

## **DNA TEST REPORT**

SOMATIC MUTATION PANEL ANALYSIS REPORT					
Subject Information		Sample Information	Sample Information		
Patient Name:	XXXXX	Order ID/Sample ID:	XXX/12345		
Gender:	Male	Collection Date & Time:	NA		
Age:	43 years	Receipt Date & Time:	11 <sup>th</sup> January, 2013, 10:00 AM		
Father-Mother:	NA	Report Date & Time:	9 <sup>tht</sup> Feb, 2013 4:00 PM		
Sample Type:	FFPE Tumor tissue Blood (2.5ml)	Requested by:			
Clinical Indication (If any): Metastatic carcinoma of bladder					

The raw sequencing fields (including the fastq as well as sequence alignment results) can be provided upon request from the referring physician.

Analysis:
-----------

# The genes sequenced and their coverage details are listed in Appendix 1.

Table 1	Genomic alterations that can be targeted with approved drugs  Table 1								
SI. No.	Gene	CDS variant#	Amino acid variant	Impact on Protein Function	Approved drugs	Drug response	Function of the gene in cancer	Hot spot mutation	Classification of variant
	NONE								

Tak	ole 2	Non-druggable, but clinically significant alterations							
SI. No	Gene	CDS variant#	Amino acid variant	Impact on Protein Function	Approved d drugs		Function of the gene in cancer	Hot spot mutation	Classification of variant
1.	PTEN	c.596delT (ENST00000 371953)	p.Met199 SerfsTer22	Yes	NA	NA	TSG	No	Pathogenic
2.	STK11	c.541delA (ENST00000 326873)	p.Asn181 ThrfsTer106	Yes	NA	NA	TSG	No	Pathogenic



TSG- Tumor Suppressor Gene

Annotation is performed against GRCh37/hg19 version of human gene.

# cDNA base is reverse compliment of genomic base in case on negative strand.

# **Test Details & Interpretation**

### **Background**

The next-generation sequencing based multi-gene analysis for cancer predisposition genes and tumor mutations will enable effective risk management and treatment by doctors. Thus, it allows us to sequence and identify variants associated with multiple genes with implications in cancer. Targeted sequencing represents a cost-effective approach with the ability to detect specific variants causing protein-coding changes in individual human genomes. These multi-gene, affordable tests will enable personalized treatment by matching the patient with the appropriate drug, based on the mutational profile.

#### Test

DNA isolated from tumor tissue and paired normal (blood) samples were used to perform enrichment and sequencing using tumor gene panel kit. The tumor tissue and the paired normal (blood) panels were covered at an average sequencing depth of 166X and 739X respectively. The sequences obtained were aligned to human reference genome (GRCh37/hg19) using BWA program [2, 3] and analyzed using Picard and GATK-Lite toolkits [4, 5]. Somatic variants unique to the tumor tissue were identified using Strelka somatic caller and annotated using our in-house annotation pipeline. Clinically relevant mutations were annotated using published variants in literature and a set of well-curated databases including ClinVar, SwissVar, OMIM, GWAS, OncoMD and COSMIC [6-11]. Only non-synonymous variants found in the sample were used for clinical interpretation.

#### Interpretation

### PTEN p.Met199SerfsTer22 (Table 2):

A single nucleotide deletion (chr10:89711978delT) that results in a frame shift and subsequent termination of the protein 22 amino acids downstream to codon 199 (p.Met199SerfsTer22) was detected in the *PTEN* gene of this subject. This deletion result in the production of a truncated PTEN protein which is functionally deficient. The *PTEN* gene encodes a ubiquitously expressed tumor suppressor protein that acts as the major cellular suppressor of PI3K signalling and AKT activation in cell proliferation. Loss of function variations in *PTEN* have been characterized in 50% of human gallbladder carcinoma (12). Targeted agents acting at the level of PI3K or downstream may be most effective for treatment of PTEN-deficient cancers. Several such drugs are currently in clinical trials for the treatment of patients with PTEN-deficient cancers (table 4). PTEN deficiency is also associated with resistance to RTK inhibitors *in vitro* (13).

#### STK11 p.Asn181ThrfsTer106 (Table 2):

A single nucleotide deletion (chr19:1220448delA) that results in a frame shift and subsequent termination of the protein 106 amino acids downstream to codon 181 (p.Asn181ThrfsTer106) was detected in the *STK11*gene of this subject. This deletion also results in the production of a truncated, functionally deficient protein. Homozygous deletions or somatic sequence mutations coupled with loss of heterozygosity in the *STK11* gene, have been identified in sporadic pancreatic and biliary adenocarcinomas (14). *STK11* acts as a tumor suppressor by negatively regulating the mTOR pathway. Hence, *STK11* loss of function results in increased mTOR signaling suggesting that therapies targeting mTOR (eg. mTOR inhibitors like FDA approved Everolimus and Temsirolimus) may be relevant for *STK11* deficient tumors (15,16). A patient with pancreatic cancer and an *STK11* mutation experienced a partial response to the mTOR inhibitor Everolimus (18). Loss of *STK11* also leads to activation of



the downstream kinase Src, suggesting that inhibitors such as dasatinib or bosutinib may be relevant for the treatment of *STK11*-deficient tumors (19).

