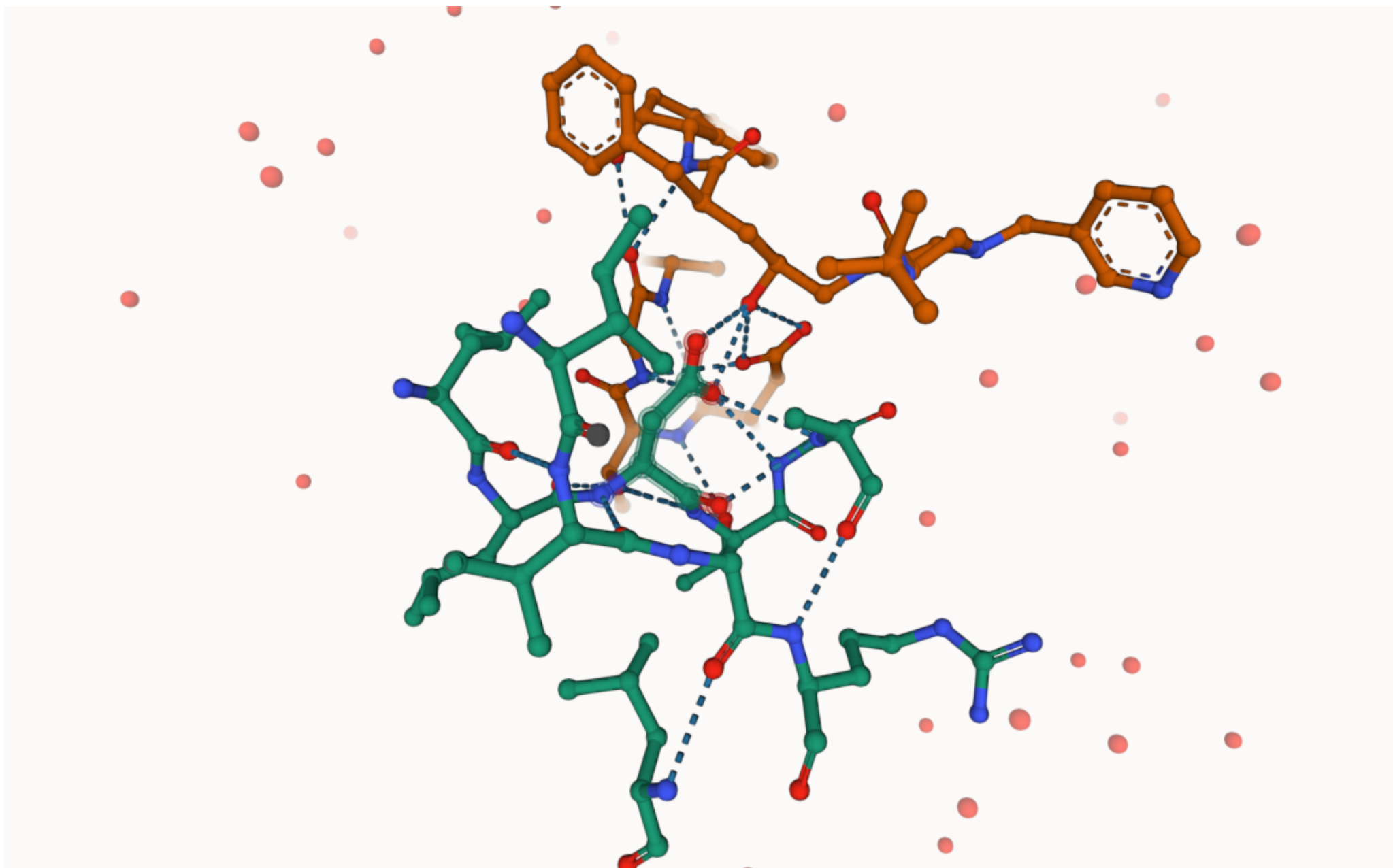


```

1 ---
2 title: "Class 9 Forrest Wang"
3 format: pdf
4 ---
5
6 ```{r}
7 setwd("~/Downloads")
8 Class9data <- read.csv("Data Export Summary.csv")
9 ```
10 1. PDB Intro
11
12 Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.
13 A1: X-Ray is 86.3%, EM is 6.58%, total is 92.9%
14
15 Q2: What proportion of structures in the PDB are protein?
16 A2: Protein accounts for 171828 out of 197512, equaling to 87.0%
17
18 Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?
19 A3: There are 240 total HIV-1 protease structures
20
21 2. PDB Visualization
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37 Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?
38 A4: We only see 1 atom/h2o molecule in this structure maybe because the environment is basic and protons are being pulled off the water
39
40 Q5: There is a critical "conserved" water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have
41 A5: The conserved H2O molecule closest to the ligand binding site is HOH 308
42
43 Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend "Ball & Stick" for these side-chains). Add this figure to your Quarto document.
44

