ImageSumming

1 Getting started

ImageSumming^A helps to create summed patches from experimental measurements (Image-Summing notebook) and visualizes DeepSTORM prediction results to support parameter adjustments (VisualizeDeepSTORM2DLocs notebook). Some smaller AddOns are collected that have been proven to be helpful for the workflow (DeepSTORM2DAddOns notebook).

DeepSTORM2D is used as Colab notebook in the ZeroCostDL4Mic framework.^{[1][2]}

Execute the code listing in an anaconda prompt to:

- create a virtual environment in anaconda and activate it
- change the directory to the *.whl file and install it
- additionally install jupyter nbextensions and activate them
- run the notebook process

Configure the notebook settings on the first run by clicking on the Nbextensions menu and selection "Collapsible Headings" and "Hide input all" (Fig. 1).

Listing 1: Commands to install *ImageSumming* and jupyter extensions in the anaconda prompt.

```
conda create --name ImageSumming
conda activate ImageSumming
cd path_to_file
conda install pip
pip install jupyterlab
pip install ImageSumming-XXXXX-py3-none-any.whl
pip install jupyter_contrib_nbextensions
jupyter notebook
```

Ahttps://github.com/JohannaRahm/ImageSumming

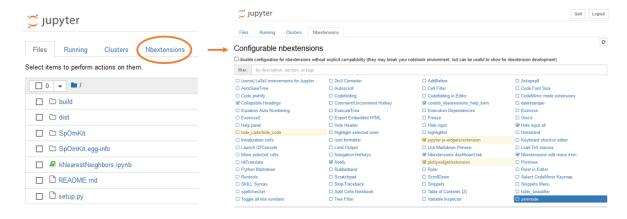


Fig. 1: Configure jupyter notebook settings.

2 General information

2.1 How to start and run a notebook

Change the directory to the notebooks and start them by typing "jupyter notebook" in the anaconda prompt. The process is opened in a browser. Select a notebook, click on restart the kernel to load the widgets (1), click on show codecell inputs to hide code (2), collapse headings if needed (3) (Fig. 2). Each notebook defines the needed input files, parameters can be adjusted, the process executed and results saved, all via userfriendly widgets.

Listing 2: Commands to run ImageSumming in the anaconda prompt. conda activate ImageSumming cd path_to_notebooks jupyter notebook kNearestNeighbors Last Checkpoint: 2 hours ago (unsaved changes) Edit View Cell Kernel Widgets Help Hide Code Insert ⟨♪ | □ | □ | ✓ ► Run ■ C Markdown pant) 10 kNearestNeighbors š

Fig. 2: Start and run a notebook.

2.2 Adjust default parameters

To adjust the default parameters "Hide input all" has to be deactivated to see the code. All adjustable settings are highlighted as comments, just change the values to your needs (Fig. 3).

```
from SpOmKit.widgets import widgetkNearestNeighbors
from SpOmKit.tools import kNearestNeighbors
from SpOmKit.save import savekNearestNeighbors
import numpy as np
import plotly
import plotly.graph_objects as go
import plotly.figure_factory as ff
import plotly.express as px
import plotly.io as pio
import matplotlib.pyplot as plt
n_centers = 2  # adjust the number of center files
n_neighbors = 2  # adjust the number of neighbor files
```

Parameters

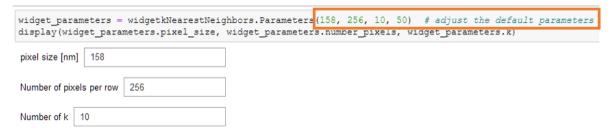


Fig. 3: Two examples of where to adjust the default parameters in the code. The places are marked with # comments.

3 Input filetypes

The notebooks require *.tif files that contain multiple frames of a measurement and corresponding localization files. We use Picasso localization files as ground truth. The files have to be converted to be used in DeepSTORM2D ZeroCostDL4Mic (=DeepSTORM file format) (fig. 4).