<u>Variants of unknown significance (VUS) in genes relevant in cancer</u> in human cancer. The variants specifically detected in this tumor have not been characterized.

Table 3 (Appendix 2) provides a list of VUS in genes known to function as oncogenes or tumor suppressor genes tly in biochemical assays and therefore their impact in this cancer remains speculative.

- The classification of variants of unknown significance can change over time. Please contact
   MedGenome at a later date for any change.
- Intronic variants are not assessed using this method.
- Large deletions or copy number variations cannot be assessed using this method.
- Certain genes may not be covered completely and few mutations could be missed.
- Only the genes that are 100% covered (Appendix-1) are included in this analysis.

#### **References:**

- 1. Richards CS, Bale S, Bellissimo DB, Das S. ACMG recommendations for standards for interpretation and reporting of sequence variations: Revisions 2007. Genet Med. 2008 Apr;10(4):294-300.
- 2. Li, H. and R. Durbin. Fast and accurate long-read alignment with Burrows-Wheeler transform. Bioinformatics, 2010. 26(5): p. 589-95.
- 3. Meyer, L.R., et al., The UCSC Genome Browser database: extensions and updates 2013. Nucleic Acids Res, 2013. 41(D1): p. D64-9.
- 4. McKenna, A., et al., The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. Genome Res, 2010. 20(9): p. 1297-303.
- 5. Li, H., et al., The Sequence Alignment/Map format and SAMtools. Bioinformatics, 2009. 25(16): p. 2078-9
- 6. http://www.ncbi.nlm.nih.gov/clinvar/
- 7. http://www.omim.org/
- 8. https://www.gwascentral.org/
- 9. http://cancer.sanger.ac.uk/cancergenome/projects/cosmic/
- 10. http://oncomd.medgenome.com/MutationViewer/
- 11. <a href="http://swissvar.expasy.org/">http://swissvar.expasy.org/</a>
- 12. Lunardi A, Webster KA, Papa A, Padmani B, Clohessy JG, Bronson RT, Pandolfi PP. Role of aberrant PI3K pathway activation in gallbladder tumorigenesis. Oncotarget. 2014 Feb 28;5(4):894-900.
- 13. Dillon LM, Miller TW.Therapeutic targeting of cancers with loss of PTEN function. Curr Drug Targets. 2014 Jan;15(1):65-79.
- 14. Su GH, Hruban RH, Bansal RK, Bova GS, Tang DJ, Shekher MC, Westerman AM, Entius MM, Goggins M, Yeo CJ, Kern SE. Germline and somatic mutations of the STK11/LKB1 Peutz-Jeghers gene in pancreatic and biliary cancers. Am J Pathol. 1999 Jun; 154(6):1835-40.
- 15. Shaw RJ, Bardeesy N, Manning BD, et al. (2004) The LKB1 tumor suppressor negatively regulates mTOR signaling. Cancer Cell 6(1):91-9
- 16. Yuan R, Kay A, Berg WJ, Lebwohl D. Targeting tumorigenesis: development and use of mTOR inhibitors in cancer therapy. Journal of Hematology & Oncology. 2009;2:45.
- 17. Zhao R-X, Xu Z-X. Targeting the LKB1 Tumor Suppressor. Current drug targets. 2014;15(1):32-52.



- 18. Klümpen HJ, Queiroz KC, Spek CA, et al. (2011) mTOR inhibitor treatment of pancreatic cancer in a patient With Peutz-Jeghers syndrome. J Clin Oncol 29(6):e150-3
- 19. 18. Carretero J, Shimamura T, Rikova K, et al. (2010) Integrative genomic and proteomic analyses identify targets for Lkb1-deficient metastatic lung tumors. Cancer Cell 17(6):547-59

