# Introduction:

Advanced Prostate Cancer and Caveolin-1**:**

Prostate cancer is the second most commonly diagnosed cancer in men (Stewart & Wild 2014). While the primary tumour can be treated and removed efficiently resulting in almost 99% survival, patients inflicted with metastatic prostate cancer possess a reduced 5-year survival rate of 29.3% (SEER 2016). Bone metastasis is the most common complication derived from advanced prostate cancer formation which severely limits the survival outcome ([Bubendorf *et al.* 2000](#_ENREF_1)). This highlights the necessity to identify therapeutic targets and underlying biological phenomena that induce the metastatic phenotype.

Caveolin-1 has been linked to prostate cancer metastasis and has been a speculated biomarker for cancer progression ([Gumulec *et al.* 2012](#_ENREF_10); [Moon *et al.* 2014](#_ENREF_22); [Hayashi *et al.* 2015](#_ENREF_12)). This protein usually functions as a cholesterol transporter where its interaction with cytoplasmic protein, cavin-1, initiates the formation of specific lipid microdomains on the plasma membrane, called Caveolae ([Hill *et al.* 2008](#_ENREF_14); [Moon et al. 2014](#_ENREF_22)). These proteins are co-expressed and co-localised in healthy human tissue, however in the case of many cancer types only caveolin-1 is expressed ([Wu *et al.* 2011](#_ENREF_31); [Moumita *et al.* 2015](#_ENREF_23)). Increased proliferation, migration and differentiation are a result of the aberrant caveolin-1 expression, yet, the mechanism that links caveolin to these phenotypes is still actively being investigated ([Grande-García *et al.* 2007](#_ENREF_9); [Chatterjee *et al.* 2015](#_ENREF_2)).

Recent studies have shown that cavin-1 is capable of reversing the pro-metastatic action of caveolin-1 ([Moon et al. 2014](#_ENREF_22)). Cavin-1 expression inhibited prostate cancer PC3 cell proliferation, migration and anchorage-independent growth *in vitro*, and tumour growth, metastasis and angiogenesis *in vivo*. Mechanistically, cavin-1 expression altered the tumour microenvironment, including reduction of fibroblasts and secretion of IL-6 (Moon 2012). This reduced IL-6 secretion was determined to be through extracellular vesicle (EV) release as opposed to the classical secretion methods (Inder). Other proteins were also found to be differentially secreted via EVs, yet their role was not extensively studied. Hereby, in addition to the previous well-known methods of cancer progression, caveolin-1 appears to modulate extracellular vesicle (EV) mediated metastasis. Intriguingly, cavin-1 expression also attenuated the EV-mediated release of microRNA-148a, which was previously reported to mediate bone metastasis through osteoclastogenesis. These studies suggest that cavin-1 attenuates the pro-metastatic action of caveolin-1 by modulating EV microRNA content.

## Horizontal Transfer of microRNAs via Extracellular Vesicles:

Secreted membrane-bound vesicles, called extracellular vesicles, are important mediators of intercellular communication ([Pegtel *et al.* 2014](#_ENREF_25)). Exosomes are defined as 40-100nm diameter extracellular vesicles which are released upon fusion of the multivesicular bodies with the plasma membrane (Gu *et al.* 2014). Whilst similar in function and biochemical markers, microvesicles (≥100nm) differ from exosomes by being released from budding of the plasma membrane (Minciacchi *et al.* 2015). This report focused on a mixed population of EVs as past research also utilized mixed populations. EV cargo consists of cytoplasmic material, functional RNA and proteins where uptake of this content had been reported to influence a range of biological processes, such as the selective export of cytokines in immunological responses, mediating homeostasis and stress response ([McKechnie *et al.* 2006](#_ENREF_19); [Wysoczynski and Ratajczak 2009](#_ENREF_32); [Hedlund *et al.* 2011](#_ENREF_13)). However, recent studies have emerged that determined cancer-derived EVs absorbed into recipient cells are able to induce the establishment of the pre-metastatic niche in cancer progression and tumour microenvironment modifications ([Costa-Silva *et al.* 2015](#_ENREF_4); [Ramteke *et al.* 2015](#_ENREF_26)). Primarily this is attributed to the proteomic EV content being introduced into the endogenous population of the target cell, such as introduction of beta-catenin, epidermal growth factor receptor and major elements of the MAPK pathway ([Dovrat *et al.* 2014](#_ENREF_6); [Kharmate *et al.* 2016](#_ENREF_16); [Song *et al.* 2016](#_ENREF_28)). Yet, more intriguing is the discovery that exported microRNAs may be associated with this function.

