

# Key ingredients for RNA-seq differential analysis

## Neutral comparison study - The importance of modeling

M-L Martin-Magniette, E. Delannoy (and G. Rigauil)



# This is not a tutorial on how to analyse RNAseq data

- A good tutorial can be found here:

<http://www.nathalievilla.org/doc/pdf/slides-rnaseq.pdf>

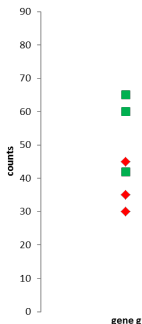
- It covers
  - Exploratory analysis
  - Normalization
  - Differential Expression analysis

# What this is about

- Here we will focus on the differential analysis
  - when only a few replicates are available
- What are the important parameters in "practise" ?
  - a strategy to identify those on your own data ?
- This is part of a course on differential analysis taught by E. Delannoy and M.L Martin-Magniette to biologists and bioinformaticians at
  - IPS2 : "Ecole chercheur SPS"
  - LIPM : "Analyse de données 'omiques"
- Details can be found in the paper:  
<https://www.ncbi.nlm.nih.gov/pubmed/27742662>
- Main conclusion **"Modeling is important"**!

# Objective of the differential analysis

- The aim is to identify a significant difference of expression between two given conditions
- It is performed with an hypothesis test based on gene expression measurements



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- It is performed with an hypothesis test based on gene expression measurements



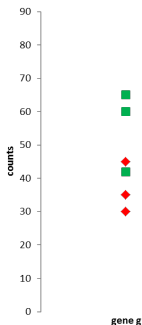
$H_0 = \{\text{There is no difference}\}: \text{red} = \text{green}$   
versus

$H_1 = \{\text{There is a difference}\}: \text{red} \neq \text{green}$

# Key steps for a test procedure

## Construction of a test

- Formulate the two hypotheses



$H_0 = \{\text{There is no difference}\}: \text{red} = \text{green}$

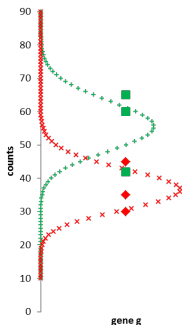
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$H_1 = \{\text{There is a difference}\}: \text{red} \neq \text{green}$

# Key steps for a test procedure

## Construction of a test

- Formulate the two hypotheses
- Construct the test statistic
- Define its distribution under the null hypothesis



$H_0 = \{\text{There is no difference}\}: \text{red} = \text{green}$

versus

$H_1 = \{\text{There is a difference}\}: \text{red} \neq \text{green}$

Estimation of the variance of expression  
for gene  $g$

# Key steps for a test procedure

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$H_0 = \{\text{There is no difference}\}$ : red = green

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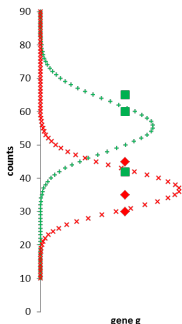
$H_1 = \{\text{There is a difference}\}$ : red  $\neq$  green

## Application of the test

- Calculate the value of test statistic
- Calculate the p-value
- Decide if  $H_0$  is rejected or not

## Definition of a p-value

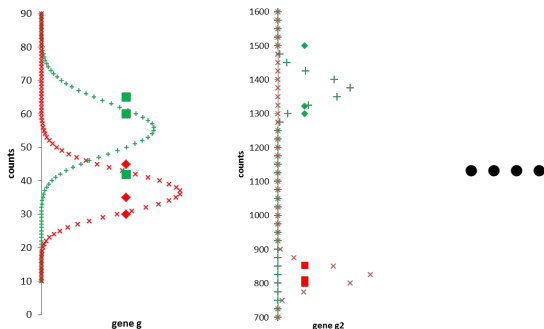
It is the probability of seeing a result at least as extreme as the observed data, when the null hypothesis is true.





