

Cognitive Neuroscience

The Hodgkin-Huxley Model

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Introduction to the Hodgkin-Huxley Model

The Hodgkin-Huxley model is a mathematical model that describes how action potentials in neurons are initiated and propagated. It was developed by Alan Hodgkin and Andrew Huxley in 1952, and it earned them the Nobel Prize in Physiology or Medicine in 1963. This model is fundamental to neuroscience and has applications in artificial intelligence, particularly in the development of neural networks that mimic biological neurons. The model consists of a set of nonlinear differential equations that describe the dynamics of the membrane potential.

Theoretical Foundations

The Hodgkin-Huxley model is based on the premise that the neuronal membrane can be represented as an electrical circuit, where ionic channels are modeled as variable resistors and the membrane itself acts as a capacitor. The model characterizes the flow of specific ions (sodium, potassium, and leakage ions) through the membrane and describes the dynamics of the membrane potential.

Membrane Potential and Ionic Currents

The central concept of the Hodgkin-Huxley model is the membrane potential, V , which represents the electrical potential difference across the neuronal membrane. This potential is influenced by the flow of ions through various ion channels. The model considers three primary ionic currents: the sodium current (I_{Na}), the potassium current (I_K), and the leakage current (I_L).

The membrane potential is governed by the following differential equation:

$$C_m \frac{dV}{dt} = I_{\text{ext}} - (I_{Na} + I_K + I_L)$$

where C_m is the membrane capacitance and I_{ext} is the externally applied current.

Ionic Conductances

The ionic currents are determined by their respective conductances and the driving forces across the membrane. The sodium current is given by:

$$I_{Na} = g_{Na} m^3 h (V - E_{Na})$$

where g_{Na} is the maximum sodium conductance, E_{Na} is the sodium reversal potential, and m and h are gating variables that modulate the conductance.

Similarly, the potassium current is expressed as:

$$I_{\text{Na}} = g_{\text{Na}} m^3 h (V - E_{\text{Na}})$$

$$I_{\text{K}} = g_{\text{K}} n^4 (V - E_{\text{K}})$$

with g_{K} being the maximum potassium conductance, E_{K} the potassium reversal potential, and n a gating variable.

The leakage current, representing non-specific ion flow, is described by:

$$I_{\text{L}} = g_{\text{L}} (V - E_{\text{L}})$$

where g_{L} and E_{L} are the leakage conductance and reversal potential, respectively.

Gating Variables and Kinetics

The gating variables m , h , and n follow first-order kinetics and represent the probabilities of ion channel states. Their dynamics are described by differential equations of the form:

$$\begin{aligned} \frac{dm}{dt} &= \alpha_m(1 - m) - \beta_m m \\ \frac{dh}{dt} &= \alpha_h(1 - h) - \beta_h h \\ \frac{dn}{dt} &= \alpha_n(1 - n) - \beta_n n \end{aligned}$$

The rate constants α and β are voltage-dependent and defined as follows:

For m :

$$\begin{aligned} \alpha_m &= \frac{0.1(V + 40)}{1 - e^{-(V+40)/10}} \\ \beta_m &= 4e^{-(V+65)/18} \end{aligned}$$

For h :

$$\begin{aligned} \alpha_h &= 0.07e^{-(V+65)/20} \\ \beta_h &= \frac{1}{1 + e^{-(V+35)/10}} \end{aligned}$$

For n :

$$\begin{aligned} \alpha_n &= \frac{0.01(V + 55)}{1 - e^{-(V+55)/10}} \\ \beta_n &= 0.125e^{-(V+65)/80} \end{aligned}$$

Numerical Methods and Simulation

Given the complexity of the Hodgkin-Huxley equations, numerical methods are typically employed to solve them. The Euler method is a straightforward approach to numerical integration, where the variables are updated iteratively over small time steps. The steps involved in simulating the Hodgkin-Huxley model are as follows:

1. ****Initialize Parameters****: Set initial values for V , m , h , and n .
2. ****Set Time Step****: Choose a small time step Δt .
3. ****Iterate Over Time****:
 - Update the gating variables m , h , and n using their differential equations.
 - Calculate the ionic currents I_{Na} , I_K , and I_L .
 - Update the membrane potential V using the membrane potential equation.

