



H3ABioNet

Pan African Bioinformatics Network for H3Africa

Introduction to Bioinformatics Online Course: IBT

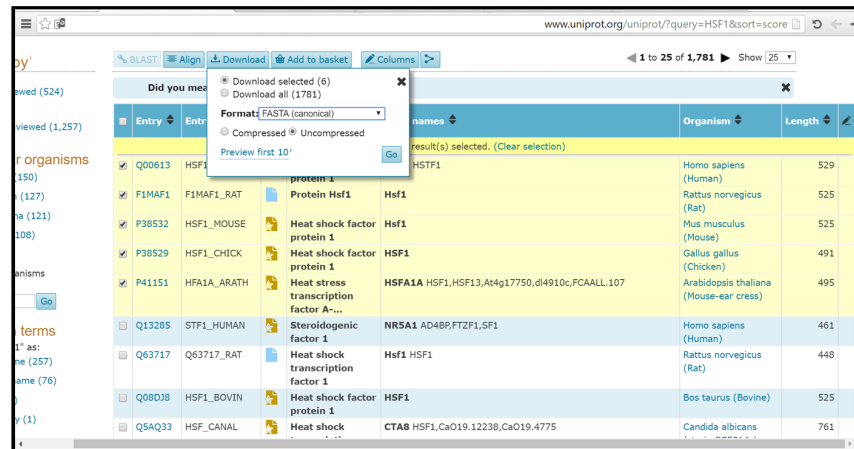
Multiple Sequence Alignment

Building Multiple Sequence Alignment

Lec2 Choosing the Right Sequences

Choosing the Right Sequences

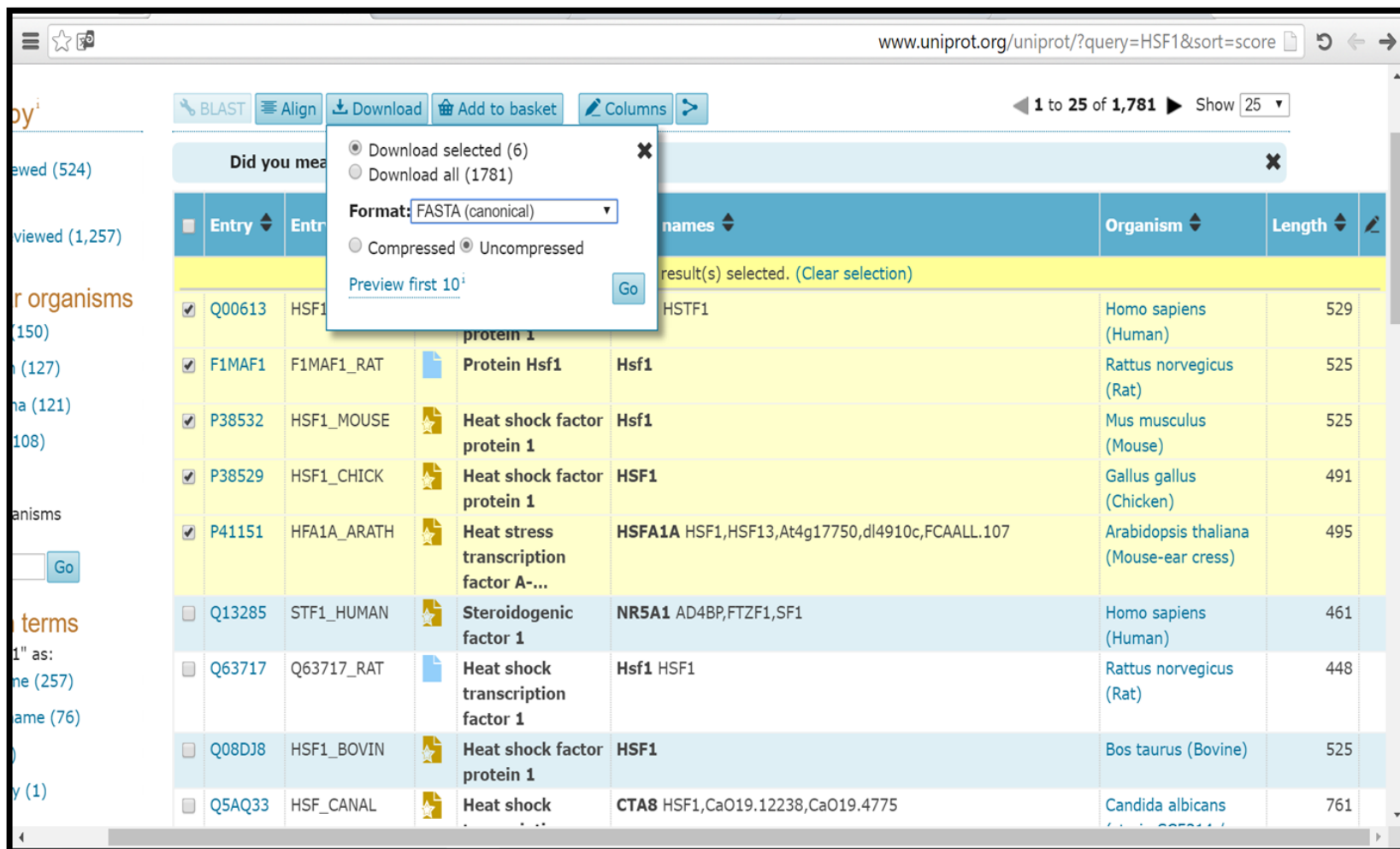
“Before you build your alignment, you must **carefully select the sequences** you want to align. These sequences are members of the **same protein family**, and they all **share a common ancestor**. The family is usually **too large** to be entirely included in your multiple alignment, and **picking the right sequences is an art.**”



Accession	Protein Name	Organism	Length
Q00613	Heat shock factor protein 1	Homo sapiens (Human)	529
F1MAF1	Heat shock factor protein 1	Rattus norvegicus (Rat)	525
P38532	Heat shock factor protein 1	Mus musculus (Mouse)	525
P38529	Heat shock factor protein 1	Gallus gallus (Chicken)	491
P41151	Heat stress transcription factor A...	Arabidopsis thaliana (Mouse-ear cress)	495
Q13285	Steroidogenic factor 1	Homo sapiens (Human)	461
Q63717	Heat shock transcription factor 1	Rattus norvegicus (Rat)	448
Q08038	Heat shock factor protein 1	Bos taurus (Bovine)	525
Q5AQ33	Heat shock	Candida albicans	761

Claverie J, Notredame C (2007). Bioinformatics for Dummies (2nd Edn). Wiley publishing, Inc. 436 pp.