```





```

45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60 3. Intro to Bio3D
61 ```{r}
62 library(bio3d)
63 ```
64
65 Read PDB File
66 ```{r}
67 pdb <- read.pdb("1hsg")
68 ```
69 File Summary:
70 ```{r}
71 pdb
72 ```

```

Call: read.pdb(file = "1hsg")

Total Models#: 1

Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)

Protein Atoms#: 1514 (residues/Calpha atoms#: 198)

Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 172 (residues: 128)

Non-protein/nucleic resid values: [HOH (127), MK1 (1)]

Protein sequence:

```

PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWKPKMIGGIGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF

```

+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call

Nucleic acid Atom# : 0 (residues/phosphate atom# : 0)

```
92 Normal Mode Analysis
```

```
93 {r}
```

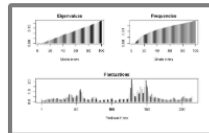
```
94 m <- nma(adk)
```

```
95 plot(m)
```

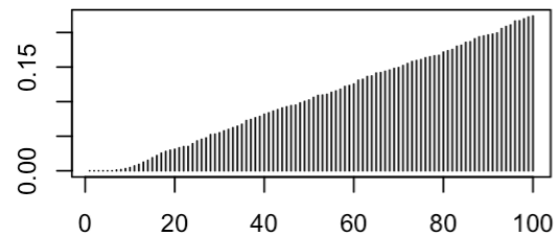
```
96
```

Building Hessian... Done in 0.072 seconds.
Diagonalizing Hessian... Done in 0.552 seconds.

