**Clustering**

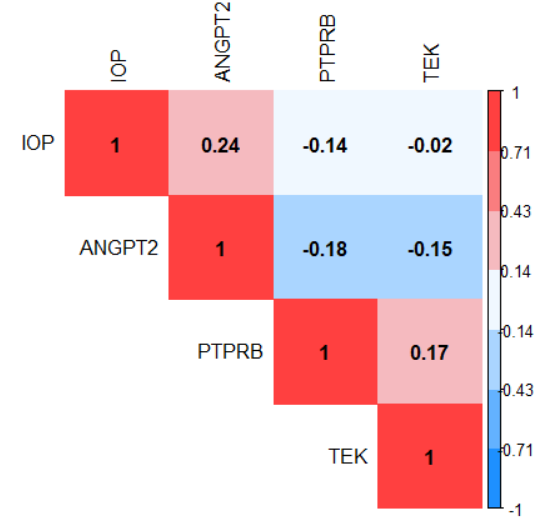


**Distribution**

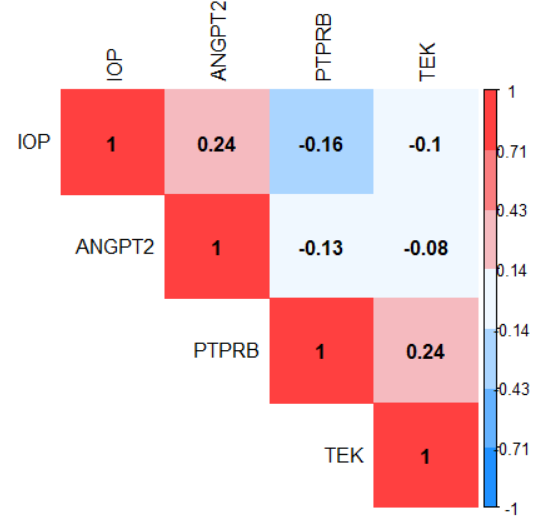


**Report date: March 22 Heatmap and Distribution Normalization method: Log2**

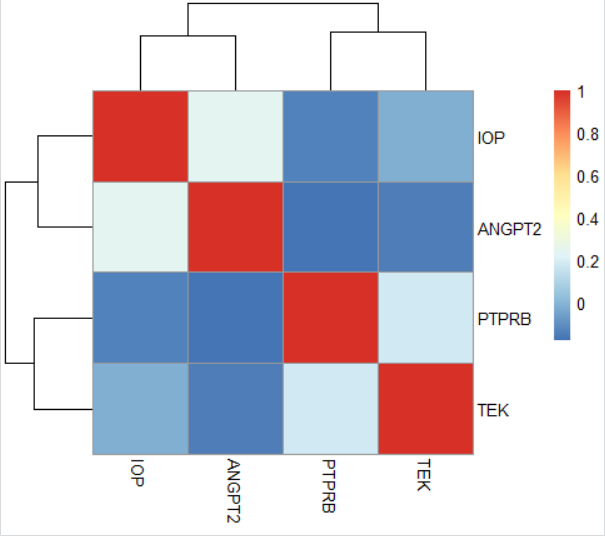
**Correlation method: Pearson**



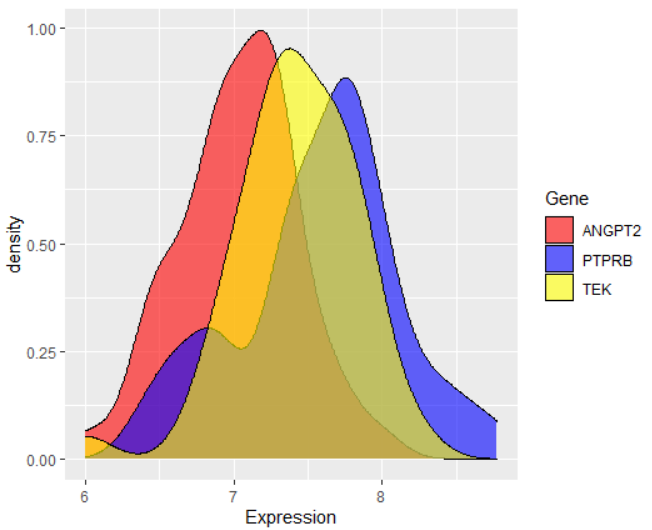
**Correlation method: Spearman**



**Clustering**

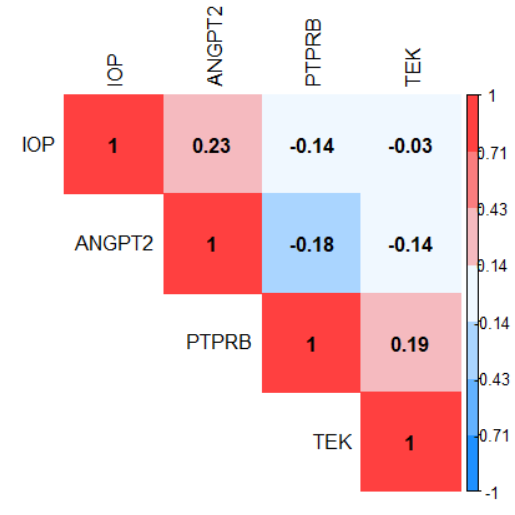


**Distribution**

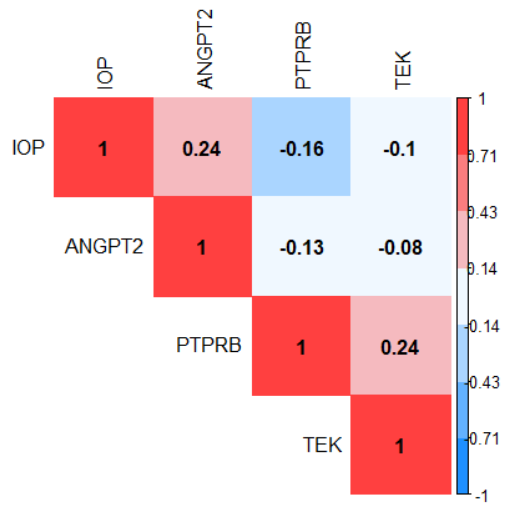


**Report date: March 22 Heatmap and Distribution Normalization method: vst**

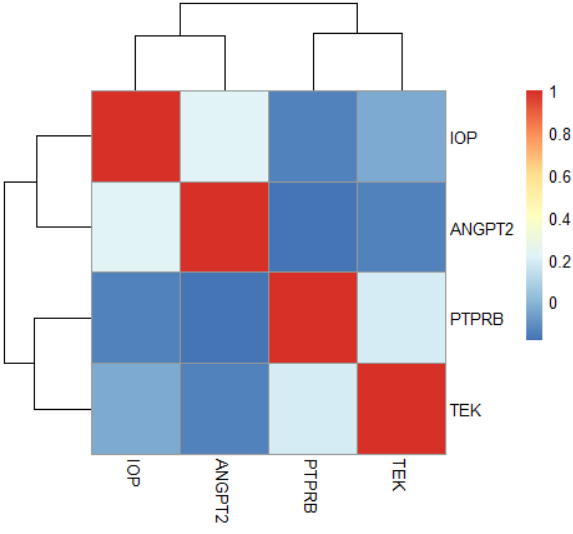
**Correlation method: Pearson**



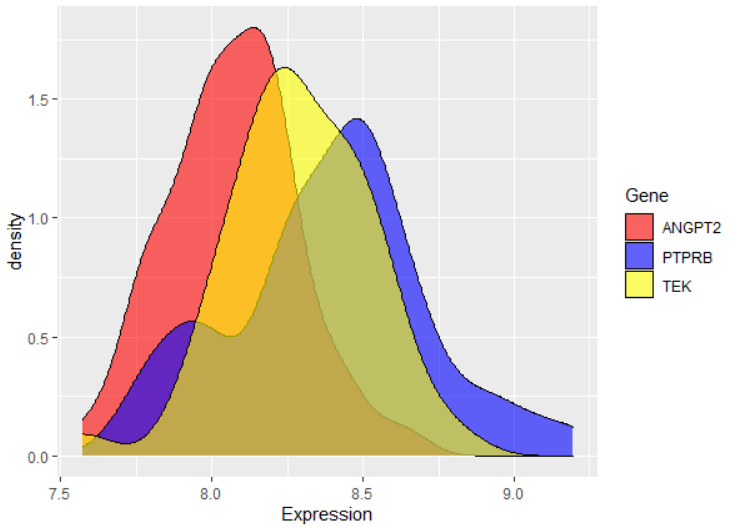
**Correlation method: Spearman**



**Clustering**

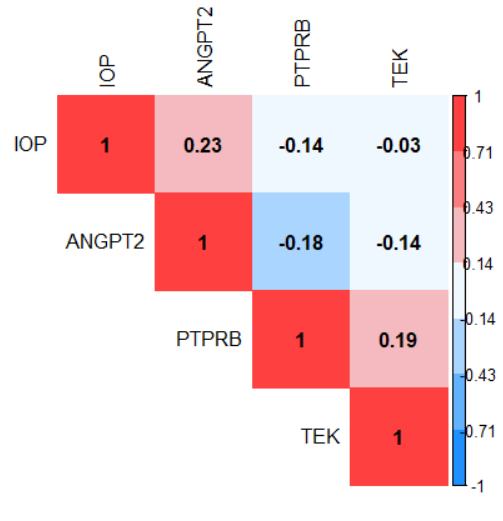


**Distribution**

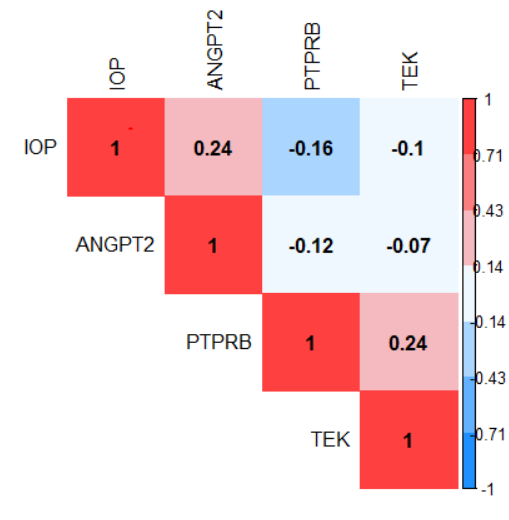


**Report date: March 22 Heatmap and Distribution Normalization method: rlog**

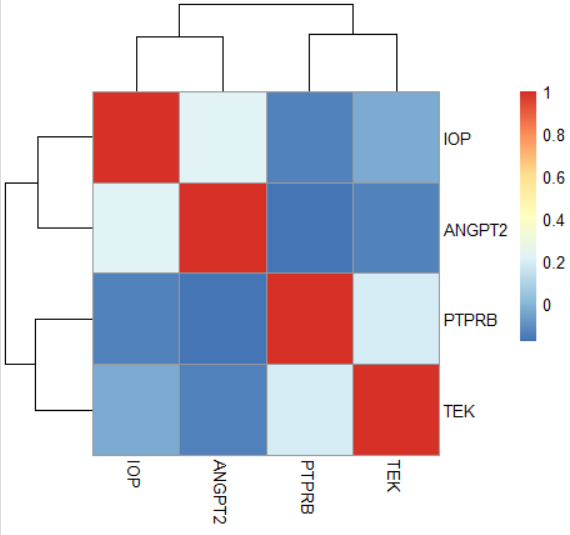
**Correlation method: Pearson**



**Correlation method: Spearman**



**Clustering**



**Distribution**

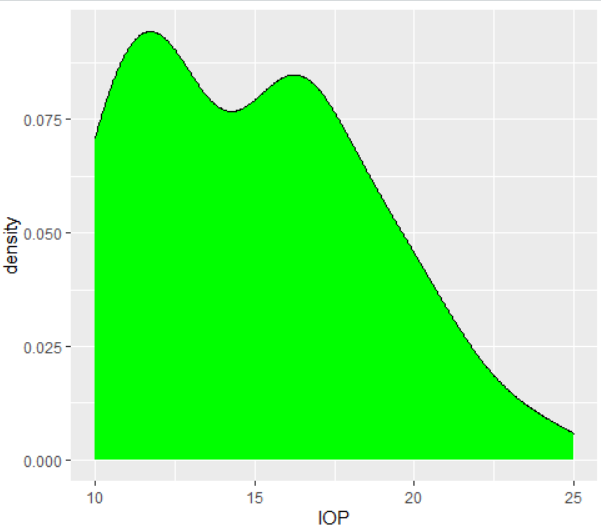


**Heatmap figures results**

Based on the Clustering and Heatmap figures, I found that two genes **PTPRB** and **TEK** act similarly. They have a negative correlation with **IOP** and **ANGPT2**. Also, **IOP** has a weak positive relationship with **ANGPT2**. Compare three normalization