

# Package ‘MutaliskR’

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**Type** Package

**Title** MutaliskR

**Description** MutaliskR is a general-purpose mutational signature identification tool that uses maximum likelihood estimation to perform linear regression.

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---

AssignIndelPCAWGClassification  
*Assigns PCAWG classification of an indel*

---

## Description

This function returns PCAWG classification data of an indel.

## Usage

```
AssignIndelPCAWGClassification(x)
```

## Arguments

x                      One row of data.frame the function PreprocessIndel.

## Value

A data.frame with the following columns:

PCAWG\_Indel\_Class  
                     PCAWG indel mutation type (e.g. 'DEL\_C\_1\_1').

Microhomology\_Class  
                     PCAWG microhomology mutation type (e.g. 'DEL\_MH\_2\_1').

Microhomology\_Size  
                     Microhomology size.

Microhomology\_Seq  
                     Microhomology sequence.

Microhomology\_Direction  
                     Microhomology direction.

---

EstimateMaximumLikelihood

*Performs maximum likelihood estimation*


---

## Description

This function performs maximum likelihood estimation of a given identification setup.

## Usage

```
EstimateMaximumLikelihood(
  mutation.frequencies,
  signature.weights,
  num.mutations,
  min.probability = 0.01
)
```

## Arguments

`mutation.frequencies` A numeric vector corresponding to the frequency of each mutation type

`signature.weights` A matrix of signature weights.

`num.mutations` An integer value that specifies the number of mutations

`min.probability` A float value that specifies the minimum probability to attribute to a signature.

## Value

A list with the following items:

`Cosine_Similarity` Cosine similarity score between `mutation.frequencies` and `Reconstructed_Spectrum`

`Par` A numeric vector of contribution weights of identified signatures.

`Par_Normalized` A normalized (0 to 1) numeric vector of contribution weights of identified signatures.

`Reconstructed_Spectrum` A numeric vector of reconstructed spectrum.

`Reconstructed_Spectrum_Normalized` A normalized (0 to 1) numeric vector of reconstructed spectrum.

`Residuals` A numeric vector of residual spectrum.

`Residuals_Normalized` A normalized (0 to 1) numeric vector of residual spectrum.

`RSS` Residual sum of squares (of `Residuals`).

`RSS_Normalized` Residual sum of squares (of `Residuals_Normalized`).

---

GetComplementBase	Returns complement base
-------------------	-------------------------

---

**Description**

This function returns the complement base of a nucleotide

**Usage**

```
GetComplementBase(nucleotide)
```

**Arguments**

nucleotide	Nucleotide base.
------------	------------------

**Value**

"A", "C", "G", or "T"

---

GetComplementBases	Returns complement bases
--------------------	--------------------------

---

**Description**

This function returns the complement bases of a nucleotide sequence

**Usage**

```
GetComplementBases(nucleotides)
```

**Arguments**

nucleotides	Nucleotide bases (e.g. "AG").
-------------	-------------------------------

**Value**

e.g. "TC"

---

`GetCosmicDbsSignaturesData`*Returns the COSMIC v3.1 DBS reference signature data*

---

**Description**

This function returns the COSMIC v3.1 doublet base substitution (DBS) mutational signatures data.

**Usage**`GetCosmicDbsSignaturesData()`**Value**

A data.frame of the PCAWG DBS mutational signatures.

---

`GetCosmicDbsSignaturesNames`*Return the COSMIC v3.1 DBS reference signature names*

---

**Description**

This function returns the COSMIC v3.1 DBS mutational signatures names.

**Usage**`GetCosmicDbsSignaturesNames()`**Value**

Returns a character vector of all COSMIC v3.1 DBS mutational signature names.

---

`GetCosmicIndelSignaturesData`*Returns the COSMIC v3.1 indel reference signature data*

---

**Description**

This function returns the COSMIC v3.1 small insertion and deletion (indel) mutational signatures data.

**Usage**`GetCosmicIndelSignaturesData()`**Value**

A data.frame of the COSMIC v3.1 indel mutational signatures.

---

`GetCosmicIndelSignaturesNames`*Returns the COSMIC v3.1 indel signature names*

---

**Description**

This function returns the COSMIC v3.1 indel mutational signatures names.

**Usage**

```
GetCosmicIndelSignaturesNames()
```

**Value**

Returns a character vector of all COSMIC v3.1 indel mutational signature names.

---

`GetCosmicSbsSignaturesData`*Returns the COSMIC v3.1 SBS reference signature data*

---

**Description**

This function returns the COSMIC v3.1 single base substitution (SBS) mutational signatures data.

**Usage**

```
GetCosmicSbsSignaturesData()
```

**Value**

A data.frame of the COSMIC v3.1 SBS mutational signatures.

---

`GetCosmicSbsSignaturesNames`*Return the COSMIC v3.1 SBS reference signature names*

---

**Description**

This function returns the COSMIC v3.1 SBS mutational signatures names.

**Usage**

```
GetCosmicSbsSignaturesNames()
```

**Value**

Returns a character vector of all COSMIC v3.1 SBS mutational signature names.

---

`GetDbsSignaturesColors`*Returns the DBS signatures colors and etiologies*

---

**Description**

This function returns the DBS mutational signatures colors and etiologies.

**Usage**

```
GetDbsSignaturesColors()
```

**Value**

A data.frame of the DBS mutational signatures colors and etiologies.

---

`GetDbsSignaturesPlotTheme`*Fetches DBS plotting theme*

---

**Description**

This function returns the DBS plotting theme elements.

