

# Imputor User Guide

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## 1. Overview

IMPUTOR is a software program written in the Python language for the purpose of identifying miscalled bases in sequence or variant data. Advances in next-generation sequencing have provided researchers with a wealth of data, but the data generated has proven variable in its fidelity to the original sequence ([Wall et al. 2014](#)).

Numerous software “pipelines” have been constructed in order to process data, with one important step being the assessment and correction of chemical or mechanical errors introduced by the NGS process. One of the challenges associated with analyzing NGS data comes from the amount of data missing from some NGS reads. To correct this problem, software pipelines and packages have been authored for the purposes of imputing missing data. In some cases, these methods have used reference sequences derived from other, slower and more reliable forms of sequencing, while some have also implemented methods to impute missing data via the principle of parsimony ([Poznik et al. 2013](#)).

## 2. Setup and installation

### 2.1. Python and its packages

IMPUTOR was written for Python 2.7 and requires a number of standard Python packages to run properly. Python distributions vary in what is installed; in the case where a needed package has not been installed, the recommended method for installing packages is the [PIP installer program](#). Following the given link should all the user to install all needed Python packages. The list of needed packages for this release of IMPUTOR is:

- biopython
- progressbar2

Two external maximum likelihood software programs can be used with IMPUTOR in order to construct phylogenetic tree from sequence. Please note that for these programs to work with IMPUTOR, they must be compiled and the executable placed in the same folder as IMPUTOR or elsewhere in the system's search path.

## 2.2. PhyML

1. Download and install PhyML for your system as per the [PhyML user guide](#).
2. Rename the resulting executable 'phym1' (no quotes).
3. Move your phym1 executable to a location that is part of your system's search path. On UNIX-like systems including macOS and Linux, you will usually want to add the line **export PATH="/your path/:\$PATH"** to the .bashrc or the .bash profile located in your home directory (where your path is the path to the directory that contains PhyML binary).

## 2.3. RAxML

1. Download and install RAxML for your system as described in the [RAxML user guide](#).
2. Compile the version best suited to your needs. Please see the RAxML docs about parallel computing, makefiles, etc.
3. Rename your compiled executable to 'raxmlHPC' (no quotes).
4. Move your raxmlHPC executable to a location that is part of your system's search path. On UNIX-like systems including macOS and Linux, you will usually want to add the line **export PATH="/your path/:\\$PATH"** to the .bashrc or the .bash profile located in your home directory (where your path is the path to the directory that contains RAxML binary).

### 3. PhyML

Unlike methods of maximum parsimony, maximum likelihood methods make use of the branch lengths of a proposed tree .

#### 3.1. How to install PhyML for use with Imputor

1. Download and install PhyML for your system as per the [PhyML user guide](#) .
2. Rename the resulting executable 'phym1' (no quotes).
3. Move your phym1 executable to a location that is part of your system's search path. On UNIX-like systems including macOS and Linux, you will usually want to add the line **export PATH="/your path/:\$PATH"** to the .bashrc or the .bash profile located in your home directory (where your path is the path to the directory that contains PhyML binary).

### 4. Input

IMPUTOR accepts input from two file types: FASTA files and VCF files. A reference sequence must also be input for either format—in the case of a FASTA file, the first sequence can simply be copied into a separate file. For VCF files, the sequence of the reference file is used to generate sequence from the defined variants inside the software, allowing either FASTA or VCF format output.

A phylogenetic tree is also necessary for the phylogeny-aware imputation performed by the software, and thus it is necessary either to import or generate such a tree.

IMPUTOR provides four options: phyloxml import, tree construction by parsimony using Biopython, maximum likelihood via the software PhyML or maximum likelihood via the software RAxML. Software options necessary to modify the output of the tree have been included in IMPUTOR, however for detailed descriptions of their functions the individual software packages' under manuals will be the user's best guide.

## 4.1. Input options

Imputor handles the following input arguments:

- **-file (required):** The raw input data, in FASTA or VCF format. Note that VCF format files must include the optional FORMAT column with the GT subheader, and include columns where individual samples are marked.
- **-ref (required):** The reference sequence, in either TXT or OBJ format. In the case of FASTA files, the first sequence can be copied into a separate reference sequence file for the purposes of generating variants.
- **-tree:** The method for importing or constructing a tree, with the following options:
  - **\*.xml:** Input of a file with extension .xml will invoke reading as a phyloxml format tree ([Han & Zmasek 2009](#)).
  - **PhyML:** Invokes the PhyML software ([Guindon et al. 2010](#)).
  - **RAxML:** Invokes the RAxML software ([Stamatakis 2014](#)).
- **-alpha:** The value of the gamma shape parameter used in PhyML. Default is  $e$ .
- **-boot:** The number of bootstrap replicates for PhyML. Please see the [PhyML user guide](#) for detailed instructions. Default is 100.
- **-rmodel:** The model type for RAxML. Please see the [RAxML user guide](#) for instructions. Default is GTRCAT.

## 5. RAxML

### 5.1. How to install RAxML for use with Imputor

1. Download and install RAxML for your system as described in the [RAxML user guide](#).
2. Compile the version best suited to your needs. Please see the RAxML docs about parallel computing, makefiles, etc.
3. Rename your compiled executable to 'raxmlHPC' (no quotes).
4. Move your raxmlHPC executable to a location that is part of your system's search path. On UNIX-like systems including macOS and Linux, you will usually want to add the line **export PATH="/your path/:\\$PATH"** to the .bashrc or the .bash profile located in your home directory (where your path is the path to the directory that contains RAxML binary).

## 6. Output

IMPUTOR provides output information to the console for the researcher to examine during run-time. After the run completes, a number of possible files can be written to storage, depending on the options chosen from the list in the next section.

