



### **Canadian Bioinformatics Workshops**

### Introduction to R Programming for Bioinformatics

Day 2- Module 3A: Introduction to Bioconductor and Genomic Data

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## Learning Objectives

Learn what Bioconductor is

Explore genomic data structures

Practice loading and exploring datasets

Prepare data for downstream bioinformatics

## What is Bioconductor?

- Open-source project for computational biology and bioinformatics
- Built on R
- Provides thousands of packages for:
  - Genomics
  - Transcriptomics
  - Proteomics
  - Epigenomics
  - And many other types of biological data



• First released in 2001, now with a large global community

## Why Bioconductor Matters

Standardized data structures → reproducibility

Rich ecosystem of packages maintained by domain experts

Integration with public data repositories (e.g., GEO, TCGA, Ensembl)

• Essential for omics analysis in biomedical research

## Installing Bioconductor Packages

- Bioconductor uses *BiocManager* for installation
- Packages are version-matched with R

```
# Install BiocManager if not installed
install.packages("BiocManager")

# Install a package, e.g. GenomicRanges
BiocManager::install("GenomicRanges")

# Load a package
library(GenomicRanges)
```

### Core Bioconductor Data Structures

#### ExpressionSet

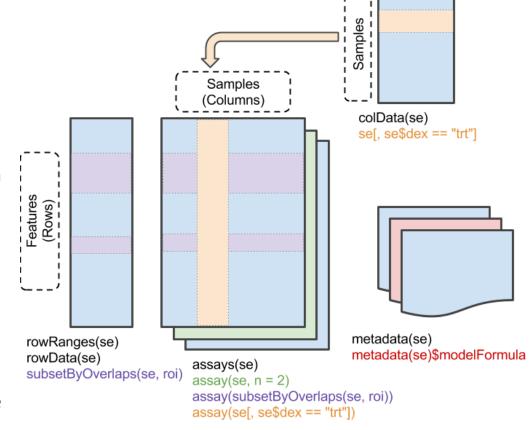
- Legacy structure for microarray & expression data
- Contains assay data + phenotypic data + feature data

#### SummarizedExperiment

- Modern replacement, widely used
- Holds assay data (e.g., counts), row metadata (genes), and column metadata (samples/patients)

#### These structures ensure data + metadata stay linked

- Assays: numeric data (counts, intensities)
- o rowData / featureData: data about features (e.g. genes)
- o colData / phenoData: data about samples/patients
- The slots ensure that subsetting keeps the assay-metadata linkage intact



- An assay matrix (features × samples)
- Linked row metadata (gene annotations)
- Linked column metadata (sample/clinical information)

## Demo: SummarizedExperiment

#### Create simple SummarizedExperiment

- Rows = genes
- Columns = samples
- Metadata describes both

```
class: SummarizedExperiment
dim: 5 4
assays(1): counts
rownames: NULL
rowData names(1): gene
colnames: NULL
colData names(1): condition
```

**dim** = 5 rows (genes) × 4 columns (samples)

**assays(1)** = the count matrix

rowData = gene names metadata

**colData** = sample conditions metadata

### Demo2: The ALL Dataset

- ALL = Acute Lymphoblastic Leukemia expression data
- Packaged in Bioconductor (ALL package)
- 128 samples of ALL patients
- Metadata: age, gender, diagnosis date, etc.

```
ExpressionSet (storageMode: lockedEnvironment)
assayData: 12625 features, 128 samples
element names: exprs
protocolData: none
phenoData
sampleNames: 01005 01010 ... LAL4 (128 total)
varLabels: cod diagnosis ... date last seen (21 total)
varMetadata: labelDescription
featureData: none
experimentData: use 'experimentData(object)'
pubMedIds: 14684422 16243790
Annotation: hau95av2
```

```
# Demo 2: ALL dataset
BiocManager::install("ALL")
library(ALL)
data(ALL)
ALL
```

**assayData**: 12,625 features (genes) × 128 samples (patients). **phenoData**: metadata about samples, such as: Patient code, Diagnosis date, Sex, Age, Immunophenotype (BT), Remission status, Relapse, Cytogenetics

**featureData**: empty here, but would contain gene annotations in some datasets.

**Annotation**: refers to the Affymetrix microarray chip used (HG-U95Av2).

## Hands-on: Exploring ALL Metadata

#### Tasks for students:

- Inspect the dataset (ALL)
  - Tip: type *ALL*
- Extract column metadata: pData(ALL)
  - Tip: use pData()
- Count and plot how many patients by gender
  - Tip: use table() on the meta data you got from the last step
- Calculate average age at diagnosis
  - Tip: use mean() and remove NA

```
# Extract and preview sample (patient) metadata
meta <- pData(ALL)
head(meta) # first 6 rows

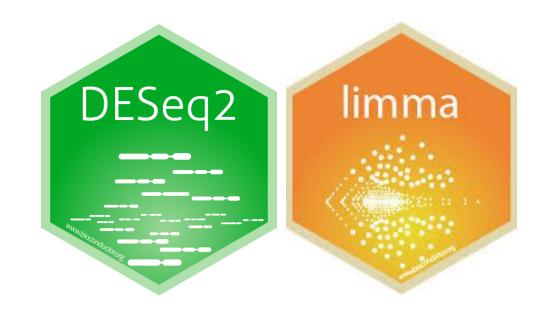
# Gender distribution
table(meta$sex)

# Mean age (ignoring missing values)
amean(meta$age, na.rm = TRUE)</pre>
```