**Usage**

```
GetDbsSignaturesPlotTheme(  
  x.axis.labels = PLOT.DBS.X.AXIS.LABELS,  
  x.axis.text.size = 10,  
  x.axis.text.angle = 90,  
  x.axis.text.hjust = 1,  
  x.axis.text.vjust = 0.5,  
  y.axis.text.size = 12,  
  y.axis.title.size = 12,  
  y.axis.min = 0,  
  y.axis.max = 1,  
  y.axis.title = "Mutation Fraction",  
  groups = DBS.MUTATION.TYPES.GROUPS,  
  strip.labels = PLOT.DBS.STRIP.LABELS,  
  strip.colors = PLOT.DBS.STRIP.COLORS,  
  strip.text.colors = PLOT.DBS.STRIP.TEXT.COLORS,  
  strip.text.size = 12,  
  legend.title = "Mutation Type",  
  legend.title.size = 12,  
  legend.text.size = 10,  
  legend.labels = PLOT.DBS.LEGEND.LABELS  
)
```



**Arguments**

<code>x.axis.labels</code>	A character vector used to label the x-axis.
<code>x.axis.text.size</code>	x-axis text size.
<code>x.axis.text.angle</code>	x-axis text angle.
<code>y.axis.text.size</code>	y-axis text size.
<code>y.axis.title.size</code>	y-axis title size.
<code>y.axis.min</code>	y-axis lower-bound (minimum range for limits).
<code>y.axis.max</code>	y-axis upper-bound (maximum range for limits).
<code>y.axis.title</code>	y-axis title text.
<code>groups</code>	A character vector of group variable values.
<code>strip.labels</code>	A character vector used to label the strip.
<code>strip.colors</code>	A character vector of hex values used to color the strip.
<code>strip.text.colors</code>	A character vector of hex values used to color the strip text.
<code>strip.text.size</code>	Strip (facet) text size.
<code>legend.title</code>	Legend title text.
<code>legend.title.size</code>	Legend title text size.
<code>legend.labels</code>	A character vector used to label the legend items.

**Value**

A data.frame of the DBS signature plot theme elements.

---

GetDeletionMicrohomologySize

*Returns microhomology size of a deletion*

---

**Description**

This function computes and returns the microhomology size of a small deletion.

**Usage**

```
GetDeletionMicrohomologySize(upstream.seq, downstream.seq, mutated.seq)
```

**Arguments**

<code>upstream.seq</code>	Upstream nucleotide sequence.
<code>downstream.seq</code>	Downstream nucleotide sequence.
<code>mutated.seq</code>	Indel nucleotide sequence

**Value**

A data.frame with the following columns:

Microhomology_Seq	Microhomology nucleotide sequence.
Microhomology_Size	Size (number of bases) of microhomology.
Microhomology_Direction	Direction of microhomology.

---

GetHsapiensEnsemblData

*Returns the hsapiens ensembl data*

---

**Description**

This function returns the hsapiens ensembl data.

**Usage**

```
GetHsapiensEnsemblData(version)
```

**Arguments**

version            Either "GRCh37" or "GRCh38".

---

GetIndelPCAWGClassification

*Returns PCAWG classification of an indel*

---

**Description**

This function classifies an indel according to the PCAWG classification.

**Usage**

```
GetIndelPCAWGClassification(type, mutated.seq, upstream.seq, downstream.seq)
```

**Arguments**

type            Indel type. Either 'INS' or 'DEL'.  
 mutated.seq    Indel nucleotide sequence.  
 upstream.seq   Upstream nucleotide sequence.  
 downstream.seq Downstream nucleotide sequence.

**Value**

A data.frame with the following columns:

PCAWG_Indel_Class	PCAWG indel mutation type (e.g. 'DEL_C_1_0').
Microhomology_Class	PCAWG microhomology mutation type (e.g. 'DEL_MH_2_1').
Microhomology_Size	Microhomology size.
Microhomology_Seq	Microhomology sequence.
Microhomology_Direction	Microhomology direction.

---

GetIndelRepeatSize	<i>Returns indel repeat size</i>
--------------------	----------------------------------

---

**Description**

This function computes and returns the repeat size of an indel variant.

**Usage**

```
GetIndelRepeatSize(downstream.seq, mutated.seq, type)
```

**Arguments**

downstream.seq	Downstream nucleotide sequence.
mutated.seq	Indel nucleotide sequence.
type	Indel type. Either 'INS' or 'DEL'.

**Value**

Repeat size (integer).

---

GetIndelSignaturesColors	<i>Returns the indel signatures colors and etiologies</i>
--------------------------	---

---

**Description**

This function returns the indel mutational signatures colors and etiologies.

**Usage**

```
GetIndelSignaturesColors()
```

**Value**

A data.frame of the indel mutational signatures colors and etiologies.

---

GetIndelSignaturesPlotTheme

*Fetches INDEL plotting theme*


---

## Description

This function returns the INDEL plotting theme elements.

## Usage

```
GetIndelSignaturesPlotTheme(
  x.axis.labels = PLOT.INDEL.X.AXIS.LABELS,
  x.axis.text.size = 10,
  x.axis.text.angle = 0,
  y.axis.text.size = 12,
  x.axis.text.hjust = 0.5,
  x.axis.text.vjust = 0.5,
  y.axis.title.size = 12,
  y.axis.min = 0,
  y.axis.max = 1,
  y.axis.title = "Mutation Fraction",
  groups = INDEL.MUTATION.TYPES.GROUPS,
  strip.labels = PLOT.INDEL.STRIP.LABELS,
  strip.colors = PLOT.INDEL.STRIP.COLORS,
  strip.text.colors = PLOT.INDEL.STRIP.TEXT.COLORS,
  strip.text.size = 12,
  legend.title = "Mutation Type",
  legend.title.size = 12,
  legend.text.size = 10,
  legend.labels = PLOT.INDEL.LEGEND.LABELS
)
```

## Arguments

x.axis.labels	A character vector used to label the x-axis.
x.axis.text.size	x-axis text size.
x.axis.text.angle	x-axis text angle.
y.axis.text.size	y-axis text size.
y.axis.title.size	y-axis title size.
y.axis.min	y-axis lower-bound (minimum range for limits).
y.axis.max	y-axis upper-bound (maximum range for limits).
y.axis.title	y-axis title text.
groups	A character vector of group variable values.
strip.labels	A character vector used to label the strip.
strip.colors	A character vector of hex values used to color the strip.

strip.text.colors      A character vector of hex values used to color the strip text.

strip.text.size      Strip (facet) text size.

legend.title      Legend title text.

legend.title.size      Legend title text size.

legend.labels      A character vector used to label the legend items.

**Value**

A data.frame of the INDEL signature plot theme elements.

---

GetKucabSbsSignaturesData

*Returns the Kucab SBS reference signature data*

---

**Description**

This function returns the Kucab single base substitution (SBS) mutational signatures data.

**Usage**

GetKucabSbsSignaturesData()

**Value**

A data.frame of the Kucab SBS mutational signatures.

---

GetKucabSbsSignaturesNames

*Return the Kucab SBS reference signature names*

---

**Description**

This function returns the Kucab SBS mutational signatures names.

**Usage**

GetKucabSbsSignaturesNames()

---

`GetMmusculusEnsemblData`*Returns the mmusculus ensembl data*

---

**Description**

This function returns the mmusculus ensembl data.