Retrieve Sequences from Uniprot (www.uniprot.org)



www.uniprot.org/uniprot/?query=HSF1&sort=score

1 to 25 of 1,781 Show 25

Download selected (6) Download all (1781) Format: FASTA (canonical) Compressed Uncompressed Preview first 10ⁱ Go

Entry	Entry	Names	Organism	Length
Q00613	HSF1_HUMAN	Heat shock factor protein 1	Homo sapiens (Human)	529
F1MAF1	F1MAF1_RAT	Protein Hsf1	Rattus norvegicus (Rat)	525
P38532	HSF1_MOUSE	Heat shock factor protein 1	Mus musculus (Mouse)	525
P38529	HSF1_CHICK	Heat shock factor protein 1	Gallus gallus (Chicken)	491
P41151	HFA1A_ARATH	Heat stress transcription factor A-...	Arabidopsis thaliana (Mouse-ear cress)	495
Q13285	STF1_HUMAN	Steroidogenic factor 1	Homo sapiens (Human)	461
Q63717	Q63717_RAT	Heat shock transcription factor 1	Rattus norvegicus (Rat)	448
Q08DJ8	HSF1_BOVIN	Heat shock factor protein 1	Bos taurus (Bovine)	525
Q5AQ33	HSF_CANAL	Heat shock	Candida albicans	761

NCBI Resources How To Sign in to NCBI

Protein Protein HSP70 Search

Create alert Advanced Help

Species

- Animals (65,606)
- Plants (30,667)
- Fungi (61,126)
- Protists (23,483)
- Bacteria (766,071)
- Archaea (5,312)
- Viruses (5,044)
- Customize ...

Source databases

- PDB (1,217)
- RefSeq (225,406)
- UniProtKB / Swiss-Prot (4,000)
- Customize ...

Genetic compartments

- Chloroplast (63)
- Mitochondrion (52)
- Plasmid (1,430)
- Plastid (69)

Sequence length

- Custom range...

Summary 20 per page Sort by Default order

See **HSP70** heat shock protein 70 in the Gene database
hsp70 reference sequences [Transcript \(1\)](#) [Protein \(1\)](#)

Items: 1 to 20 of 961097

1. [HSP70, partial \[Cryptosporidium hominis\]](#)
629 aa protein
Accession: AKL59498.1 GI: 829026803
[GenPept](#) [Identical Proteins](#) [FASTA](#) [Graphics](#)

2. [HSP70 \[Mytilus galloprovincialis\]](#)
654 aa protein
Accession: AAW52766.1 GI: 57635269
[GenPept](#) [Identical Proteins](#) [FASTA](#) [Graphics](#)

3. [hsp70, partial \[Drosophila auraria\]](#)
633 aa protein
Accession: CAA04699.1 GI: 3005909
[GenPept](#) [Identical Proteins](#) [FASTA](#) [Graphics](#)

Send to: Filters: [Manage Filters](#)

Choose Destination

- ☒ File
- ☐ Clipboard
- ☐ Collections

Download 3 items.

Format

FASTA

Sort by

Default order

Show GI ☐

Create File

Find items

Search details

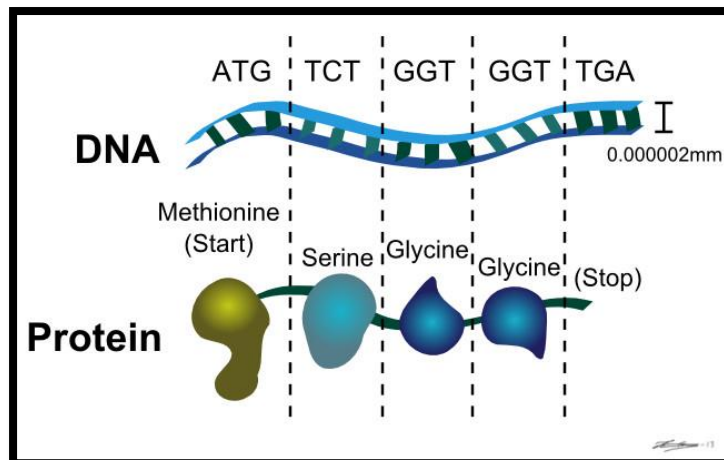
HSP70[All Fields]

A Few Guidelines for Selecting Sequences

Proteins or DNA

Use **proteins whenever possible**. You can turn them back into DNA after doing the multiple alignment.

If the sequences are **non-coding sequences**, you must **use DNA**

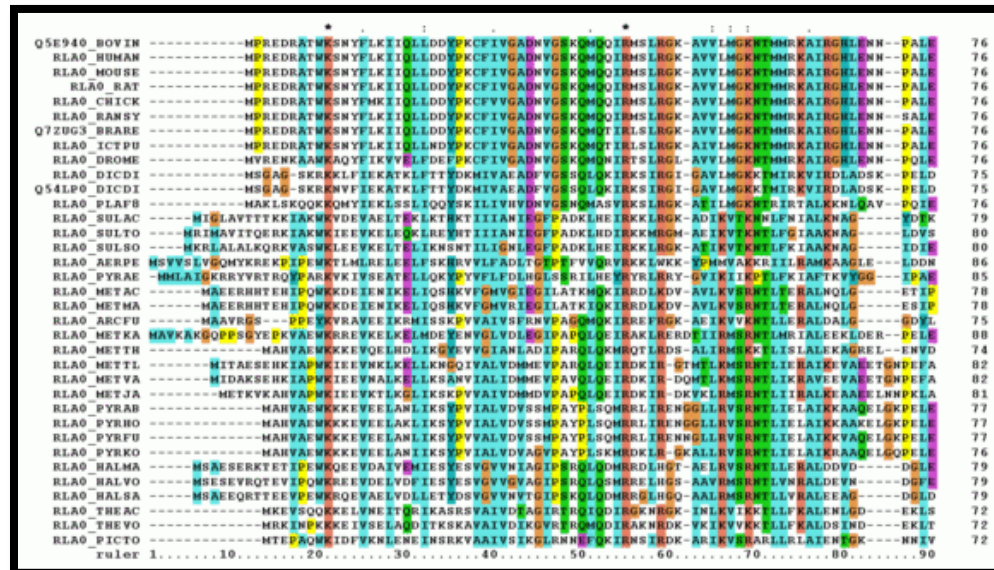


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A Few Guidelines for Selecting Sequences

Many sequences

Start with 10–15 sequences; **avoid** aligning **more than 50** sequences.