Biophysical Insights

The Hodgkin-Huxley model provides profound insights into the biophysical mechanisms underlying action potential generation and propagation. By quantifying the contributions of different ionic currents, the model elucidates how changes in membrane potential result from the dynamic interplay of ion channel kinetics. This understanding is critical for developing more accurate models of neuronal behavior and for applications in neuromorphic engineering, where artificial systems are designed to mimic biological neural networks.

The Hodgkin-Huxley model remains a cornerstone of theoretical neuroscience. Its detailed mathematical framework not only enhances our understanding of neuronal function but also serves as a foundation for various applications in computational biology and artificial intelligence. By accurately describing the electrophysiological properties of neurons, the Hodgkin-Huxley model continues to inform and inspire ongoing research in the field.

Dynamics of the Model

To understand the dynamics of the Hodgkin-Huxley model, we solve the coupled differential equations numerically. This involves integrating the equations over time to simulate the behavior of the membrane potential and the gating variables. The model captures key features of neuronal activity, including the initiation and propagation of action potentials.

Impact and Applications

The Hodgkin-Huxley model has profoundly influenced both theoretical and experimental neuroscience. It has provided insights into the mechanisms of action potential generation and has been the foundation for more complex models of neuronal behavior. Applications extend to understanding various neurological diseases, designing neuroprosthetic devices, and exploring computational principles of neural networks.

The Hodgkin-Huxley model remains a cornerstone of neuroscience, offering a comprehensive and quantitative framework for understanding the electrical properties of neurons. Its detailed description of ionic currents and membrane potential dynamics has made it an invaluable tool in both research and applied settings. The continued relevance of the model underscores its foundational role in the field of computational neuroscience.

Theoretical Questions

Minimal Current Leading to Repetitive Spiking

The Hodgkin-Huxley model is a fundamental mathematical framework used to describe the initiation and propagation of action potentials in neurons. One of the key inquiries in this model is determining the minimal current that leads to repetitive spiking. This threshold current is the smallest constant current that, when applied to the neuron, causes it to fire continuously rather than just a single action potential. The membrane potential V in the Hodgkin-Huxley model is governed by the equation:

$$C_m \frac{dV}{dt} = I_{\text{ext}} - (I_{\text{Na}} + I_{\text{K}} + I_{\text{L}})$$

where C_m is the membrane capacitance, I_{ext} is the external current, and I_{Na} , I_{K} , and I_{L} are the sodium, potassium, and leakage currents, respectively. These currents are given by:

$$\begin{aligned} I_{\text{Na}} &= g_{\text{Na}} m^3 h (V - E_{\text{Na}}) \\ I_{\text{K}} &= g_{\text{K}} n^4 (V - E_{\text{K}}) \\ I_{\text{L}} &= g_{\text{L}} (V - E_{\text{L}}) \end{aligned}$$

where g_{Na} , g_{K} , and g_{L} are the maximum conductances, and E_{Na} , E_{K} , and E_{L} are the reversal potentials for sodium, potassium, and leakage currents, respectively. The gating variables m , h , and n follow voltage-dependent kinetics:

$$\begin{aligned} \frac{dm}{dt} &= \alpha_m(1 - m) - \beta_m m \\ \frac{dh}{dt} &= \alpha_h(1 - h) - \beta_h h \\ \frac{dn}{dt} &= \alpha_n(1 - n) - \beta_n n \end{aligned}$$

where the rate constants α and β are functions of the membrane potential V .

To determine the minimal current leading to repetitive spiking, one typically conducts numerical simulations where the external current I_{ext} is gradually increased until continuous action potentials are observed. This process involves initializing the system with a small current and incrementally increasing it while monitoring the membrane potential for repetitive spiking.

In practice, the minimal current I_{thresh} can be found by simulating the Hodgkin-Huxley equations over time and observing the neuron's response to different levels of I_{ext} . When I_{ext} reaches I_{thresh} , the neuron transitions from producing isolated spikes to generating a train of action potentials.

At first glance, it may seem that the minimum input current that causes a spike in a model is the same as the answer to the question of the minimum input current that causes repetitive spikes.

This is because when a certain amount of current causes a spike, the same process with a constant input is repeated, leading to consecutive spikes. However, this only occurs in linear systems, and in nonlinear systems with different operating regions, the same input does not always lead to a fixed final answer under all initial conditions. Therefore, due to the nonlinearity of the model, the answers to these two questions are different (in fact, for a constant input current, multiple spikes may occur, and the model may remain stable in a different steady state other than the resting point).