R Console

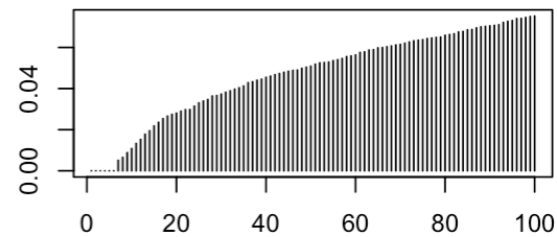


Eigenvalues



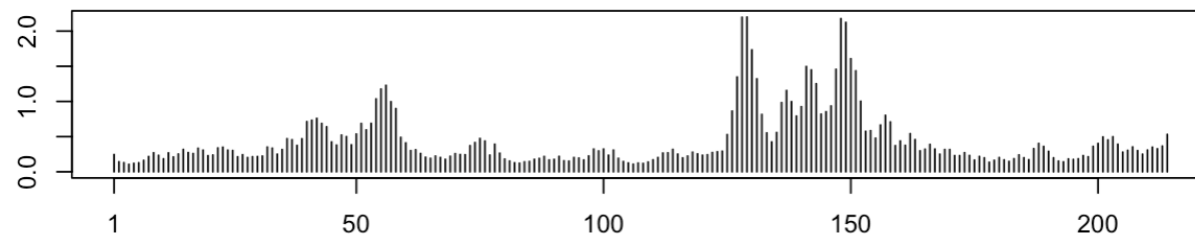
Mode index

Frequencies



Mode index

Fluctuations



Residue index

```
97 View like movie
```

```
98 {r}
```

```
99 mktrj(m, file="adk_m7.pdb")
```

```
100
```

```
101
```

```
102
```

```
103 4. Comparative structure analysis of Adenylate Kinase
```

```
104
```

```
105 Q10. Which of the packages above is found only on BioConductor and not CRAN?
```

```
106 A10: "msa"
```

```
107
```

```

107
108 Q11. Which of the above packages is not found on BioConductor or CRAN?
109 A11: "Grantlab/bio3d-view"
110
111 True or False? Functions from the devtools package can be used to install packages from GitHub and BitBucket?
112 A; True
113
114 ```{r}
115 library(bio3d)
116 aa <- get.seq("lake_A")
117 ```
118
119 Visualize AA
120 ```{r}
121 aa
122 ```
123
124 Q13. How many amino acids are in this sequence, i.e. how long is this sequence?
125 A13: 214 AAs
126
127 BLAST Search
128 ```{r}
129 # Blast or hmmer search
130 b <- blast.pdb(aa)
131 ```
132
133 ```{r}
134 b
135 ```

