

Co-regulation of NER repair factor expression

Third TAC-meeting

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Introduction

The recently published model-aided analysis of the DNA repair process revealed the link between the emergent phenomenon of rapidly exchanging and transiently interacting NER components with the experimentally observed slow first-order kinetics of repair [1]. An important functional consequence of this kinetic design is that the control of the repair rate is shared by all repair factors. This manifests in the mathematical prediction of uniformly distributed response coefficients, which quantify the relative change of the repair rate in answer to changes in the nuclear repair protein concentration. Exploiting the natural variability in NER factor expression we experimentally corroborated the moderate control of the repair components on the repair rate. However, these findings were made under the assumption of a functional independence of the individual NER components. Downstream effects regulating the repair rate due to NER factor co-expression were so far not considered.

To test this assumption, we experimentally investigate the potential cross-correlation between five repair factors (XPC, TFIIH, XPA, XPF and RPA). Surprisingly, we find that the nuclear expression of these pairwise measured repair factors is indeed strong positively correlated, whereas there is no correlation with the repair-independent cell cycle marker Ki67. This result suggests an additional control mechanism orchestrating NER factor expression on the transcriptional or translational level.

Nuclear expression of NER factors is strongly correlated

Using fluorescence microscopy for the quantitative analysis of the NER process proved to result in accurate measurements of the nuclear repair factor expression and their UV-induced repair dynamics. This applies for the detection of stably transfected fluorescently tagged repair proteins as well as for an immunocytochemistry approach with an indirect antibody-labeling of the measured NER factors. The latter has the additional advantage that it allows for a flexible combinatorial tagging of multiple antigens simultaneously.

We established such a protocol for seven single cell double stainings (XPC-XPC; XPC-TFIIH; XPC-XPA; XPC-XPF; XPC-RPA; TFIIH-XPF; XPA-XPF; XPA-RPA) in human diploid female fibroblasts and subsequently performed a cross-correlation analysis. The pairwise co-expression was quantified by 3-dimensional imaging with a confocal microscope. To test whether the antibody co-staining experiment is suitable for the cross-correlation analysis we measured XPC expression with a directly labeled antibody together with an indirect immuno-staining and correlated both signals (cf. Figure 1A). The measurement error was determined with a principal components analysis. ... Hence, this approach is sufficiently accurate to exploit the natural variability in protein expression for the cross-correlation analysis.

As it turns out, all pairwise correlations of the measured co-staining experiments are strong positively correlated with correlation coefficients between 0.74 and 0.88 (cf. Figure 1B-D (only

