

VAR-Seq Workflow Report for Compute Cluster

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1 Introduction

1.1 Background and objectives

This report describes the analysis of a VAR-Seq project studying the genetic differences among several strains ... from *organism*

1.2 Experimental design

Typically, users want to specify here all information relevant for the analysis of their NGS study. This includes detailed descriptions of FASTQ files, experimental design, reference genome, gene annotations, etc.

2 Load workflow environment

2.1 Load packages and sample data

The *systemPipeR* package needs to be loaded to perform the analysis steps shown in this report ([Girke, 2014](#)).

```
library(systemPipeR)
```

Load workflow environment with sample data into your current working directory. The sample data are described [here](#).

```
library(systemPipeRdata)
genWorkenvir(workflow="varseq")
setwd("varseq")
```

In the workflow environments generated by *genWorkenvir* all data inputs are stored in a *data/* directory and all analysis results will be written to a separate *results/* directory, while the *systemPipeVARseq.Rnw* script and the *targets* file are expected to be located in the parent directory. The R session is expected to run from this parent directory. Additional parameter files are stored under *param/*.

To work with real data, users want to organize their own data similarly and substitute all test data for their own data. To rerun an established workflow on new data, the initial *targets* file along with the corresponding FASTQ files are usually the only inputs the user needs to provide.

If applicable users can load custom functions not provided by *systemPipeR*. Skip this step if this is not the case.

```
source("systemPipeVARseq_Fct.R")
```

2.2 Experiment definition provided by targets file

The `targets` file defines all FASTQ files and sample comparisons of the analysis workflow.

```
targetspath <- system.file("extdata", "targetsPE.txt", package="systemPipeR")
targets <- read.delim(targetspath, comment.char = "#")[,1:5]
targets
```

	FileName1	FileName2	SampleName	Factor	SampleLong
1	./data/SRR446027_1.fastq	./data/SRR446027_2.fastq	M1A	M1	Mock.1h.A
2	./data/SRR446028_1.fastq	./data/SRR446028_2.fastq	M1B	M1	Mock.1h.B
3	./data/SRR446029_1.fastq	./data/SRR446029_2.fastq	A1A	A1	Avr.1h.A
4	./data/SRR446030_1.fastq	./data/SRR446030_2.fastq	A1B	A1	Avr.1h.B
5	./data/SRR446031_1.fastq	./data/SRR446031_2.fastq	V1A	V1	Vir.1h.A
6	./data/SRR446032_1.fastq	./data/SRR446032_2.fastq	V1B	V1	Vir.1h.B
7	./data/SRR446033_1.fastq	./data/SRR446033_2.fastq	M6A	M6	Mock.6h.A
8	./data/SRR446034_1.fastq	./data/SRR446034_2.fastq	M6B	M6	Mock.6h.B
9	./data/SRR446035_1.fastq	./data/SRR446035_2.fastq	A6A	A6	Avr.6h.A
10	./data/SRR446036_1.fastq	./data/SRR446036_2.fastq	A6B	A6	Avr.6h.B
11	./data/SRR446037_1.fastq	./data/SRR446037_2.fastq	V6A	V6	Vir.6h.A
12	./data/SRR446038_1.fastq	./data/SRR446038_2.fastq	V6B	V6	Vir.6h.B
13	./data/SRR446039_1.fastq	./data/SRR446039_2.fastq	M12A	M12	Mock.12h.A
14	./data/SRR446040_1.fastq	./data/SRR446040_2.fastq	M12B	M12	Mock.12h.B
15	./data/SRR446041_1.fastq	./data/SRR446041_2.fastq	A12A	A12	Avr.12h.A
16	./data/SRR446042_1.fastq	./data/SRR446042_2.fastq	A12B	A12	Avr.12h.B
17	./data/SRR446043_1.fastq	./data/SRR446043_2.fastq	V12A	V12	Vir.12h.A
18	./data/SRR446044_1.fastq	./data/SRR446044_2.fastq	V12B	V12	Vir.12h.B