```

\$hit.tbl

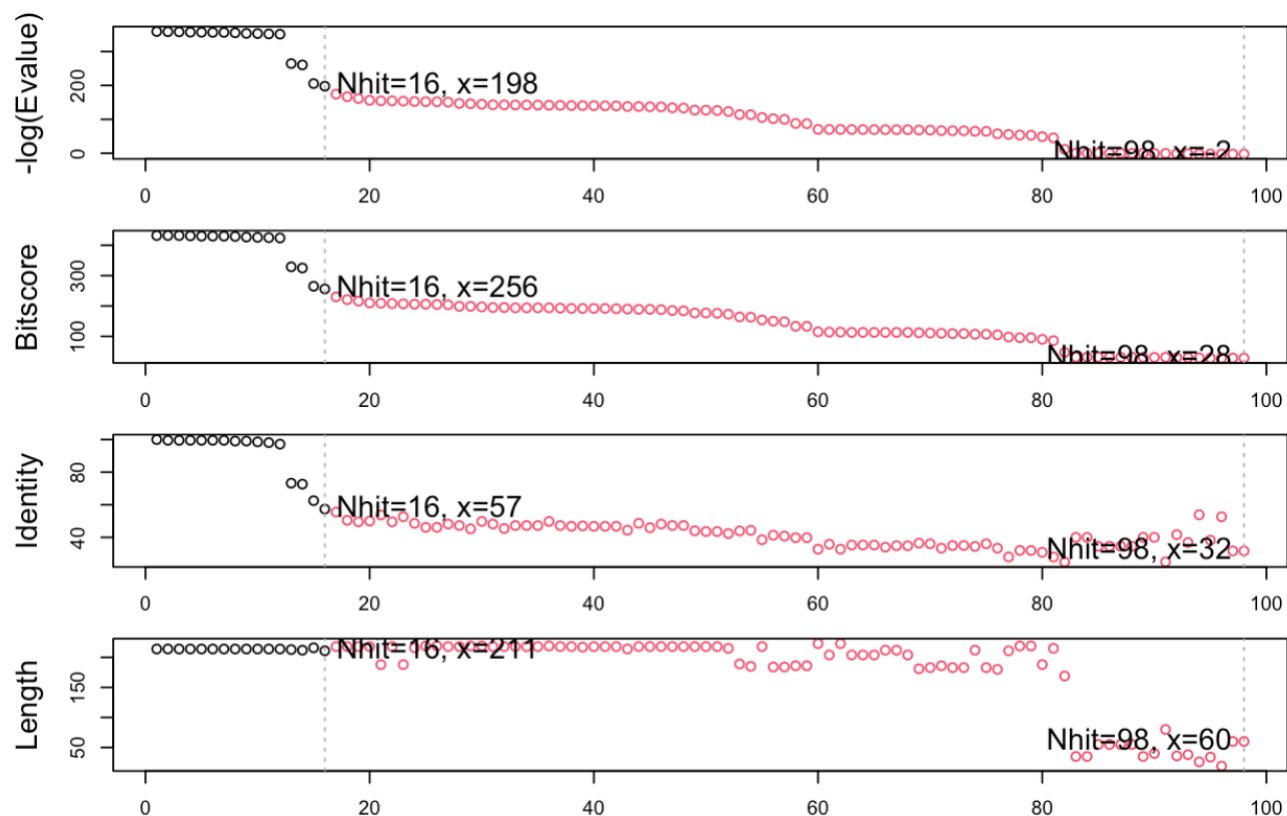
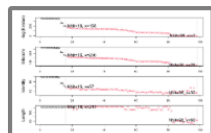
	queryid	subjectids	identity	alignmentlength	mismatches	gapopens	q.start	q.end	s.start	s.end	evaluate	bitscore
1	Query_44619	1AKE_A	100.000	214	0	0	1	214	1	214	1.32e-156	432
2	Query_44619	4X8M_A	99.533	214	1	0	1	214	1	214	2.36e-156	432
3	Query_44619	6S36_A	99.533	214	1	0	1	214	1	214	3.47e-156	432
4	Query_44619	6RZE_A	99.533	214	1	0	1	214	1	214	9.94e-156	431
5	Query_44619	4X8H_A	99.533	214	1	0	1	214	1	214	1.31e-155	430
6	Query_44619	3HPR_A	99.533	214	1	0	1	214	1	214	1.86e-155	430
7	Query_44619	1E4V_A	99.533	214	1	0	1	214	1	214	1.96e-155	430
8	Query_44619	5EJE_A	99.065	214	2	0	1	214	1	214	5.88e-155	429
9	Query_44619	1E4Y_A	99.065	214	2	0	1	214	1	214	3.40e-154	427
10	Query_44619	3X2S_A	98.598	214	3	0	1	214	1	214	5.70e-154	426
11	Query_44619	6HAP_A	98.131	214	4	0	1	214	1	214	1.69e-153	425
12	Query_44619	6HAM_A	97.196	214	6	0	1	214	1	214	3.44e-153	424
13	Query_44619	4K46_A	73.239	213	57	0	1	213	1	213	1.53e-115	329
14	Query_44619	4NP6_A	72.642	212	58	0	2	213	5	216	8.49e-114	325
15	Query_44619	3GMT_A	62.500	216	75	1	2	211	10	225	6.64e-90	265
16	Query_44619	4PZL_A	57.346	211	86	2	2	209	26	235	1.58e-86	256
17	Query_44619	5G3Y_A	55.505	218	88	2	1	214	1	213	2.34e-76	230

```

136 Plot!
137 ```{r}
138 hits <- plot(b)
139 hits
140 ```

```

R Console



```

141 Top Hits:
142 ```{r}
143 # List out some 'top hits'
144 head(hits$pdb.id)
145 ```

```

```
[1] "1AKE_A" "6S36_A" "6RZE_A" "3HPR_A" "1E4V_A" "5EJE_A"
```

```

146
147 ```{r}
148 hits <- NULL
149 hits$pdb.id <- c('1AKE_A', '6S36_A', '6RZE_A', '3HPR_A', '1E4V_A', '5EJE_A', '1E4Y_A', '3X2S_A', '6HAP_A', '6HAM_A', '4K46_A', '3GMT_A', '4P

```



```

150 ^ ``
151
152
153 Download related files
154 ^ ``{r}
155 files <- get.pdb(hits$pdb.id, path="pdbs", split=TRUE, gzip=TRUE)
156 ^ ``