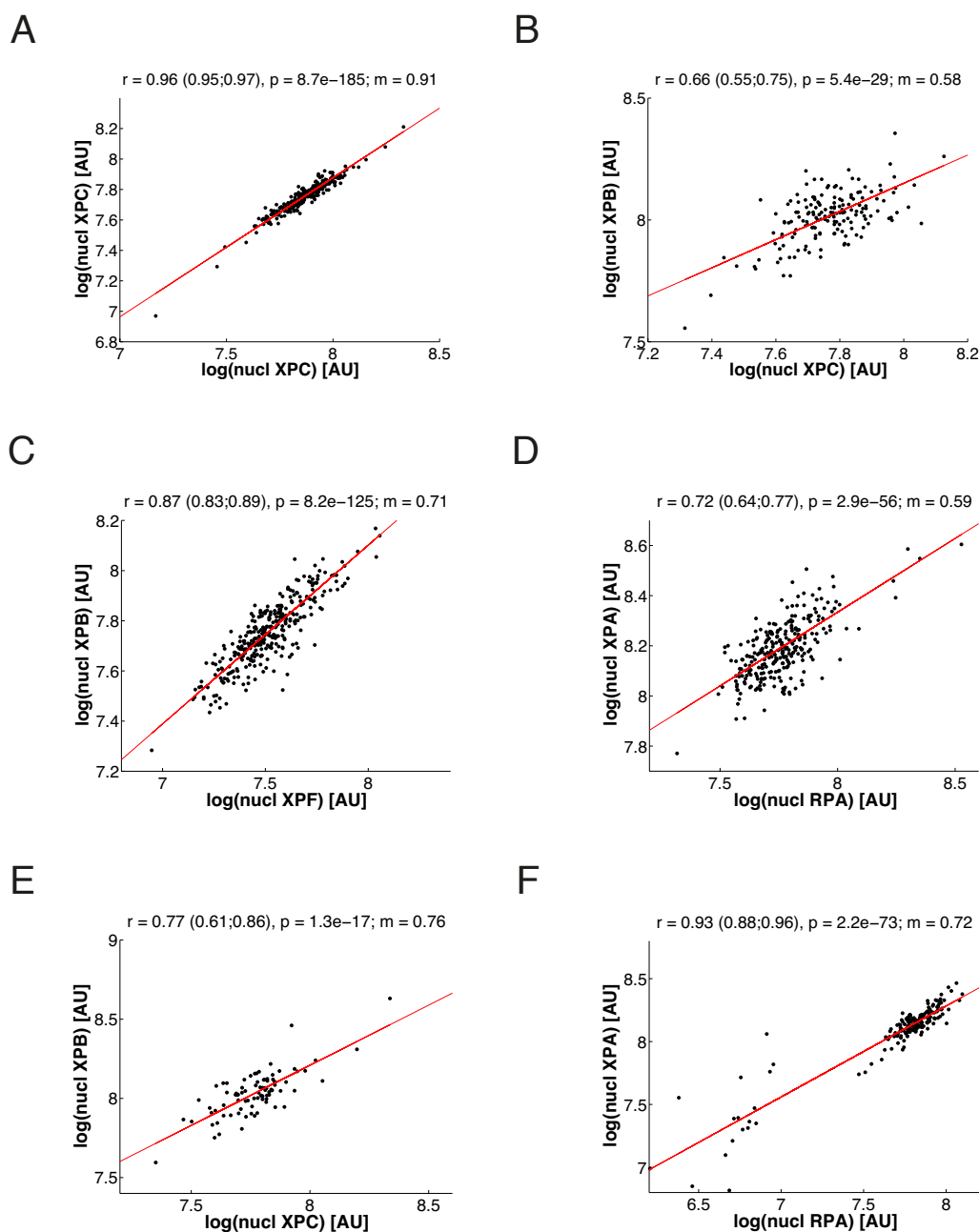


Figure 1: NER factor cross-correlation is repair independent. A) Scatter plot of indirectly antibody stained XPC against a directly labeled antibody recognizing XPC ($n=$) as determined by quantitative (immuno) fluorescence microscopy. B-D) Pairwise correlations of indirectly antibody-labeled XPB against XPC (B), XPB against XPF (C) and XPA against RPA (D) in locally damaged cells. Expression values represent fluorescence intensities originating from the nucleus including the damaged region (signal quantification was performed analogous to [2]). E-F) Scatter plots of the nuclear expression of XPB vs. XPC and XPA vs. RPA in undamaged cells.

selected pairs)). Notably, the result is indifferent whether the acquired fluorescence signal is taken from the whole nucleus including the locally damaged area or only from the undamaged chromatin region (cf. Figure 1B-D and 1E-F). This suggests that the correlation of nuclear NER factors is independent of the ongoing repair. As a consequence, we suspect the regulatory mechanism determining the protein concentrations on a preceding level like transcription or translation.

To further pursue this question we repeated the experiment by flow cytometry, this time, investigating the NER-factor expression in human brain pericytes. We established a double-staining protocol for XPA and RPA and observed that also in this cell type the two repair factors strongly correlate (cf. Figure 2A and B).