**Usage**

```
GetMmusculusEnsemblData(version)
```

**Arguments**

version            Either "NCBIM37" or "GRCm38".

---

`GetPcawgDbsSignaturesData`*Returns the PCAWG DBS reference signature data*

---

**Description**

This function returns the PCAWG doublet base substitution (DBS) mutational signatures data.

**Usage**

```
GetPcawgDbsSignaturesData()
```

**Value**

A data.frame of the PCAWG DBS mutational signatures.

---

`GetPcawgDbsSignaturesNames`*Return the DBS reference signature names*

---

**Description**

This function returns the PCAWG DBS mutational signatures names.

**Usage**

```
GetPcawgDbsSignaturesNames()
```

**Value**

Returns a character vector of all PCAWG DBS mutational signature names.

---

`GetPcawgIndelSignaturesData`*Returns the PCAWG indel reference signature data*

---

**Description**

This function returns the PCAWG small insertion and deletion (indel) mutational signatures data.

**Usage**

```
GetPcawgIndelSignaturesData(version)
```

**Arguments**

`version`                Either "SigProfiler" or "SignatureAnalyzer".

**Value**

A data.frame of the PCAWG indel mutational signatures.

---

`GetPcawgIndelSignaturesNames`*Returns the PCAWG indel signature names*

---

**Description**

This function returns the PCAWG indel mutational signatures names.

**Usage**

```
GetPcawgIndelSignaturesNames(version)
```

**Arguments**

`version`                Either "SigProfiler" or "SignatureAnalyzer".

**Value**

Returns a character vector of all PCAWG indel mutational signature names.

---

`GetPcawgPlatinumSbsSignaturesData`*Returns the PCAWG Platinum SBS reference signature data*

---

**Description**

This function returns the PCAWG Platinum single base substitution (SBS) mutational signatures data (from Petljak et al., Cell 2019).

**Usage**

```
GetPcawgPlatinumSbsSignaturesData()
```

**Value**

A data.frame of the PCAWG Platinum SBS mutational signatures.

---

`GetPcawgPlatinumSbsSignaturesNames`*Return the PCAWG Platinum SBS reference signature names*

---

**Description**

This function returns the PCAWG Platinum SBS mutational signatures names.

**Usage**

```
GetPcawgPlatinumSbsSignaturesNames()
```

**Value**

Returns a character vector of all PCAWG Platinum SBS mutational signature names.

---

`GetPcawgSbsPentanucleotideSignaturesData`*Returns the PCAWG SBS penta-nucleotide reference signature data*

---

**Description**

This function returns the PCAWG single base substitution (SBS) pentanucleotide context mutational signatures data.

**Usage**

```
GetPcawgSbsPentanucleotideSignaturesData()
```

**Value**

A data.frame of the PCAWG SBS penta-nucleotide mutational signatures.



---

`GetPcawgSbsPentanucleotideSignaturesNames`*Return the PCAWG SBS penta-nucleotide reference signature names*

---

**Description**

This function returns the PCAWG SBS penta-nucleotide mutational signatures names.

**Usage**

```
GetPcawgSbsPentanucleotideSignaturesNames()
```

**Value**

Returns a character vector of all PCAWG SBS penta-nucleotide mutational signature names.

---

`GetPcawgSbsSignaturesData`*Returns the PCAWG SBS reference signature data*

---

**Description**

This function returns the PCAWG single base substitution (SBS) mutational signatures data.

**Usage**

```
GetPcawgSbsSignaturesData(sequencing.type)
```

**Arguments**

`sequencing.type`

Either 'WES' for whole-exome sequencing or 'WGS' for whole-genome sequencing.

**Value**

A data.frame of the PCAWG SBS mutational signatures.

---

`GetPcawgSbsSignaturesNames`*Return the PCAWG SBS reference signature names*

---

**Description**

This function returns the PCAWG SBS mutational signatures names.

**Usage**

```
GetPcawgSbsSignaturesNames(sequencing.type)
```

**Arguments**

`sequencing.type`

Either 'WES' for whole-exome sequencing or 'WGS' for whole-genome sequencing.

**Value**

Returns a character vector of all PCAWG SBS mutational signature names.

---

`GetPetljakSbsSignaturesData`*Returns the Petljak SBS reference signature data*

---

**Description**

This function returns the Petljak single base substitution (SBS) mutational signatures data.

**Usage**

```
GetPetljakSbsSignaturesData()
```

**Value**

A data.frame of the Petljak SBS mutational signatures.

---

`GetPetljakSbsSignaturesNames`*Return the Petljak SBS reference signature names*

---

**Description**

This function returns the Petljak SBS mutational signatures names.

**Usage**`GetPetljakSbsSignaturesNames(sequencing.type)`**Value**

Returns a character vector of all Petljak SBS mutational signature names.

---

`GetPleguezuelosManzanoIndelSignaturesData`*Returns the Pleguezuelos-Manzano indel reference signature data*

---

**Description**

This function returns the Pleguezuelos-Manzano small insertion and deletion (indel) mutational signatures data.

**Usage**`GetPleguezuelosManzanoIndelSignaturesData()`**Value**

A data.frame of the Pleguezuelos-Manzano indel mutational signatures.

---

`GetPleguezuelosManzanoIndelSignaturesNames`*Returns the Pleguezuelos-Manzano indel signature names*

---

**Description**

This function returns the Pleguezuelos-Manzano indel mutational signatures names.

**Usage**`GetPleguezuelosManzanoIndelSignaturesNames()`**Value**

Returns a character vector of all Pleguezuelos-Manzano indel mutational signature names.

---

`GetPleguezuelosManzanoSbsSignaturesData`*Returns the Pleguezuelos-Manzano SBS reference signature data*

---

**Description**

This function returns the Pleguezuelos-Manzano single base substitution (SBS) mutational signatures data.

**Usage**`GetPleguezuelosManzanoSbsSignaturesData()`**Value**

A data.frame of the Petljak Pleguezuelos-Manzano mutational signatures.

---

`GetPleguezuelosManzanoSbsSignaturesNames`*Return the Pleguezuelos-Manzano SBS reference signature names*

---

**Description**

This function returns the Pleguezuelos-Manzano SBS mutational signatures names.

**Usage**`GetPleguezuelosManzanoSbsSignaturesNames()`

---

`GetSbsSignaturesColors`*Returns the SBS signatures colors and etiologies*

---

**Description**

This function returns the SBS mutational signatures colors and etiologies.

