

ChIP-Seq project report template: Some Descriptive Title

Project ID: ChIPseq-PI_Name_Organism_Jun2015

Project PI: First Last (first.last@inst.edu)

Author of Report: First Last (first.last@inst.edu)

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1 Introduction

This report describes the analysis of an ChIP-Seq project from Dr. First Last's lab which studies the gene expression changes of ... in *organism* The experimental design is as follows...

2 Sample definitions and environment settings

2.1 Environment settings and input data

Typically, the user wants to record here the sources and versions of the reference genome sequence along with the corresponding annotations. In the provided sample data set all data inputs are stored in a data subdirectory and all results will be written to a separate results directory, while the `systemPipeChIPseq.Rnw` script and the `targets` file are expected to be located in the parent directory. The R session is expected to run from this parent directory.

To run this sample report, mini sample FASTQ and reference genome files can be downloaded from [here](#). The chosen data set [SRP010938](#) contains 18 paired-end (PE) read sets from *Arabidopsis thaliana* [Howard et al. \(2013\)](#). To minimize processing time during testing, each FASTQ file has been subsetting to 90,000-100,000 randomly sampled PE reads that map to the first 100,000 nucleotides of each chromosome of the *A. thaliana* genome. The corresponding reference genome sequence (FASTA) and its GFF annotation files (provided in the same download) have been truncated accordingly. This way the entire test sample data set is less than 200MB in storage space. A PE read set has been chosen for this test data set for flexibility, because it can be used for testing both types of analysis routines requiring either SE (single end) reads or PE reads.

2.2 Required packages and resources

The `systemPipeR` package needs to be loaded to perform the analysis steps shown in this report ([Girke, 2014](#)).

```
library(systemPipeR)
```

If applicable load custom functions not provided by `systemPipeR`

```
source("systemPipeChIPseq_Fct.R")
```

2.3 Experiment definition provided by targets file

The `targets` file defines all FASTQ files and sample comparisons of the analysis workflow.

```
targets <- read.delim("targets_chip.txt", comment.char = "#")[,1:4]
targets
```

	FileName	SampleName	Factor	SampleLong
1	./data/SRR446027_1.fastq.gz	M1A	M1	Mock.1h.A
2	./data/SRR446028_1.fastq.gz	M1B	M1	Mock.1h.B
3	./data/SRR446029_1.fastq.gz	A1A	A1	Avr.1h.A
4	./data/SRR446030_1.fastq.gz	A1B	A1	Avr.1h.B
5	./data/SRR446031_1.fastq.gz	V1A	V1	Vir.1h.A
6	./data/SRR446032_1.fastq.gz	V1B	V1	Vir.1h.B
7	./data/SRR446033_1.fastq.gz	M6A	M6	Mock.6h.A
8	./data/SRR446034_1.fastq.gz	M6B	M6	Mock.6h.B
9	./data/SRR446035_1.fastq.gz	A6A	A6	Avr.6h.A
10	./data/SRR446036_1.fastq.gz	A6B	A6	Avr.6h.B
11	./data/SRR446037_1.fastq.gz	V6A	V6	Vir.6h.A

12	./data/SRR446038_1.fastq.gz	V6B	V6	Vir.6h.B
13	./data/SRR446039_1.fastq.gz	M12A	M12	Mock.12h.A
14	./data/SRR446040_1.fastq.gz	M12B	M12	Mock.12h.B
15	./data/SRR446041_1.fastq.gz	A12A	A12	Avr.12h.A
16	./data/SRR446042_1.fastq.gz	A12B	A12	Avr.12h.B
17	./data/SRR446043_1.fastq.gz	V12A	V12	Vir.12h.A
18	./data/SRR446044_1.fastq.gz	V12B	V12	Vir.12h.B

3 Read preprocessing

3.1 FASTQ quality report

The following `seeFastq` and `seeFastqPlot` functions generate and plot a series of useful quality statistics for a set of FASTQ files including per cycle quality box plots, base proportions, base-level quality trends, relative k-mer diversity, length and occurrence distribution of reads, number of reads above quality cutoffs and mean quality distribution. The results are written to a PDF file named `fastqReport.pdf`.

```
args <- systemArgs(sysma="param/bowtieSE.param", mytargets="targets_chip.txt")
fqlist <- seeFastq(fastq=infile1(args), batchsize=100000, klength=8)
pdf("./results/fastqReport.pdf", height=18, width=4*length(fqlist))
seeFastqPlot(fqlist)
dev.off()
```