methods: **Log2** (Log2 Normalized Counts Transformation), **rlog** (Regularized Log Transformation), and **vst** (Variance Stabilizing Transformation), and based on the distribution plots of three genes, it shows that **rlog** normalization gives good performance for our analysis in this step. Therefore, I consider the **rlog** function as a normalization method.

Also, between Pearson (linear correlation) method and Spearman (linear-skewed correlation) method, I found that the probability density function of TEK (yellow) is between ANGPT2 (red) and PTPRB (blue). It means that there is an important role or relation between ANGPT2 and TEK. I think Spearman correlation is better than Pearson and states that ANGPT2 and TEK are almost uncorrelated. I mean, I believe that ANGPT2 and PTPRB have a negative relationship, PTPRB and TEK have a positive correlation, and ANGPT2 and TEK are uncorrelated.

Based on the density function of IOP, in the figure below, I found that **maybe** its distribution is not normal, and I need to normalize it first, then compare with other three genes. There are several way to check the normality of a distribution like plot the probability density function (like figure below), plot the quartile-quartile plot (QQplot), and Shapiro-Wilk normality test. Among them, I preferred the confident Shapiro-Wilk normality test. In the output of this test, the p-value > 0.05 implying that the distribution of the data are not significantly different from normal distribution. In other words, I can assume the normality for p-value > 0.05. After run this test for three normalized genes and IOP data, I found the for sure the IOP data is not normal and need to normalization. In fact, it is right skewed data. Common transformations of this data include **square root, cube root, and log.**



data: corrTable$ANGPT2

W = 0.9904, p-value = 0.9686 -------- as p-value > 0.05 then ANGPT2 is normal.

data: corrTable$PTPRB

W = 0.97918, p-value = 0.587 -------- as p-value > 0.05 then PTPRB is normal.

data: corrTable$TEK

W = 0.98158, p-value = 0.6841 -------- as p-value > 0.05 then TEK is normal.

data: corrTable$IOP

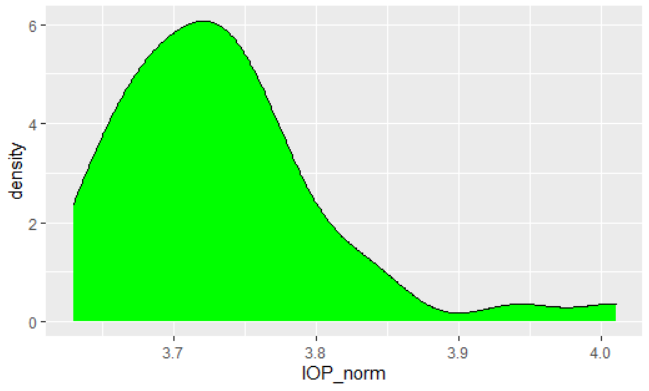
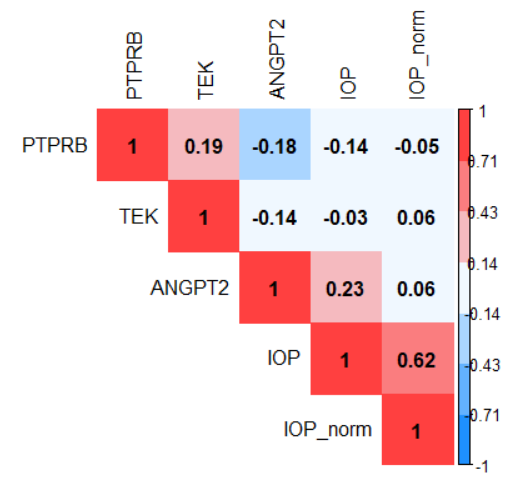
W = 0.93549, p-value = 0.01474 ------- as p-value < 0.05 then IOP is not normal.

1. **Normalization of IOP using rlog**

First, I tried to normalize the IOP with other 32883 genes and used logarithmic geometry mean method. However, after checking the Shapiro-Wilk normality test, I found that it is not normal again. It has some skewed in the righte side. So, as I thought, this is not a good method for normalization.

data: corrTable$IOP\_norm

W = 0.88815, p-value = 0.0004137 ------- as p-value < 0.05 then IOP\_norm is not normal.