**Usage**`GetSbsSignaturesColors()`**Value**

A data.frame of the SBS mutational signatures colors and etiologies.

---

```
GetSbsSignaturesPlotTheme
    Fetches SBS plotting theme
```

---

## Description

This function returns the SBS plotting theme elements.

## Usage

```
GetSbsSignaturesPlotTheme(
  x.axis.labels = PLOT.SBS.X.AXIS.LABELS,
  x.axis.text.size = 10,
  x.axis.text.angle = 90,
  x.axis.text.hjust = 0.5,
  x.axis.text.vjust = 0.5,
  y.axis.text.size = 12,
  y.axis.title.size = 12,
  y.axis.min = 0,
  y.axis.max = 1,
  y.axis.title = "Mutation Fraction",
  groups = SBS.MUTATION.TYPES.GROUPS,
  strip.labels = PLOT.SBS.STRIP.LABELS,
  strip.colors = PLOT.SBS.STRIP.COLORS,
  strip.text.colors = PLOT.SBS.STRIP.TEXT.COLORS,
  strip.text.size = 12,
  legend.title = "Mutation Type",
  legend.title.size = 12,
  legend.text.size = 10,
  legend.labels = PLOT.SBS.LEGEND.LABELS
)
```

## Arguments

<code>x.axis.labels</code>	A character vector used to label the x-axis.
<code>x.axis.text.size</code>	x-axis text size.
<code>x.axis.text.angle</code>	x-axis text angle.
<code>y.axis.text.size</code>	y-axis text size.
<code>y.axis.title.size</code>	y-axis title size.
<code>y.axis.min</code>	y-axis lower-bound (minimum range for limits).
<code>y.axis.max</code>	y-axis upper-bound (maximum range for limits).
<code>y.axis.title</code>	y-axis title text.
<code>groups</code>	A character vector of group variable values.
<code>strip.labels</code>	A character vector used to label the strip.
<code>strip.colors</code>	A character vector of hex values used to color the strip.

strip.text.colors      A character vector of hex values used to color the strip text.

strip.text.size      Strip (facet) text size.

legend.title      Legend title text.

legend.title.size      Legend title text size.

legend.labels      A character vector used to label the legend items.

### Value

A data.frame of the SBS signature plot theme elements.

---

IdentifyDbsSignatures    *Identify doublet base substitution mutational signatures*

---

### Description

This function identifies DBS mutational signatures.

### Usage

```
IdentifyDbsSignatures(
  input,
  bsg,
  sample.id = "Sample",
  reference = GetPcawgDbsSignaturesData(),
  target.signatures = GetPcawgDbsSignaturesNames(),
  plot.theme = GetDbsSignaturesPlotTheme(),
  analyze.variants.column.gene,
  analyze.variants.column.group,
  analyze.variants = FALSE,
  n.cores = 2,
  combn.m = 3,
  n.max.signatures = 7,
  min.probability = 0.01,
  zeta.value = 1e-10,
  save = TRUE,
  save.dir = NULL
)
```

### Arguments

input      Either VCF file path or data.frame. If the input is a data.frame, it must include the following columns: Chr, Pos, Ref, Alt.

bsg      BSgenome object.

sample.id      Sample ID that will be used to name output files (default: 'Sample').

reference      A data.frame with the following columns: Mutation\_Type, and names of DBS signatures (default: a data.frame returned from [GetPcawgDbsSignaturesData](#)).

<code>target.signatures</code>	Signatures to be considered for identification (default: an array returned from <a href="#">GetPcawgDbSignaturesNames</a> ).
<code>plot.theme</code>	A data.frame returned from <a href="#">GetDbSignaturesPlotTheme</a> .
<code>analyze.variants.column.gene</code>	Name of column in the data.frame corresponding to the gene name or ID (e.g. "Gene.refGene" if using ANNOVAR data).
<code>analyze.variants.column.group</code>	Name of column in the data.frame corresponding to the variant group for plotting purposes (e.g. "Func.refGene" if using ANNOVAR data).
<code>n.cores</code>	Number of cores to use.
<code>combn.m</code>	Number of signatures to consider in each step. 'm' parameter in <code>combn</code> function (default: 3).
<code>n.max.signatures</code>	Maximum number of signatures to identify. Recommended: <code>n.max.signatures</code> $\geq$ <code>initial.exploration.combn.m</code> (default: 7).
<code>min.probability</code>	Minimum probability to attribute to a signature (default: 0.01).
<code>zeta.value</code>	A float value that is added to the data frequency (default: 1e-10).
<code>save</code>	Save resulting files if TRUE, otherwise do not save (default: TRUE).
<code>save.dir</code>	Save directory path (default: NULL).

**Value**

A list with the following elements:

`results`: a data.frame with the following columns:

<code>Mutations_Count</code>	Number of mutations.
<code>Signatures</code>	Identified mutational signatures separated by comma.
<code>Signatures_Count</code>	Number of identified mutational signatures.
<code>Signatures_Weights</code>	Normalized (0 to 1) weights of identified mutational signatures separated by comma.
<code>Mutation_Types</code>	Mutation types separated by comma.
<code>Mutation_Types_Groups</code>	Mutation type groups separated by comma.
<code>Observed_Spectrum</code>	Normalized spectrum (frequency) of observed mutations separated by comma.
<code>Reconstructed_Spectrum</code>	Normalized spectrum (frequency) of MLE reconstructed mutations separated by comma.
<code>Residual_Spectrum</code>	Normalized spectrum (frequency) of residual mutations separated by comma.
<code>Cosine_Similarity</code>	Cosine similarity score between <code>Observed_Spectrum</code> and <code>Reconstructed_Spectrum</code> .
<code>RSS</code>	Raw residual sum of squares (derived from <code>Residual_Spectrum</code> ).

RSS\_Normalized Normalized residual sum of squares (derived from Residual\_Spectrum).

BIC Bayesian information criterion of the identified model.

results.variants: a data.frame with the following columns:

---

IdentifyIndelSignatures

*Identify indel mutational signatures*

---

## Description

This function identifies indel mutational signatures.