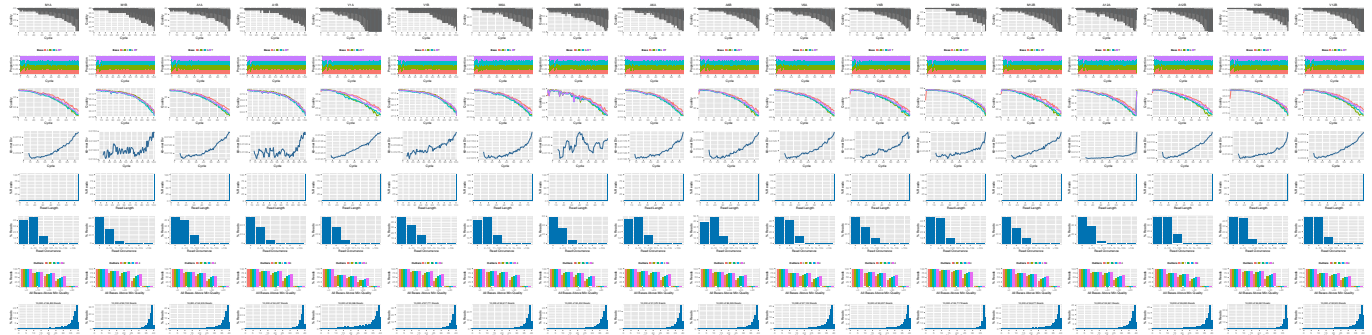


Figure 1: QC report for 18 FASTQ files.

4 Alignments

4.1 Read mapping with Bowtie2

The NGS reads of this project will be aligned against the reference genome sequence using Bowtie2 ([Langmead and Salzberg, 2012](#)). The parameter settings of the aligner are defined in the `tophat.param` file.

```
args <- systemArgs(sysma="param/bowtieSE.param", mytargets="targets_chip.txt")
sysargs(args)[1] # Command-line parameters for first FASTQ file
```

Submission of alignment jobs to compute cluster, here using 72 CPU cores (18 qsub processes each with 4 CPU cores).

```
moduleload(modules(args))
system("bowtie2-build ./data/tair10.fasta ./data/tair10.fasta")
resources <- list(walltime="20:00:00", nodes=paste0("1:ppn=", cores(args)), memory="10gb")
reg <- clusterRun(args, conffile=".BatchJobs.R", template="torque.tmpl", Njobs=18, runid="01",
                  resourceList=resources)
runCommandline(args)
writeTargetsout(x=args, file="targets_bam.txt", overwrite=TRUE)
```

Check whether all BAM files have been created

```
file.exists(outpaths(args))
```

4.2 Read and alignment stats

The following provides an overview of the number of reads in each sample and how many of them aligned to the reference.

```
read_statsDF <- alignStats(args=args)
write.table(read_statsDF, "results/alignStats.xls", row.names=FALSE, quote=FALSE, sep="\t")
read.delim("results/alignStats.xls")
```

4.3 Create symbolic links for viewing BAM files in IGV

The symLink2bam function creates symbolic links to view the BAM alignment files in a genome browser such as IGV. The corresponding URLs are written to a file with a path specified under urlfile, here [IGVurl.txt](#).

```
symLink2bam(sysargs=args, htmlDir=c("~/html/", "somedir/"),
            urlbase="http://biocluster.ucr.edu/~tgirke/",
            urlfile="./results/IGVurl.txt")
```

5 Peak calling with MACS2

5.1 Merge BAM files of replicates prior to peak calling

Merging BAM files of technical and/or biological replicates can improve the sensitivity of the peak calling by increasing the depth of read coverage. The mergeBamByFactor function merges BAM files based on grouping information specified by a factor, here the Factor column of the imported targets file. It also returns an updated SYSargs object containing the paths to the merged BAM files as well as to any unmerged files without replicates. This step can be skipped if merging of BAM files is not desired.

```
args <- systemArgs(sysma=NULL, mytargets="targets_bam.txt")
args_merge <- mergeBamByFactor(args, overwrite=TRUE)
writeTargetsout(x=args_merge, file="targets_mergeBamByFactor.txt", overwrite=TRUE)
```

5.2 Peak calling without input/reference sample

```
args <- systemArgs(sysma="param/macs2_noinput.param", mytargets="targets_mergeBamByFactor.txt")
sysargs(args)[1] # Command-line parameters for first FASTQ file
runCommandline(args)
```

```
file.exists(outpaths(args))
writeTargetsout(x=args, file="targets_mac3.txt", overwrite=TRUE)
```