# Multiple genes, multiple testing

Apply the previous procedure to every tested gene



P-value and multiple testing

- By definition,  $P(\text{to be a false positive}) = \alpha$
- If 10.000 tests are performed at level  $\alpha$ , then the averaged number of false-positives is 500

# Contingency table for multiple hypothesis testing

	True null hypotheses	False null hypotheses	
Declared non-significant	True Negatives	False Negatives	Negatives
Declared significant	False Positives	True Positives	Positives

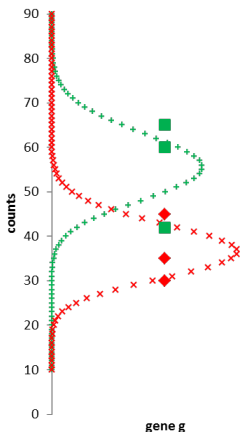
## Adjustment of the raw p-values

- $FWER = P(FP > 0)$  (Bonferroni procedure)
- $FDR = E(FP/P)$  if  $P > 0$  or 1 otherwise (Benjamini-Hochberg procedure)

## Decision rule

A gene is declared differentially expressed if its adjusted p-value is lower than a given threshold

# How to model RNA-seq data ?



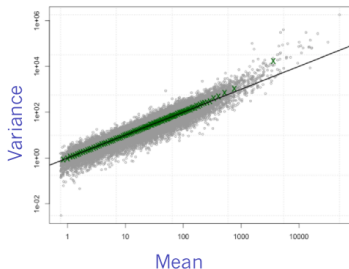
Estimation of the variance of gene g expression  
Not enough measurements (replicates)



Modeling of RNA-seq data

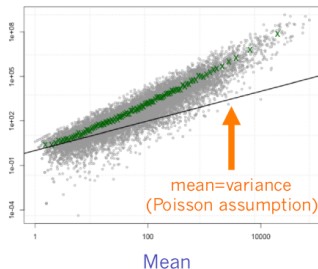
# How to model RNA-seq data ?

Technical replicates



data from Marioni et al. *Gen Res* 2008

Biological replicates



data from Parikh et al. *Genome Bio* 2010

- Overdispersion between biological replicates
- Negative binomiale distribution is often assumed:  $Y \sim NB(\mu, \phi)$

$$E(Y) = \mu$$

$$V(Y) = \mu(1 + \phi\mu)$$

# Three statistical frameworks

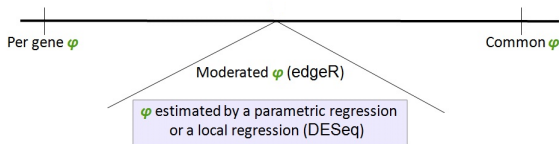
- A negative binomiale distribution (2008)
  - Expression = library size  $\times \lambda_{condition}$
- A NB generalized linear model (2012)
  - allows us to decompose the expression
  - each condition is described by several factors

$$\log(\lambda_{condition}) = Cst + \alpha_{genotype} + \beta_{stress} + \gamma_{genotype, stress}$$

- Effect of each factor is tested
- A linear model (2014)
  - data are transformed to work with a Gaussian
  - allows us to decompose the expression



- Do we filter genes with low expression (yes or no)
- How to model the gene expression (NB, GLM or LM)
- Which method to estimate the variance of the gene expression (several methods)



# Neutral comparison study

We want to answer these questions with a large evaluation study

- How the statistical models fit RNA-seq data ?  
→ study of the p-value distribution
- Do p-values well discriminate DE and NDE genes ?  
→ ROC curves
- Are the false-positives controlled ?  
→ proportion of truly NDE declared DE
- Are the methods powerful (able to find the truly DE genes)  
→ proportion of truly DE declared DE

# Which kind of data is relevant for an evaluation ?

- **Real data:**

- More realistic
- ... but no extensively validated data yet available

- **Simulated data:**

- Truth is well-controlled
- ... but what model should be used to simulate data? How realistic are the simulated data? How much do results depend on the model used?