```

```

=====
100%ZE.pdb.gz exists. Skipping downloadWarning: pdbs/3HPR.pdb.gz exists. Skipping downloadWarning: pdbs/1E4V.pdb.gz exists.
Skipping downloadWarning: pdbs/5EJE.pdb.gz exists. Skipping downloadWarning: pdbs/1E4Y.pdb.gz exists. Skipping
downloadWarning: pdbs/3X2S.pdb.gz exists. Skipping downloadWarning: pdbs/6HAP.pdb.gz exists. Skipping downloadWarning:
pdbs/6HAM.pdb.gz exists. Skipping downloadWarning: pdbs/4K46.pdb.gz exists. Skipping downloadWarning: pdbs/3GMT.pdb.gz
exists. Skipping downloadWarning: pdbs/4PZL.pdb.gz exists. Skipping download

```

```

157
158 Align & Repurpose Structures
159 Align
160 ^ ``{r}
161 # Align related PDBs
162 pdbs <- pdbaln(files, fit = TRUE, exe="msa")
163 ^ ``

```

```

Reading PDB files:
pdbs/split_chain/1AKE_A.pdb
pdbs/split_chain/6S36_A.pdb
pdbs/split_chain/6RZE_A.pdb
pdbs/split_chain/3HPR_A.pdb
pdbs/split_chain/1E4V_A.pdb
pdbs/split_chain/5EJE_A.pdb
pdbs/split_chain/1E4Y_A.pdb
pdbs/split_chain/3X2S_A.pdb
pdbs/split_chain/6HAP_A.pdb
pdbs/split_chain/6HAM_A.pdb
pdbs/split_chain/4K46_A.pdb
pdbs/split_chain/3GMT_A.pdb
pdbs/split_chain/4PZL_A.pdb
  PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
..  PDB has ALT records, taking A only, rm.alt=TRUE
.... PDB has ALT records, taking A only, rm.alt=TRUE
.   PDB has ALT records, taking A only, rm.alt=TRUE
...

```

Extracting sequences

```

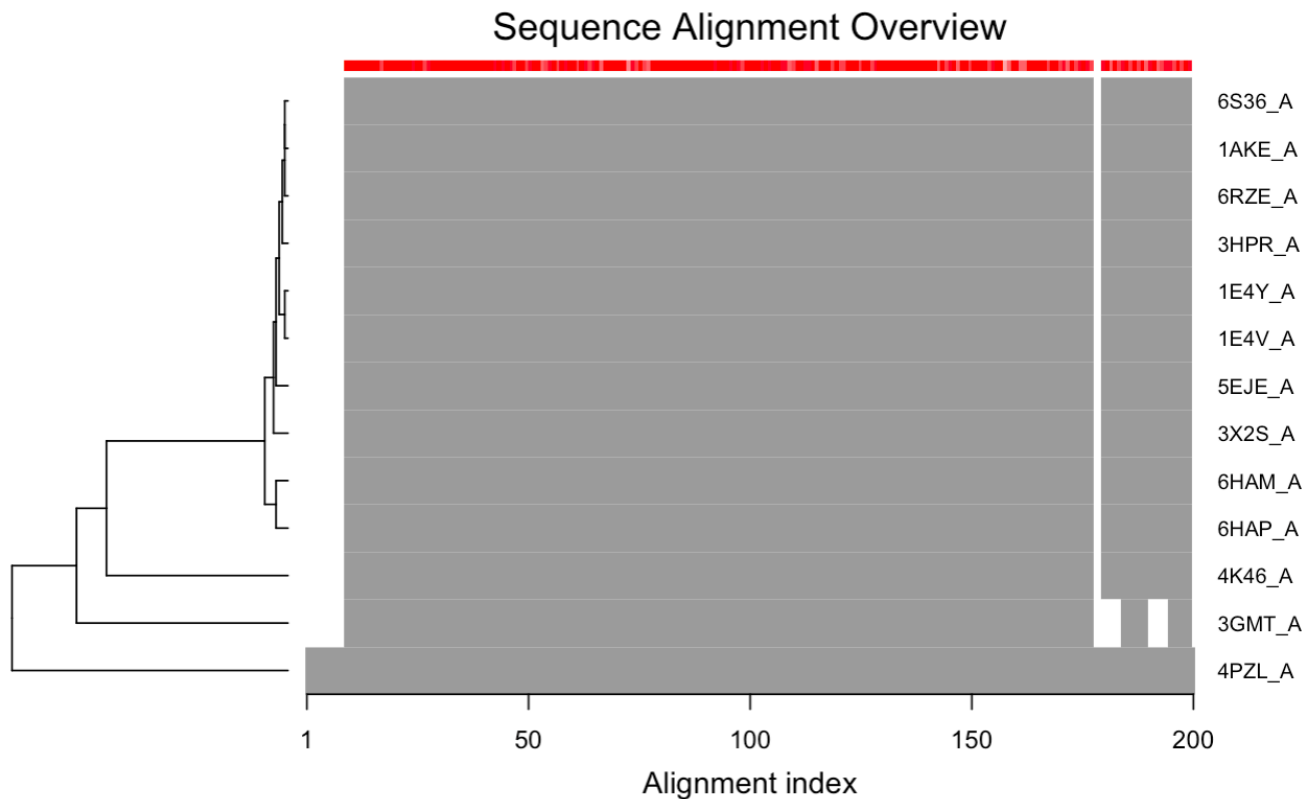
pdb/seq:1  name: pdbs/split_chain/1AKE_A.pdb