MicroRNAs (miRNAs, miRs) are small non-coding RNAs found to be involved in most developmental and pathological processes due to its ubiquitous gene regulatory function. The functional miRNA sequences (~19-24 nt) are derived from longer transcripts that undergo processing and shuttling events to give rise to functional mature sequences, known to induce RNA degradation ([Ha and Kim 2014](#_ENREF_11)). Typically, the mature miRNA sequence interact with the 3’ untranslated region (3’-UTR) of its target transcripts and guides a multi-protein RNA induced silencing complex (RISC) to destine these molecules for degradation or translational inhibition ([Djuranovic *et al.* 2012](#_ENREF_5)). A 2009 estimate predicted that approximately 60% of the mammalian genome are able to be directly mediated by the miRNA RISC mechanism where a single miRNA can target hundreds of transcripts ([Friedman *et al.* 2009](#_ENREF_8)). This indicates the necessity of tight temporal and spatial control over miRs to prevent dysregulation of vital pathways. This is thought to be maintained by the high content of RNases in the extracellular serum which would rapidly degrade any miRNAs that attempt translocation across extracellular space ([Reddi and Holland 1976](#_ENREF_27); [Tsui *et al.* 2002](#_ENREF_29)). However, EV-contained miRNAs were found to bypass this degradation which allows for the absorption of these molecules into recipient cells, thus evoking their canonical function in a potentially diverse cell type ([Kosaka *et al.* 2010](#_ENREF_17); [Montecalvo *et al.* 2012](#_ENREF_21)).

Earlier work from our lab utilizes the caveolin-1/cavin-1 system to investigate the role of caveolin-1 in prostate cancer ([Inder *et al.* 2014](#_ENREF_15)). Interestingly, the cellular modification inflicted by comparing between PC3 and PC3 cavin-1+ cells modified extracellular vesicle (EV) content, a pathway unrelated to the function of caveolin or cavin-1. In addition to limiting adhesion independent growth, hyper-proliferation and EV protein content of PC3 cells, the ectopic expression of putative tumour suppressor, cavin-1, modified miRNAs found within EVs; specifically miR-148a ([Inder et al. 2014](#_ENREF_15)). Expression of miR-148a in bone marrow was reported to induce osteoclastogenesis by targeting an inhibitory transcription factor, MAFB, of the RANKL-induced osteoclastogenesis pathway, where the inverse was observed upon miR-148a inhibition ([Cheng *et al.* 2013](#_ENREF_3)). Bone fracture, pain and fragility are common co-morbidities associated with the bone metastasis-mediated prostate cancer due to increased bone resorption ([Luz and Aprikian 2010](#_ENREF_18)). Therefore the export of miR-148a from pro-metastatic prostate cancer cell line is consistent with clinical findings and may be a regulator of metastatic progression. However upon closer investigation, the addition of cavin-1 does not modify the cellular expression of miR-148a, only the EV content. This suggests that there may be selective export over the EV exported miRNAs, truncated by cavin-1 expression. Selective EV export of miRNAs had been observed in other studies, some of which links these miRNAs with disease states, particularly cancer metastasis ([Palma *et al.* 2012](#_ENREF_24); [Zhou *et al.* 2014](#_ENREF_33)). Yet, the mechanism that governs this selectively is mostly unknown.

A recent clue was provided by Villarroya-Beltri *et at*, who reported that sumoylated ribonucleoprotein, hnRNPA2B1 mediate the transport and subcellular localization of particular miRNAs in T-lymphocytes ([Villarroya-Beltri *et al.* 2013](#_ENREF_30)). Typically, the hnRNP family are involved in mRNA processing within the nucleus for translational control, mRNA stability and subcellular localisation, yet this is the first reported case of EV/multivesicular body localisation occurring from this mechanism and one of the first reports of its ability to bind to miRNAs ([Mili *et al.* 2001](#_ENREF_20); [Dreyfuss *et al.* 2002](#_ENREF_7)). Further questions arise due to this finding, such as the use of other hnRNP proteins for miRNA subcellular localization, how hnRNPs are targeted to the EVs and whether this protein family could be responsible for miRNA EV export in other cell types and stimuli.

Hypothesis and Aims:

Based on the above, we hypothesised that cavin-1 expressed in PC3 cells attenuates the EV export of oncogenic miR-148a by modulating export of RNA-binding proteins, similar to the mechanism identified by Villarroya-Beltri *et al* (2014). Given that RNA-binding proteins select for targets by binding conserved RNA sequences, known as motifs, miR-148a and other RNA targets will share a motif that allows specifically for their export over other microRNAs. Therefore, this differentially exported RNA-binding protein can regulate the export of motif-containing miRNAs due to their presence or absence in the forming EVs. The following aims were devised to address this hypothesis:

1. Assess the microRNA species that are modified by the PC3/cavin-1 model.
2. Identify candidate export proteins that participate in microRNA EV export.
3. Verify the interaction between candidate protein and microRNA by *in situ* and *ex vivo* experimental methods.

Methods and Materials:

## Reagents:

Trypsin-EDTA (Gibco), Roswell Park Memorial Institute (RPMI) 1640 media (Sigma), Fetal Bovine Serum (FBS) (Bovogen), Phosphate Buffered Saline (PBS) (Amresco Inc), Geneticin G418 Antibiotic (Invitrogen). Rabbit anti-hnRNP K (Abcam).