**Our idea was to create synthetic data**



# Creation of synthetic datasets

Leaves vs Leaves

$H_0$  full  
dataset

$H_0$  genes

Buds vs Leaves

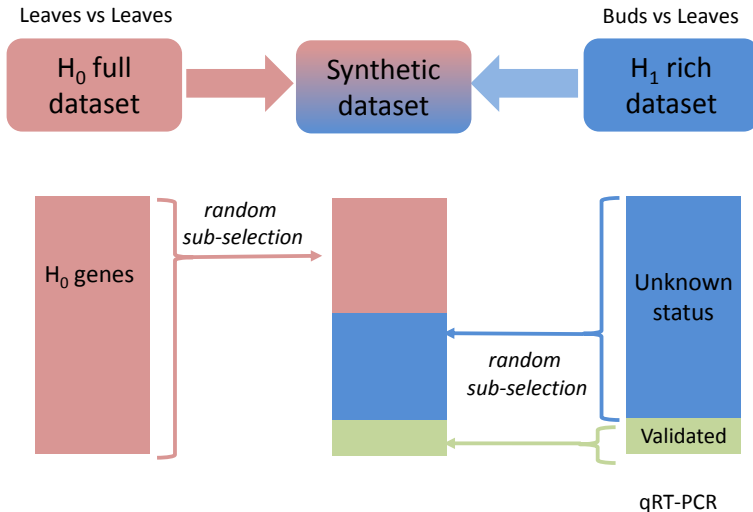
$H_1$  rich  
dataset

Unknown  
status

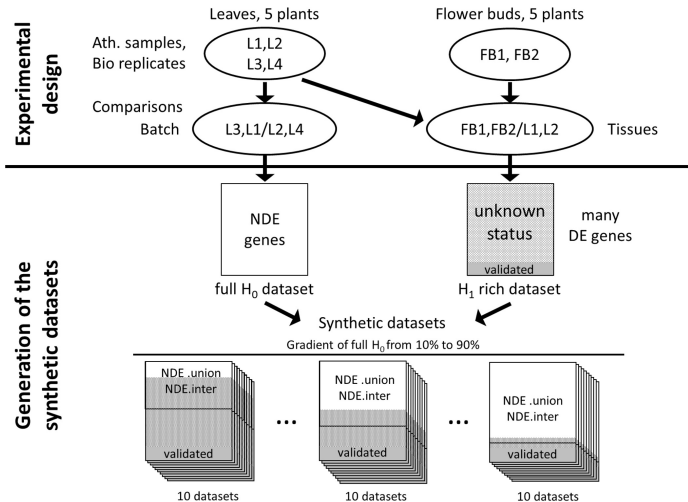
Validated

qRT-PCR

# Creation of synthetic datasets



# Creation of synthetic datasets



# Definition of the truth

## the set of truly DE genes

251 DE genes identified by qRT-PCR among 332 randomly chosen genes

## the set of truly NDE genes

- The proper identification is not straightforward  
Definition of two sets
- NDE.union: may include some genes that are not truly NDE
- NDE.inter: may exclude some truly NDE genes.

# The 3 frameworks described by 9 methods

- **edgeR** and **DESeq** are **NB-based method**

$$\text{Expression} = \text{library size} \times \lambda_{\text{condition}}$$

- **glm edgeR** and **DESeq2** are **GLM approaches**

$$\log(\lambda_{\text{condition}}) = Cst + \alpha_{\text{tissue}} + \beta_{\text{biological replicate}}$$

- **limma-voom** is **a linear model**

Data are transformed with the voom method

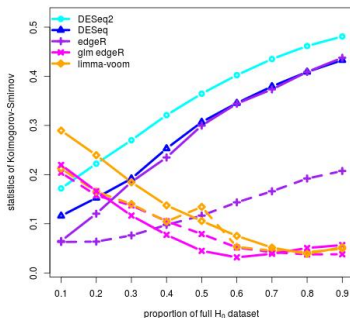
$$\text{Expression} = Cst + \alpha_{\text{tissue}} + \beta_{\text{biological replicate}}$$

- \* All methods except DESeq are also applied on filtered data
- \* In each method, nominal value of FDR is 5 %

# Distribution of the p-values

## Method

- When no difference is expected, histogram of the p-values are expected to be uniform histogram
- For each synthetic dataset, 100 evaluations of the uniform distribution of 1000 genes randomly chosen in the full  $H_0$  dataset are performed



- the raw p-values are not properly calculated (67% of tests are rejected after a strict FP control)
- test statistic values are smaller for linear or generalized linear models

# Definition of a ROC curve

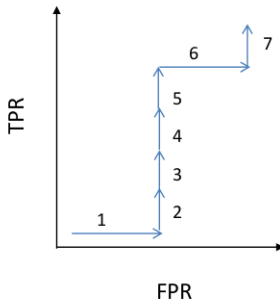
Drawing a ROC curve:

- 1- sort genes by increasing raw p-value
- 2- knowing the truth (DE or NDE) for each gene, go down the sorted list counting the proportion of all the DE genes encountered so far (TPR) and the proportion of all the NDE genes encountered so far in the list (FPR)