3 Read preprocessing

3.1 Read quality filtering and trimming

```
library(BiocParallel); library(BatchJobs)
args <- systemArgs(sysma="param/trimPE.param", mytargets="targetsPE.txt")
f <- function(x) {
  library(systemPipeR)
  source("systemPipeVARseq_Fct.R")
  args <- systemArgs(sysma="param/trimPE.param", mytargets="targetsPE.txt")
  preprocessReads(args=args[x], Fct="filterFct(fq, cutoff=20, Nexceptions=0)", batchsize=100000)
}
funs <- makeClusterFunctionsTorque("torque.tmpl")
param <- BatchJobsParam(length(args), resources=list(walltime="20:00:00", nodes="1:ppn=1", memory="16gb"),
register(param)
d <- bplapply(seq(along=args), f)
writeTargetsout(x=args, file="targets_PETrim.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_PETrim.txt
```

3.2 FASTQ quality report

The following `seeFastq` and `seeFastqPlot` functions generate and plot a series of useful quality statistics for a set of FASTQ files including per cycle quality box plots, base proportions, base-level quality trends, relative k-mer diversity,

length and occurrence distribution of reads, number of reads above quality cutoffs and mean quality distribution. The results are written to a PDF file named `fastqReport.pdf`.

```
library(BiocParallel); library(BatchJobs)
args <- systemArgs(sysma="param/bwa.param", mytargets="targets_PETrim.txt")
f <- function(x) {
  library(systemPipeR)
  args <- systemArgs(sysma="param/bwa.param", mytargets="targets_PETrim.txt")
  seeFastq(fastq=infile1(args)[x], batchsize=100000, klength=8)
}
funs <- makeClusterFunctionsTorque("torque.tpl")
param <- BatchJobsParam(length(args), resources=list(walltime="20:00:00", nodes="1:ppn=1", memory="16gb"),
register(param)
fqlist <- bplapply(seq(along=args), f)
pdf("./results/fastqReport.pdf", height=18, width=4*length(fqlist))
seeFastqPlot(unlist(fqlist, recursive=FALSE))
dev.off()

pdf
2
```

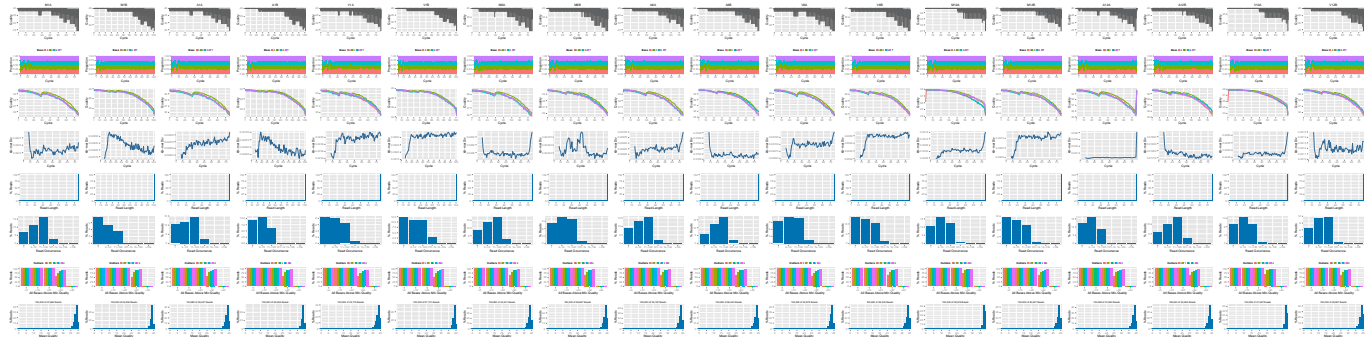


Figure 1: QC report for 18 FASTQ files.