## Usage

```
IdentifyIndelSignatures(
  input,
  bsg,
  sample.id = "Sample",
  reference = GetPcawgIndelSignaturesData(version = "SigProfiler"),
  target.signatures = GetPcawgIndelSignaturesNames(version = "SigProfiler"),
  plot.theme = GetIndelSignaturesPlotTheme(),
  analyze.variants.column.gene,
  analyze.variants.column.group,
  analyze.variants = FALSE,
  n.cores = 2,
  max.len = 25,
  padding.len = 80,
  combn.m = 3,
  n.max.signatures = 7,
  min.probability = 0.01,
  zeta.value = 1e-10,
  save = TRUE,
  save.dir = NULL
)
```

## Arguments

input	Either VCF file path or data.frame. If the input is a data.frame, it must include the following columns: Chr, Pos, Ref, Alt.
bsg	BSgenome object.
sample.id	Sample ID that will be used to name output files (default: 'Sample').
reference	A data.frame with the following columns: Mutation_Type, and names of indel signatures (default: a data.frame returned from <a href="#">GetPcawgIndelSignaturesData(version = "SigProfiler")</a> ).
target.signatures	Signatures to be considered for identification (default: an array returned from <a href="#">GetPcawgIndelSignaturesNames(version = "SigProfiler")</a> ).
plot.theme	A data.frame returned from <a href="#">GetIndelSignaturesPlotTheme</a> .



<code>analyze.variants.column.gene</code>	Name of column in the data.frame corresponding to the gene name or ID (e.g. "Gene.refGene" if using ANNOVAR data).
<code>analyze.variants.column.group</code>	Name of column in the data.frame corresponding to the variant group for plotting purposes (e.g. "Func.refGene" if using ANNOVAR data).
<code>analyze.variants</code>	A boolean value that indicates whether variant-level signature analysis should be performed (default: FALSE).
<code>n.cores</code>	Number of cores to use.
<code>max.len</code>	Maximum number of bases allowed for a small insertion and deletion (indels bigger than this will be excluded; default: 25).
<code>padding.len</code>	Number of bases to use for upstream and downstream sequences (default: 80).
<code>combn.m</code>	Number of signatures to consider in each step. 'm' parameter in <code>combn</code> function (default: 3).
<code>n.max.signatures</code>	Maximum number of signatures to identify. Recommended: <code>n.max.signatures</code> $\geq$ <code>initial.exploration.combn.m</code> (default: 7).
<code>min.probability</code>	Minimum probability to attribute to a signature (default: 0.01).
<code>zeta.value</code>	A float value that is added to the data frequency (default: 1e-10).
<code>save</code>	Save resulting files if TRUE, otherwise do not save (default: TRUE).
<code>save.dir</code>	Save directory path (default: NULL).

**Value**

A list with the following elements:

`results`: a data.frame with the following columns:

<code>Mutations_Count</code>	Number of mutations.
<code>Signatures</code>	Identified mutational signatures separated by comma.
<code>Signatures_Count</code>	Number of identified mutational signatures.
<code>Signatures_Weights</code>	Normalized (0 to 1) weights of identified mutational signatures separated by comma.
<code>Mutation_Types</code>	Mutation types separated by comma.
<code>Mutation_Types_Groups</code>	Mutation type groups separated by comma.
<code>Observed_Spectrum</code>	Normalized spectrum (frequency) of observed mutations separated by comma.
<code>Reconstructed_Spectrum</code>	Normalized spectrum (frequency) of MLE reconstructed mutations separated by comma.
<code>Residual_Spectrum</code>	Normalized spectrum (frequency) of residual mutations separated by comma.
<code>Cosine_Similarity</code>	Cosine similarity score between <code>Observed_Spectrum</code> and <code>Reconstructed_Spectrum</code> .

RSS                      Raw residual sum of squares (derived from Residual\_Spectrum).  
 RSS\_Normalized      Normalized residual sum of squares (derived from Residual\_Spectrum).  
 BIC                      Bayesian information criterion of the identified model.

results.variants: a data.frame with the following columns:

---

IdentifySbsSignatures    *Identify single-base substitution mutational signatures*

---

## Description

This function identifies SBS mutational signatures.

## Usage

```
IdentifySbsSignatures(
  input,
  bsg,
  sample.id = "Sample",
  reference = GetPcawgSbsSignaturesData(sequencing.type = "WGS"),
  target.signatures = GetPcawgSbsSignaturesNames(sequencing.type = "WGS"),
  plot.theme = GetSbsSignaturesPlotTheme(),
  analyze.variants.column.gene,
  analyze.variants.column.group,
  analyze.variants = FALSE,
  context.length = 3,
  n.cores = 2,
  combn.m = 3,
  n.max.signatures = 7,
  min.probability = 0.01,
  zeta.value = 1e-10,
  save = TRUE,
  save.dir = NULL
)
```

## Arguments

input	Either VCF file path or data.frame. If the input is a data.frame, it must include the following columns: Chr, Pos, Ref, Alt.
bsg	BSgenome object.
sample.id	Sample ID that will be used to name output files (default: 'Sample').
reference	A data.frame with the following columns: Mutation_Type, and names of SBS signatures (default: a data.frame returned from <code>GetPcawgSbsSignaturesData(sequencing.type = "WGS")</code> ).
target.signatures	Signatures to be considered for identification (default: an array returned from <code>.{GetPcawgSbsSignaturesNames(sequencing.type = "WGS")}</code> ).

**Value**

A list with the following elements:

results: a data.frame with the following columns:

Mutations_Count	Number of mutations.
Signatures	Identified mutational signatures separated by comma.
Signatures_Count	Number of identified mutational signatures.
Signatures_Weights	Normalized (0 to 1) weights of identified mutational signatures separated by comma.
Mutation_Types	Mutation types separated by comma.
Mutation_Types_Groups	Mutation type groups separated by comma.
Observed_Spectrum	Normalized spectrum (frequency) of observed mutations separated by comma.
Reconstructed_Spectrum	Normalized spectrum (frequency) of MLE reconstructed mutations separated by comma.
Residual_Spectrum	Normalized spectrum (frequency) of residual mutations separated by comma.
Cosine_Similarity	Cosine similarity score between Observed_Spectrum and Reconstructed_Spectrum.
RSS	Raw residual sum of squares (derived from Residual_Spectrum).
RSS_Normalized	Normalized residual sum of squares (derived from Residual_Spectrum).
BIC	Bayesian information criterion of the identified model.

results.variants: a data.frame with the following columns:

---

IdentifySignatures	<i>Identify contribution weights of underlying mutational signatures</i>
--------------------	--

---

**Description**

This function identifies which mutational signatures are underlying a given data.frame and contribution weights of each mutational signature.