Example:

7 genes: 5 DE and 2 NDE

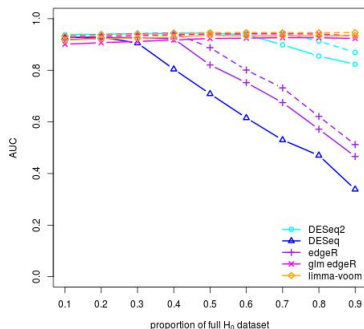
rank	gene	p-value	truth	TPR	FPR
1	G1	p1	NDE	0/5	1/2
2	G2	p2 (>p1)	DE	1/5	1/2
3	G3	p3(>p2)	DE	2/5	1/2
4	G4	p4(>p3)	DE	3/5	1/2
5	G5	p5(>p4)	DE	4/5	1/2
6	G6	p6(>p5)	NDE	4/5	2/2
7	G7	p7(>p6)	DE	5/5	2/2



# Discrimination of DE and NDE genes

## Method

- sort raw p-values into ascending order
- compare them with the truth
- construct a ROC curve and calculate AUC
- AUC close to 1 indicates a good discrimination



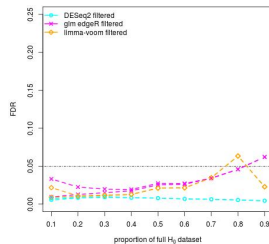
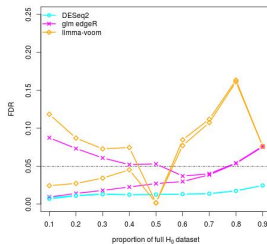
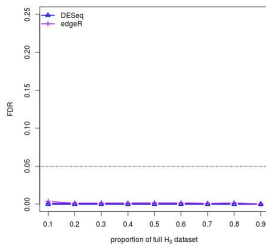
- For linear model or glm, the AUC is high and independent of the proportion of full  $H_0$  datasets
- For NB-based method, the AUC steadily decrease with the increase of the proportion of full  $H_0$  dataset when it is larger than 0.3-0.4



# FDR estimation

## Method

FDR estimation by the proportion of truly NDE among the declared DE  
Comparison with the expected value 0.05

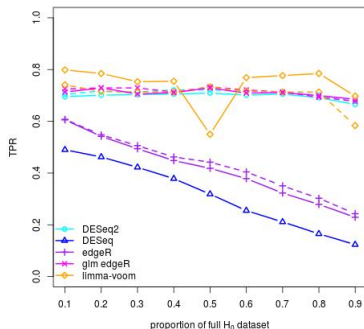


- For NB-based method, both bounds are close to 0
- For DESeq2, the FDR is always lower than 5%
- For glm edgeR, the interval generally contains 5%
- For limma-voom, the FDR control is more variable but the filtering step stabilizes its behavior

# Are truly DE declared DE ?

## Method

Proportion of truly DE genes among the declared DE genes



- LM or GLM based-methods show a high TPR
- For NB-based methods, the TPR is a function of the full  $H_0$  dataset proportion.
- The variance-mean relationship modeling and the data filtering seem to have only a limited impact.

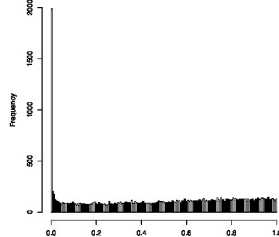
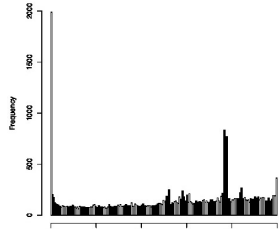
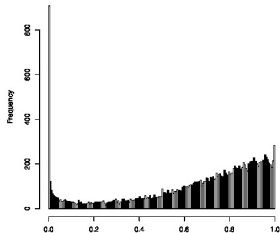
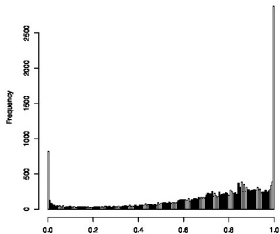
modeling  $\geq$  filtering  $\geq$  dispersion

## Synthetic data are a relevant framework

- Forget edgeR and DESeq
- use glm edgeR, DESeq2 or limma-voom
- include biological replicate as a factor
- filtering allows methods to control FDR

# Definition of an indicator of quality

An histogram with a peak at the right side = analysis of bad quality  
Let's play a game : which analysis is correct ?