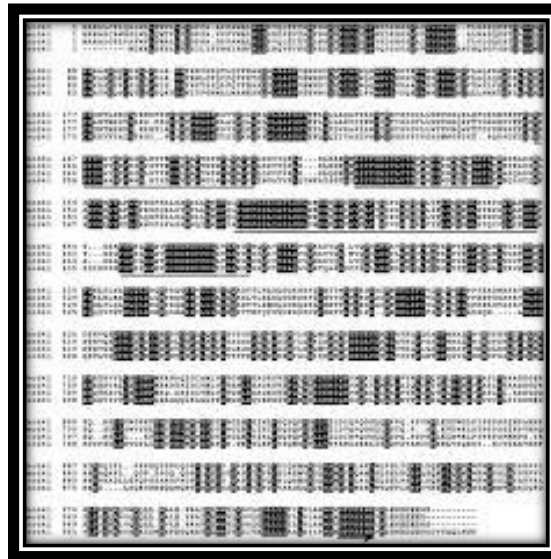


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A Few Guidelines for Selecting Sequences

Very different sequences

Sequences that are **less than 30 percent** identical to more than half the other sequences in the set often **cause troubles**.



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A Few Guidelines for Selecting Sequences

Identical sequences

They **never help**. Unless you have a very good reason to do so, **avoid incorporating** into your multiple alignment any sequence that's more than **90 percent identical** to another sequence in the set.

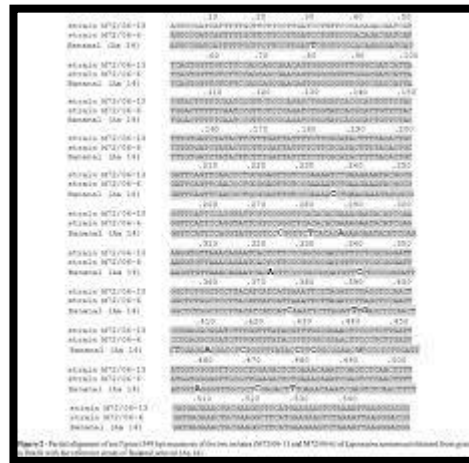


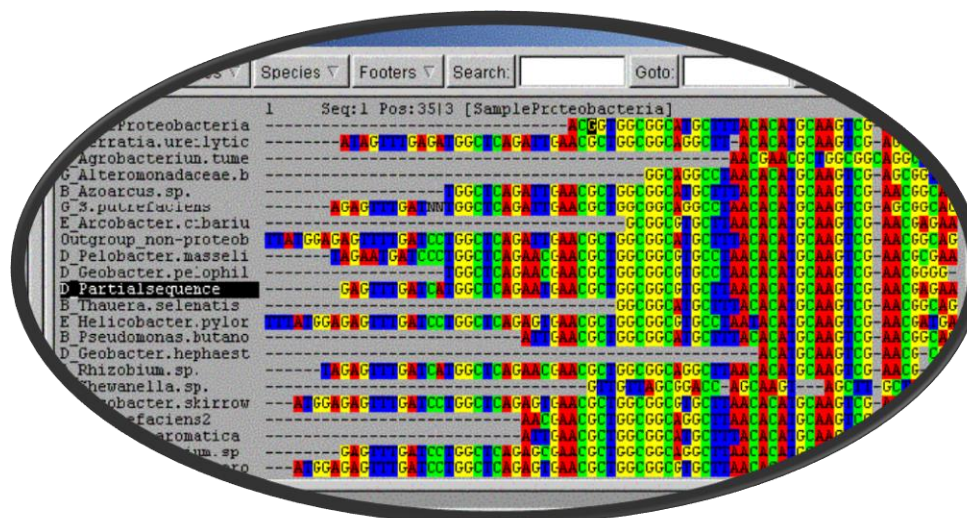
Figure 2. Partial alignment of sequences 349 bp upstream of the two isolates (MT209-11 and MT209-12) of *Legionella pneumophila* obtained from a tick (6) with a reference strain of *Legionella* serovar 14g (4).

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A Few Guidelines for Selecting Sequences

Partial sequences

Multiple-sequence-alignment programs prefer sequences that are roughly the same length. Programs often have **difficulties comparing** items in a **mixture of complete sequences and shorter fragments**.

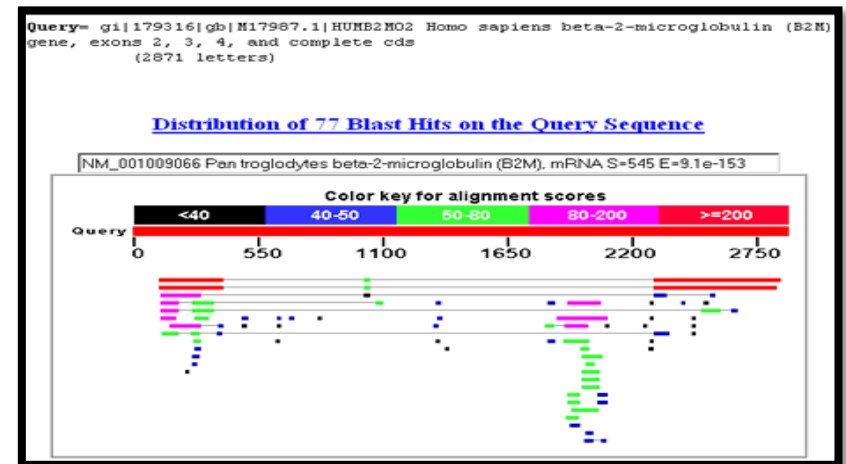
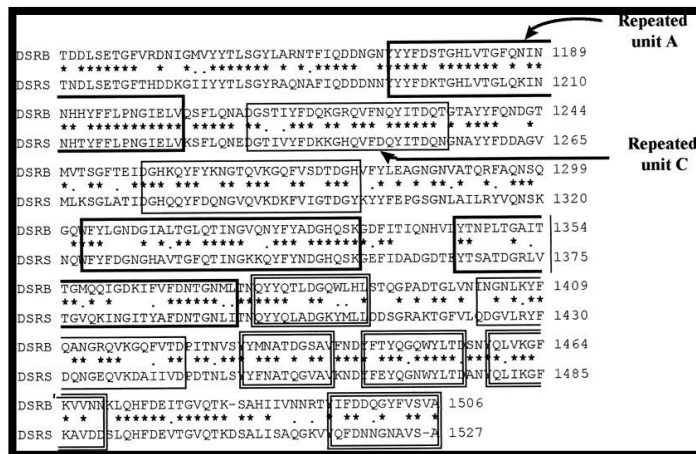


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A Few Guidelines for Selecting Sequences

Repeated domains

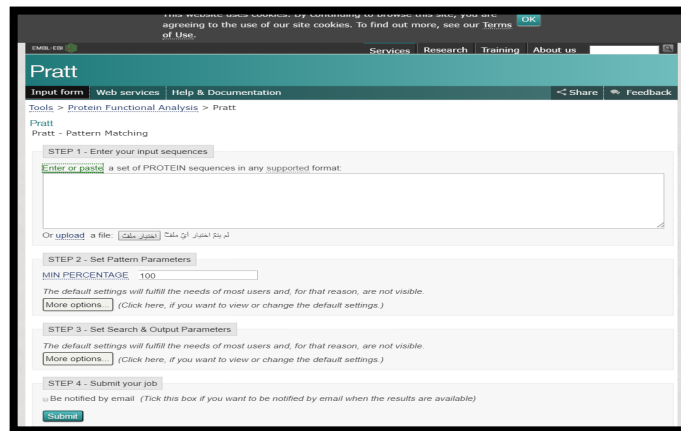
Sequences with repeated domains **cause trouble** for most multiple-alignment programs — especially if the **number of domains is different**. When this happens, you may be better off extracting the domains yourself with Dotlet or Lalign and making a multiple alignment of those segments.