**Usage**

```
IdentifySignatures(
  data,
  reference,
  target.signatures,
  n.cores = 2,
  combn.m = 3,
  n.max.signatures = 7,
  min.probability = 0.01,
  zeta.value = 1e-10
)
```

**Arguments**

<code>data</code>	A data.frame prepared by either <code>PrepareSbsVcfFile</code> , <code>PrepareDbsVcfFile</code> , <code>PrepareIdVcfFile</code> , or <code>PrepareSvVcfFile</code> depending on <code>signature.type</code> .
<code>reference</code>	A data.frame where the first column is the list of mutation types and where 2nd to k-th column headers are the names of the signatures.
<code>target.signatures</code>	A character vector that lists all desired signatures to consider for identification.
<code>n.cores</code>	Number of cores to use (default: 2).
<code>combn.m</code>	An integer value that specifies the number of signatures to consider in each step. 'm' parameter in <code>combn</code> function.
<code>n.max.signatures</code>	An integer value that specifies the maximum number of signatures to consider. It is recommended that <code>n.max.signatures &gt;= initial.exploration.combn.m</code> .
<code>min.probability</code>	A float value that specifies the minimum probability to attribute to a signature.
<code>zeta.value</code>	A float value that is added to the data frequency.

**Value**

A data.frame with the following columns:

<code>Mutations_Count</code>	Number of mutations.
<code>Signatures</code>	Identified mutational signatures separated by comma.
<code>Signatures_Count</code>	Number of identified mutational signatures.
<code>Signatures_Weights</code>	Normalized (0 to 1) weights of identified mutational signatures separated by comma.
<code>Mutation_Types</code>	Mutation types separated by comma.
<code>Mutation_Types_Groups</code>	Mutation type groups separated by comma.
<code>Observed_Spectrum</code>	Normalized spectrum (frequency) of observed mutations separated by comma.
<code>Reconstructed_Spectrum</code>	Normalized spectrum (frequency) of MLE reconstructed mutations separated by comma.
<code>Residual_Spectrum</code>	Normalized spectrum (frequency) of residual mutations separated by comma.
<code>Cosine_Similarity</code>	Cosine similarity score between <code>Observed_Spectrum</code> and <code>Reconstructed_Spectrum</code> .
<code>RSS</code>	Raw residual sum of squares (derived from <code>Residual_Spectrum</code> ).
<code>RSS_Normalized</code>	Normalized residual sum of squares (derived from <code>Residual_Spectrum</code> ).
<code>BIC</code>	Bayesian information criterion of the identified model.

---

**IdentifyVariantSignatures***Identifies mutational signatures associated with each variant*

---

**Description**

This function identifies mutational signatures associated with each variant.

**Usage**

```
IdentifyVariantSignatures(prepared.data, identified.model, reference)
```

**Arguments**

prepared.data	A data.frame returned from <a href="#">PrepareSbsDataFrame</a> .
identified.model	A data.frame returned from <a href="#">IdentifySignatures</a> .
reference	A data.frame with the following columns: Mutation_Type, and names of SBS signatures.

**Value**

A data.frame with the following columns:

Chr	Chromosome.
Pos	Genomic position.
Ref	Reference allele.
Alt	Alternate allele.
Mutation_Type	Mutation type.
Variant_Gene	Variant gene.
Variant_Group	Variant group.
Signature	Signature.
Probability	Probability the mutation is the result of the signature.

---

**LinearRegressionOptimFn***Computes and returns the residual sum of squares*

---

**Description**

This function estimates the residual sum of squares.

**Usage**

```
LinearRegressionOptimFn(par, mutation.frequencies, signature.weights)
```

**Arguments**

<code>par</code>	A numeric vector of parameters to optimize.
<code>mutation.frequencies</code>	A numeric vector corresponding to the frequency of each mutation type.
<code>signature.weights</code>	A matrix of signature weights.

**Value**

Residual sum of squares.

---

PlotIdentifiedModel	<i>Plots identified model</i>
---------------------	-------------------------------

---

**Description**

This function plots an identified model.

**Usage**

```
PlotIdentifiedModel(identified.model, reference, plot.theme)
```

**Arguments**

<code>identified.model</code>	A row of the data.frame returned from the function <code>IdentifySignatures</code> .
<code>reference</code>	A data.frame with the following columns: <code>Mutation_Type</code> , and names of signatures.
<code>plot.theme</code>	A data.frame returned from <a href="#">GetSbsSignaturesPlotTheme</a> , <a href="#">GetDbsSignaturesPlotTheme</a> or <a href="#">GetIndelSignaturesPlotTheme</a> .

**Value**

A list with the following items:

<code>plot.merged</code>	Merged ( <code>plot.observed</code> , <code>plot.reconstructed.spectrum</code> , <code>plot.residual.spectrum</code> , <code>plot.legend</code> ) plot.
<code>plot.observed</code>	Observed mutation frequencies plot.
<code>plot.reconstructed.spectrum</code>	Reconstructed mutation frequencies plot.
<code>plot.residual.spectrum</code>	Residual mutation frequencies plot.
<code>plot.legend</code>	Legend plot.

---

PlotIdentifiedModels    *Plots identified models*


---

## Description

This function plots identified models for a group of samples IDs.

## Usage

```
PlotIdentifiedModels(
  df.models,
  df.signatures.colors,
  signature.group.order = c(),
  sample.ids.order = c(),
  apply.facet = FALSE,
  df.sample.ids.groups = c(),
  sample.ids.groups.order = c(),
  multiply.by.mutations.count = FALSE,
  y.axis.max = NULL,
  plot.title = "",
  title.size = 12,
  x.axis.text.size = 8,
  y.axis.text.size = 10,
  y.axis.title.size = 12,
  legend.title.size = 12,
  legend.text.size = 10
)
```

## Arguments

<code>df.models</code>	A data.frame of all best models appended by rbind with <code>Sample_ID</code> added as a column.
<code>df.signatures.colors</code>	A data.frame returned from <a href="#">GetSbsSignaturesColors</a> , <a href="#">GetDbsSignaturesColors</a> , or <a href="#">GetIndelSignaturesColors</a> .
<code>signature.group.order</code>	A character vector specifying the order of the signature groups.
<code>x.axis.text.size</code>	A numeric value that sets the x.axis.text size.
<code>y.axis.text.size</code>	A numeric value that sets the y.axis.text size.
<code>y.axis.title.size</code>	A numeric value that sets the y.axis.title size.
<code>legend.title.size</code>	A numeric value that sets the legend title size.
<code>legend.text.size</code>	A numeric value that sets the legend text size.