Picasso hdf5 format

	frame	х	у	photons	SX	sy	bg	lpx	lpy	ellipticity	net_gradient
0	0	1.4413762	254.61617	23015.182	1.6556486	1.3295391	371.85214	0.01742	0.02595	0.19696	15884.189
1	0	1.6262058	329.20697	12868.999	1.0678529	1.2171849	305.56125	0.01796	0.01505	0.12268	17199.262
2	0	1.8233835	439.0251	4496.8906	1.1016792	1.220813	210.9673	0.03491	0.03094	0.09758	5861.6675

Picasso csv format

"id", "frame", "x [nm]", "y [nm]", "sigma [nm]", "intensity [photon]", "offset [photon]", "bkgstd [photon]", "uncertainty_xy [nm]" 0,0,201.31,6689.55,141.18,67220,557,0,0.85 1,0,224.16,11655.38,130.04,15519,299,01.66 2,0,252.65,9010.93,123.40,15052,489,0,1.76

DeepSTORM csv format

,frame,x [nm],y [nm],Photon #,Sigma [nm] 1,1.0,201.31,6689.55,141.18,67220 2,1.0,224.16,11655.38,130.04,15519 3,1.0,252.65,9010.93,123.4,15052

Fig. 4: Example of hdf5 file format of Picasso (top) csv file format of Picasso (middle) csv file format of DeepSTORM (bottom).

4 Notebooks

4.1 ImageSumming

This notebook takes movies and localization files of multiple measurements, cuts patches and randomly summs them. It is compatible with DeepSTORM and Picasso file formats.

4.1.1 Parameters

Following parameters have to be defined. The pixel size in nm. The camera offset in px intensitiy (average px intensity with closed shutter), as the offset is only regarded once and is subtracted from the final summed image (number of summed images - 1 time). The final patch size in px, the DeepSTORM notebook handles input images with a maximum size of final patch size - 1, if a patch size of 26x26 px² should be the input in DeepSTORM notebook, the patch size has to be set to 27 in the ImageSumming notebook. The number of patches created defines the number of randomly cropped patches from the input measurement *.tif files. Number of summed patches created defines the final number of summed patches saved as *.tif movies, where each patch is a frame and corresponding localizations are saved as csv in DeepSTORM format, the patches for summed are drawn from the created patches and only used once per summed patch. Define the number of patches to be summed together. Min emitters per patch defines the minimum number of emitters a patch has to contain (before summed) to be used, because empty patches do not contain information for the network.

Parameters

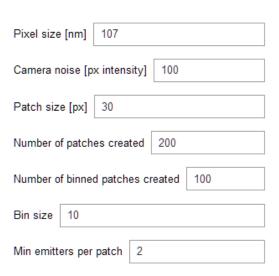


Fig. 5: Parameters of ImageSumming Notebook.

4.1.2 Input

Visualization of the measurement inputs (fig. 6). A chosen measurement is visualized framewise and localizations are marked with crosses. The mean density per frame is calculated to help determining the amount of images to be summed.

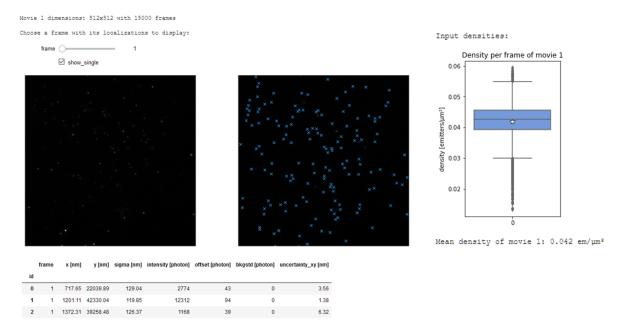


Fig. 6: Visualization of the measurement input in the ImageSumming notebook. A chosen measurement is visualized framewise and the localizations are marked with blue crosses. Average frame density is calculated and displayed as boxplot.

4.1.3 Patches

Patches are created from frames of measurement files and summed. Visualization of the created summed patches (fig. 7). Summed patches are visualized framewise and localizations are marked with crosses. The mean density per patch is calculated to show the final density. Summed patches and a corresponding localization list are saved and are ready to use in the DeepSTORM notebook as input.

Fig. 7: Visualization of the created patches in the ImageSumming notebook. Summed patches are visualized framewise and the localizations are marked with blue crosses. Average summed patch density is calculated and displayed as boxplot. By enabling show_single, single patches are shown that contribute to the summed image (left) single patch (middle) final summed patch (right) orange region markes the patch on the measurement frame.

4.2 DeepSTORM2DAddOns

DeepSTORM2DAddOns are small add ons that have proven helpful for the workflow.

4.2.1 Convert Picasso csv file to DeepSTORM2D csv format

A Picasso csv file is converted into DeepSTORM csv file format.

Input file: Picasso localization file csv Output file: DeepSTORM2D file csv

4.2.2 Convert Picasso hdf5 file to DeepSTORM2D csv format

A Picasso hdf5 file is converted into DeepSTORM csv file format.

Input file: Picasso localization file hdf5

Output file: DeepSTORM2D file csv

4.2.3 Split tif movie and corresponding localization file at defined frame

A *.tif movie and corresponding localization file are split into two files at defined frame.