4 Alignments

4.1 Read mapping with BWA

The NGS reads of this project are aligned against the reference genome sequence using the highly variant tolerant short read aligner BWA (Li, 2013; Li and Durbin, 2009). The parameter settings of the aligner are defined in the `bwa.param` file.

```
args <- systemArgs(sysma="param/bwa.param", mytargets="targets_PETrim.txt")
sysargs(args)[1] # Command-line parameters for first FASTQ file

"bwa mem -t 4 -M -R '@RG\tID:group1\tSM:sample1\tPL:illumina\tLB:lib1\tPU:unit1' /bigdata/girkelab/tg
```

Runs the alignments sequentially (e.g. on a single machine)

```
bampaths <- runCommandline(args=args)
```

Alternatively, the alignment jobs can be submitted to a compute cluster, here using 72 CPU cores (18 qsub processes

each with 4 CPU cores).

```
args <- systemArgs(sysma="param/bwa.param", mytargets="targets_PETrim.txt")
moduleload(modules(args))
system("bwa index -a bwtsw ./data/tair10.fasta")
resources <- list(walltime="20:00:00", nodes=paste0("1:ppn=", cores(args)), memory="4gb")
reg <- clusterRun(args, conffile=".BatchJobs.R", template="torque.tmpl", Njobs=18, runid="01",
  resourceList=resources)
waitForJobs(reg)

[1] TRUE

writeTargetsout(x=args, file="targets_bam.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_bam.txt
```

Check whether all BAM files have been created

```
file.exists(outpaths(args))
```

4.2 Read mapping with gsnap

```
library(gmapR); library(BiocParallel); library(BatchJobs)
gmapGenome <- GmapGenome(systemPipeR::reference(args), directory="data", name="gmap_tair10chr", create=TRUE)
args <- systemArgs(sysma="param/gsnap.param", mytargets="targets_PETrim.txt")
f <- function(x) {
  library(gmapR); library(systemPipeR)
  args <- systemArgs(sysma="param/gsnap.param", mytargets="targets_PETrim.txt")
  gmapGenome <- GmapGenome(reference(args), directory="data", name="gmap_tair10chr", create=FALSE)
  p <- GsnapParam(genome=gmapGenome, unique_only=TRUE, molecule="DNA", max_mismatches=3)
  o <- gsnap(input_a=infile1(args)[x], input_b=infile2(args)[x], params=p, output=outfile1(args)[x])
}
funs <- makeClusterFunctionsTorque("torque.tmpl")
param <- BatchJobsParam(length(args), resources=list(walltime="20:00:00", nodes="1:ppn=1", memory="6gb"),
register(param)
d <- bplapply(seq(along=args), f)
writeTargetsout(x=args, file="targets_gsnap_bam.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_gsnap_bam.txt

file.exists(outpaths(args))

[1] TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE TRUE
```

4.3 Read and alignment stats

The following generates a summary table of the number of reads in each sample and how many of them aligned to the reference.

```
args <- systemArgs(sysma="param/bwa.param", mytargets="targets_PETrim.txt")
read_statsDF <- alignStats(args=args)
write.table(read_statsDF, "results/alignStats.xls", row.names=FALSE, quote=FALSE, sep="\t")
```

4.4 Create symbolic links for viewing BAM files in IGV

The `symLink2bam` function creates symbolic links to view the BAM alignment files in a genome browser such as IGV. The corresponding URLs are written to a file with a path specified under `urlfile`, here [IGVurl.txt](#).

```
symLink2bam(sysargs=args, htldir=c("~/html/", "somedir/"),
            urlbase="http://biocluster.ucr.edu/~tgirke/",
            urlfile="./results/IGVurl.txt")
```

5 Variant calling

The following performs variant calling with GATK, BCFtools and *VariantTools* in parallel mode on a compute cluster (McKenna et al., 2010; Li, 2011). If a cluster is not available, the `runCommandline()` function can be used to run the variant calling with GATK and BCFtools for each sample sequentially on a single machine, or `callVariants` in case of *VariantTools*. Typically, the user would choose here only one variant caller rather than running several ones.