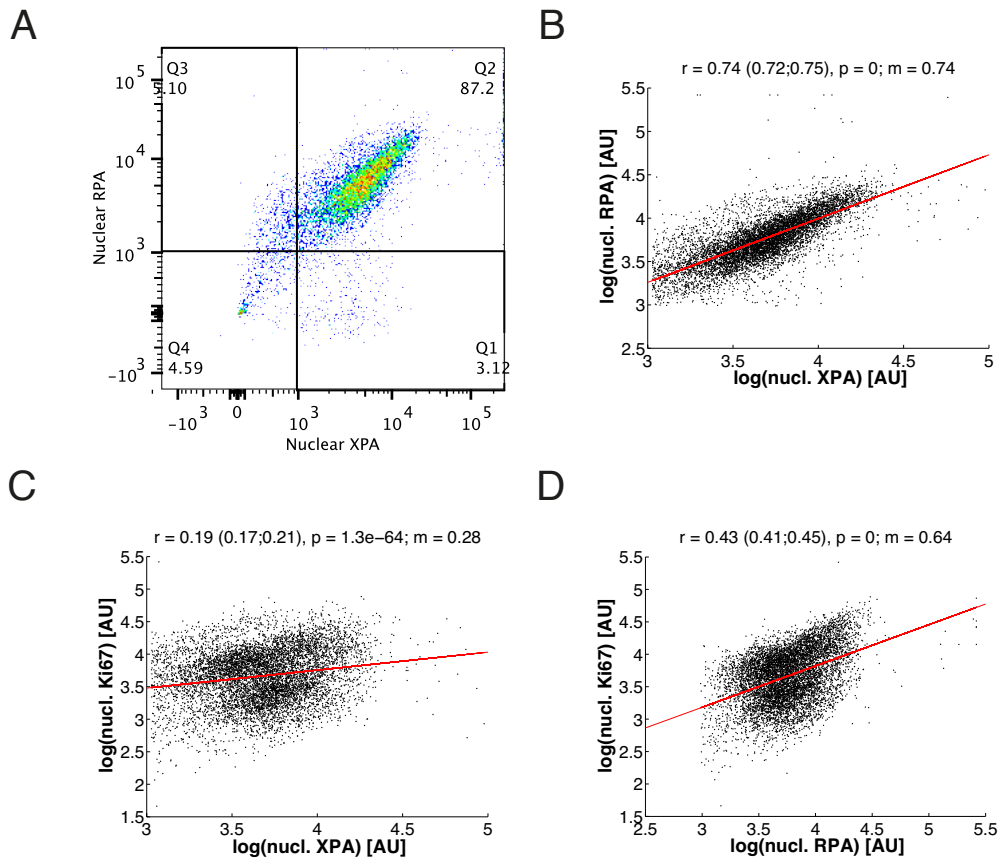


Figure 2: Correlated expression of RPA and XPA in human brain pericytes A) Selection of XPA and RPA positive human brain pericytes (Q2) as determined by flow cytometry ($n=$). B-D) Nuclear expression of indirectly antibody-labeled RPA against XPA (B), Ki67 against XPA (C) and Ki67 against RPA (D).

Quantitatively, the correlation coefficients of the double-staining signals in both cell types have the same order of magnitude. In particular, each value falls into the confidence interval of the other (cf. Figure 2B and Figure 1D). To test, whether this correlation is specific for proteins involved in DNA repair we also measured the expression of the proliferation marker Ki67. Surprisingly, although the correlation between both repair factors and Ki67 is visibly reduced (XPA vs. Ki67 = 0.19 and RPA vs. Ki67 = 0.43 against XPA-RPA = 0.73) it is still significantly positive (cf. Figure 2C and D). Especially the correlation between RPA and Ki67 remains at over 50% of the XPA-RPA cross-correlation (cf. Figure 2D).

Cell cycle independent cross-correlation of NER factor expression

On closer inspection of the RPA-Ki67 correlation cloud, one could anticipate two distinct regimes characterized by a discrete increase in correlation at higher Ki67 values (cf. Figure 2D - red circle). We asked, whether this differential correlation is related to different cell cycle phases as one would expect for increased Ki67 expression during cells division? To answer this question, we stained the cell's DNA using FxCycle violet and gated them according to their DNA content (cf. Figure 3A). Two distinct peaks denote the portion of cells traversing the G1 or the synthesis and G2 phase, respectively. By sorting the protein expression values in accordance with this classification we identified for each protein correlation two contiguous regions revealing a general trend of increased protein expression during the S1 and G2 phase in comparison to the G1 phase (cf. Figure 3B-D).