Input file: Tif movie and csv file (Picasso or DeepSTORM2D format).

Parameter split at frame: Number of frames of the first file.

Output file: Two tif movies and csv files, split at defined frame.

4.2.4 Merge multiple movies and localization files

Merge multiple movies and their localization files to one.

Input files: Define multiple paths to movies and their localization files (Picasso or De-epSTORM format).

Output file: Tif movie with frames of movies stacked and localization file with continous frame numbering.

4.2.5 Split into single frames

Split a movie and localization file into single frames.

Input file: Tif movie and csv file (Picasso or DeepSTORM2D format).

Output files: Single frames of tif movies with corresponding csv file saved in defined directory.

4.2.6 Sum frames to create high density movie

Input file: Tif movie.

Camera offset: Camera offset in px intensitiy. The n summed frames are corrected for multiple camera offsets by subtracting the value n-1 times.

Number of summing frames: Define how many frames should be summed to create a high density frame.

Output files: Summed frames, each frame corresponds to a high density frame, containing summed information from n frames.

4.3 VisualizeDeepSTORM2DLocs

This notebooks visualizes the found localizations after the post-processing in the Deep-STORM notebook and helps to tune post-processing parameters (fig. 8-10).

Prediction directory: Define the directory to test file single frames and localization files (prediction output of DeepSTORM2D, sec 6.1), optionally this directory contains the ground truth localization file.

Confidence threshold: All localizations below this threshold will be filtered out. If no filtering should be applied, set the value to 0.

Save: The localizations are filtered by the confidence threshold and saved as new csv file.

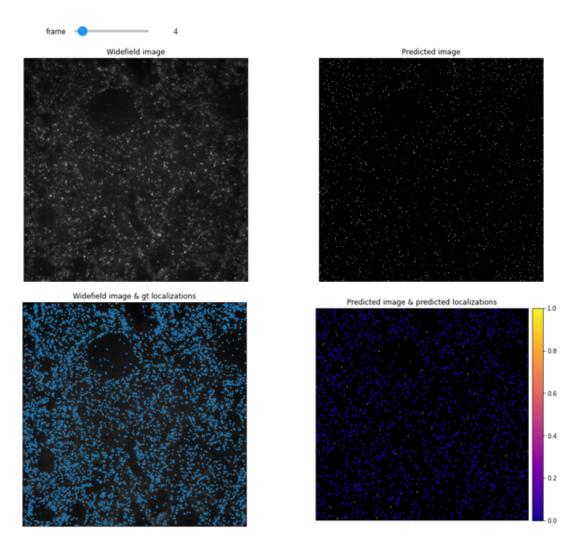


Fig. 8: Framewise visualization of recorded widefield image, predicted image, widefield image and ground truth localizations (if available) as blue crosses and predicted image and predicted localizations as crosses color coded by their confidence.

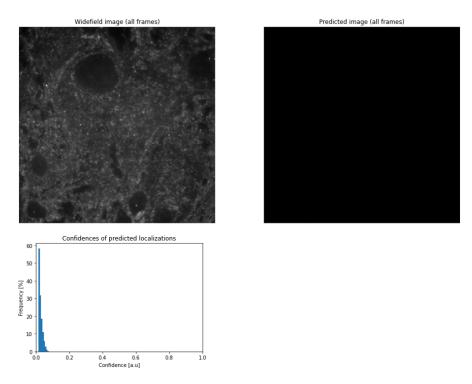


Fig. 9: Visualization of the merged widefield image (all frames are displayed as once) and the merged prediction image. Histogram of confidences of found localizations.

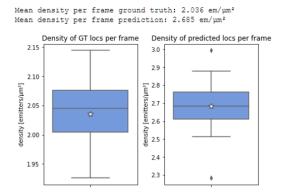


Fig. 10: Mean density of ground truth localizations and predicted localizations.

5 DeepSTORM2D notebook workflow and code adjustments

In Section 3.2 Generate training patches, patch_size is set to the patch_size created in ImageSumming - 1 (smaller patch sizes are possible as well). To consider each summed patch once, the parameter num_patches_per_frame is set to 1.

In Section 5.2 Error mapping and quality metrics estimation, the upsampling factor has to be hard coded, because image sizes are calculated based on the inputs in Section 3.2 that differ from the test image dimensions.

```
if pixel_size_INPUT == None:
    pixel_size, N, M = getPixelSizeTIFFmetadata(os.path.join(QC_image_folder,imageFilename))

#upsampling_factor = int(Mhr/M)

upsampling_factor = 8

print('Upsampling factor: '+str(upsampling_factor))

pixel_size_hr = pixel_size/upsampling_factor # in nm
```

Fig. 11: Code change in section 5.3: Hardcode the upsampling factor in this section to run quality control on differently shaped test images.