5.3 Peak calling with input/reference sample

```
writeTargetsRef(infile="targets_mergeBamByFactor.txt", outfile="targets_bam_ref.txt", silent=FALSE, overwr
args <- systemArgs(sysma="param/mac3.param", mytargets="targets_bam_ref.txt")
sysargs(args)[1] # Command-line parameters for first FASTQ file
runCommandline(args)
file.exists(outpaths(args))
writeTargetsout(x=args, file="targets_mac3.txt", overwrite=TRUE)
```

6 Annotate peaks with genomic context

6.1 Annotation with ChIPpeakAnno package

```
library(ChIPpeakAnno); library(GenomicFeatures)
args <- systemArgs(sysma="param/annotate_peaks.param", mytargets="targets_mac3.txt")
txdb <- loadDb("./data/tair10.sqlite")
ge <- genes(txdb, columns=c("tx_name", "gene_id", "tx_type"))
for(i in seq(along=args)) {
  peaksGR <- as(read.delim(infile1(args)[i], comment="#"), "GRanges")
  annotatedPeak <- annotatePeakInBatch(peaksGR, AnnotationData=genes(txdb))
  df <- data.frame(as.data.frame(annotatedPeak), as.data.frame(values(ge[values(annotatedPeak)$feature,]
  write.table(df, outpaths(args[i]), quote=FALSE, row.names=FALSE, sep="\t")
}
writeTargetsout(x=args, file="targets_peakanno.txt", overwrite=TRUE)
## Perform previous step with full genome annotation from Biomart
# txdb <- makeTxDbFromBiomart(biomart="ENSEMBL_MART_PLANT", dataset="athaliana_eg_gene")
# tx <- transcripts(txdb, columns=c("tx_name", "gene_id", "tx_type"))
# ge <- genes(txdb, columns=c("tx_name", "gene_id", "tx_type")) # works as well
# seqlevels(ge) <- c("Chr1", "Chr2", "Chr3", "Chr4", "Chr5", "ChrC", "ChrM")
# table(mcols(tx)$tx_type)
# tx <- tx[!duplicated(unstrsplit(values(tx)$gene_id, sep=","))] # Keeps only first transcript model for ea
# annotatedPeak <- annotatePeakInBatch(mac3Output, AnnotationData = tx)
```

6.2 Annotation with ChIPseeker package

Same as in previous step but using the *ChIPseeker* package for annotating the peaks.

```
library(ChIPseeker)
txdb <- loadDb("./data/tair10.sqlite")
for(i in seq(along=args)) {
  peakAnno <- annotatePeak(infile1(args)[i], TxDb=txdb, verbose=FALSE)
  df <- as.data.frame(peakAnno)
  write.table(df, outpaths(args[i]), quote=FALSE, row.names=FALSE, sep="\t")
}
writeTargetsout(x=args, file="targets_peakanno.txt", overwrite=TRUE)
```

Summary plots provided by the *ChIPseeker* package. Here applied to only one sample.

```
peak <- readPeakFile(outpaths(args)[1])
covplot(peak, weightCol="X.log10.pvalue.")
peakHeatmap(outpaths(args)[1], TxDb=txdb, upstream=1000, downstream=1000, color="red")
plotAvgProf2(outpaths(args)[1], TxDb=txdb, upstream=1000, downstream=1000, xlab="Genomic Region (5'→3')",
```

7 Count reads overlapping peak regions

```
library(GenomicRanges)
args <- systemArgs(sysma="param/count_rangesets.param", mytargets="targets_macs.txt")
args_bam <- systemArgs(sysma=NULL, mytargets="targets_bam.txt")
bfl <- BamFileList(outpaths(args_bam), yieldSize=50000, index=character())
countDFnames <- countRangeset(bfl, args, mode="Union", ignore.strand=TRUE)
writeTargetsout(x=args, file="targets_countDF.txt", overwrite=TRUE)
```

8 Differential binding analysis of peaks

To be continued ...

```
args_bam <- systemArgs(sysma=NULL, mytargets="targets_bam.txt")
cmp <- readComp(file=args_bam, format="matrix")
dbrlist <- runDiff(countfiles=countDFnames, diffFct=run_edgeR, targets=targetsin(args_bam), extension="_ed
```

9 GO term enrichment analysis

The following performs GO term enrichment analysis for all annotated peak sets disregarding the differential binding results.

```
args <- systemArgs(sysma="param/mac2.param", mytargets="targets_bam_ref.txt")
args_anno <- systemArgs(sysma="param/annotate_peaks.param", mytargets="targets_chip_anno.txt")
annofiles <- paste0(outpaths(args), ".annotated.xls"); names(annofiles) <- names(outpaths(args))
gene_ids <- sapply(names(annofiles), function(x) unique(as.character(read.delim(annofiles[x])[, "gene_id"])))
load("data/GO/catdb.RData")
BatchResult <- GOCluster_Report(catdb=catdb, setlist=gene_ids, method="all", id_type="gene", CLSZ=2, cutoff
```