## Preprocessing Data

#### **Common tasks in genomic analysis:**

- Filtering low-quality samples or features
- Normalization of expression values
- Transformation (e.g., log2)
- **Batch correction** for technical artifacts



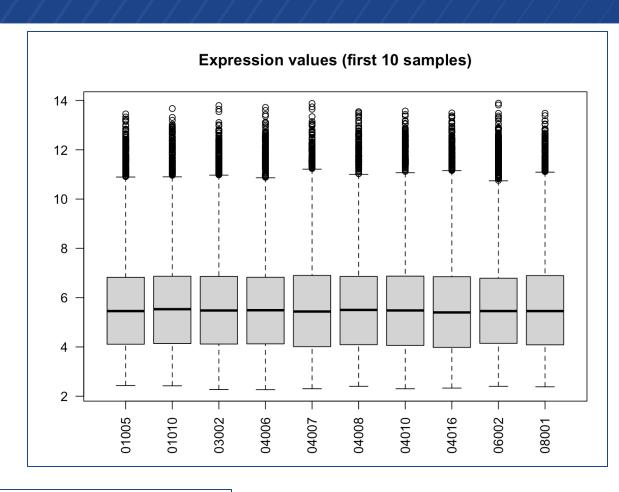
Many Bioconductor packages provide pipelines (e.g., limma, DESeq2)

### Visualization in Bioconductor

Visual checks are essential before analysis

- Common plots:
  - Boxplots for sample distributions
  - Heatmaps for expression matrices
  - PCA plots for sample clustering

Example (Boxplot of ALL expression data):



```
# Visualization in Bioconductor
boxplot(exprs(ALL)[,1:10], las=2, main="Expression values (first 10 samples)")
```

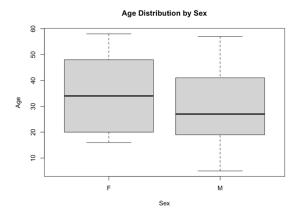
### Hands-on Exercise

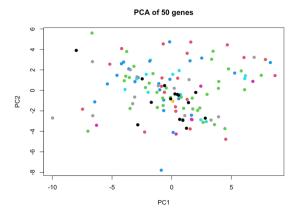
#### **Tasks for Students:**

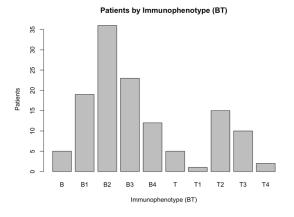
- Subset the dataset: Select only patients younger than 20 years old.
  - Tip: use dim()
- Summarize by group: Plot how many patients are in each Immunophenotype group (BT).
  - Tip: use barplot(), table()
- Create a PCA plot of expression data for the first 50 genes across all patients.
  - Tip: use prcomp(), plot()
- Visualize gender differences: Create a boxplot of patient age split by sex.
  - Tip: use boxplot()
- (Challenge) Filter out patients with missing age values and re-run the PCA.
  - Tip: explor the use of ! and is.na()

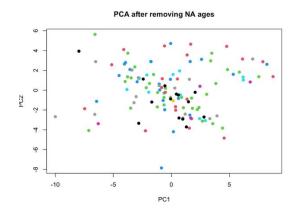
### Hands-on Exercise

```
# Subset patients < 20
young_patients <- ALL[, pData(ALL)$age < 20]</pre>
dim(young_patients)
# Count patients by Immunophenotype (BT)
table(pData(ALL)$BT)
# PCA on first 50 genes
expr <- exprs(ALL)[1:50, ]
pca <- prcomp(t(expr), scale. = TRUE)</pre>
plot(pca$x[,1:2], col = as.factor(pData(ALL)$BT),
     pch=19, main="PCA of 50 genes")
#Boxplot of Age by Sex
boxplot(age \sim sex, data = pData(ALL),
        main="Age Distribution by Sex", xlab="Sex", ylab="Age")
# Challenge (Filter missing age & re-run PCA)
ALL_clean <- ALL[, !is.na(pData(ALL)$age)]
expr_clean <- exprs(ALL_clean)[1:50, ]
pca_clean <- prcomp(t(expr_clean), scale. = TRUE)</pre>
plot(pca\_clean$x[,1:2], col = as.factor(pData(ALL\_clean)$BT),
     pch=19, main="PCA after removing NA ages")
```









# THANK YOU