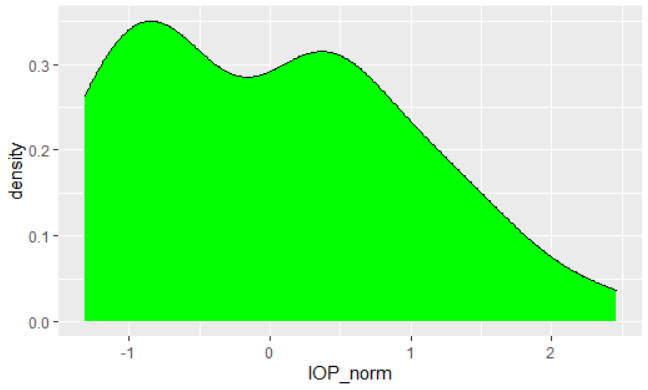
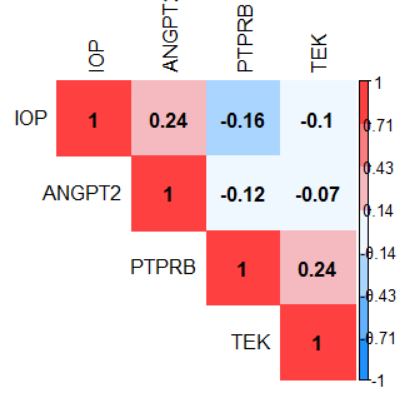
1. **Normalization of IOP using standard method**

Then uses the standard method for normalization of IOP data to convert its distribution into the normal distribution.

(x - mean(x)) / std(x)

data: corrTable$IOP\_norm

W = 0.93549, p-value = 0.01474 ------- as p-value < 0.05 then IOP\_norm is not normal.

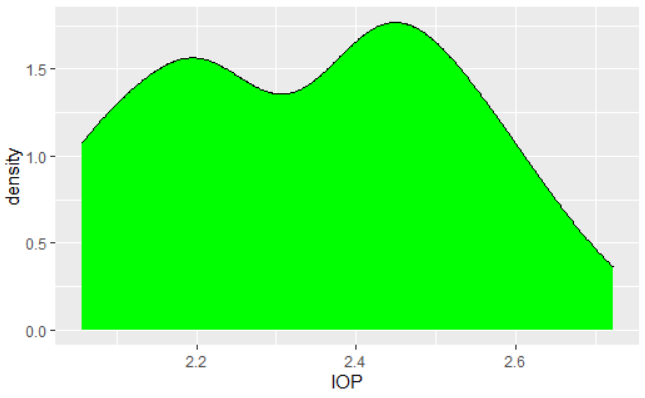
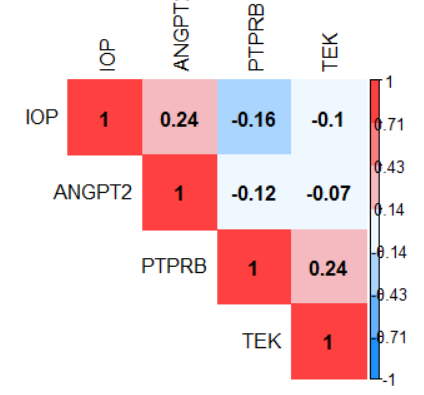
 

1. **Normalization of IOP using Box-Cox power transformation**

Lambda = -0.1

data: corrTable$IOP

W = 0.93785, p-value = 0.01795 ------- as p-value < 0.05 then IOP is not normal.

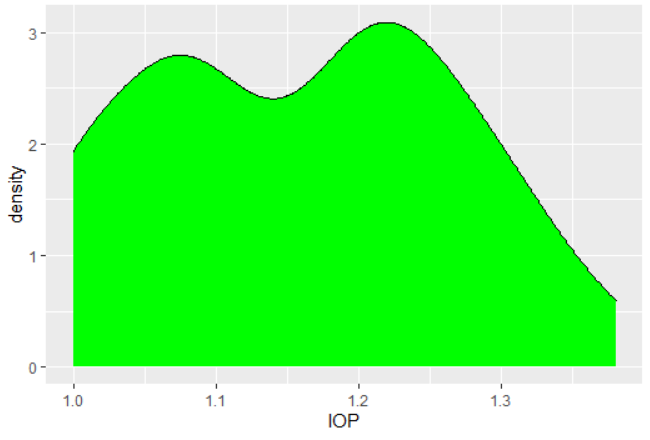
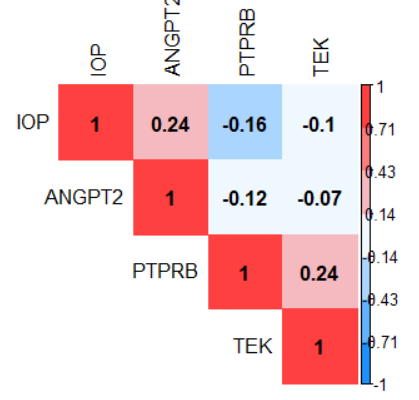
 

1. **Normalization of IOP using Log transformation**

Log(x), Log2(x), Log10(x)

data: corrTable$IOP

W = 0.93874, p-value = 0.01934 ------- as p-value < 0.05 then IOP is not normal.

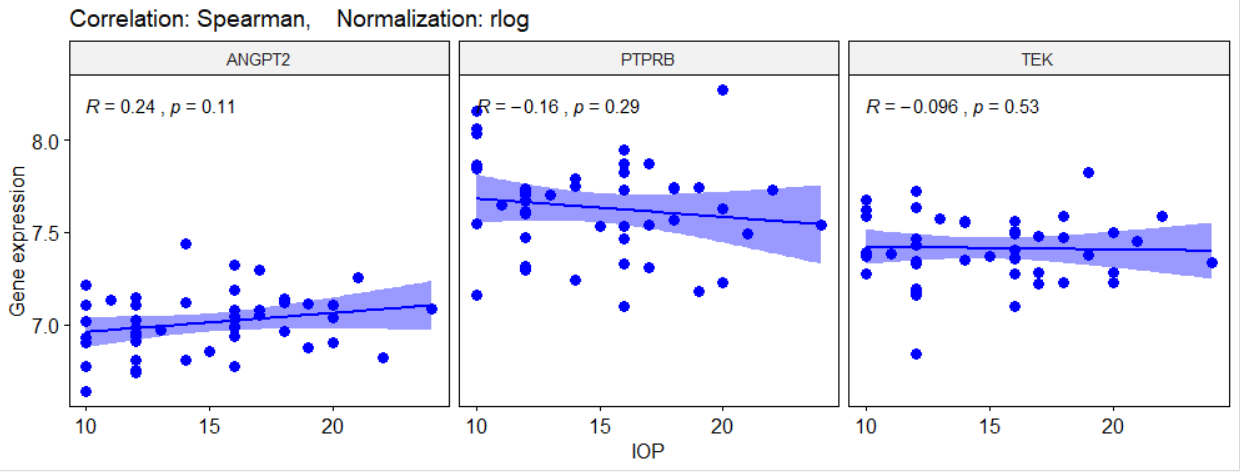
 

Among these four normalization methods, none of them cannot convert the distribution of IOP to a good normal distribution. However, I have choosed the **Log10 transformation normalization**, as its correlation analysis’ results did not changed. Also, the density of IOP is semi-normal compare to other methods.