5.1 Variant calling with GATK

The following creates in the initial step a new targets file (`targets_bam.txt`). The first column of this file gives the paths to the BAM files created in the alignment step. The new targets file and the parameter file `gatk.param` are used to create a new `Sysargs` instance for running GATK. Since GATK involves many processing steps, it is executed by a bash script `gatk_run.sh` where the user can specify the detailed run parameters. All three files are expected to be located in the current working directory. Samples files for `gatk.param` and `gatk_run.sh` are available in the subdirectory `./inst/extdata/` of the source file of the *systemPipeR* package. Alternatively, they can be downloaded directly from [here](#).

```
#system("java -jar CreateSequenceDictionary.jar R=./data/tair10.fasta O=./data/tair10.dict")
system("java -jar /opt/picard/1.81/CreateSequenceDictionary.jar R=./data/tair10.fasta O=./data/tair10.dict")
args <- systemArgs(sysma="param/gatk.param", mytargets="targets_bam.txt")
resources <- list(walltime="20:00:00", nodes=paste0("1:ppn=", 1), memory="10gb")
reg <- clusterRun(args, conffile=".BatchJobs.R", template="torque.tpl", Njobs=18, runid="01",
                  resourceList=resources)
waitForJobs(reg)

[1] TRUE

unlink(outfile1(args), recursive = TRUE, force = TRUE)
writeTargetsout(x=args, file="targets_gatk.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_gatk.txt
```

5.2 Variant calling with BCFtools

The following runs the variant calling with BCFtools. This step requires in the current working directory the parameter file `sambcf.param` and the bash script `sambcf_run.sh`.

```
args <- systemArgs(sysma="param/sambcf.param", mytargets="targets_bam.txt")
resources <- list(walltime="20:00:00", nodes=paste0("1:ppn=", 1), memory="10gb")
reg <- clusterRun(args, conffile=".BatchJobs.R", template="torque.tpl", Njobs=18, runid="01",
                  resourceList=resources)
waitForJobs(reg)

[1] TRUE
```

```
unlink(outfile1(args), recursive = TRUE, force = TRUE)
writeTargetsout(x=args, file="targets_sambcf.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_sambcf.txt
```

5.3 Variant calling with VariantTools

```
library(gmapR); library(BiocParallel); library(BatchJobs)
args <- systemArgs(sysma="param/vartools.param", mytargets="targets_gsnap_bam.txt")
f <- function(x) {
  library(VariantTools); library(gmapR); library(systemPipeR)
  args <- systemArgs(sysma="param/vartools.param", mytargets="targets_gsnap_bam.txt")
  gmapGenome <- GmapGenome(systemPipeR::reference(args), directory="data", name="gmap_tair10chr", create=TRUE)
  tally.param <- TallyVariantsParam(gmapGenome, high_base_quality = 23L, indels = TRUE)
  bfl <- BamFileList(infile1(args)[x], index=character())
  var <- callVariants(bfl[[1]], tally.param)
  sampleNames(var) <- names(bfl)
  writeVcf(asVCF(var), outfile1(args)[x], index = TRUE)
}
funs <- makeClusterFunctionsTorque("torque.tpl")
param <- BatchJobsParam(length(args), resources=list(walltime="20:00:00", nodes="1:ppn=1", memory="6gb"),
register(param)
d <- bplapply(seq(along=args), f)
file.exists(outpaths(args))
writeTargetsout(x=args, file="targets_vartools.txt", overwrite=TRUE)
```

6 Filter variants

The function `filterVars` filters VCF files based on user definable quality parameters. It sequentially imports each VCF file into R, applies the filtering on an internally generated `VRanges` object and then writes the results to a new subsetted VCF file. The filter parameters are passed on to the corresponding argument as a character string. The function applies this filter to the internally generated `VRanges` object using the standard subsetting syntax for two dimensional objects such as: `vr[filter,]`. The parameter files (`filter_gatk.param`, `filter_sambcf.param` and `filter_vartools.param`), used in the filtering steps, define the paths to the input and output VCF files which are stored in new `SYSargs` instances.