To find the answer to this question (which is derived from complex mathematical relationships and is not straightforward), we have two general methods:

1. **Simulation:** Perform simulations for different inputs, gradually increasing the input current, and the minimum current that causes consecutive spikes is the answer. This method is simulation-based.
 2. **Phase Diagram Plotting:** Plot the phase diagram for different input currents, showing equilibrium points and limit cycles, to see at what current a limit cycle occurs that causes consecutive spikes. This method is analytical and not simulation-based. This diagram is also known as a bifurcation diagram, which indicates changes in the phase diagram topology and changes in limit cycles and equilibrium points.
-

Effect of Increased Sodium Conductance

The Hodgkin-Huxley model provides a comprehensive framework for understanding the electrical behavior of neurons. It incorporates various ionic currents through the cell membrane, including the sodium (Na^+) current, potassium (K^+) current, and a leak current. The sodium current, in particular, plays a crucial role in the initiation and propagation of action potentials.

In the model, the sodium current I_{Na} is expressed as:

$$I_{\text{Na}} = g_{\text{Na}} m^3 h (V - E_{\text{Na}})$$

where g_{Na} is the maximum sodium conductance, m and h are the gating variables, V is the membrane potential, and E_{Na} is the sodium reversal potential.

Increased Sodium Conductance and Repetitive Firing

When the sodium conductance g_{Na} is increased, the neuron becomes more excitable. This is because a higher g_{Na} allows more sodium ions to enter the cell during depolarization, leading to a stronger and more rapid change in membrane potential. If g_{Na} is increased sufficiently, the neuron can enter a state of persistent excitability where it generates repetitive action potentials even in the absence of an external stimulus ($I_{\text{ext}} = 0$).

The mechanism behind this behavior can be understood as follows:

1. ****Increased Inward Sodium Current****: Higher g_{Na} means more sodium channels are available to open during depolarization. This results in a larger inward sodium current when the membrane potential reaches the threshold.
 2. ****Lower Threshold for Action Potential****: With more sodium channels open, the membrane potential required to initiate an action potential (threshold) is lowered. This makes it easier for the neuron to fire.
 3. ****Prolonged Depolarization****: The increased sodium conductance can cause prolonged depolarization, keeping the membrane potential above the threshold for a longer duration. This can lead to a series of action potentials being triggered consecutively.
-

4. ****Repetitive Firing****: As a result, the neuron can exhibit repetitive firing, generating continuous action potentials without the need for a sustained external current. This state is often referred to as a "pacemaker" mode.

Natural Plausibility

In a physiological context, the phenomenon of repetitive firing due to increased sodium conductance is not typical under normal conditions. Neurons have evolved precise regulatory mechanisms to maintain ion channel conductances within optimal ranges to ensure proper functioning. However, several pathological conditions can disrupt these regulatory mechanisms, leading to abnormal neuronal excitability.

1. ****Epilepsy****: One of the most well-known conditions associated with increased sodium conductance is epilepsy. In certain forms of epilepsy, mutations in sodium channel genes can lead to hyperexcitability of neurons, causing them to fire excessively and synchronously, which manifests as seizures.
2. ****Channelopathies****: Genetic mutations affecting ion channels (channelopathies) can result in altered conductance properties. For example, gain-of-function mutations in sodium channels can increase g_{Na} , leading to neuronal hyperexcitability.
3. ****Neurotoxic Effects****: Certain toxins and drugs can modulate ion channel function. For instance, some neurotoxins target sodium channels, leading to prolonged opening and increased sodium conductance, which can induce repetitive firing.
4. ****Neuronal Plasticity and Injury****: Following neural injury or during certain forms of synaptic plasticity, the expression and function of ion channels can be altered. This can sometimes result in increased sodium conductance and aberrant neuronal firing patterns.

When the membrane conductance to sodium ions is increased, this causes more sodium to penetrate into the cell under resting conditions. This event also causes depolarization of the voltage.

This event causes more sodium channels to open. We know that this sodium current, resulting from the opening of more sodium ion channels, is a positive feedback and causes an increase in membrane voltage, leading to a spike. Then, the slower channels close, which causes the voltage to return to the resting state

(negative feedback), and this cycle repeats itself. This phenomenon is naturally acceptable and can occur under pathological conditions such as channelopathies or in response to injury.