## Cell culture:

Previously generated PC3 cell lines, containing GFP or GFP tagged cavin-1 (referred to as PC3-cavin-1), were assessed through western immunoblotting for GFP to determine stable GFP expressivity. These cell lines were cultured in 5% FBS/RPMI1640 media in a 5% CO2 incubator set to 37°C. G418 antibiotic drug was added to these cultured cells to select for GFP expressing cells, making a total concentration of 0.1mg/mL. Detachment of the cells during passaging was completed using 0.25% Trypsin-EDTA solution.

## Differential miRNA expression:

Previously collected RNA-seq data for small RNAs, generated by Illumina sequencing, had been aligned and assessed for raw counts for miRNAs in cell and EVs. An R package, DESeq2, had normalized these counts to fit a negative binomial distribution and excluded microRNA data that possessed low to no counts (≤10 counts) for miRNA species across the triplicates and conditions (GFP and cavin-1), allowing for only relevant microRNAs to be assessed. Applying the function makes comparisons of expression between GFP and cavin-1 cell conditions and returns this in the form of log2 fold change (FC), Wald test p-value and a false discovery rate corrected p-value. This analysis was completed separately for cell and EV miRNA content. By using the log2FC values for each miR, comparisons were made between cell and EV expression by taking the difference in the form of FCcell-FCexo. Frequency disruption graphs were plotted by measuring the frequency of FC-FC in increments of 0.05. GraphPad Prism was used to generate this graph and line of best fit added by analysis of ‘Sum of two Gaussians’.

## Extracellular Vesicle Extraction and RNA extraction:

Cells were grown to 70% confluency prior to the addition of fresh serum free RPMI1640 media on 15cm Petri dishes. The conditioned media was collected after 24hrs of incubation and concentrated in a 10kDa ultracentrifugation filter tube (Sigma) until 1mL of concentrated media was achieved. This was processed through an exoRNeasy midi kit (Qiagen) to extract the total EV RNA as per manufacturer’s instruction. A sample of the cognate cells were also collected for comparison. The total cellular RNA was collected using the MiRvana kit as per manufactures’ instruction (Invitrogen). Nanodrop was used to assess to the purity and concentration of the RNA, where samples with an A260/280 approximating 1.8 were be used for further experimentation.

## Reverse Transcription quantitative Polymerase Chain Reaction (RT-qPCR) and preparation:

Poly-adenylation was completed using the E.coli polyadenylation enzyme and associated buffers (NEB) using a standard protocol (Balcells *et al.* 2011). This was immediately followed by cDNA conversion using the Superscript II Reverse Transcriptase and 0.1μg/μL oligo DT (Invitrogen) as per standard Invitrogen protocol. RT-qPCR was performed on the samples with primers specific to miR-363-3p, 148a-3p, 200a-3p, 30a-3p and 574-5p (IDT). Mir-125a-3p was used as the reference gene due to producing the same level of expression in EV derived from both GFP and cavin-1 PC3 cells based on the RNA-seq data. Delta delta CT statistics were completed by comparing between GFP and cavin-1 cell lines for the target and reference genes. Bar graphs generated by GraphPad Prism 6 and statistics calculated using a non-parametric two-sided T-test (Mann-Whitney U-test).

Motif Discovery and Assessment:

TAMO (Tools for Analysis of MOtifs) unix package was used to determine shared RNA-motifs within the differentially exported miRNA data set. The MEME algorithm was used to find a motif 4 to 10 nucleotides in length mapped amongst the inputted miRNA sequences with at least 70% conservation. The resulting motif was compared to the sequences of all expressed miRNAs in the PC3 cells to determine specificity to the differentially exported miRNAs using the sitemap algorithm. Sequence logos were generated using WebLogo (http://weblogo.berkeley.edu/).

## Proteomic Analysis:

Previously published liquid chromatography tandem mass spectrometry results for GFP and cavin-1 cell lines analyzed the proteomic content of the EVs excreted from these cells (Inder 2012). The fold change difference inflicted by cavin-1 was generated (mean GFP/Cavin-1) by Kerry Inder for each protein detected and p-value determined by two-sided paired Student t-test. This data were filtered to identify significantly modified proteins (p ≤ 0.05). Each protein in this data set were analyzed using the biomaRt R package for Gene Ontology (GO) annotation to determine RNA-binding ability (GO:0003723).

## Co-localization by Immunofluorescence Confocal Microscopy:

Cell were grown to 70% confluency on coverslips prior to fixation with 4% PFA for 30minutes and washing with PBS. 0.1% Triton-X in 1% BSA in PBS was added to the coverslips to block and permeabilize the cells. After 30 minutes of incubation, the coverslips were washed and primary antibodies in blocking solution (1% BSA/PBS) were then incubated with the coverslips for 1 hour at room temperature. Coverslips were then washed 3 times with PBS prior to incubation with secondary antibodies in blocking buffer for 1 hour in the dark at room temperature. After washing 3 times in PBS, 1:1000 dilution of DAPI in blocking solution was incubated with coverslips for 10minutes in the dark, followed by additional PBS and MilliQ water washing. Excess water was removed by Kimwipe prior to mounting on slides with 8μL Prolong Diamond (Invitrogen). Slides were dried for 24 hours at 37°C prior to imaging with the Olympus Confocal microscope. Pseudocolour and scale bar added by the FluorView software for the Olympus microscope.