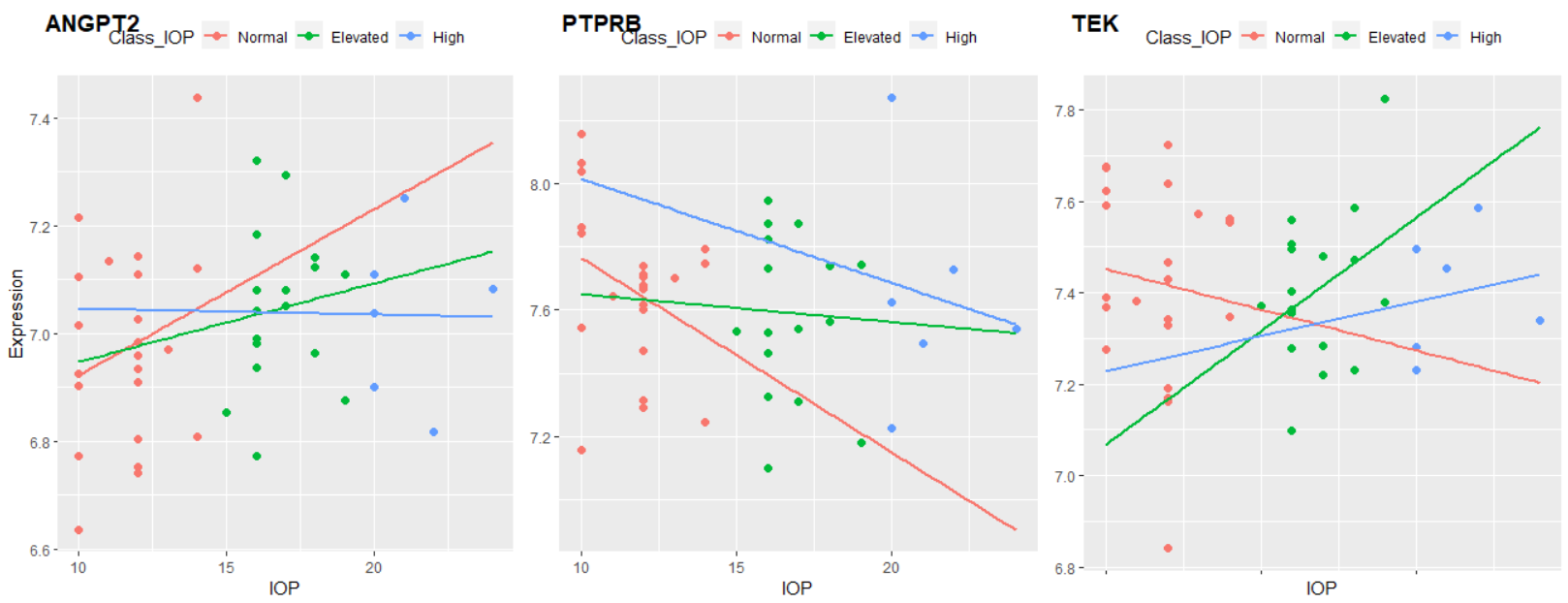
Therefore, I have considered **rlog** as a normalization method for genes and **Log10** as a normalization method for IOP. Aslo, considered **Spearman** as a correlation method for further analysis.