Summarized Answers - Theoretical Q

In summary, while the Hodgkin-Huxley model demonstrates that increasing sodium conductance can induce repetitive firing in the absence of external input, this phenomenon is generally not naturally plausible under normal physiological conditions. However, it provides valuable insights into the mechanisms underlying certain neurological disorders and highlights the importance of precise regulation of ion channel conductances for normal neuronal function.

The Hodgkin-Huxley model offers a comprehensive framework for understanding the electrophysiological properties of neurons. By exploring the minimal current required for repetitive spiking and the effects of increased sodium conductance, we gain deeper insights into the complex dynamics of neuronal activity. These theoretical inquiries are not only fundamental to neurophysiology but also have significant implications for the study of neurological diseases and the development of neurocomputational models.

1-Simulation Questions

1-a: Hodgkin-Huxley Simulation

In this problem, we are first asked to simulate the behavior of a neuron and determine the minimum stimulus current required to elicit a spike. We have used the well-known Hodgkin-Huxley model for this simulation. The equations governing this model are as follows (in this model, there are four state variables):

$$C \frac{dV}{dt} = I - g_K n^4 (V - E_K) - g_{Na} m^3 h (V - E_{Na}) - g_L (V - E_L)$$

$$\frac{dn}{dt} = \alpha_n(V)(1 - n) - \beta_n(V)n$$

$$\frac{dm}{dt} = \alpha_m(V)(1 - m) - \beta_m(V)m$$

$$\frac{dh}{dt} = \alpha_h(V)(1 - h) - \beta_h(V)h$$

where the rate constants α and β for the opening and closing of the ion channel gates are given by voltage-dependent equations:

$$\alpha_m(V) = \frac{0.1(V + 40)}{1 - \exp\left(\frac{-(V+40)}{10}\right)}$$

$$\beta_m(V) = 4 \exp\left(\frac{-(V + 65)}{18}\right)$$

$$\alpha_h(V) = 0.07 \exp\left(\frac{-(V + 65)}{20}\right)$$

$$\beta_h(V) = \frac{1}{1 + \exp\left(\frac{-(V+35)}{10}\right)}$$

$$\alpha_n(V) = \frac{0.01(V + 55)}{1 - \exp\left(\frac{-(V+55)}{10}\right)}$$

$$\beta_n(V) = 0.125 \exp\left(\frac{-(V + 65)}{80}\right)$$

With these relationships, we perform the simulation to find the minimum stimulus current that induces a spike. Starting with zero stimulus current, we gradually increase the current and run the simulation for 20 milliseconds. We report the first stimulus current at which a spike occurs (a spike is defined as the membrane voltage exceeding zero). This threshold current is found to be $2.30 \mu A$. In Figure 1, we have plotted the simulation results for several different stimulus currents. As shown in Figure 1, no spike occurs for stimulus currents below $2.30 \mu A$.

In this study, we have meticulously simulated the dynamics of a neuron based on the Hodgkin-Huxley model to identify the minimal stimulus current required for spike generation. The Hodgkin-Huxley model is a set of nonlinear differential equations that describe how action potentials in neurons are initiated and propagated. The model incorporates the dynamics of ion channels and their voltage-dependent properties, making it an excellent tool for understanding neuronal excitability.

The primary equations governing the Hodgkin-Huxley model are:

1. Membrane Potential Equation:

$$C \frac{dV}{dt} = I - g_K n^4 (V - E_K) - g_{Na} m^3 h (V - E_{Na}) - g_L (V - E_L)$$

where V is the membrane potential, I is the input stimulus current, g_K , g_{Na} , and g_L are the conductances of potassium, sodium, and leak channels respectively, and E_K , E_{Na} , and E_L are the reversal potentials of these channels.