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A Few Guidelines for Selecting Sequences

If you still **cannot generate a proper alignment** from sequences that you know are related, you could **use a local** multiple alignment method, such as the **Gibbs sampler**, or a pattern extraction motif, such as **Pratt**.



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EMBL-EBI [Services](#) [Research](#) [Training](#) [About us](#)

Pratt

[Input form](#) [Web services](#) [Help & Documentation](#) [Share](#) [Feedback](#)

[Tools](#) > [Protein Functional Analysis](#) > Pratt

Pratt

Pratt - Pattern Matching

STEP 1 - Enter your input sequences

[Enter or paste](#) a set of PROTEIN sequences in any supported format:

Paste your sequences here

Or [upload](#) a file: [اختيار ملف](#) [لم يتم اختيار أي ملف](#)

STEP 2 - Set Pattern Parameters

[MIN PERCENTAGE](#)

The default settings will fulfill the needs of most users and, for that reason, are not visible.

[More options...](#) *(Click here, if you want to view or change the default settings.)*

STEP 3 - Set Search & Output Parameters

The default settings will fulfill the needs of most users and, for that reason, are not visible.

[More options...](#) *(Click here, if you want to view or change the default settings.)*

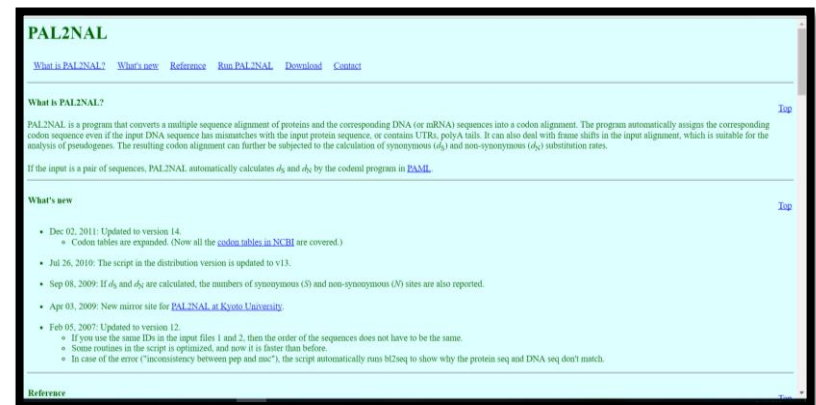
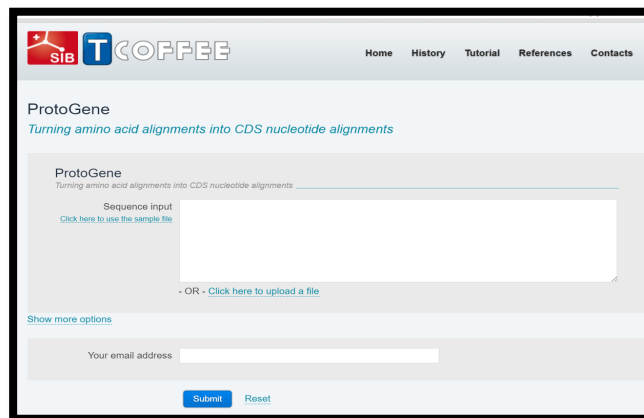
STEP 4 - Submit your job

☐ Be notified by email *(Tick this box if you want to be notified by email when the results are available)*



[Submit](#)

For carrying out a **phylogenetic analysis** on a set of **coding DNA** sequences, do the following:

1. **Translate** your **DNA** sequences into **proteins**.
2. **Perform** the **multiple alignments** on the proteins.
3. **Thread** the **DNA** back onto the protein multiple sequence alignment framework using **pal2nal** (coot.embl.de/pal2nal) or **Protogene** if you do not have the original DNA sequence (www.tcoffee.org).



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ProtoGene

Turning amino acid alignments into CDS nucleotide alignments

ProtoGene

Turning amino acid alignments into CDS nucleotide alignments

Sequence input

[Click here to use the sample file](#)

Paste your sequences here

- OR - [Click here to upload a file](#)

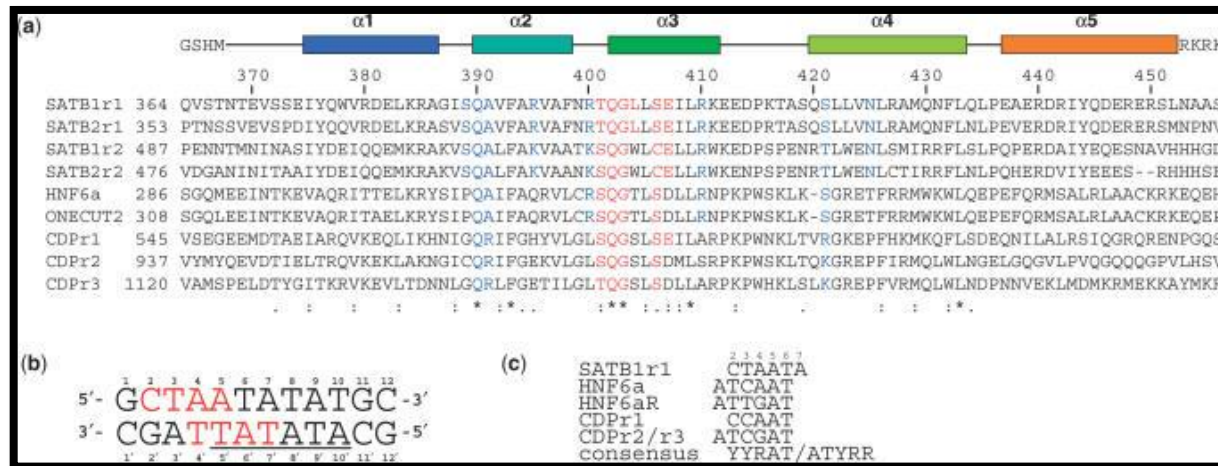
[Show more options](#)