## MicroRNA *In situ* Hybridization:

Cells were grown to 70% confluency on coverslips prior to fixation with cold 100% methanol. Coverslips were then washed thrice with PBS and incubated in the dark overnight at room temperature in 50pmole of Cy5 conjugated antimiR in oligo hybridization buffer; 50mM NaCl, 1mM Tris-Cl (pH 8.0), 0.1mM EDTA (pH 8). Cy5-scrambled oligo was used as a negative control and anti-miR-589 was used as a biological control as this miR should not bind or co-localize with hnRNPK. Excess antimiR was removed by washing thrice in PBS before 4% PFA fixation for 30minutes. Subsequent steps were performed as per immunofluorescence protocol for hnRNPK localisation with Alexa Fluor 568 secondary antibody. Spectral bleed through of Cy5 and Alexa Fluor was checked prior to data collection by individually staining with antibody or hybridizing fluorophore-antimiR and visualizing neighbouring channels.

Immunoprecipitation:

Protein G DynaBeads were washed thrice in washing and antibody binding buffer (W&B buffer) to remove storage reagents using the DynaMag2 magnet to separate and fix beads. W&B contains XXXX. 1uL of Anti-hnRNPK (Cat. No) were added to 200uL of W&B buffer, added to the beads and incubated on a rotating wheel for 40minutes. Excess antibody were removed by washing with W&B buffer thrice. 1% formaldehyde were added to plated cells for 8 minutes. This were subsequently washed with PBS and pellets scraped into tubes. Cell pellets were lysed by 20 minute incubation, on ice, using modified lysis buffer; 1% Triton-X, 20mM Tris (pH7.5), 150mM NaCl, 1x Protein Inhibitor complex, 0.5mM MAEBSF, 0.5mM Na3VO4 and 10mM NaF +NaPyropyrate (what []??). After incubation, lysates were centrifuged at maximum speed, 5 minutes at 4°C and supernatant moved to new tube. Cell lysate was added to diluted lysis buffer and added to antibody-bead tube. This was incubated for 45minutes at 4°C on a rotating wheel prior to washing 5 times with diluted lysis buffer. Protein-RNA crosslinked sample were eluted by incubation with SDS-PAGE buffer for 5 minutes at 95°C. This were then analyzed with Western blot. Alternatively, RNA pulled down by hnRNPK were purified by Trizol extraction. First, Trizol were added to the beads and incubated at 95°C for 5minutes to reverse the crosslink. Subsequent steps were performed as per standard Trizol extraction protocol. Quantification of RNA was performed by Nanodrop analysis.

## Western blotting:

Sample buffer were added to whole cell lysate or EV lysate sample to reach a final 1X concentration and protein denatured by incubation at 95° C for 5minutes if denaturation was not already performed. BioRad Precession Plus protein ladder were loaded into a 12.5% SDS-PAGE gel with 4% stacking gel. Blanks (10uL of SDS-PAGE buffer) were placed in wells either side of the ladder. Samples were added in equal amounts to the wells. SDS-PAGE buffer were added to any unfilled wells to maintain consistent salt concentrations across the gel. Gels were ran at 80V until sample stacked, then increased to 100V until the dye front reached the end of the gel. Wet transfer was completed following a standard procedure and reagents to transfer protein to a fluorescent nitrocellulose membrane (refereence). Membrane was subsequently blocked in 3% BSA/PBS with 0.1% Triton-X for 30minutes to prevent non-specific antibody binding. hnRNPK antibody was diluted 1:1000 in blocking buffer in a 50mL falcon tube. Membrane was added face up into the tube, avoiding air bubbles and incubated for 1.5hrs at room temperature on a roller. This was followed by washing thrice in TBS-Tween-20. IRdye800W anti-mouse was added to blocking solution to a final concentration of 1:7,500, kept in the dark and incubated with the membrane for 1hr. After subsequent washing thrice with TBS-tween, membrane was washed again with PBS and dried in the dark. Odessyse Li-Cor visualized the bands in two channels with the ImageStudio software.