# Practical case

## Context

An experiment is performed to evaluate whether a given mutant behaves as the wild-type plant. Transcriptome of both plants is measured on a three-point time series at four different dates.

## Biological questions

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**Genotype effect** what genes are differentially expressed between the two genotypes ?

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**Genotype effect** what genes are differentially expressed between the two genotypes ?

**Time effect** what genes are differentially expressed between two consecutive times ?

**Genotype x Time effect** what genes are impacted in their transcription by an interaction between the genotype and the time ?



# Statistical translation of the context

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An experiment is performed to evaluate whether a given mutant behaves as the wild-type plant. Transcriptome of both plants is measured on a three-point time series at four different dates.

## Factor identification

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- Genotype  $\in \{\text{wild-type, mutant}\}$
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- Replicate (date)  $\in \{1, 2, 3, 4\}$

2 biological factors and one technical factor

# Statistical modelling

## Factor identification

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## Statistical modelling

Let  $Y_{gtr}^{\tilde{g}}$  be the expression of gene  $\tilde{g}$  for the genotype  $g$  at time  $t$  in the  $r^{\text{th}}$  experiment and  $\mu_{gtr}^{\tilde{g}} = \log\{E(Y_{gtr}^{\tilde{g}})\}$ .

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Assume

$$\mu_{gtr}^{\tilde{g}} = \mu^{\tilde{g}} + \text{Genotype}_g + \text{Time}_t + \text{Replicate}_r + \text{Genotype}_g : \text{Time}_t$$

# Statistical modelling

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$\rightsquigarrow$  interactions between technical and biological factors are not considered



# First remarks

$$\mu_{gtr}^{\tilde{g}} = \mu^{\tilde{g}} + \textit{Genotype}_g + \textit{Time}_t + \textit{Replicate}_r + \textit{Genotype}_g : \textit{Time}_t$$

## Statistical modelling

The logarithm of the mean expression of gene  $\tilde{g}$  is modeled as

$$\mu_{gtr}^{\tilde{g}} = \mu^{\tilde{g}} + G_g + T_t + R_r + GT_{gt}$$

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## Some calculations

$$\mu_{mtr}^{\tilde{g}} - \mu_{wtr}^{\tilde{g}} = (G_m - G_w) + (GT_{mt} - GT_{wt})$$

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$$\mu_{gtr}^{\tilde{g}} - \mu_{gt'r}^{\tilde{g}} = (T_t - T_{t'}) + (GT_{gt} - GT_{gt'})$$

These quantities are independent of the factor Replicate

# Statistical translation of the biological questions

$$\mu_{mtr}^{\tilde{g}} - \mu_{wtr}^{\tilde{g}} = (G_m - G_w) + (GT_{mt} - GT_{wt})$$

## Genotype effect

- What genes are differentially expressed between the two genotypes ?

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$$\mu_{mtr}^{\tilde{g}} - \mu_{wtr}^{\tilde{g}} = (G_m - G_w) + (GT_{mt} - GT_{wt})$$

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- What genes are differentially expressed between the two genotypes ?
- what genes show a time-averaged difference in their expression between the two genotypes independently from the experiments?

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- What genes are differentially expressed between the two genotypes ?
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$$\Delta_{genotype}^{\tilde{g}} = \frac{1}{3} \sum_t (\mu_{mtr}^{\tilde{g}} - \mu_{wtr}^{\tilde{g}})$$

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$$H_0 = \{\Delta_{genotype}^{\tilde{g}} = 0\} \text{ vs } H_1 = \{\Delta_{genotype}^{\tilde{g}} \neq 0\}$$



$$\mu_{g\textcolor{red}{t}r}^{\tilde{g}} - \mu_{g\textcolor{red}{t}'r}^{\tilde{g}} = (T_{\textcolor{red}{t}} - T_{\textcolor{red}{t}'}) + (GT_{g\textcolor{red}{t}} - GT_{g\textcolor{red}{t}'})$$

## Time effect

- what genes are differentially expressed between two consecutive times ?

$$\mu_{gtr}^{\tilde{g}} - \mu_{gt'r}^{\tilde{g}} = (T_t - T_{t'}) + (GT_{gt} - GT_{gt'})$$

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- **For a given genotype, whatever the experiment**, what genes are differentially expressed between two consecutive times ?