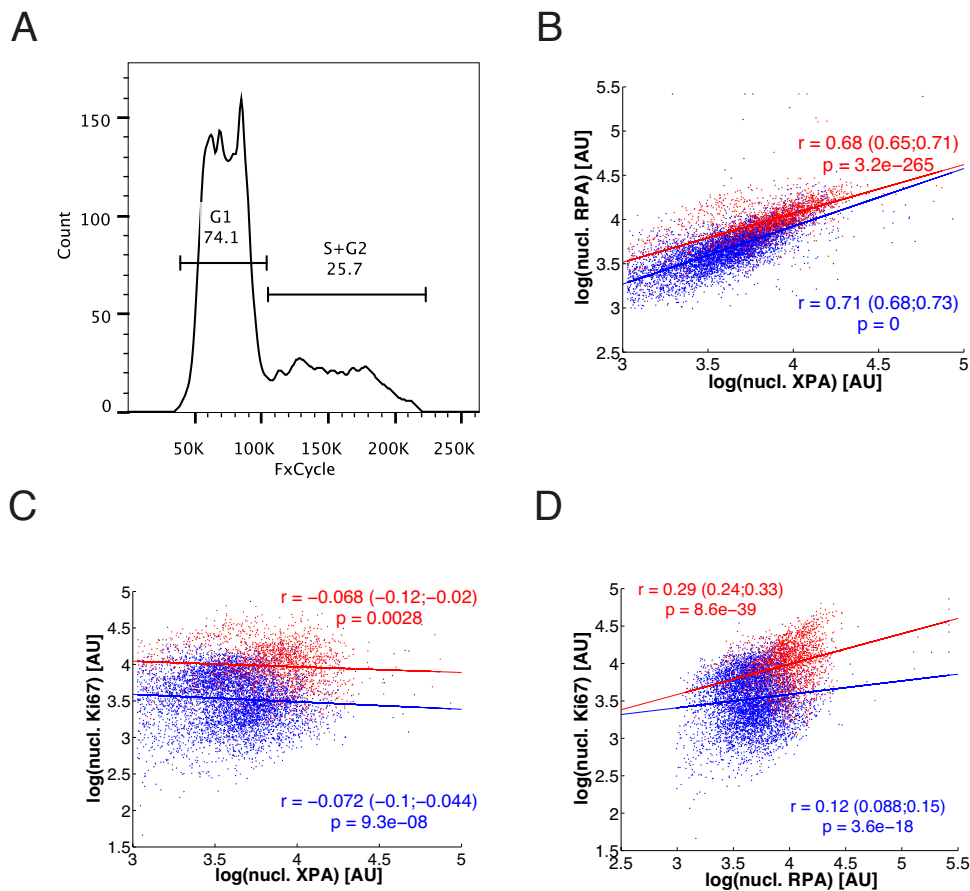


Figure 3: NER factor cross-correlation is robust against cell cycle progression. A) Fluorescence Distribution of fluorescently labeled DNA proportional to the DNA content. Horizontal bars indicate the fractions of cells assigned to G1 (left peak) and S+G2 (right peak). B-D) Individual regression analysis of the nuclear expression values sorted according to the corresponding cell-cycle phase (G1 - blue; S+G2 - red) for RPA vs. XPA (B), Ki67 vs. XPA (C) and Ki67 vs. RPA (D).

Remarkably, whereas the correlation coefficients between XPA and RPA remain constant for both regimes (G1: 0.71, S+G2: 0.68, all: 0.74) the correlation between XPA and Ki67 disappear (G1: -0.072, S+G2: -0.068). Between RPA and Ki67 there is close to no correlation in the G1 phase but a significant small positive correlation in the S and G2 phase (G1: 0.12, S+G2: 0.29). This result strengthens the impression derived by the cross-correlation analysis in fibroblasts

that NER factor expression is functionally co-regulated. In particular, the missing correlation between the nuclear factors and the cell cycle marker after the reassignment in accordance with the cell-cycle phase suggests that the measured correlation is specific to NER proteins.

Conclusion

To determine potential mutual dependencies in NER factor expression we performed two independent cross-correlation experiments, one in human fibroblast under the microscope and the other in human brain pericytes using flow cytometry. For both approaches we found conclusive evidence for a positive pairwise correlation of the nuclear repair factor concentration. We could show that the correlation is independent of the repair process and the cell cycle phase despite the general elevation of the protein expression levels in the S and G2 phase compared to G1. This result indicates a so far unknown regulatory step organizing the repair factor co-expression, which has to be considered when determining the control on the rate of repair.

References

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