## Value

A ggplot object.

---

**PlotIdentifiedModelSignatureWeights**

*Plots signature weights of an identified model*

---

**Description**

This function plots signature weights of an identified model.

**Usage**

```
PlotIdentifiedModelSignatureWeights(
  identified.model,
  df.signatures.colors,
  plot.theme
)
```

**Arguments**

`identified.model`

A row of the data.frame returned from the function `IdentifySignatures`.

`df.signature.colors`

A data.frame with the following columns: `Mutation_Type`, and names of signatures.

**Value**

ggplot object.

---

**PlotIdentifiedModelVariantSignatures**

*Plots a stacked barplot of signatures probabilities of variants*

---

**Description**

This function plots a stacked barplot of signature probabilities of variants.

**Usage**

```
PlotIdentifiedModelVariantSignatures(df.sigs.probs, df.signatures.colors)
```

**Value**

A list with the following items:

`plots.merged`      Merged rainfall plots.

`plots`              Rainfall plot for each chromosome.



---

PlotSpectrum	<i>Plots spectrum</i>
--------------	-----------------------

---

**Description**

This function plots spectrum of mutation frequencies.

**Usage**

```
PlotSpectrum(y, plot.theme)
```

**Arguments**

y	A numeric vector corresponding to the y-axis values.
plot.theme	Plot theme (returned from GetSbsSignaturesPlotTheme).

**Value**

A list with the following items:

plot	plot.
legend	legend plot.

---

PrepareAnnovarFile	<i>Fetches the closest Gene.refGene from an ANNOVAR file</i>
--------------------	--

---

**Description**

This function prepares an ANNOVAR file by fetching the closest Gene.refGene from an ANNOVAR file. If the genomic distance information cannot be parsed, the original 'Gene.refGene' value is returned.

**Usage**

```
PrepareAnnovarFile(annovar.file)
```

**Arguments**

annovar.file	ANNOVAR file including path.
--------------	------------------------------

**Value**

A data.frame with the following items:

Chr	Chromosome name.
Pos	Genomic position.
Ref	Nucleotide(s).
Alt	Nucleotide(s).
Gene.refGene	Closest Gene.refGene.
Func.refGene	Functional consequence of variant.

---

PrepareDbsDataFrame	<i>Prepares DBS data.frame</i>
---------------------	--------------------------------

---

### Description

This function prepares a DBS data.frame and returns relevant data to run IdentifySignatures function.

### Usage

```
PrepareDbsDataFrame(df, bsg, analyze.variants)
```

### Arguments

df	A data.frame with the following columns: Chr, Pos, Ref, Alt.
bsg	BSgenome object.
analyze.variants	A boolean value that indicates whether variant-level signature analysis should be performed.

### Value

A list with the following items:

df.dbs	A data.frame of DBS data.
df.dbs.frequencies	A data.frame of DBS mutation type frequencies.

---

PrepareDbsVcfFile	<i>Prepares DBS VCF file</i>
-------------------	------------------------------

---

### Description

This function prepares a DBS VCF file and returns relevant data to run IdentifySignatures function.

### Usage

```
PrepareDbsVcfFile(vcf.file, bsg)
```

### Arguments

vcf.file	VCF file including path.
bsg	BSgenome object.

### Value

A list with the following items:

df.dbs	A data.frame of DBS data.
df.dbs.frequencies	A data.frame of DBS mutation type frequencies.

---

PrepareIndelDataFrame    *Prepares indel data.frame*

---

### Description

This function prepares an indel data.frame and returns relevant data to run IdentifySignatures function.

### Usage

```
PrepareIndelDataFrame(  
  df,  
  bsg,  
  analyze.variants,  
  max.len = 25,  
  padding.len = 80  
)
```

### Arguments

df	A data.frame with the following columns: Chr, Pos, Ref, Alt.
bsg	BSgenome object.
analyze.variants	A boolean value that indicates whether variant-level signature analysis should be performed.
max.len	Maximum length of an indel to include. Indels longer than this length will be excluded.
padding.len	Number of bases to pad upstream and downstream of each indel.

### Value

A list with the following items:

df.indel	A data.frame of the indel data.
df.indel.frequencies	A data.frame of the PCAWG indel class frequencies.

---

PrepareIndelVcfFile    *Prepares indel VCF file*

---

### Description

This function prepares an indel VCF file and returns relevant data to run IdentifySignatures function.

### Usage

```
PrepareIndelVcfFile(vcf.file, bsg, max.len = 25, padding.len = 80)
```

**Arguments**

<code>vcf.file</code>	VCF file including path.
<code>bsg</code>	BSgenome object.
<code>max.len</code>	Maximum length of an indel to include. Indels longer than this length will be excluded.
<code>padding.len</code>	Number of bases to pad upstream and downstream of each indel.

**Value**

A list with the following items:

<code>df.indel</code>	A data.frame of the indel data.
<code>df.indel.frequencies</code>	A data.frame of the PCAWG indel class frequencies.

---

PrepareSbsDataFrame	<i>Prepares SBS data.frame</i>
---------------------	--------------------------------

---

**Description**

This function prepares a SBS data.frame and returns relevant data to run IdentifySignatures function.

**Usage**

```
PrepareSbsDataFrame(df, bsg, reference, context.length, analyze.variants)
```

**Arguments**

<code>df</code>	A data.frame with the following columns: Chr, Pos, Ref, Alt.
<code>bsg</code>	BSgenome object.
<code>reference</code>	A data.frame with the following columns: Mutation_Type, and names of SBS signatures (default: a data.frame returned from <a href="#">GetPcawgSbsSignaturesData</a> ).
<code>context.length</code>	Number of context nucleotides.
<code>analyze.variants</code>	A boolean value that indicates whether variant-level signature analysis should be performed (default: FALSE).

**Value**

A list with the following items:

<code>df.sbs</code>	A data.frame of SBS data.
<code>df.sbs.frequencies</code>	A data.frame of SBS mutation type frequencies.

---

PrepareSbsVcfFile	<i>Prepares SBS VCF file</i>
-------------------	------------------------------

---

**Description**

This function prepares a SBS VCF file and returns relevant data to run IdentifySignatures function.