6.1 Filter variants called by GATK

The below example filters for variants that are supported by $\geq x$ reads and $\geq 80\%$ of them support the called variants. In addition, all variants need to pass $\geq x$ of the soft filters recorded in the VCF files generated by GATK. Since the toy data used for this workflow is very small, the chosen settings are unreasonably relaxed. A more reasonable filter setting is given in the line below (here commented out).

```
library(VariantAnnotation)
library(BBmisc) # Defines suppressAll()
args <- systemArgs(sysma="param/filter_gatk.param", mytargets="targets_gatk.txt")
filter <- "totalDepth(vr) >= 2 & (altDepth(vr) / totalDepth(vr) >= 0.8) & rowSums(softFilterMatrix(vr))>=1"
# filter <- "totalDepth(vr) >= 20 & (altDepth(vr) / totalDepth(vr) >= 0.8) & rowSums(softFilterMatrix(vr))>=1"
suppressAll(filterVars(args, filter, varcaller="gatk", organism="A. thaliana"))
writeTargetsout(x=args, file="targets_gatk_filtered.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_gatk_filtered.txt
```


6.2 Filter variants called by BCFtools

The following shows how to filter the VCF files generated by *BCFtools* using similar parameter settings as in the previous filtering of the GATK results.

```
args <- systemArgs(sysma="param/filter_sambcf.param", mytargets="targets_sambcf.txt")
filter <- "rowSums(vr) >= 2 & (rowSums(vr[,3:4])/rowSums(vr[,1:4]) >= 0.8)"
# filter <- "rowSums(vr) >= 20 & (rowSums(vr[,3:4])/rowSums(vr[,1:4]) >= 0.8)"
suppressAll(filterVars(args, filter, varcaller="bcftools", organism="A. thaliana"))
writeTargetsout(x=args, file="targets_sambcf_filtered.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_sambcf_filtered.txt
```

6.3 Filter variants called by VariantTools

The following shows how to filter the VCF files generated by *VariantTools* using similar parameter settings as in the previous filtering of the GATK results.

```
args <- systemArgs(sysma="param/filter_vartools.param", mytargets="targets_vartools.txt")
filter <- "(values(vr)$n.read.pos.ref + values(vr)$n.read.pos) >= 2 & (values(vr)$n.read.pos / (values(vr)$n.read.pos + values(vr)$n.read.pos.ref) >= 0.8)"
# filter <- "(values(vr)$n.read.pos.ref + values(vr)$n.read.pos) >= 20 & (values(vr)$n.read.pos / (values(vr)$n.read.pos + values(vr)$n.read.pos.ref) >= 0.8)"
suppressAll(filterVars(args, filter, varcaller="vartools", organism="A. thaliana"))
writeTargetsout(x=args, file="targets_vartools_filtered.txt", overwrite=TRUE)

Written content of 'targetsout(x)' to file: targets_vartools_filtered.txt
```

7 Annotate filtered variants

The function `variantReport` generates a variant report using utilities provided by the *VariantAnnotation* package. The report for each sample is written to a tabular file containing genomic context annotations (e.g. coding or non-coding SNPs, amino acid changes, IDs of affected genes, etc.) along with confidence statistics for each variant. The parameter file `annotate_vars.param` defines the paths to the input and output files which are stored in a new `SYSargs` instance.

7.1 Annotate filtered variants called by GATK

```
library("GenomicFeatures")
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_gatk_filtered.txt")
txdb <- loadDb("./data/tair10.sqlite")
fa <- FaFile(systemPipeR::reference(args))
suppressAll(variantReport(args=args, txdb=txdb, fa=fa, organism="A. thaliana"))
```

7.2 Annotate filtered variants called by BCFtools

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
txdb <- loadDb("./data/tair10.sqlite")
fa <- FaFile(systemPipeR::reference(args))
suppressAll(variantReport(args=args, txdb=txdb, fa=fa, organism="A. thaliana"))
```


7.3 Annotate filtered variants called by VariantTools

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_vartools_filtered.txt")
txdb <- loadDb("./data/tair10.sqlite")
fa <- FaFile(systemPipeR::reference(args))
suppressAll(variantReport(args=args, txdb=txdb, fa=fa, organism="A. thaliana"))
```