### 6.1. Output options

- -out: The type of output file requested. Options are: fasta or vcf. Defaults to fasta.
- -outimpute: Write the list of imputed mutations to <inputfile minus extension> + "impute.txt". Defaults to true.

## 7. Imputation

A major challenge for Next Generation Sequencing is the separation of actual variation from errors that arise from the mechanical process of the method (DePristo et al. 2011). Imputation methods have been used to identify mistakenly called sites due to mechanical or laboratory error (Wang et al. 2012). These efforts are often made in order to “fill in” calls marked “missing” by the sequencing process (Lippold et al. 2014). One of the primary methods by which imputation is carried out for Next-Generation Sequencing (NGS) is by comparing NGS sequence reads to that of a set of reference sequences (Marchini & Howie 2010). An example of such a reference sequence set is Sanger sequencing, which is slower than NGS but has a lower error rate (Wall et al. 2014).

### 7.1. Imputation by parsimony

IMPUTOR imputes mutations for an input set of data via comparison of variants amongst near neighbors, via the principle of parsimony, wherein neighboring samples on a phylogenetic tree that are identical by descent (IBD) for a derived allele are unlikely to experience a reversion to the ancestral allele amongst one of their members. Under the principle of parsimony, originally introduced as the “principle of minimum evolution”, the course taken in evolutionary history is most likely to match the course that requires the fewest changes (Edwards & Cavalli-Sforza 1964). The principle of parsimony or minimum evolution has been applied via a phylogenetic tree for the purposes of imputing missing data (Poznik et al. 2013). In addition performing to this function, IMPUTOR searches for likely candidates for falsely negatively called mutations, which can appear in NGS as reversions aka back mutations. Previous studies imputing missing mutations have avoided introducing the possibilities for reversions due to their rarity (Wei et al. 2013).

One possible scenario for a false negative call is an apparent back mutation on a the phylogenetic tree. On such a tree, a back mutation will appear, from the point of view of any site if, when following its descendants we encounter a different allele, and then, continuing on the same descending path, encounter the original allele again **(Requeno & Colom 2016)**. The same logic holds in the reverse direction, that if while ascending toward the root of a phylogenetic tree, one finds a different allele than the target allele, after which, on continuing to ascend, one finds a reversion to the target allele. Back mutations are not impossible, but rare, since the occurrence of a mutation on a lineage must be compounded with that of another on a branch of that lineage that happens to revert to the ancestral allele.

According to the principle of parsimony, should a target site differ from its three nearest neighbors, and should those three neighbors possess the same allele, and furthermore that target site be found among more distant neighbors (implying a possible reversion), the more likely scenario requiring fewer mutations would be the one generating the putative reversion than the mutations necessary to separately generate the mutations in its non-matching near neighbors. The length of the private branch upon which the target sequence sits represents the proportion of changes in that sequence relative to the sequence size. That length, or distance, multiplied by the rate of mutation, is the likelihood that a reversion has truly occurred, since at a constant rate of mutation the branch implies time. A short private branch is less likely to have provided the opportunity for a true reversion to have occurred, and is thus more likely that an identified reversion is the result of NGS sequencing error or an error of some other form.

The decision rule for imputation involves each sequence and its closest neighbors on the constructed or imported phylogenetic tree. IMPUTOR searches site by site for each sample, iterating through the list of samples in random order and implementing the following decision rule:

1. For each target, iterated in random order:
  1. Step back toward the root of the tree collecting nearest siblings of the target
  2. Keep stepping back until root is reached or threshold number of siblings found (default = 3)
2. For each variant in the set of all variants in the input file:
  1. If the target is marked as missing:
    1. If the nearest two neighbors are the same and not missing, impute their state for the target (**Poznik et al. 2013**)
  2. If the target is not marked as missing:
    1. If the target does not match its three nearest neighbors and the three neighbors match each other and are non-missing
      1. If the target state is found farther away than the three nearest neighbors (i.e. a potential reversion (**Requeno & Colom 2016**))
        1. Calculate the likelihood that a back mutation could have occurred under these circumstances (Poisson):
        2. If the chance of back mutation multiplied by the given transition or transversion ratio does not exceed the user-supplied threshold chance (default = 0.05) impute the target to be the same as its neighbor

### 7.1.1. Probability of Mutation



The rate of point mutation has been assessed for various compartments of the human genome, e.g.  $8.71 \times 10^{-10}$  for much of the Y chromosome (Helgason et al. 2015).

The chance that a particular mutational event might occur is governed by the Poisson distribution. The probability of observing  $k$  events during a period of time is given by the probability mass function of the Poisson distribution:

$$P = (\lambda t)^k e^{-\lambda t} / k! \quad (1)$$

## 7.2. Imputation using a reference sequence (under development!)

In many studies involving NGS, output sequence from many individuals are compared to a reference sequence which has been generated by a slower method but one that generates sequence with a lower rate of error (Wang et al. 2012). The reference sequence does not necessarily occupy a particular position on the phylogenetic tree, such as a branch proximal to the root. Instead, such sequences are often generated from conveniently accessible sample material and used in subsequent studies for the sake of consistency. Thus the reference sequence does not need to contain information about the ancestral state of an allele—the reference might easily possess a derived allele.

The decision rule IMPUTOR uses when searching for false negatives is as follows:

1. If the target is marked as missing data at a site
  1. Locate the target's two nearest neighbors with non-missing data at the site
  2. If the two nearest neighbors match each other, impute their state for the target (essentially uses parsimony)
2. If the target is not marked as missing data at a site
  1. Locate the target's two nearest neighbors with non-missing data at the site
  2. If the target state matches the reference sequence but does not match its nearest two neighbors, and the two nearest neighbors match each other, impute the state of the neighbors for the target

## 8. References

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