Your email address

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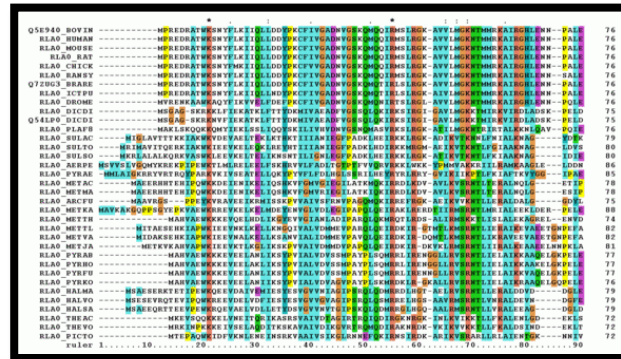
Choosing the right number of sequences

- 1- you should start with a relatively small number of sequences — between **10 and 15 sequences** would be suitable for most cases.
- 2- After you get **something interesting** happening with this small set, you can always **increase its size**.
- 3- it's **hard to** see any reason for generating a multiple alignment with **more than 50 sequences**, **unless** you're interested in building some **extensive phylogenetic tree**.



Why you should not use too much sequences to align?

- 1- **Computing** big alignments is difficult.
- 2- **Building** big alignments is difficult
- 3- **Displaying** big alignments is difficult.
- 4- **Using** big alignments is difficult.
- 5- **Making accurate** big alignments is difficult



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Should you choose sequences that are very similar or very different?

- Make the right **compromise** between **similarity** and **new information**
- An alignment that only contains **very similar** sequences brings **little information**.
- You can use it to **extrapolate annotations**, but you **can't do phylogeny, structure prediction, function prediction**, or any of the other useful applications that we mentioned before

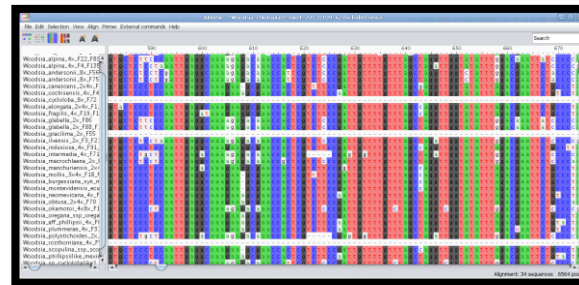
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The general rule is that you want them to be **as distantly related as possible** - without requiring too many **gaps in order to be properly aligned.**

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Two things multiple-sequence-alignment programs *really* don't like are

- 1- Sequences that are very different from every other sequence in the group
- 2- Sequences that need long insertions/deletions to be properly aligned

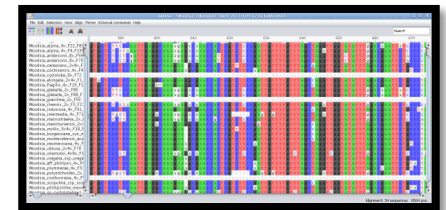


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Gathering your sequences with online BLAST servers

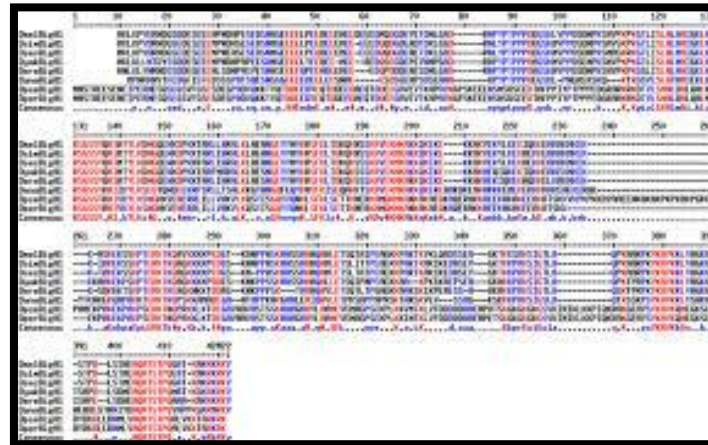
Characterized sequences: Try to include sequences with **good annotations and experimental information** in your alignment because they bring biological information with them — and also **allow feature propagation**.

Uncharacterized sequences: including them in your multiple alignment is to **distinguish between the conserved positions** that cannot mutate and the other, less-important columns. They help in **getting some contrast** on your sequence of interest.



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Sequences that are **so similar** to the query are **probably homologous**. We commonly refer to such sequences **as hits** or **matches**.

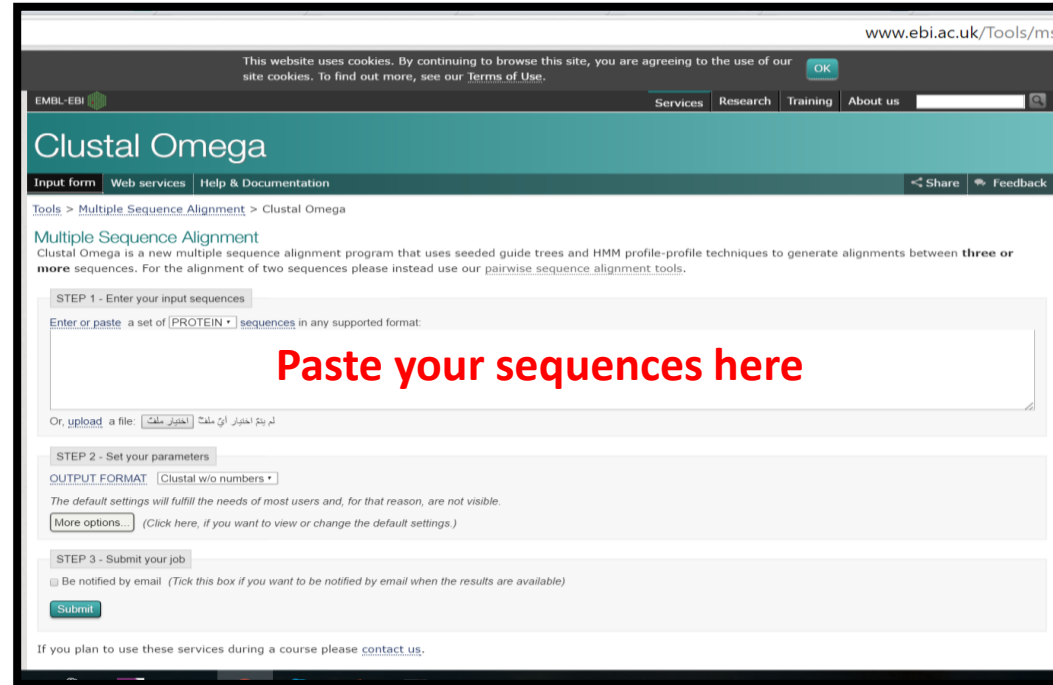


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Choosing the Right Method of Multiple Sequence Alignment