**Usage**

```
PrepareSbsVcfFile(vcf.file, bsg, reference, context.length)
```

**Arguments**

vcf.file	VCF file including path.
bsg	BSgenome object.
reference	A data.frame with the following columns: Mutation_Type, and names of SBS signatures (default: a data.frame returned from <a href="#">GetPcawgSbsSignaturesData</a> ).
context.length	Number of context nucleotides.

**Value**

A list with the following items:

df.sbs	A data.frame of SBS data.
df.sbs.frequencies	A data.frame of SBS mutation type frequencies.

---

PrepareVcfFile	<i>Prepares a VCF file</i>
----------------	----------------------------

---

**Description**

This function prepares a VCF file.

**Usage**

```
PrepareVcfFile(vcf.file)
```

**Arguments**

vcf.file	VCF file including path.
----------	--------------------------

**Value**

A data.frame with the following items:

Chr	Chromosome name.
Pos	Genomic position.
Ref	Genomic position.
Alt	Genomic position.

---

PreprocessDbs	<i>Preprocess doublet base substitutions</i>
---------------	--

---

**Description**

This function preprocesses a data.frame of DBS.

**Usage**

```
PreprocessDbs(df, bsg, analyze.variants)
```

**Arguments**

df	A data.frame with the following columns: Chr, Pos, Ref, Alt.
bsg	BSgenome object.
analyze.variants	A boolean value that indicates whether variant-level signature analysis should be performed.

**Value**

A data.frame with the following columns:

Chr	Chromosome name.
Pos	Genomic position.
Ref	Reference nucleotide sequence.
Alt	Alternate nucleotide sequence.
Mutation_Type	One of the 96 mutation sub types (e.g. AC>CA).

---

PreprocessIndels	<i>Preprocess indels</i>
------------------	--------------------------

---

**Description**

This function preprocesses a data.frame of indels.

**Usage**

```
PreprocessIndels(df, bsg, padding.len, analyze.variants)
```

**Arguments**

df	A data.frame with the following columns: Chr, Pos, Ref, Alt.
bsg	BSgenome object.
padding.len	Number of bases to fetch upstream and downstream of each indel.
analyze.variants	A boolean value that indicates whether variant-level signature analysis should be performed.

**Value**

A data.frame with the following columns:

Chr	Chromosome name.
Pos	Genomic position.
Ref	Reference nucleotide sequence.
Alt	Alternate nucleotide sequence.
Type	Either 'INS' (insertion) or 'DEL' (deletion).
Len	Length (number of bases) of this indel.
Mutated_Seq	Nucleotide sequence of this indel.
Upstream_Seq	Upstream nucleotide sequence of this indel.
Upstream_Start	Upstream nucleotide sequence start position.
Upstream_End	Upstream nucleotide sequence end position.
Downstream_Seq	Downstream nucleotide sequence of this indel.
Downstream_Start	Downstream nucleotide sequence start position..
Downstream_End	Downstream nucleotide sequence end position.

---

PreprocessSbs

*Preprocess single base substitutions*


---

**Description**

This function preprocesses a data.frame of SBS.

**Usage**

```
PreprocessSbs(df, bsg, context.length, analyze.variants)
```

**Arguments**

df	A data.frame with the following columns: Chr, Pos, Ref, Alt.
bsg	BSgenome object.
context.length	Number of context nucleotides.
analyze.variants	A boolean value that indicates whether variant-level signature analysis should be performed.

**Value**

A data.frame with the following columns:

Chr	Chromosome name.
Pos	Genomic position.
Ref	Reference nucleotide sequence.
Alt	Alternate nucleotide sequence.

Type	One of the six mutation types (e.g. C>A).
Sub_Type	One of the 32 trinucleotide subtypes (e.g. ACA).
Somatic_Mutation_Type	One of the 96 mutation sub types (e.g. A[C>A]A).
Upstream_Seq	Upstream nucleotide sequence of this SBS.
Downstream_Seq	Downstream nucleotide sequence of this SBS.

---

PrintLog	<i>Prints log</i>
----------	-------------------

---

### Description

This function prints a log message.

### Usage

```
PrintLog(message, type = "INFO")
```

### Arguments

message	String value message to print along with log type and date.
type	String value that represents type of this message. 'INFO' by default.

---

SortDataFrameForStackedBarPlot	<i>Sorts a data.frame before plotting a stacked barplot</i>
--------------------------------	---

---

### Description

This function sorts a data.frame before plotting a stacked barplot.

### Usage

```
SortDataFrameForStackedBarPlot(df, ordered.features, x.axis.var)
```

### Arguments

df	A data.frame.
ordered.features	An ordered character vector of columns to sort in df.
x.axis.var	String value for x axis variable.

### Value

A sorted data.frame.



---

UnwrapIdentifiedModel    *Unwraps an identified model*

---

### Description

This function returns an unwrapped list of an identified model.

### Usage

```
UnwrapIdentifiedModel(identified.model)
```

### Arguments

identified.model

A data.frame returned from [IdentifySbsSignatures](#), [IdentifyDbsSignatures](#), or [IdentifyIndelSignatures](#).

### Value

A data.frame.

---

WrapIdentifiedModel    *Wraps an identified model*

---

### Description

This function returns a (wrapped) data.frame of an identified model

### Usage

```
WrapIdentifiedModel(  
  mutations.count,  
  signatures,  
  signatures.weights,  
  mutation.types,  
  mutation.types.groups,  
  observed.spectrum,  
  reconstructed.spectrum,  
  residual.spectrum,  
  cosine.similarity,  
  rss,  
  rss.normalized,  
  bic  
)
```

**Arguments**

<code>mutations.count</code>	Mutations count.
<code>signatures</code>	A character vector of signature names.
<code>signatures.weights</code>	A numeric vector of signature weights.
<code>mutation.types</code>	A character vector of mutation types.
<code>mutation.types.groups</code>	A character vector of mutation type groups.
<code>observed.spectrum</code>	A numeric vector of observed spectrum.
<code>reconstructed.spectrum</code>	A numeric vector of reconstructed spectrum.
<code>residual.spectrum</code>	A numeric vector of residual spectrum.
<code>cosine.similarity</code>	Cosine similiarity score.
<code>rss</code>	Residual sum of squares.
<code>rss.normalized</code>	Normalized residual sum of squares.
<code>bic</code>	Bayesian information criterion.

**Value**

A data.frame with the following columns:

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