## **APPENDIX-1**

Genecoding region coveredGenecoding region coveredGenecoding coveredELF399.03HIST1H2BD100.00SIN3A100.00	ntage of g region vered 00.00 00.00 8.46
covered         covered         covered           ELF3         99.03         HIST1H2BD         100.00         SIN3A         10	vered 00.00 00.00
ELF3 99.03 HIST1H2BD 100.00 SIN3A 10	00.00
	00.00
PPOX         100.00         EPHB6         100.00         CTCF         10	8.46
CDKN2C         100.00         PIK3CG         99.51         CDH1         9	
NRAS 100.00 NFE2L3 100.00 CBFB 10	00.00
RPL5 100.00 KMT2C 100.00 CTD-2012K14.6 10	00.00
ARID1A 100.00 NDUFB2 100.00 CDK12 10	00.00
RPL22 91.58 HGF 100.00 WRAP53 10	00.00
MTOR         100.00         BRAF         100.00         MAP2K4         9	9.20
B4GALT3 100.00 EZH2 100.00 BRCA1 9	9.38
PCBP1-AS1 100.00 EGFR 100.00 MIR142 10	00.00
ACVR2A 100.00 RAD21-AS1 100.00 SOX9 10	00.00
IDH1 100.00 EGR3 100.00 CTD-2535L24.2 10	00.00
ERBB4 100.00 RP11-459E5.1 100.00 NCOR1 9	8.62
SF3B1 100.00 SOX17 100.00 AXIN2 10	00.00
DNMT3A 98.79 EPPK1 100.00 BZRAP1-AS1 10	00.00
IDH1-AS1 100.00 UTP23 100.00 SPOP 10	00.00
PCBP1 100.00 RAD21 100.00 VEZF1 9	9.45
NFE2L2 100.00 RP11-145E5.5 100.00 NF1 10	00.00
TGFBR2 100.00 CDKN2A 100.00 TP53 9	8.74
TBL1XR1 100.00 NOTCH1 100.00 SMAD2 10	00.00
ATR 100.00 TLR4 89.52 SETBP1 10	00.00
PBRM1 100.00 PTEN 100.00 RP11-729L2.2 10	00.00
POLQ 100.00 GATA3 100.00 RP11-456K23.1 10	00.00
PIK3CA 99.49 FGFR2 100.00 SMAD4 10	00.00
VHL 86.66 SMC3 100.00 PPP2R1A 10	00.00
CTNNB1 100.00 ARID5B 100.00 KMT2B 10	00.00
BAP1 100.00 ATM 100.00 PRX 10	00.00
GNL3 100.00 ORAOV1 100.00 KEAP1 10	00.00
AK095242 100.00 C11ORF65 100.00 ARHGAP35 10	00.00
AK311005 100.00 MALAT1 100.00 CEBPA 10	00.00
RP11-641D5.2 100.00 WT1 100.00 TSHZ3 10	00.00
MECOM 100.00 MAPK8IP1 100.00 ERCC2 10	00.00
EPHA3 98.61 CCND1 100.00 STK11 10	00.00
EIF4A2 100.00 LRRK2 100.00 FOXA2 10	00.00
SETD2 100.00 NAV3 100.00 ASXL1 9	9.43
FBXW7 100.00 H3F3C 100.00 AL354993.1 9	9.07
TET2 99.67 LYRM5 100.00 TSHZ2 10	00.00
KIT 100.00 ACVR1B 100.00 RUNX1 10	00.00
FGFR3 100.00 PTPN11 99.86 U2AF1 10	00.00
CRIPAK 100.00 CDKN1B 100.00 EP300 10	00.00
FIP1L1 100.00 TBX3 100.00 CHEK2 10	00.00



Gene	Percentage of coding region covered	Gene	Percentage of coding region covered	Gene	Percentage of coding region covered
PDGFRA	100.00	KMT2D	100.00	TAF1	95.81
NSD1	100.00	KRAS	100.00	RP1-315G1.3	100.00
PIK3R1	99.36	RB1	100.00	ATRX	100.00
LIFR	100.00	BRCA2	100.00	AR	99.84
LIFR-AS1	100.00	FLT3	100.00	SMC1A	87.67
MAP3K1	100.00	AKT1	100.00	KDM5C	97.70
APC	100.00	AJUBA	100.00	PHF6	100.00
CTC-554D6.1	100.00	RP11-298I3.5	100.00	USP9X	100.00
NPM1	100.00	FOXA1	100.00	KDM6A	100.00
CDKN1A	100.00	IDH2	100.00	STAG2	100.00
HIST1H1C	100.00				

## **APPENDIX -2**

Table 3 V			Variants (	of unknown significance in oncogo	enes and tumor suppressor gen	es
	SI. No.	Gene		CDS variant	Amino acid variant	Function of the gene in cancer
	1.	KMT2D		c.3379_3383delGGGAT (ENST00000301067)	p.Gly1127Ter	TSG

TSG- Tumor Suppressor Gene

Table 4		Approved/ Investigational drugs against variants in oncogenes and tumor suppressor genes of this subject					
SI. No.	Gene	Drug (Generic name)	Approved in patient's cancer	Approved in other cancer			
1.	PTEN	Temsirolimus, Everolimus (Approved)	No	Renal cell carcinoma			
		Ridaforolimus (under clinical development)	-	-			
2.	STK11	Temsirolimus, Everolimus (Approved)	No	Renal cell carcinoma			
		Several under clinical development (Ref.13)	-	-			

TSG- Tumor Suppressor Gene