Example 3. Analysis pipeline (Using RU-Net and WAC-Net in sequence) Open in Colab This example demonstrates how to use a pre-trained RU-Net and WAC-Net in sequence to get the dry mass values NOTE: If you're running this notebook on your local machine, please comment the code in the cell below In [1]: !git clone https://github.com/softmatterlab/Quantitative-Microplankton-Tracker.git %cd Quantitative-Microplankton-Tracker/examples/ %matplotlib inline In [2]: import sys sys.path.append("..") 1. Setup Imports the needed dependencies to run this tutorial. In [3]: import numpy as np import pandas as pd import tensorflow as tf import matplotlib as mpl import matplotlib.pyplot as plt from tqdm.notebook import tqdm from skimage.measure import label, regionprops import deeptrack as dt 2. Loading experimental holographic image We use a 50 frame experimental image sequence containing species in this tutorial. Experimental images are located in sample-data folder. exp data = np.load(In [4]: "../data/data figure2/Oxyrrhis data.npy" np.shape(exp_data) Out[4]: (50, 1024, 1280) 2.1. Visualising experimental image First frame of the experimental holographic sequence. In [5]: plt.figure(figsize=(12, 8)) plt.imshow(exp data[0], cmap="gray") plt.show() 200 400 600 800 1000 800 1200 200 400 600 1000 0 3. Using RU-Net on experimental images We use RU-Net model to segment planktons from the background, and to extract their properties. Pre-trained models are located in pretrained models folder 3.1. Load pre-trained RU-Net model In [6]: RUNet model = tf.keras.models.load model("../pre-trained-models/RUNet.h5", compile=False 3.2. Predictions followed by extracting properties predict image function extracts the 2d positions, dry mass and z-distance from predictions of RU-Net In [7]: def predict image(predicted image, threshold): 11 11 11 Outputs the predicted channels for regression U-nets Inputs: image and image threshold, Eg., 0.1 Outputs: xpositions, ypositions, zpositions, drymass, and eccentricity 11 11 11 Data = []X, Y = np.meshgrid(range(0, predicted_image.shape[2]), range(0, predicted_image.shape[1]) X = np.reshape(X, (predicted_image.shape[1] * predicted_image.shape[2])) Y = np.reshape(Y, (predicted_image.shape[1] * predicted_image.shape[2])) for i in range(predicted image.shape[0]): zpos = []mass = []xpos = []ypos = []ecc = []xyri = []predX = predicted image[i, :, :, 3] predY = predicted image[i, :, :, 4] gx1, gy1 = np.gradient(predX)gx2, gy2 = np.gradient(predY)I = gy1 + gx2I2 = np.zeros(I.shape) I2[I > threshold] = 1I3 = label(I2)prop = regionprops(I3) zs = predicted_image[i, :, :, 1] masses = predicted_image[i, :, :, 2] I3 = np.reshape(I3, (predicted_image.shape[2] * predicted_image.shape[1]) zs = np.reshape(zs, (predicted_image.shape[2] * predicted_image.shape[1]) masses = np.reshape(masses, (predicted image.shape[2] * predicted image.shape[1]) for j in range(np.amax(I3)): if prop[j].eccentricity < 1.2:</pre> ecc.append(prop[j].eccentricity) inds = np.where(I3 == j + 1)zpos.append(np.mean(zs[inds])) mass.append(np.mean(masses[inds])) xpos.append(np.mean(X[inds])) ypos.append(np.mean(Y[inds])) xyri.append([np.mean(X[inds]), np.mean(Y[inds]), np.mean(zs[inds]), np.mean(masses[inds]), xyri = np.array(xyri) Data.append(xyri) return Data 3.3. Preparing the images for prediction Split the 50 frame sequence into batches of 5 for prediction In [8]: batchsize = 5 data in batches = [exp_data[i : i + batchsize] for i in range(0, int(len(exp_data)), batchsize) Function to obtain RU-Net predictions on a batch of images In [9]: def get_prediction_batch(batch, model): return the prediction of a batch of images batch_reshape = np.expand_dims(batch, axis=3) prediction = model.predict(batch_reshape) return prediction 3.4. Predicting positions Get the positions of planktons from the RU-Net predictions. The positions are stored in position data. This process should take about a minute. In [10]: position_data = [] threshold = 0.1median_val = np.median(exp_data[0]) for i in tqdm(range(len(data in