In Section 2 save the prediction not only as a merged image over all frames, but also as a stacked *.tif with each frame accessible. This helps to adjust post-processing parameters, but needs a lot of disk space.

```
Prediction = np.seros((M*upsampling_factor, N*upsampling_factor), dtype=np.float32)
Widefield = np.seros((M, N), dtype=np.float22)

Prediction_stacked = np.asarray([np.seros((M*upsampling_factor, N*upsampling_factor), dtype=np.float22) for i in range(nFrames)])
# run model in batches
n_batches = math.ceil(nFrames/batch_size)
for b in tqdm(range(n_batches)):
   nF = min(batch_sise, nFrames - b*batch_sise)
   Images_norm = np.seros((nF, M, N),dtype=np.float32)
   Images_upsampled = np.seros((nF, M*upsampling_factor, N*upsampling_factor), dtype=np.float32)
     Images_norm(f,:,:) = project_01(Images[b^batch_sise+f,:,:])
     Images\_norm\{f,:,:\} = normalise\_im(Images\_norm\{f,:,:\}, \ test\_mean, \ test\_std)
      Images\_upeampled(f,:,:) = np.kron(Images\_norm(f,:,:), np.ones((upeampling\_factor, upeampling\_factor)))
     Widefield += Images(b*batch_sise+f,:,:)
   # Reshaping
Images_upsampled = np.expand_dims(Images_upsampled,axis=4)
  # Run prediction and local ammxima finding
predicted_density = model.predict_on_batch(Images_upsampled)
   predicted_density[predicted_density < 0] = 0

Prediction_stacked[b] = predicted_density.sum(axis = 3).sum(axis = 0) 

Prediction += predicted_density.sum(axis = 3).sum(axis = 0)
  bind, wind, yind, confidence = max_layer(predicted_density)
   confidence /= L2_weighting_factor
   # turn indices to mms and append to the results
   xind, yind = xind*pixel_sise_hr, yind*pixel_sise_hr
frmind = (bind.numpy() + b*batch_sise + 1).tolist()
   xind = xind.numpy().tolist()
  yind = yind.numpy().tolist()
confidence = confidence.numpy().tolist()
   frame_number_list += frmind
  x_nm_list += xind
y_nm_list += yind
   confidence_au_list += confidence
if use_local_avg:
 ext = '_avg
with open(os.path.join(saveFath, 'Localizations_' + os.path.splitext(filename)[0] + ext + '.csv'), "w", newline*'') as file:
  writer = csv.writer(file)
  writer.writerow(('frame', 'x [nm]', 'y [nm]', 'confidence [a.u]'])
locs = list(xip(frame_number_list, x,nm_list, y,nm_list, confidence_au_list))
   writer.writerows(locs)
# Save the prediction and widefield image
Widefield = np.kron(Widefield, np.ones((upsampling_factor, upsampling_factor)))
Widefield = np.float32(Widefield)
# io.imsave(os.path.join(savePath, 'Predicted_'+os.path.splitext(filename)[0]+'.tif'), Prediction
# io.imsave(os.path.join(savePath, 'Widefield_'+os.path.splitext(filename)[0]+'.tif'), Widefield)
tifffile.imwrite(savePath + '/' + 'Predicted_stacked_'+os.path.splitext(filename)[0] + ".bif", Prediction_stacked) = saveAsTIF(savePath, 'Predicted_'+os.path.splitext(filename)[0], Prediction, pixel_size_hr)
saveAsTIF(savePath, 'Widefield_'+os.path.splitext(filename)[0], Widefield, pixel_size_hr)
```

Fig. 12: Code change in section 2: Save prediction as stacked tif with single frames additionally to merged prediction output.

6 Literatur

- [1] E. Nehme, L. E. Weiss, T. Michaeli, Y. Shechtman, Optica 2018, 5, 458–464.
- [2] L. Chamier, R. F. Laine, J. Jukkala, C. Spahn, D. Krentzel, E. Nehme, M. Lerche, S. Herández-Pérez, P. K. Mattila, E. Karinou, S. Holden, A. C. Solak, A. Krull, T.-O. Buchholz, M. L. Jones, L. Royer, C. Leterrier, Y. Shechtman, F. Jug, M. Heilemann, G. Jacquemet, R. Henriques, *Nat. Comm.* 2021, 12:2276.