8 Combine annotation results among samples

To simplify comparisons among samples, the `combineVarReports` function combines all variant annotation reports referenced in a `SYSargs` instance (here `args`). At the same time the function allows to consider only certain feature types of interest. For instance, the below setting `filtercol=c(Consequence="nonsynonymous")` will include only nonsynonymous variances listed in the `Consequence` column of the annotation reports. To omit filtering, one can use the setting `filtercol="All"`

8.1 Combine results from GATK

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_gatk_filtered.txt")
combinedF <- combineVarReports(args, filtercol=c(Consequence="nonsynonymous"))
write.table(combinedF, "./results/combinedF_nonsyn_gatk.xls", quote=FALSE, row.names=FALSE, sep="\t")
```

8.2 Combine results from BCFtools

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
combinedF <- combineVarReports(args, filtercol=c(Consequence="nonsynonymous"))
write.table(combinedF, "./results/combinedF_nonsyn_sambcf.xls", quote=FALSE, row.names=FALSE, sep="\t")
```

8.3 Combine results from VariantTools

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_vartools_filtered.txt")
combinedF <- combineVarReports(args, filtercol=c(Consequence="nonsynonymous"))
write.table(combinedF, "./results/combinedF_nonsyn_vartools.xls", quote=FALSE, row.names=FALSE, sep="\t")
```

9 Summary statistics of variants

The function `varSummar` counts the number of variants for each feature type included in the annotation reports.

9.1 Summary for GATK

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_gatk_filtered.txt")
write.table(varSummary(args), "./results/variantStats_gatk.xls", quote=FALSE, col.names = NA, sep="\t")
```

9.2 Summary for BCFtools

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
write.table(varSummary(args), "./results/variantStats_sambcf.xls", quote=FALSE, col.names = NA, sep="\t")
```

9.3 Summary for VariantTools

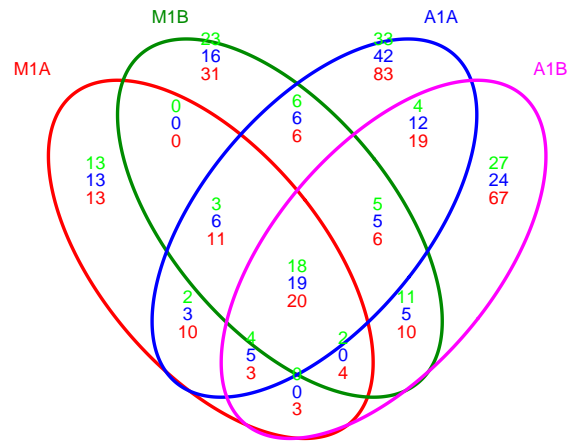
```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_vartools_filtered.txt")
write.table(varSummary(args), "./results/variantStats_vartools.xls", quote=FALSE, col.names = NA, sep="\t")
```

10 Venn diagram of variants

The venn diagram utilities defined by the *systemPipeR* package can be used to identify common and unique variants reported for different samples and/or variant callers. The below generates a 4-way venn diagram comparing four samples for each of the two variant callers.

```
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_gatk_filtered.txt")
varlist <- sapply(names(outpaths(args))[1:4], function(x) as.character(read.delim(outpaths(args)[x])$VARID))
vennset_gatk <- overLapper(varlist, type="vennsets")
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_sambcf_filtered.txt")
varlist <- sapply(names(outpaths(args))[1:4], function(x) as.character(read.delim(outpaths(args)[x])$VARID))
vennset_bcf <- overLapper(varlist, type="vennsets")
args <- systemArgs(sysma="param/annotate_vars.param", mytargets="targets_vartools_filtered.txt")
varlist <- sapply(names(outpaths(args))[1:4], function(x) as.character(read.delim(outpaths(args)[x])$VARID))
vennset_vartools <- overLapper(varlist, type="vennsets")
pdf("./results/vennplot_var.pdf")
vennPlot(list(vennset_gatk, vennset_bcf, vennset_vartools), mymain="", mysub="GATK: red; BCFtools: blue; V",
dev.off())
```