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$$H_0 = \{\Delta_{g,t,t'}^{\tilde{g}} = 0\} \text{ vs } H_1 = \{\Delta_{g,t,t'}^{\tilde{g}} \neq 0\}$$

# Genotype x Time effect

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$$\Delta_{m,w,t,t'}^{\tilde{g}} = (\mu_{\textcolor{red}{m}\textcolor{blue}{t}r}^{\tilde{g}} - \mu_{\textcolor{red}{w}\textcolor{blue}{t}r}^{\tilde{g}}) - (\mu_{\textcolor{red}{m}\textcolor{blue}{t'}r}^{\tilde{g}} - \mu_{\textcolor{red}{w}\textcolor{blue}{t'}r}^{\tilde{g}})$$

# Genotype x Time effect

- what genes are impacted in their transcription by an interaction between the genotype and the time ?
- what genes show a **genotype** difference between **two time points** ?

$$\Delta_{m,w,t,t'}^{\tilde{g}} = (\mu_{m\textcolor{red}{t}r}^{\tilde{g}} - \mu_{w\textcolor{blue}{t}r}^{\tilde{g}}) - (\mu_{m\textcolor{red}{t}'r}^{\tilde{g}} - \mu_{w\textcolor{blue}{t}'r}^{\tilde{g}})$$

$$\mu_{m\textcolor{red}{t}r}^{\tilde{g}} - \mu_{w\textcolor{blue}{t}r}^{\tilde{g}} = (G_m - G_w) + (GT_{m\textcolor{blue}{t}} - GT_{w\textcolor{blue}{t}})$$

## Genotype x Time interaction



# Genotype x Time effect

- what genes are impacted in their transcription by an interaction between the genotype and the time ?
- what genes show a **genotype** difference between **two time points** ?

$$\Delta_{m,w,t,t'}^{\tilde{g}} = (\mu_{\textcolor{red}{m}\textcolor{blue}{t}r}^{\tilde{g}} - \mu_{\textcolor{red}{w}\textcolor{blue}{t}r}^{\tilde{g}}) - (\mu_{\textcolor{red}{m}\textcolor{blue}{t}'r}^{\tilde{g}} - \mu_{\textcolor{red}{w}\textcolor{blue}{t}'r}^{\tilde{g}})$$

$$\mu_{\textcolor{red}{m}\textcolor{blue}{t}r}^{\tilde{g}} - \mu_{\textcolor{red}{w}\textcolor{blue}{t}r}^{\tilde{g}} = (G_{\textcolor{red}{m}} - G_{\textcolor{red}{w}}) + (GT_{\textcolor{red}{m}\textcolor{blue}{t}} - GT_{\textcolor{red}{w}\textcolor{blue}{t}})$$

## Genotype x Time interaction

$$\Delta_{m,w,t,t'}^{\tilde{g}} = (GT_{\textcolor{red}{m}\textcolor{blue}{t}} - GT_{\textcolor{red}{w}\textcolor{blue}{t}}) - (GT_{\textcolor{red}{m}\textcolor{blue}{t}'} - GT_{\textcolor{red}{w}\textcolor{blue}{t}'})$$

# Genotype x Time effect

- what genes are impacted in their transcription by an interaction between the genotype and the time ?
- what genes show a **genotype** difference between **two time points** ?

$$\Delta_{m,w,t,t'}^{\tilde{g}} = (\mu_{mtr}^{\tilde{g}} - \mu_{wtr}^{\tilde{g}}) - (\mu_{mt'r}^{\tilde{g}} - \mu_{wt'r}^{\tilde{g}})$$

$$\mu_{mtr}^{\tilde{g}} - \mu_{wtr}^{\tilde{g}} = (G_m - G_w) + (GT_{mt} - GT_{wt})$$

## Genotype x Time interaction

$$\Delta_{m,w,t,t'}^{\tilde{g}} = (GT_{mt} - GT_{wt}) - (GT_{mt'} - GT_{wt'})$$

$$H_0 = \{\Delta_{m,w,t,t'}^{\tilde{g}} = 0\} \text{ vs } H_1 = \{\Delta_{m,w,t,t'}^{\tilde{g}} \neq 0\}$$

Remark : when the two factors have only two modalities, it becomes to test the interaction coefficient of the model

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