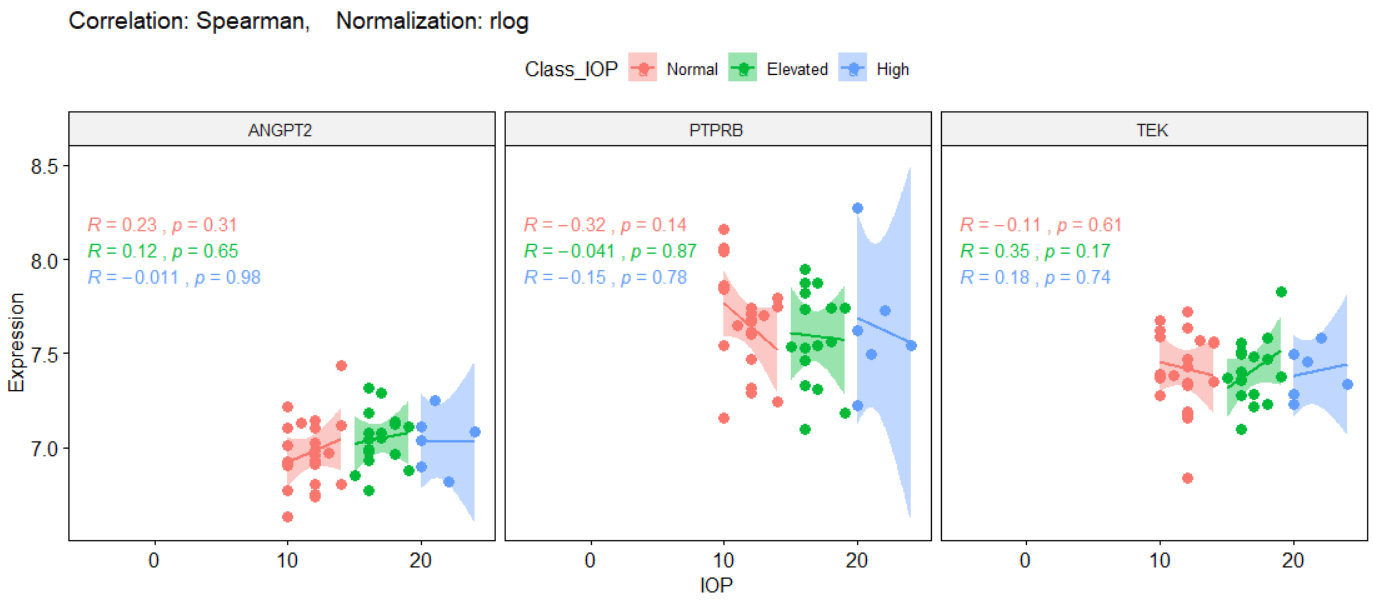
**Correlation between IOP and ANGPT2**

In this section, I want to calculate the **Spearman** correlation of IOP (and all subgroups of IOP) with ANGPT2 gene expression (based on the **rlog** normalization method). Based on the correlation coefficient R and P values in the figure below, I found that IOP has a positive relationship with ANGPT2 and a negative relationship with PTPRB and almost not correlated with TEK. However, none of them are significantly correlated. Based on the best R and P-values, I focused on the week relationship of IOP and ANGPT2.

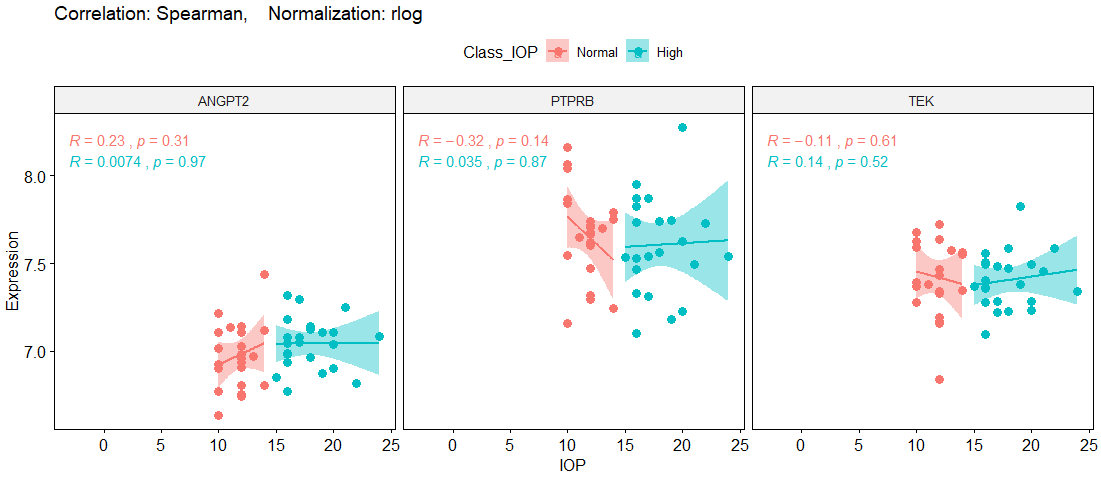


In the two figures below, I plot the correlation of three genes with three subgroups of IOP. As I can see, ANGPT2 has the highest positive correlation with Normal\_IOP (R = 0.23) subgroup. Also, PTPRB has the highest negative correlation with Normal\_IOP (R = -0.32) subgroup. It means that the Normal subgroup has a major role in the IOP feature.





Although I am looking for a high correlation between the High\_IOP subgroup and ANGPT2, however, as there are low samples in the high\_IOP subgroup, it is not a significant relationship. Therefore, I decided to classified the IOP feature into two subgroups: Normal and High. In the figure below, I look for the correlation of IOP subgroups with three genes. Again, here there are more samples with High\_IOP; around 50% of samples (23 samples). However, the High\_IOP subgroup is not correlated with ANGPT2 and PTPRB. I mean, Normal\_IOP has important role in the correlation of IOP with ANGPT2 or PTPRB.



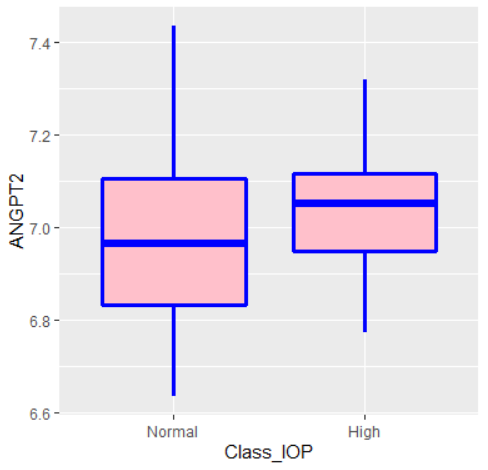
As a conclusion and based on figures above, ANGPT2 has week correlation, but not significant, with whole IOP data (R=0.24, P-value=0.11), which related to the Normal\_IOP (R=0.23, P-value=0.31). However, the ANGPT2 gene has the maximum (week) correlation with IOP among other genes. Therefore, we need to look at this gene deeply. Maybe other features like Age, Sex, or Batch give some clues. To discuss statistically about the correlation between ANGPT2 and IOP subgroups, I did t-test and ANOVA analysis too.

**Analysis of t-test for IOP subgroups**

In this analysis, I want to test that the mean of expression level of ANGPT2 is equal in two separated groups of Normal\_IOP and High\_IOP? (with almost same sample sizes). If not, then the expression level of ANGPT2 in each subgroup of IOP are not similar together, and the expression level are varied in different subgroupe of IOP.

**H0:** The mean of ANGPT2 expression level in Normal\_IOP group is equal to the mean of ANGPT2 expression level in High\_IOP group.