batches)), desc="Predicting batch "): batch = data in batches[i] prediction = get_prediction_batch(batch / median val, RUNet model positions = predict image(prediction, threshold for coords in positions: position_data.append(coords) 3.5. Highlighting the plankton positions Change frame no to higlight plankton positions in other frames In [11]: frame no = 10fig, ax = plt.subplots(figsize=(15,15)) ax.imshow(exp_data[frame_no], cmap='gray') X = position data[frame no][:, 0] Y = position_data[frame_no][:, 1] for i in range (0, len(X), 1): x0 = X[i]y0 = Y[i]ax.plot(x0, y0,'o', ms=50, markerfacecolor="None", markeredgecolor="#e377c2", markeredgewidth=4, al pha=0.7)plt.axis("off") plt.show() 0 O O O 0 0 4. Plankton trajectories 4.1. Getting plankton traces We use get particle trace function to link the plankton positions in all the frames to individual trajectories. In [12]: particles, completed traces = dt.get particle trace(position data) print('Particles', np.shape(particles), 'completed traces', np.shape(completed traces)) all_traces = completed_traces + particles[0] print("Number of particle traces ", len(all_traces)) Particles (1, 110) completed traces (30,) Number of particle traces 140 4.2. Formatting trajectories into a dataframe Formatting trajectories into dataframes makes it easy to read the trajectories In [13]: list of dataframes = [] for i in range(len(all traces)): df = pd.DataFrame(all traces[i], columns = ['frame', 'x', 'y', 'z', 'drymass']) df = df.assign(particle = [i]*len(all traces[i])) list of dataframes.append(df) particle_data = pd.concat(list_of_dataframes, ignore_index = True) particle data = particle data[particle data.x != 0] #deletes all rows where x=0 and y=0, because that's probably some weird prediction at the edge particle_data = particle_data[particle_data.y != 0] particle data Out[13]: z drymass particle frame 0 1145.125000 601.875000 -0.054241 0.187524 601.833333 -0.081731 0.178887 1 1144.833333 602.285714 -0.081766 0.184561 2 1145.000000 3 1145.500000 601.500000 -0.061431 0.125581 0 4 1145.333333 602.333333 -0.109081 0.147072 4201 1043.500000 783.000000 0.079965 0.048296 135 136 4202 49 230.000000 838.500000 0.121690 0.077743 0.190252 0.102897 49 10.200000 861.400000 137 4203 1013.250000 4204 49 565.000000 0.169295 0.102271 138 1023.000000 -0.015778 0.117703 4205 49 767.000000 139 4206 rows × 6 columns 4.3. Processing trajectories Trajectories are processed and relevant information is stored in traj data array. In [14]: trajs = []trajs_lengths = [] positions=particle_data.set_index(['particle', 'frame'])[['x', 'y']].unstack() for index, rows in positions.iterrows(): traj_len=np.count_nonzero(~np.isnan(rows[['x', 'y'][0]])) trajs_lengths.append(traj_len) trajs.append(np.transpose(['x','y'][0]].to_numpy(), rows[['x', 'y'][1]].to_numpy()])) In [15]: # Appending the frame number before the nodes traj data = [] for i in range(len(trajs)): non_nan_indices = np.where(np.isnan(trajs[i][:,0]) == False)[0] frame_no = np.reshape(non_nan_indices, (len(non_nan_indices),1)) frame_and_centroids = np.append(frame_no, trajs[i][non_nan_indices], axis=1) traj_data.append(frame_and_centroids) 4.4. Visualising the trajectories Trajectories are plotted over the final frame of the sequence for easy visualization. The index of the trajectory helps to isloate individual single plankton traces. In [16]: fig, ax=plt.subplots(figsize=(18, 12)) ax.imshow(exp_data[-1], cmap="gray") i = 0for trajectory in traj_data: ax.plot(trajectory[:,1], trajectory[:,2], "red", alpha=0.7, linewidth=1.5) ax.text(trajectory[:,1][-1], trajectory[:,2][-1], str(i), color="white", fontsize=16) i += 123 125 11426 115 **3B**27 200 97 400 94 600 63 133 65 21/09 134 70 13690 83 **82** 1000 200 400 600 800 1000 1200 5. Dry mass prediction by WAC-Net We now crop 64 x 64 pixel images around the single plankton traces, and predict a improved dry mass values with a pre-trained WAC-Net. 5.1. Selecting planktons Pick a plankton trace by their number in the above figure for the dry mass prediction. Alternatively, dry