10 Motif analysis

10.1 Parse DNA sequences of peak regions from genome

```
library(Biostrings); library(seqLogo); library(BCRANK)
for(i in seq(along=rangefiles)) {
  df <- read.delim(rangefiles[i], comment="#")
  peaks <- as(df, "GRanges")
  names(peaks) <- paste0(as.character(seqnames(peaks)), "_", start(peaks), "-", end(peaks))
  pseq <- getSeq(FaFile("./data/tair10.fasta"), peaks)
  names(pseq) <- names(peaks)
  writeXStringSet(pseq, paste0(rangefiles[i], ".fasta"))
}
```

```
}
```

10.2 Motif discovery with BCRANK

```
set.seed(0)
BCRANKout <- bcrank(paste0(rangefiles[1], ".fasta"), restarts=25, use.P1=TRUE, use.P2=TRUE)
toptable(BCRANKout)
topMotif <- toptable(BCRANKout, 1)
weightMatrix <- pwm(topMotif, normalize = FALSE)
weightMatrixNormalized <- pwm(topMotif, normalize = TRUE)
seqLogo(weightMatrixNormalized)
```

11 Version Information

```
toLatex(sessionInfo())
```

- R version 3.2.1 (2015-06-18), x86_64-unknown-linux-gnu
- Locale: C
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, stats4, utils
- Other packages: Biobase 2.29.1, BiocGenerics 0.15.6, BiocParallel 1.3.48, Biostrings 2.37.4, DBI 0.3.1, GenomInfoDb 1.5.10, GenomicAlignments 1.5.12, GenomicRanges 1.21.18, IRanges 2.3.18, RSQLite 1.0.0, Rsamtools 1.21.14, S4Vectors 0.7.12, ShortRead 1.27.5, SummarizedExperiment 0.3.3, XVector 0.9.1, knitr 1.10.5, systemPipeR 1.3.22
- Loaded via a namespace (and not attached): AnnotationDbi 1.31.17, AnnotationForge 1.11.16, BBmisc 1.9, BatchJobs 1.6, BiocStyle 1.7.6, Category 2.35.1, GO.db 3.1.2, GOstats 2.35.1, GSEABase 1.31.3, MASS 7.3-43, Matrix 1.2-2, RBGL 1.45.1, RColorBrewer 1.1-2, Rcpp 0.12.0, XML 3.98-1.3, annotate 1.47.4, base64enc 0.1-3, bitops 1.0-6, brew 1.0-6, checkmate 1.6.2, colorspace 1.2-6, digest 0.6.8, edgeR 3.11.2, evaluate 0.7, fail 1.2, formatR 1.2, futile.logger 1.4.1, futile.options 1.0.0, genefilter 1.51.0, ggplot2 1.0.1, graph 1.47.2, grid 3.2.1, gtable 0.1.2, highr 0.5, hwriter 1.3.2, lambda.r 1.1.7, lattice 0.20-33, latticeExtra 0.6-26, limma 3.25.15, magrittr 1.5, munsell 0.4.2, pheatmap 1.0.7, plyr 1.8.3, proto 0.3-10, reshape2 1.4.1, rjson 0.2.15, scales 0.2.5, sendmailR 1.2-1, splines 3.2.1, stringi 0.5-5, stringr 1.0.0, survival 2.38-3, tools 3.2.1, xtable 1.7-4, zlibbioc 1.15.0

12 Funding

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13 References

- Thomas Girke. systemPipeR: NGS workflow and report generation environment, 28 June 2014. URL <https://github.com/tgirke/systemPipeR>.
- Brian E Howard, Qiwen Hu, Ahmet Can Babaoglu, Manan Chandra, Monica Borghi, Xiaoping Tan, Luyan He, Heike Winter-Sederoff, Walter Gassmann, Paola Veronese, and Steffen Heber. High-throughput RNA sequencing of pseudomonas-infected arabidopsis reveals hidden transcriptome complexity and novel splice variants. *PLoS One*, 8 (10):e74183, 1 October 2013. ISSN 1932-6203. doi: 10.1371/journal.pone.0074183. URL <http://dx.doi.org/10.1371/journal.pone.0074183>.

Ben Langmead and Steven L Salzberg. Fast gapped-read alignment with bowtie 2. *Nat. Methods*, 9(4):357–359, April 2012. ISSN 1548-7091. doi: 10.1038/nmeth.1923. URL <http://dx.doi.org/10.1038/nmeth.1923>.