ClustalOmega

The **most commonly** used multiple sequence alignment package. Clustal Omega is a new multiple sequence alignment program that uses seeded guide trees and **HMM profile-profile** techniques to generate alignments between **three or more** sequences



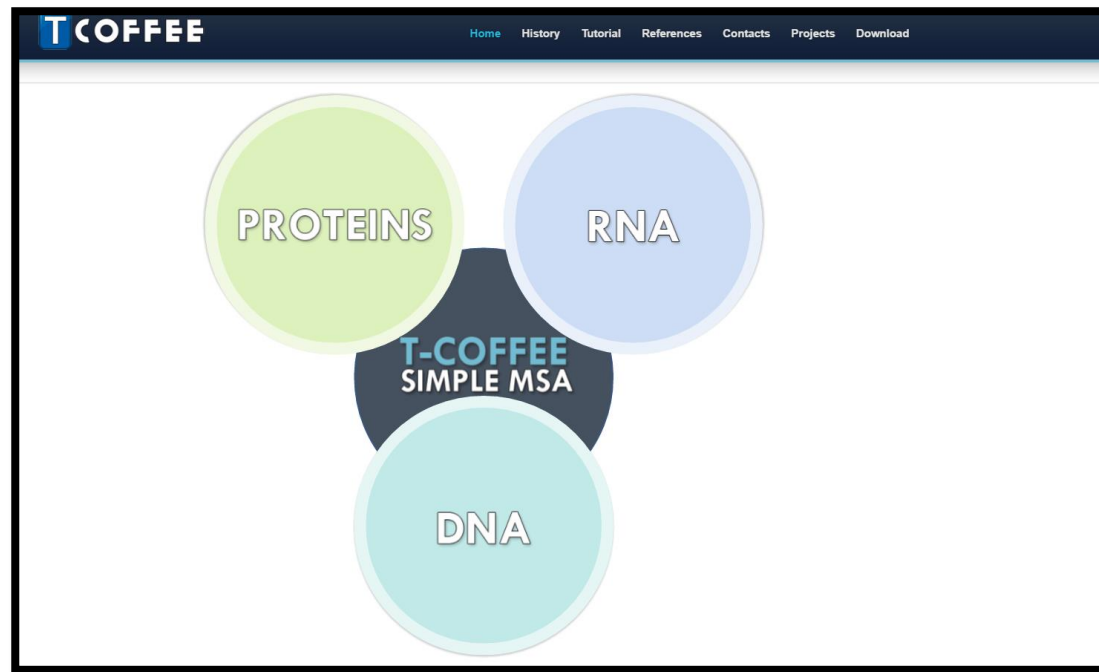
The screenshot shows the Clustal Omega web interface at www.ebi.ac.uk/Tools/ms. The page has a teal header with the "Clustal Omega" title. Below the header, there are tabs for "Input form", "Web services", and "Help & Documentation". The "Input form" tab is active. The main content area is titled "Multiple Sequence Alignment" and includes a description: "Clustal Omega is a new multiple sequence alignment program that uses seeded guide trees and HMM profile-profile techniques to generate alignments between three or more sequences. For the alignment of two sequences please instead use our pairwise sequence alignment tools." The interface is divided into three steps: "STEP 1 - Enter your input sequences", "STEP 2 - Set your parameters", and "STEP 3 - Submit your job". In STEP 1, there is a large text area with the red text "Paste your sequences here" and a button "Enter or paste". Below this, there is a section for "Or, upload a file" with a button "Upload". In STEP 2, there is a section for "OUTPUT FORMAT" with a dropdown menu set to "Clustal w/o numbers". In STEP 3, there is a checkbox "Be notified by email" and a "Submit" button. At the bottom, there is a link "contact us" and a note: "If you plan to use these services during a course please contact us."

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Choosing the Right Method of Multiple Sequence Alignment

2-Tcoffee

One of the latest multiple-sequence-alignment packages that you can use. With Tcoffee, you **can combine sequences** and **structures**, **evaluate an alignment**, or **merge** several alternative multiple alignments into a single unified result.



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Choosing the Right Method of Multiple Sequence Alignment

Continue.....

tcoffee.crg.cat/apps/

T
COFFEE

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T-Coffee
A collection of tools for Computing, Evaluating and Manipulating Multiple Alignments of DNA, RNA, Protein Sequences and Structures

Alignment

- [T-Coffee](#) Aligns DNA, RNA or Proteins using the default T-Coffee >> Cite
- [M-Coffee](#) Aligns DNA, RNA or Proteins by combining the output of popular aligners >> Cite
- [R-Coffee](#) Aligns RNA sequences using predicted secondary structures >> Cite
- [SARA-Coffee](#) Aligns RNA sequences using tertiary structure NEW >> Cite
- [Expresso](#) Aligns protein sequences using structural information >> Cite
- [PSI-Coffee](#) Aligns distantly related proteins using homology extension (slow and accurate) >> Cite
- [PSI/TM-Coffee](#) Align Proteins using Homology Extension against Reduced Databases >> Cite
- [Pro-Coffee](#) Aligns homologous promoter regions NEW >> Cite
- [Accurate](#) Automatically combine the most accurate modes for DNA, RNA and Proteins (experimental!)
- [Combine](#) Combines two (or more) multiple sequence alignments into a single one >> Cite

Evaluation

- [TCS](#) Evaluates your Alignment and outputs a Colored version indicating the local reliability. >> Cite
- [iRMSD-APDB](#) Evaluates Multiple Sequence Alignment using structural information with APDB and iRMSD. >> Cite
- [T-RMSD](#) Allows fine-grained structural clustering of a given group of related protein domains NEW >> Cite
- [Strike](#) Evaluation of protein MSAs using a single 3D structure >> Cite

