Rarefaction curve

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Read the biom file for the mines Pmt and Topaz.

##

```
library("phyloseq")
library("ggplot2")
library("RColorBrewer")
library("patchwork")
library(vegan)
## Loading required package: permute
## Loading required package: lattice
## This is vegan 2.6-2
setwd("~/metagenomas")
raw_metagenomes <- import_biom("metagenomes_Pmt_S1.biom")</pre>
raw_metagenomes@tax_table@.Data <- substring(raw_metagenomes@tax_table@.Data, 4)
colnames(raw_metagenomes@tax_table@.Data)<- c("Kingdom", "Phylum", "Class", "Order", "Family", "Genus",</pre>
unique(raw_metagenomes@tax_table@.Data[,"Kingdom"])
## [1] "Bacteria" "Archaea"
                                 "Eukaryota" "Viruses"
raw_metagenomes <- subset_taxa(raw_metagenomes, Kingdom == "Bacteria")</pre>
Extract the OTUs and obtain the maximun number of counts between the samples.
otu <- otu_table(raw_metagenomes)</pre>
otu <- as.data.frame(t(otu))</pre>
sample_names <- rownames(otu)</pre>
S <- specnumber(otu)
## Pmt2_S287
               S1_S286
        7887
                   7981
```

```
raremax <- min(rowSums(otu))
raremax</pre>
```

[1] 2184595

We apply rarefy with the following parameters:

- Database: the transpose of the otu database.
- sample: a vector from 100000 to maximun number of counts with step 100000.
- Standar error: TRUE
- MARGIN: 2

With the rarefy function, we obtain the species richness in random samples of some specified size. This was calculated for a vector of samples from 100000 to the maximum of the two samples with steps of 100000. In this way we obtained the expected richness for each element of the vector and the standard error.

```
rare1 <- rarefy(t(otu), sample= c(seq(100000, 2000000), by = 100000), raremax), se = T, MARG = 2) rare_t <- t(rare1)
```

Save the data obtained by the rarefy function and compute the standar deviasion for each sample.

Now we can use the library ggplot to obtain the rarefaction curve with the database obtained.