Results:

## MicroRNAs are selectively exported from prostate cancer cells.

MicroRNA-148a was previously found to be selectively exported from the PC3 cell line where this export was truncated by ectopic expression of cavin-1. However, this analysis had not considered other miRs that may also be moderated by this system. To determine all miRNAs that are being modified in the PC3-EVs by cavin-1 expression, our lab conducted a comprehensive RNA-seq analysis to quantify the miRNAs in EV and cognate cells. 95 miRs were detected in EVs from PC3 cell lines. Comparison between GFP and cavin-1 cell lines through differential expression analysis (DESeq2) revealed a total of 12 significantly (p ≤ 0.05) modified miRs in the EVs (fig. 1a), including miR-148a (-3p). The previous study revealed that reduction of EV-contained miR-148a was not reflected by a global cellular change, but rather decreases independent of the cellular expression. Here, I wanted to determine if this trend persists with additional miRs. Comparison between cellular and EV modifications induced by cavin-1 reveals a subset of miRs that are dramatically reduced in the EVs with little modification of total cellular expression. These are the miRs likely to be acted upon by the proposed protein mediated export, attenuated by cavin-1 expression. In contrast, 5 of these miRs present with proportionate cellular expression change that could explain the decrease or increase in EV miR content. This process is known as sampling, where miRs in the cytoplasm are taken into the forming EVs due to proximity as opposed to protein mediated export that would confer some selectivity. This establishes that both sampling and selective export of miRs can occur in this system.

6 significantly modified miRs were selected for validation across the sampling and selective export groups established above; miR-30a-5p, miR-148a-3p, miR-200a-3p, miR-10b-5p, miR-574-5p and miR-363-3p. RT-qPCR of these targets confirms whether the cavin-1 induced modification of these miRs were reproducible. Here, the trend first displayed by the

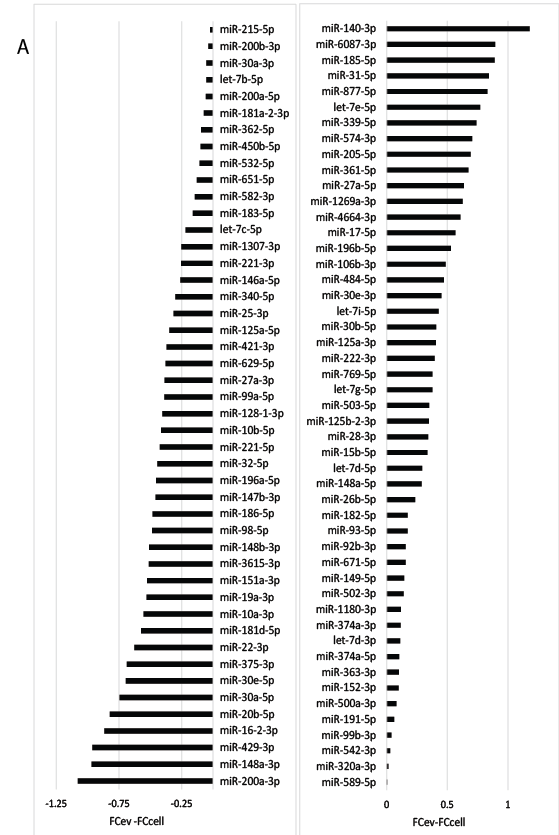


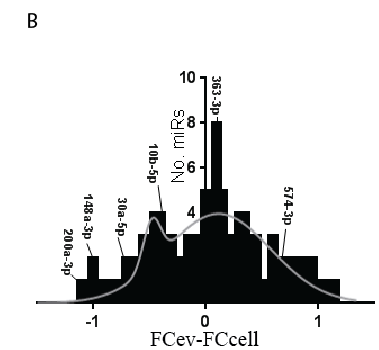
**Figure 1**: *Addition of cavin-1 to PC3 cells modifies EV microRNAs.* **A**) RNA-seq was performed previously on microRNAs contained in the EV and cellular content of PC3-GFP and PC3-cavin-1 cells. DESeq2 analysis compared PC3-GFP to PC3-cavin-1 EV content (black), expressed as log2FC, to determine the effect of cavin-1 on EV miR content. miRs that were significantly modified (\* p ≤ 0.05) in the EVs were plotted. Analysis was repeated on cellular content (grey) for each of the miRs significantly modified in the EV. This reveals that cavin-1 has an effect of miR EVs where some effected miRs are not modified in the cell. **B**) RT-qPCR was performed to validate the RNA-seq data. This was performed on EV and cellular RNA content extracted from GFP and cavin-1 PC3 cells to determine relative amount of miR-200a-3p, 148a-3p, 30a-5p, 10b-5p, 574-3p and 363-3p (n > 3). Delta-delta CT (ddCT) was calculated and plotted (ddCT + SEM) by comparing expression of targets to miR-125a-3p. This miR was unchanged in the EV and cell by cavin-1 expression. A Mann-Whitney U test compared EV change to the cellular change for each of the miRs. This analysis confirms the trends found from the RNA-seq data.

RNA-seq data is maintained, shown by a decreased presence of miR-30a-5p, -148a-3p, -10b-5p and -200a-3p in EVs between the cell lines compared to the cellular content. Inversely, miR-574-3p was increased due to the presence of cavin-1 in the EVs, and miR-363-3p is confirmed to be not selectively exported by cavin-1. This establishes that some miRs are indeed selectively exported from PC3 cells, where cavin-1 modulated this export.