Other

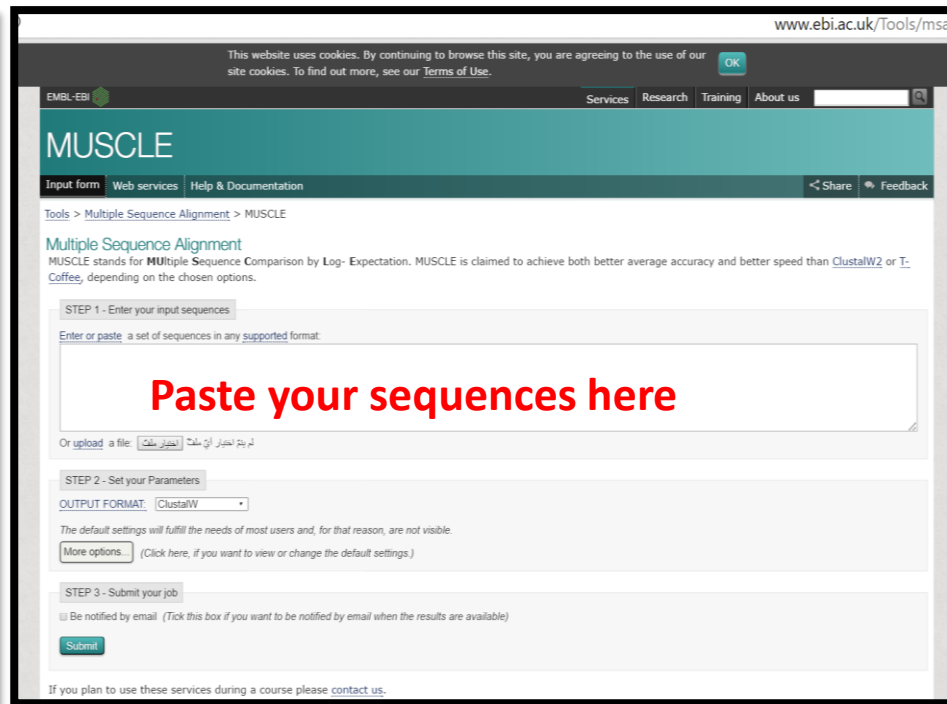
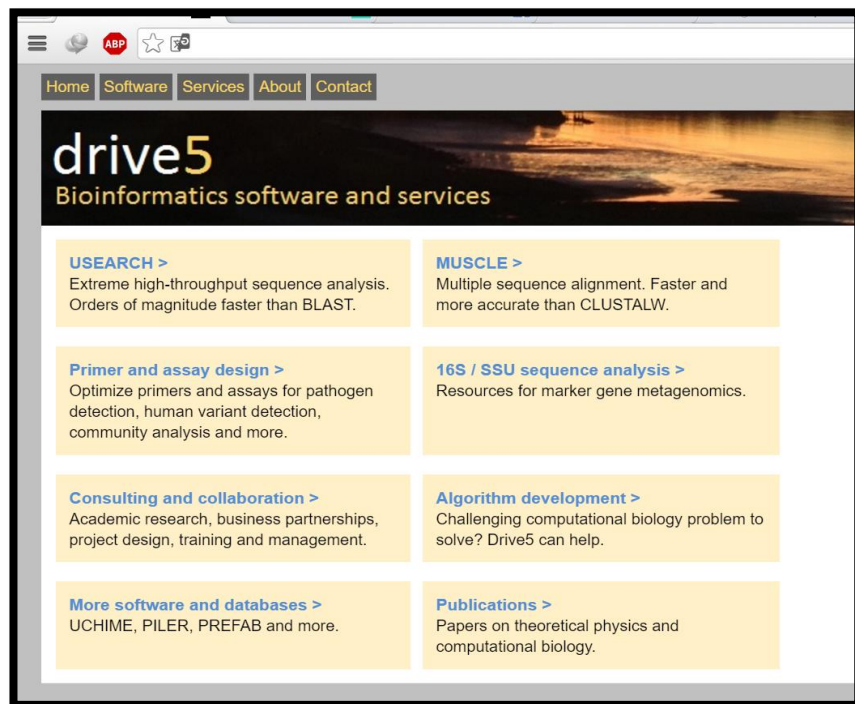
- [Advanced](#) Run your alignment using full featured T-Coffee options. >> Cite

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Choosing the Right Method of Multiple Sequence Alignment

3- MUSCLE

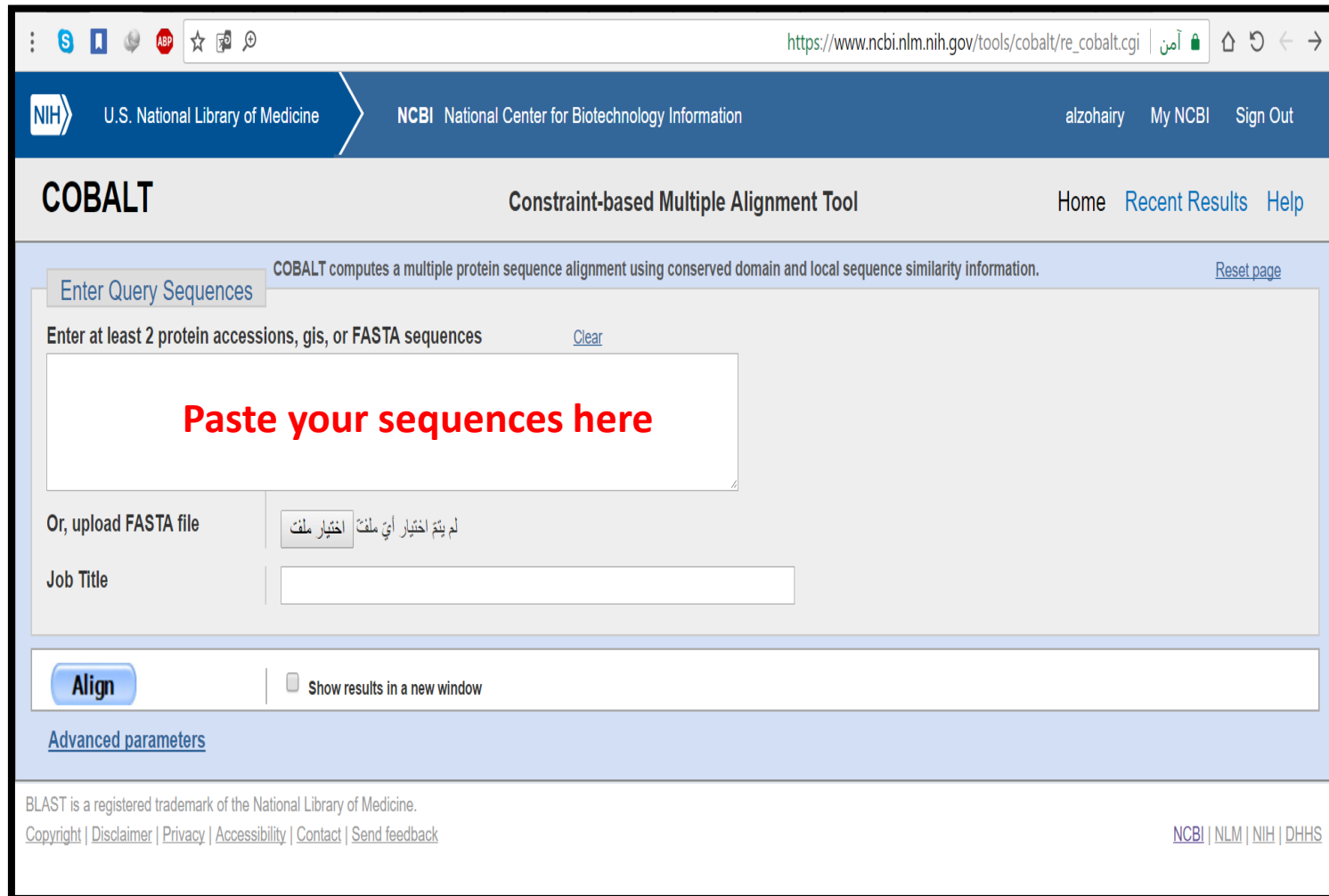
One of **the fastest** alignment methods around for aligning large set of sequences