```
pdf
2
```



GATK: red; BCFtools: blue; VariantTools: green

Figure 2: Venn Diagram for 4 samples from GATK, BCFtools and VariantTools.

11 Version Information

```
toLatex(sessionInfo())
```

- R version 3.2.1 (2015-06-18), x86_64-unknown-linux-gnu
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, stats4, utils
- Other packages: Biobase 2.29.1, BiocGenerics 0.15.6, BiocParallel 1.3.48, Biostrings 2.37.5, DBI 0.3.1, GenomInfoDb 1.5.10, GenomicAlignments 1.5.12, GenomicRanges 1.21.18, IRanges 2.3.18, RSQLite 1.0.0, Rsamtools 1.21.15, S4Vectors 0.7.13, ShortRead 1.27.5, SummarizedExperiment 0.3.3, XVector 0.9.1, knitr 1.10.5, systemPipeR 1.3.25
- Loaded via a namespace (and not attached): AnnotationDbi 1.31.17, AnnotationForge 1.11.16, BBmisc 1.9, BatchJobs 1.6, BiocStyle 1.7.6, Category 2.35.1, GO.db 3.1.2, GOstats 2.35.1, GSEABase 1.31.3, MASS 7.3-43, Matrix 1.2-2, RBGL 1.45.1, RColorBrewer 1.1-2, Rcpp 0.12.0, XML 3.98-1.3, annotate 1.47.4, base64enc 0.1-3, bitops 1.0-6, brew 1.0-6, checkmate 1.6.2, colorspace 1.2-6, digest 0.6.8, edgeR 3.11.2, evaluate 0.7.2, fail 1.2, formatR 1.2, futile.logger 1.4.1, futile.options 1.0.0, genefilter 1.51.0, ggplot2 1.0.1, graph 1.47.2, grid 3.2.1, gtable 0.1.2, highr 0.5, hwriter 1.3.2, lambda.r 1.1.7, lattice 0.20-33, latticeExtra 0.6-26, limma 3.25.15, magrittr 1.5, munsell 0.4.2, pheatmap 1.0.7, plyr 1.8.3, proto 0.3-10, reshape2 1.4.1, rjson 0.2.15, scales 0.2.5, sendmailR 1.2-1, splines 3.2.1, stringi 0.5-5, stringr 1.0.0, survival 2.38-3, tools 3.2.1, xtable 1.7-4, zlibbioc 1.15.0

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13 References

- Thomas Girke. systemPipeR: NGS workflow and report generation environment, 28 June 2014. URL <https://github.com/tgirke/systemPipeR>.
- H Li and R Durbin. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*, 25(14): 1754–1760, July 2009. ISSN 1367-4803. doi: 10.1093/bioinformatics/btp324. URL <http://dx.doi.org/10.1093/bioinformatics/btp324>.
- Heng Li. A statistical framework for SNP calling, mutation discovery, association mapping and population genetical parameter estimation from sequencing data. *Bioinformatics*, 27(21):2987–2993, 1 November 2011. ISSN 1367-4803. doi: 10.1093/bioinformatics/btr509. URL <http://bioinformatics.oxfordjournals.org/content/27/21/2987.abstract>.
- Heng Li. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. 03 2013. URL <http://arxiv.org/abs/1303.3997>.
- Aaron McKenna, Matthew Hanna, Eric Banks, Andrey Sivachenko, Kristian Cibulskis, Andrew Kernytsky, Kiran Garimella, David Altshuler, Stacey Gabriel, Mark Daly, and Mark A DePristo. The genome analysis toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.*, 20(9):1297–1303, 19 July 2010. ISSN 1088-9051. doi: 10.1101/gr.107524.110. URL <http://dx.doi.org/10.1101/gr.107524.110>.