## Distinct sequence motifs are overrepresented in differentially exported microRNAs.

Protein to RNA interactions are dictated by specific nucleic acid sequences, or motifs, that are conserved across the targeted RNAs. Here, we attempted to assess whether the selectively exported miRNAs share a sequence motif to explain their selective export. Unfortunately, motif discovery with only 5 miRs was not adequate to establish a significant motif, so additional miRs were considered for this analysis. All 95 miRs recorded in the EVs from the RNA-seq data were compared to their cellular change in the form of log2FCexo- log2FCcell. This provides a single value that reflects how different the EV modifications are from the cell, where selective export results in a large negative (cavin-1 attenuates export) or positive (cavin-1 enhances export) value and sampling approximates 0. Plotting this value as a frequency distribution plot reveals how prevalent each form of export is. This yielded a large population of miRs that undergo sampling, around 0.1, but also a small peak at -0.45. The miRs that were validated in RT-qPCR were noted on the graph to demonstrate where they fit into this distribution. This shows the selectively exported 200a-3p, 148a-3p and 30a-5p to the far left of the graph, with 363-3p in the sampling population (FC-FC≈0.1) and 574-3p to the right of the graph. This corresponds to the groupings established previously (Fig.1a). While miR-10b-5p (FC-FC ≈ -0.4) did show a decrease in the EV more than the cell, the difference between them was not considered adequate enough to presume it were under action of protein





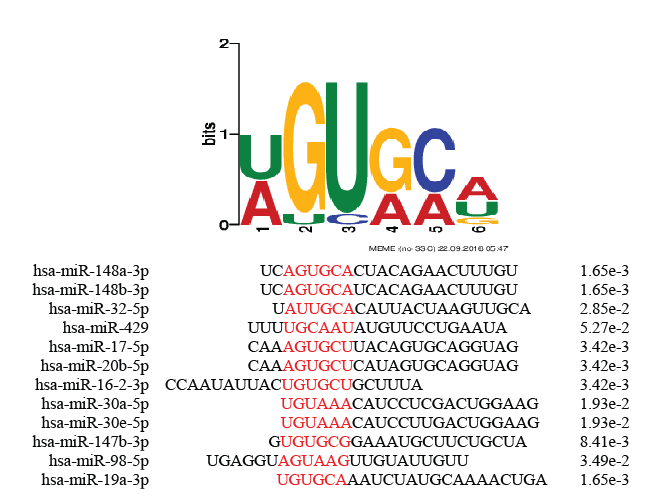
**Figure 2**. *Comparison of microRNA EV and cellular changes inflicted by cavin-1 expression.* **A**) All 95 miRs detected in the EVs were analysed to determine selective or sampling mediated export. Bar graph displays the difference between FCev and FCcell for each miR. **B**) Frequency distribution graph of the FC-FC value, by increments of 0.05, and experimentally validated microRNAs. Line of best fit modelled by sum of two Gaussians.

mediated export. Hereby, miRs that possess a FC-FC of -0.45 or lower are considered selectively exported for this motif discovery. 19 miRs fulfill this criteria.

Motif discovery was used to define stretches of RNA that are shared amongst the differentially exported miRNAs that would be targeted by the miR export protein. This analysis returned two distinct motifs that are enriched in the miR group that possess reduced export upon cavin-1 expression (fig.3). These motifs are present within 14 of the 19 miRs within this group with minimal (n=1) hits in the non-differentially exported miR group. This indicates potential binding sites the export protein may be able to bind to evoke specificity and selectivity of the targets.

## Candidate proteins are present in EVs with RNA binding ability.

To identify candidate proteins that could mediate this selective miR export, EV protein content was assessed for differential export upon cavin-1 expression and RNA-binding ability. Previously published proteomic data determined the identity of EV proteins from PC3 GFP and PC3 cavin-1 cell lines. Comparing between the cell lines returned a total of 120 significantly differentially exported proteins, where 109 of these proteins were decreased upon cavin-1 expression (Fig.4). Additionally, these proteins were further analyzed by GO analysis to determine whether these proteins had previously reported RNA-binding capacities. Together, this yields a total of 5 differentially exported RNA-binding proteins. Interestingly, further investigation into the RNA binding specificity of hnRNPK revealed, through mutagenesis assays, its affinity to bind to sequence AGUGUG in miR-122. Upon comparison to the known motif, using the FIMO algorithm, hnRNPK matches adequately to the predicted binding motif (p=0.0435). Hereby, hnRNPK was considered a viable candidate protein to mediate the selective export of miRs.



**Figure 3**. *Shared motif contained within a subset of the selectively exported microRNAs.*

Motif discovery using the MEME algorthim revealed a motif from the exported miR

group. Table shows the miRs containing the motif, position of motif (red) and p-value

calculated from the sitemap algorthm. Motif matches to 12 of the 19 miRs in this group.