```

```

165 Visualize:
166 ```{r}
167 # Vector containing PDB codes for figure axis
168 ids <- basename.pdb(pdb$id)
169
170 # Draw schematic alignment
171 plot(pdb, labels=ids)
172 ```

```



```

173
174 Annotate PDB Structures
175 ```{r}
176 anno <- pdb.annotate(ids)
177 unique(anno$source)
178 anno
179 ```

```

R Console

data.frame
13 x 17

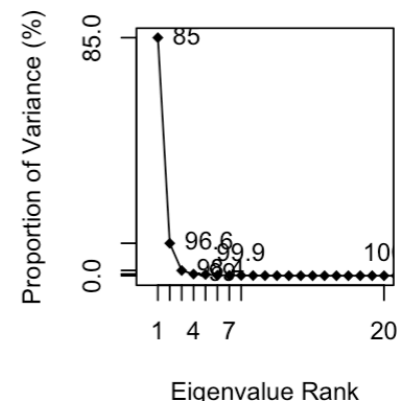
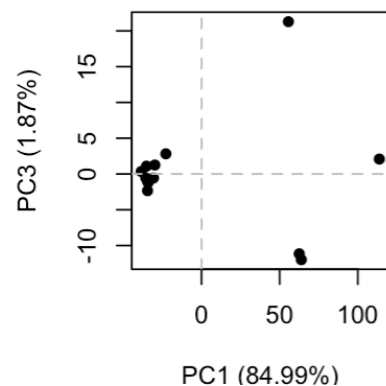
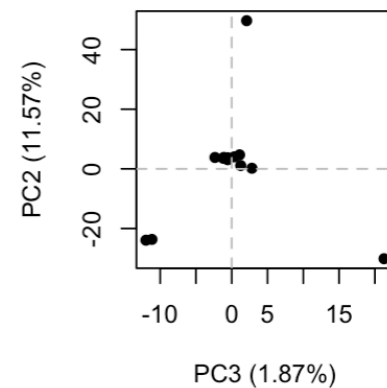
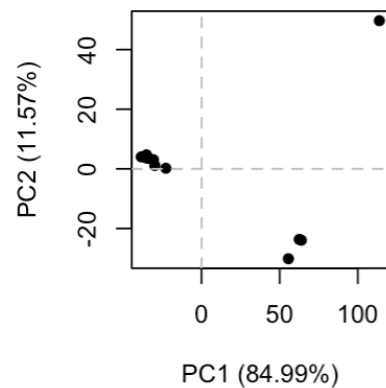
	structureId <chr>	chainId <chr>	macromoleculeType <chr>	chainLength <int>	experimentalTechnique <chr>	resolution <dbl>
1AKE_A	1AKE	A	Protein	214	X-ray	2.00
6S36_A	6S36	A	Protein	214	X-ray	1.60
6RZE_A	6RZE	A	Protein	214	X-ray	1.69
3HPR_A	3HPR	A	Protein	214	X-ray	2.00
1E4V_A	1E4V	A	Protein	214	X-ray	1.85
5EJE_A	5EJE	A	Protein	214	X-ray	1.90
1E4Y_A	1E4Y	A	Protein	214	X-ray	1.85
3X2S_A	3X2S	A	Protein	214	X-ray	2.80
6HAP_A	6HAP	A	Protein	214	X-ray	2.70
6HAM_A	6HAM	A	Protein	214	X-ray	2.55