**Ha:** Both means are not equal.



The results of two sided t-test with 95% confidential is:

t.test(ANGPT2 ~ Class\_IOP, data=corrTable\_IOP2, mu=0, alt="two.sided", conf=0.95, var.eq=F, paired=F)

Welch Two Sample t-test

data: ANGPT2 by Class\_IOP

t = -1.4124, df = 39.885, p-value = 0.1656

alternative hypothesis: true difference in means is not equal to 0

95 percent confidence interval:

-0.1705142 0.0302361

sample estimates:

mean in group Normal mean in group High

6.973136 7.043275

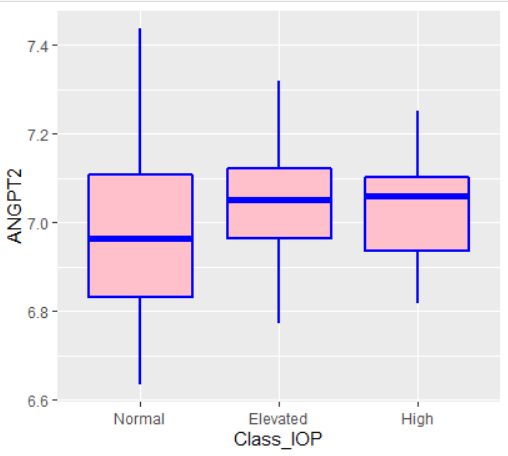
The P-value of the test is 0.1656, which is greater than the significant level alpha = 0.05. Therefore, I cannot reject the Null-Hypothesis and accept the alternative hypothesis. I conclude that the average expression level of ANGPT2 gene in samples with Normal\_IOP is not significantly different with the samples with High\_IOP; the expression level of ANGPT2 gene are almost same in two gropus. It means that different IOP subgroups have not important role in the expression level of ANGPT2.

**Analysis of ANOVA for IOP subgroups**

As in the real state, we have three subgroups of IOP, so we need to run ANOVA analysis insteed of t-test for them.

**H0:** The mean of ANGPT2 expression level in all three groups of IOP (Normal, Elevated, and High) is equal together.

**Ha:** They are not equal.



Anova\_results <- aov(ANGPT2 ~ Class\_IOP, data=corrTable)

summary(Anova\_results)

Df Sum Sq Mean Sq F value Pr(>F)

Class\_IOP 2 0.0561 0.02806 1 0.377

Residuals 42 1.1789 0.02807

F(2,42) = 1, P-value>0.05

The P-value is greater thatn 0.05, so the null hypothesis is not rejected again. Therefore, I cannot claime that the expression level of ANGPT2 is significantly varied in three different IOP groups.

**Partial-Correlation between IOP and ANGPT2 based on Age feature**

In this section, I am going to calculate the correlation between IOP and ANGPT2 based on different Age groups. I classified samples into four subgroups: **Adolescent** (113<=Age<135), **Adult** (135<=Age<154), **Middle\_Aged**(154<=Age<183), and **Aged** (183<=Age<=346). Then, plot the correlation based on each subgroups:

****

346

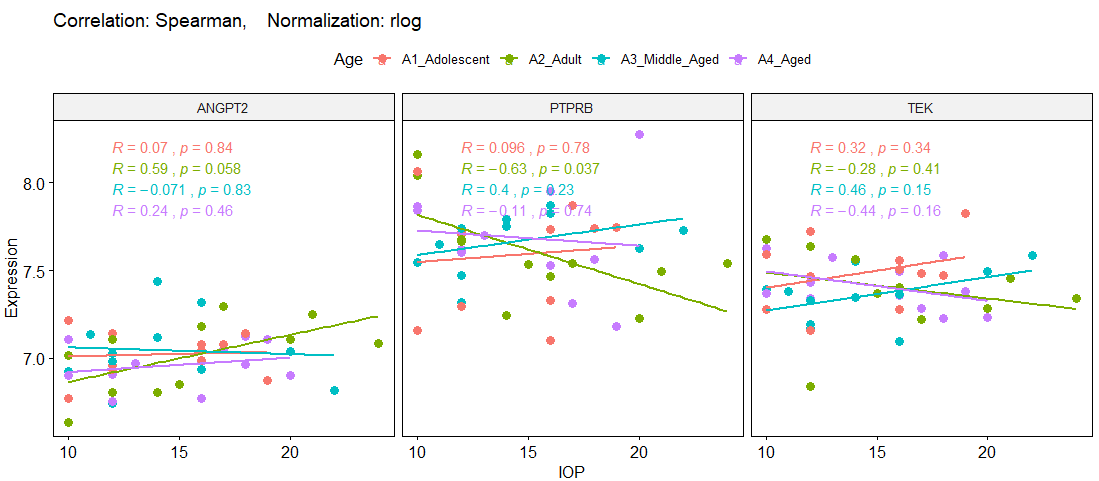
238

183

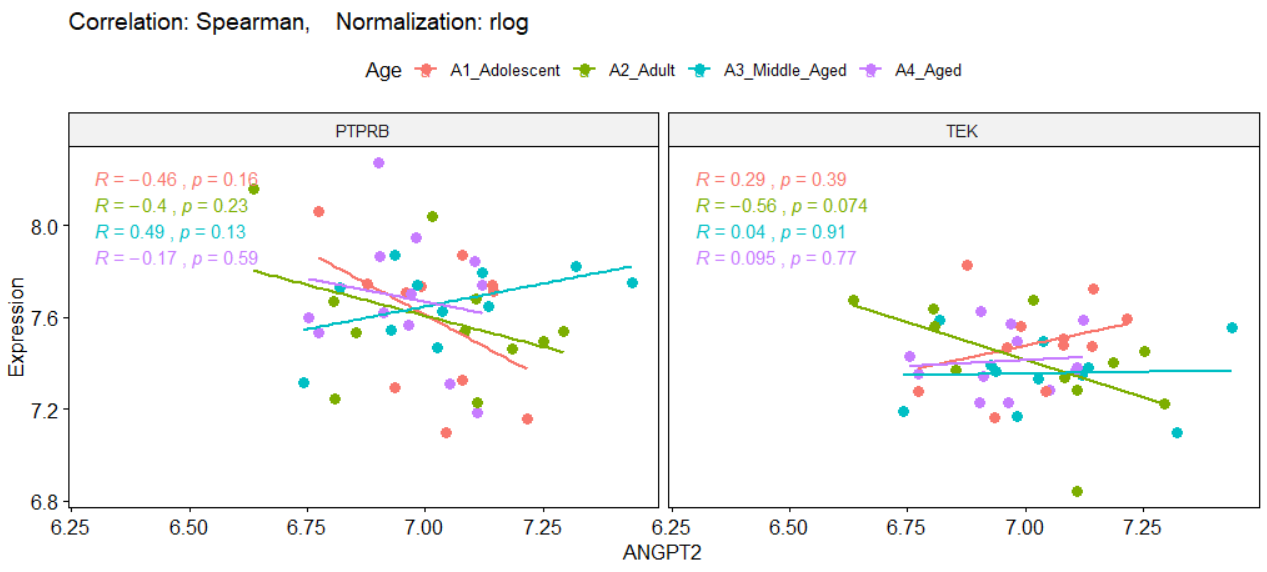
154

135

113



**Good news:** Based on the results, I can see there is a significant relationship in the Adult subgroup between IOP and ANGPT2 (R=+0.59, P-value=0.05), and PTPRB (R=-0.63, P-value=0.03). Also, there is semi-significant relationship between ANGPT2 and TEK genes in this subgroup (Adult); figure below.

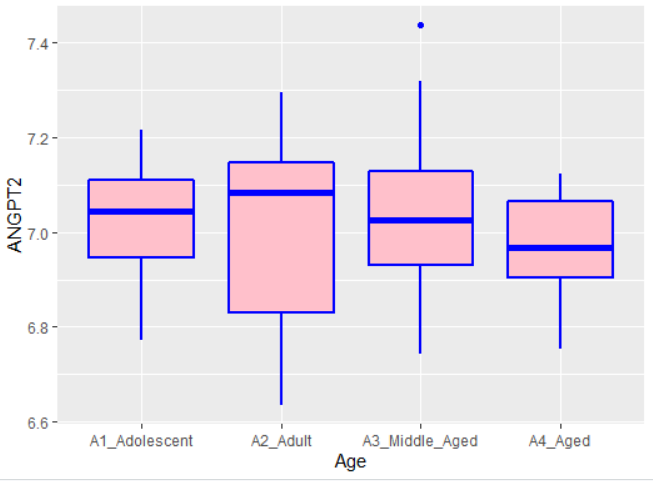


**Analysis of ANOVA for Age subgroups**

There is four subgroups of Age, and I run ANOVA analysis in for ANGPT2 expression along these subgroups.

**H0:** The mean of ANGPT2 expression level in all four subgroups of Age is equal together.

**Ha:** They are not equal.



Anova\_results <- aov(ANGPT2 ~ Age, data=corrTable)

summary(Anova\_results)

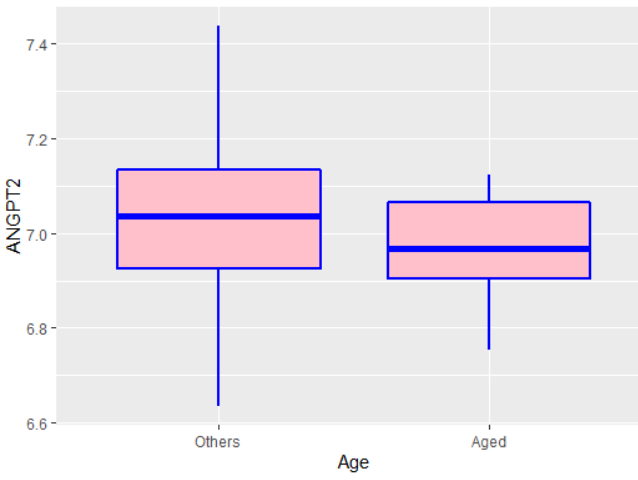
Df Sum Sq Mean Sq F value Pr(>F)

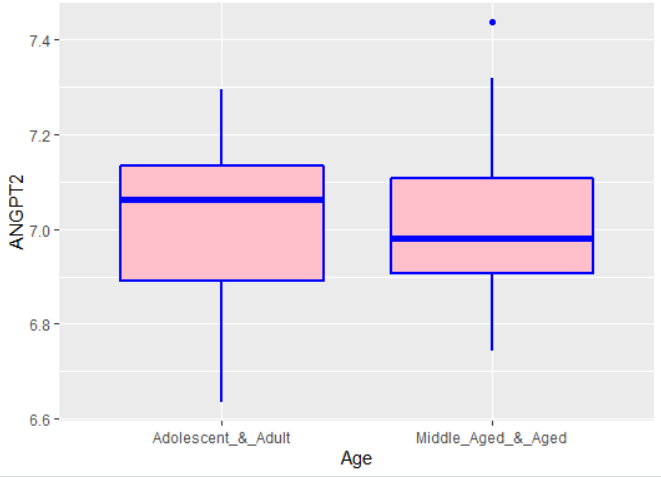
Age 3 0.0415 0.01384 0.476 0.701

Residuals 41 1.1935 0.02911

F(3,41) = 0.476, P-value > 0.05

The P-value is greater thatn 0.05, so the null hypothesis is not rejected again. Therefore, I cannot claime that the expression level of ANGPT2 is significantly varied in four different Age groups.

Then, I decided to devide the Age into just two subgroups as belows. However, in none of them I did not get significant P-value in ANOVA analysis.



Anova\_results <- aov(ANGPT2 ~ Age, data=corrTable\_Age2) Anova\_results <- aov(ANGPT2 ~ Age, data=corrTable\_Age2)

summary(Anova\_results) summary(Anova\_results)

Df Sum Sq Mean Sq F value Pr(>F) Df Sum Sq Mean Sq F value Pr(>F)

Age 1 0.003 0.002999 0.105 0.748 Age 1 0.0361 0.03613 1.296 0.261

Residuals 43 1.232 0.028651 Residuals 43 1.1989 0.02788

F(1,43) = 0.105, P-value > 0.05 F(1,43) = 1.296, P-value > 0.05

**Partial-Correlation between IOP and ANGPT2 based on Sex feature**