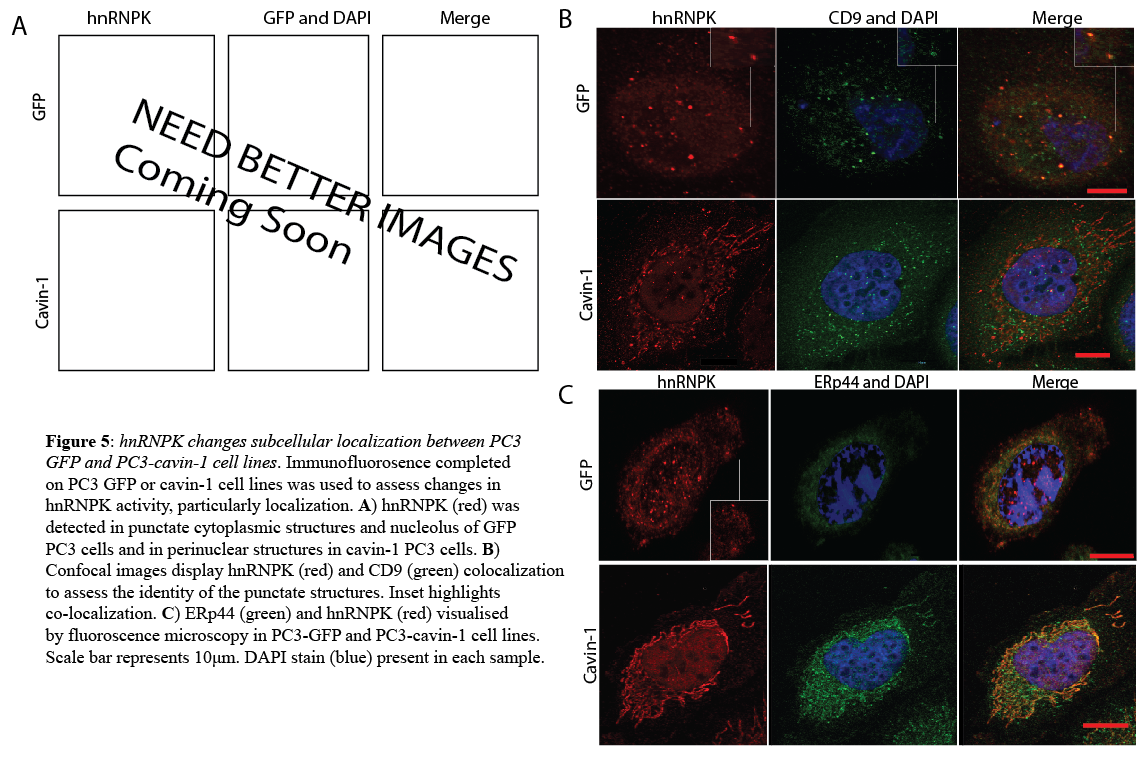
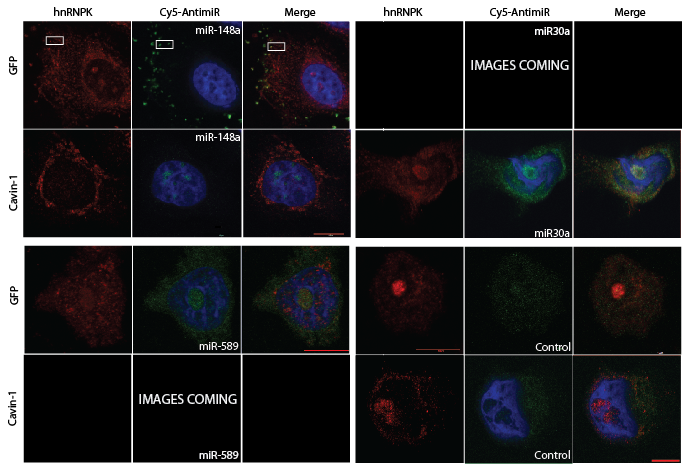
**Figure 4**: *Cavin-1 attenuates the export of RNA binding proteins*. MS/MS data compared the protein content of EVs between PC3 and PC3-cavin-1 cells to determine significantly differentially exported proteins. Gene Ontology analysis revealed whether these exported proteins bind RNAs. 5 RNA binding proteins had reduced exported upon cavin-1 expression.

## hnRNPK sub-cellular localization modified in cavin-1 PC3 line.

Immunofluorescence was performed using hnRNP K specific antibodies to determine cellular localization changed between GFP and cavin-1 cell lines which may explain differential export. An initial observation of hnRNPK localization revealed a distinct change between cell lines, from punctate like structures in PC3 GFP to a perinuclear focus in cavin-1+ cell lines. Further co-localization studies were performed to determine what these structures were. CD9 is a commonly used marker for multivesicular bodies and exosome formation. hnRNPK appears to co-localize somewhat with the CD9 protein in GFP PC3 cells, which indicates presence in the multivesicular bodies that is not occurring in the PC3 cavin-1 cell line. However, the hnRNPK in PC3 cavin-1 was found present in endoplasmic reticulum, shown by strong overlap with ER resident protein, ERp44. Therefore, change in subcellular localization modified by cavin-1 could explain the differential export of hnRNPK and miRNAs.