1-10 of 13 rows | 1-7 of 17 columns

Previous 1 2 Next

180
181 Principal Component Analysis

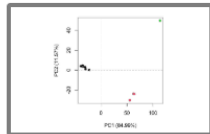
```
182 ```{r}
183 pc.xray <- pca(pdbb)
184 plot(pc.xray)
185 ```
```



```

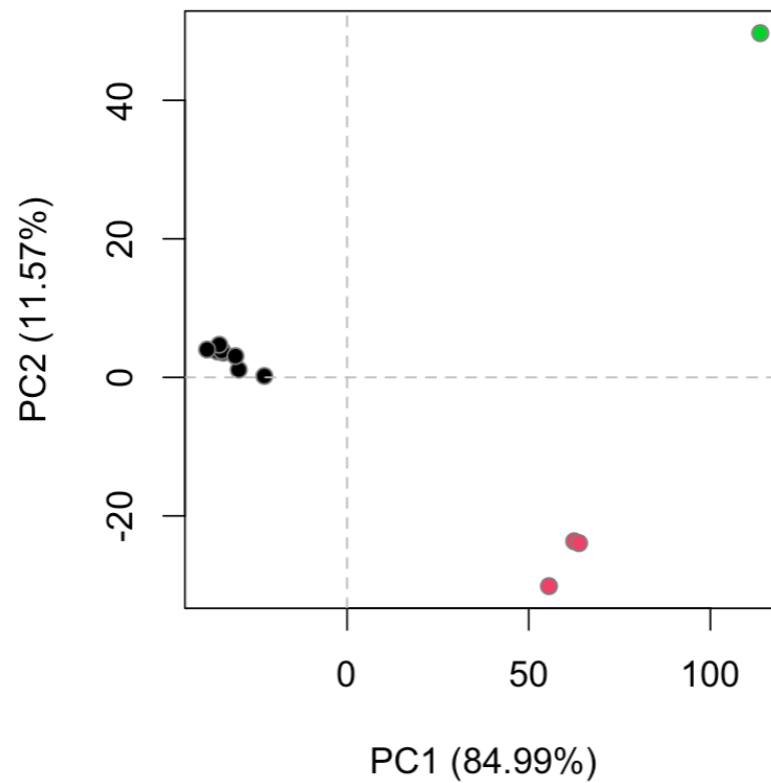
186
187 ```{r}
188 # Calculate RMSD
189 rd <- rmsd(pdbx)
190
191 # Structure-based clustering
192 hc.rd <- hclust(dist(rd))
193 grps.rd <- cutree(hc.rd, k=3)
194
195 plot(pc.xray, 1:2, col="grey50", bg=grps.rd, pch=21, cex=1)
196 ```

```



Warning: No indices provided, using the 284 non NA positions

R Console



```

197
198
199
200
201
202

```