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4- COBALT

Constraint-based Multiple Alignment Tool



The screenshot shows the COBALT web interface. At the top, there's a navigation bar with NIH and NCBI logos, and links for 'alzoairy', 'My NCBI', and 'Sign Out'. The main header displays 'COBALT' and 'Constraint-based Multiple Alignment Tool', with links for 'Home', 'Recent Results', and 'Help'. Below the header, a description states: 'COBALT computes a multiple protein sequence alignment using conserved domain and local sequence similarity information.' A 'Reset page' link is available. The main input area is titled 'Enter Query Sequences' and contains a large text box with the instruction 'Paste your sequences here' in red. Below this, there's a section for 'Or, upload FASTA file' with a file selection button labeled 'لم يتم اختيار أي ملف' and 'اختيار ملف'. A 'Job Title' input field is also present. At the bottom of the input section, there's an 'Align' button and a checkbox for 'Show results in a new window'. A link for 'Advanced parameters' is located below the input section. The footer contains a disclaimer about BLAST, copyright information, and links for 'NCBI', 'NLM', 'NIH', and 'DHHS'.

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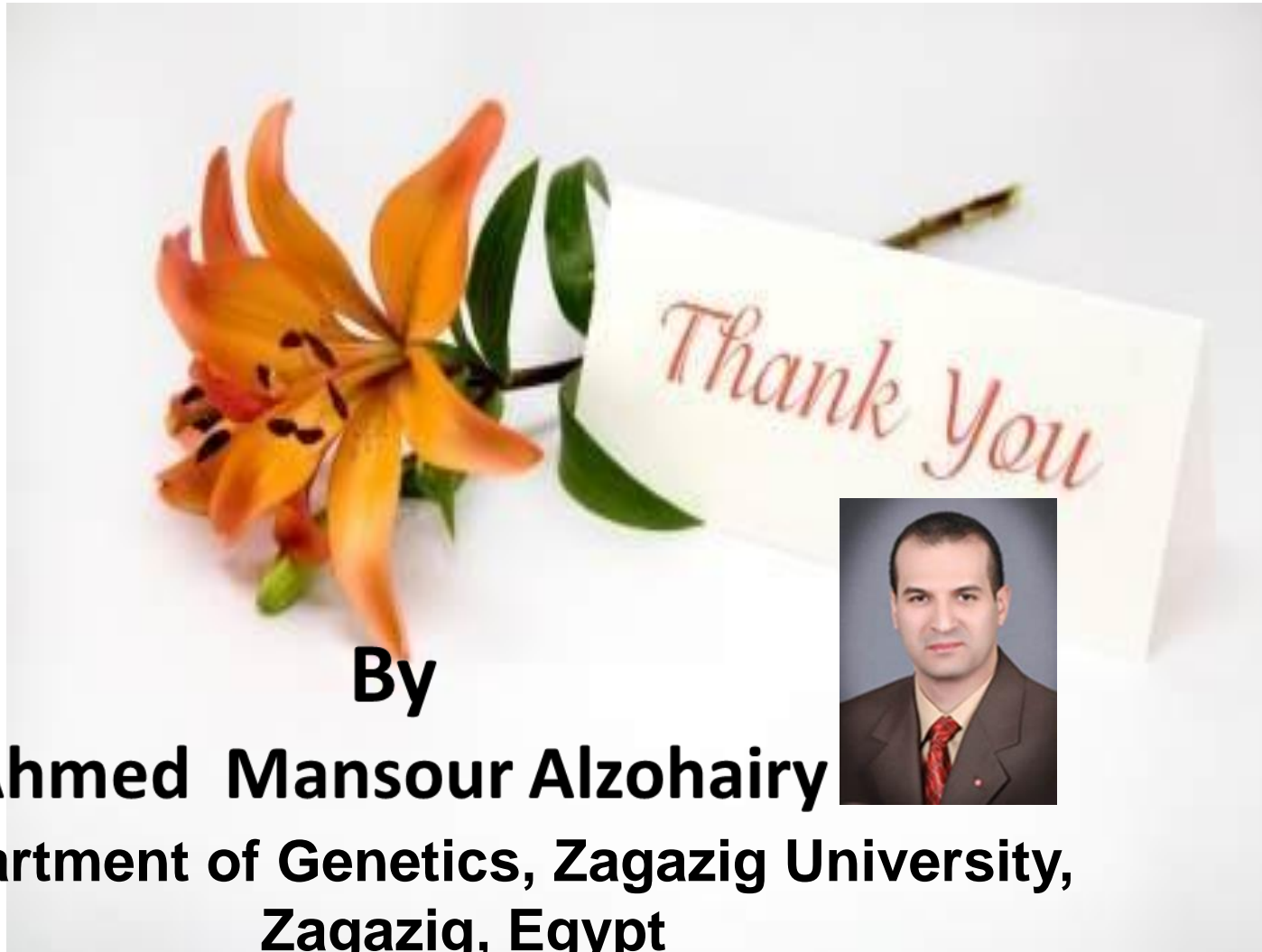


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- 1st Module: What Bioinformatics Can Do for You
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- 4th Module: How to Build a Multiple Sequence Alignment?
- 5th Module: Inferring Phylogenetic analysis using Jellview
- 6th Module: Advanced Molecular Concepts
- 7th Module: Inferring Protein Sequence (Structure & Function)
- 8th Module: RNAanalysis and Function
- 9th Module: Editing and Publishing Alignments in your Manuscript
- 10th Module: Building and Publishing Phylogenetic Trees
- 11th Module: Working with Protein 3-D Structures
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