mass can also be predicted for all the planktons In [17]: $trace_no = 58$ Function to obtain the crop around the plankton position def croppedimage(image, position, window=32): In [18]: crops a region of 2*window pixels around the position [a, b, c, d] = [int(position[0] - window), int(position[0] + window), int(position[1] - window), int(position[1] + window), return image[c:d, a:b] 5.2. Obtaining sequence of crops for a single plankton trajectory Crops are stored in cropped sequence In [19]: cropped sequence = [] single_plankton_traj = traj_data[trace_no] for i in range(len(single_plankton_traj)): crop = croppedimage(exp data[int(single plankton traj[i][0]) single plankton traj[i][1:3] cropped_sequence.append(crop) Visuazlise first frame in the cropped plankton sequence In [20]: plt.imshow(cropped_sequence[0], cmap="gray") plt.title("Predator hologram (\$\it{Oxyrrhis\ Marina}\$)") plt.show() Predator hologram (Oxyrrhis Marina) 10 20 30 40 50 60 5.3. Loading pre-trained WAC-Net model for dry mass model We define the WAC-Net model and load the weights from a pre-trained model. In [21]: from tensorflow import keras Sequential = keras.models.Sequential Model = keras.models.Model Dense = keras.layers.Dense Conv = keras.layers.Conv2D Conv1D = keras.layers.Conv1D ConvL = keras.layers.LocallyConnected2D Pool = keras.layers.MaxPooling2D Input = keras.layers.Input Concat = keras.layers.Concatenate TimeDistributed = keras.layers.TimeDistributed Flatten = keras.layers.Flatten Lambda = keras.layers.Lambda K = keras.backendmodel = Sequential() model.add((Conv (32, kernel_size=3, strides=1, activation="relu", input_shape=(64, 64, 1), model.add(Pool(2)) model.add((Conv(64, kernel_size=3, strides=1, activation="relu"))) model.add(Pool(2)) model.add((Conv(128, kernel_size=3, strides=1, activation="relu"))) model.add(Pool(2)) model.add((Conv(256, kernel size=3, strides=1, activation="relu"))) model.add((Flatten())) model.add((Dense(128, activation="relu"))) model.add((Dense(128, activation="relu"))) stack = Input (model.input shape) vectors = TimeDistributed(model)(stack # time distributed applies a layer to every temporal slice of the input weights = Conv1D(128, 1, padding="same") (vectors) weights = Conv1D(128, 1, padding="same") (weights) weights = Conv1D(1, 1, padding="same") (weights) def merge_function(tensors): x = tensors[0]weights = tensors[1] weights = K.softmax(weights, axis=1) merged = K.sum(x * weights, axis=1) return merged merge_layer = Lambda(merge_function) merged = merge_layer([vectors, weights]) merged = Dense(32, activation="relu") (merged) merged = Dense(32, activation="relu") (merged) out = Dense(2) (merged) model = Model(stack, out) model.compile(tf.keras.optimizers.Adam(1r=0.0001, amsgrad=True), loss="mae") model.load weights ("../pre-trained-models/WACNet dry mass.h5") 5.4. Normalising the crops Normalise and reshape the images for WAC-Net prediction In [22]: | def Normalise(images, batch = 15): Normalised = [] for i in range(len(images)): Normalised.append(images[i]/np.median(images[i])) Normalised = np.array(Normalised)-1 proc = [] #sliding window for i in range(len(Normalised)-batch+1): proc.append(Normalised[i:i+batch]) return np.expand dims(proc, axis = -1) In [23]: # Funtion to convert the predicted dry mass to real dry mass units (pico grams) **def** real dm(p, a=209.16, b=0.28, sp ri inc = 0.21):return (p*a+b)/sp ri inc 5.5. Prediction on cropped sequences Experimental sequences are normalized with batch size of 15. batch value can be increased or decreased to generate a sliding window of the experimental sequence. The WAC-Net assigns weights to the images with in sliding windows to predict the best possible value for dry mass considering all the frames in a sliding window. In [25]: | prediction = model.predict(Normalise(cropped sequence, batch = 155.6. Convert the predictions to real dry mass units In [26]: drymass prediction = real dm(prediction[:,0]) 5.7. Visualising dry mass predictions

240 - 220 - 200 - 5	10 15 20 Time (seconds)	25 30 35		