## hnRNPK co-localizes with selectively exported microRNAs.

The interaction between the selectively exported microRNAs and hnRNPK was assessed in two separate ways: by assessing co-localization by microRNA *in situ* hybridization immunofluorescence (miR-ISH IF) and binding ability by RNA immunoprecipitation (RIP). miR-ISH IF methodology was established by modifying the existing Fluorescence *In Situ* Hybridization (FISH) methods and IF. The anti-miR probe highlights the target miRs based on RNA-RNA hybridization. Here, I assessed the subcellular co-localization of miR-148a-3p, -30a-5p, 589-5p, a scrambled miR and hnRNPK. miR-148, 30a and 589-5p all possess nucleoli localization, which confirms that the ISH was successful.

**Figure 6***. miR-148a and miR-30a co-localize with hnRNPK in situ*. Images show localization of hnRNPK (red) and fluorophore tagged (Cy5) anti-miR (pseudocoloured green) in PC3-GFP and PC3-cavin-1 cell lines, with DAPI stain (blue). MicroRNA targets include selectively exported miRs, 148a-3p (top left) and 30a-5p (top right), miR-589-3p biological control, microRNA unchanged by cavin-1 expression and not known to bind to hnRNPK, (bottom left) and technical control; scrambled-148a oligo (bottom right). Yellow overlap between red hnRNPK and green anti-miR and boxed areas indicate co-localization. Scale bar represents 10μm.

## hnRNPK binds RNAs in the PC3 cell line.

Co-localization between hnRNPK and selectively exported microRNAs only confirms their ability to be compartmentalized simultaneously. However whether these interact though direct binding needed to be investigated. After optimization of various immunoprecipitation methods, I completed an IP using anti-hnRNPK to pull down hnRNPK and all of it binding partners, including RNA. As the RNA-protein interaction can be fairly transient, cellular material was crosslinked by formaldehyde prior to the IP. After elution from the IP beads, a western blot was performed to determine if IP conditions were suitable to pull down the targets of hnRNPK. This is observed as a band approximating 58-62kDa that reflects the native weight of hnRNPK. Additional bands at 70, 100, 125 and approximately 140kDa show that hnRNPK is bound to various partners in this IP. However, this could include proteins, RNAs and microRNAs. Hereby, I attempted to purify the RNAs that hnRNPK binds. After purification by Trizol extraction, the RNA was quantified using nanodrop. This yielded a consistent increase of RNA identified from the hnRNPK pull down compared to the IgG control. This is consistent with past research which shows hnRNPK binding to RNAs and reports of it binding microRNAs. While this is instrumental in determining whether hnRNPK binds to microRNAs, the low yield indicates the need to scale up the experiments. Further assessment is needed to determine whether this population of RNA contains microRNAs and whether these miRs are the ones also predicted.

# Discussion:

This study determined that a subset of microRNAs are selectively exported from the PC3 cell line, where addition of cavin-1 attenuated this export. Furthermore, a protein predicted to mediate this export proved to be a viable candidate due to being selectively exported, predicted to bind to many of those exported miRs and co-localizing to miR-148a.

Points to discuss:

* RNA-seq data: microRNAs aren’t so dramatically modified by the proteins. System is definitely a gradient rather than a switch. May suggest synergy of proteins working together, differences between EV types being modified by cavin-1 eg mechanism in exosomes but not microvesicles. Additionally there is nothing to prevent sampling from occurring, where if the export protein isn’t there there still isn’t anything preventing miRs going into forming EVs.
* Validation with rt-qpcr, while confirms trends, the analysis struggles to yield significant results; discuss why and use of ddPCR.
  + Relevance with the field: IE relation of these microRNAs to disease and what that means for this system. Eg. What does export of miR-429 even mean for the recipient cell? How does past data fit in with this?
  + Do these differentially exported miRs relate to clinical findings? Eg. Exported miR-148a from prostate cancer makes sense as its role with oestoclastogensis is consistent with advanced prostate cancer.
* Motifs: discuss that some do not contain the motif but seem to be differentially exported. Ie. This would come back to the multiple proteins involved with same process theory.
* hnRNPK information: family members associated with the function, ability to bind to the motif/miRs. Role in cancer on its own, usually related to increase or decrease in cytoplasm/nucleus (Emerging roles of heterogeneous nuclear ribonucleoprotein K (hnRNP K) in cancer progression)
* hnRNPK localization to cd9 positive MVB and whether that’s been detected before. What it could mean (duh, export) etc.
* Discus co-localization results and how that help the hypothesis. Discus concerns with methodology set up: eg unlikely for the probe to be binding the target miR if in active site so maybe we aren’t visualizing miRs actively being bound and therefore missing some hard hitting evidence. Then suggest alternative methods.
* Discuss binding of hnRNPK to RNA. This is consistent with previous results however doesn’t establish anything new. Need to scale up, rethink approach, and try specific techniques. Limitation in this method, IE. Binding is probably only transient to the protein, hence the low pull down amounts. Additionally, it gives all information about hnRNPK binding partners but nothing of the microRNA ability to bind. Ideally, fixing microRNA to beads and pulling down its binding partners would provide more information about the system.