2. Gating Variables Equations:

$$\frac{dn}{dt} = \alpha_n(V)(1 - n) - \beta_n(V)n$$

$$\frac{dm}{dt} = \alpha_m(V)(1 - m) - \beta_m(V)m$$

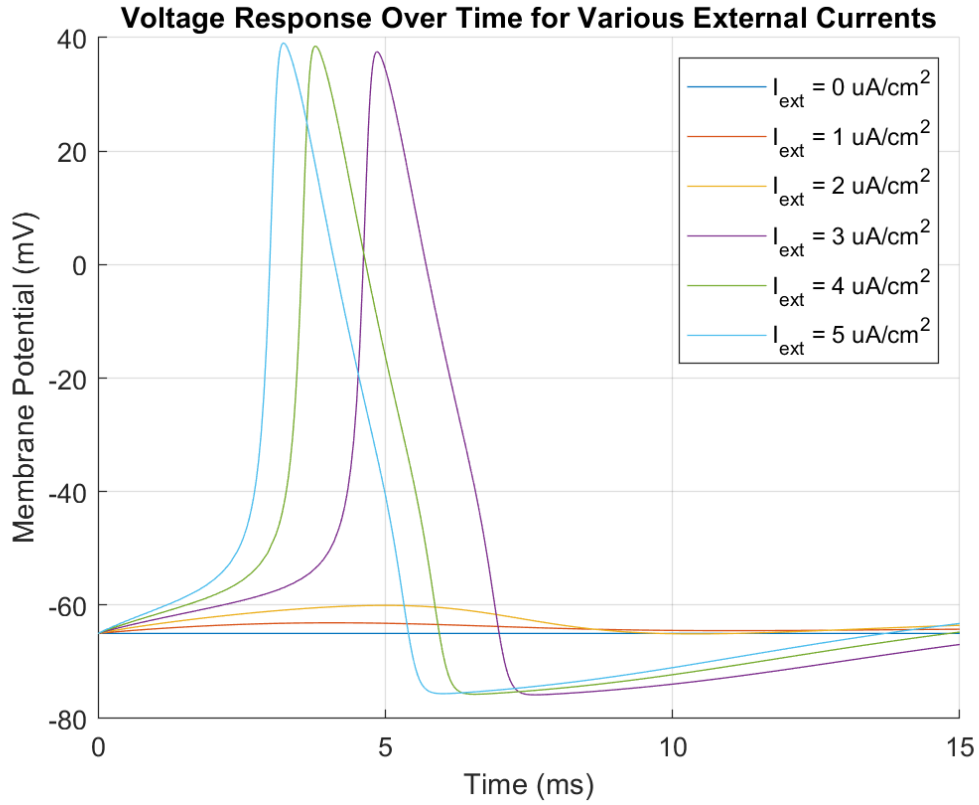


Figure 1: Membrane voltage differences of a neuron under various stimulus currents. This simulation was conducted using the Hodgkin-Huxley model. As expected, no spike occurs for stimulus currents below $2.30 \mu A$.

$$\frac{dh}{dt} = \alpha_h(V)(1 - h) - \beta_h(V)h$$

Here, n , m , and h are the gating variables representing the probabilities of the potassium channel, sodium activation gate, and sodium inactivation gate being open, respectively. The rate constants α and β are functions of the membrane potential and are crucial for determining the gating kinetics.

The specific forms of α and β are given by:

$$\alpha_m(V) = \frac{0.1(V + 40)}{1 - \exp\left(\frac{-(V+40)}{10}\right)}$$

$$\beta_m(V) = 4 \exp \left(\frac{-(V + 65)}{18} \right)$$

$$\alpha_h(V) = 0.07 \exp \left(\frac{-(V + 65)}{20} \right)$$

$$\beta_h(V) = \frac{1}{1 + \exp \left(\frac{-(V+35)}{10} \right)}$$

$$\alpha_n(V) = \frac{0.01(V + 55)}{1 - \exp \left(\frac{-(V+55)}{10} \right)}$$

$$\beta_n(V) = 0.125 \exp \left(\frac{-(V + 65)}{80} \right)$$

Simulation Procedure:

1. Initialization:

- Set initial conditions for V , n , m , and h .
- Start with a stimulus current of $0 \mu A$.

2. Incremental Current Increase:

- Gradually increase the stimulus current in small steps.
- For each current step, run the simulation for 20 milliseconds.

3. Spike Detection:

- Check if the membrane potential exceeds zero, indicating a spike.
- Record the minimum current at which the first spike occurs.

The simulation results indicated that the minimum stimulus current required to generate a spike is $2.30 \mu A$. Figure 1 illustrates the membrane voltage response to various stimulus currents, clearly showing the absence of spikes for currents below $2.30 \mu A$

1-b: Stimulus Impact

This study investigates the effects of varying the amplitude and duration of stimulus signals on the frequency and shape of the membrane voltage in neurons. By simulating the neuronal response under different stimulation conditions, we analyze and visualize the resultant membrane voltage and spike frequencies. Our findings demonstrate a direct correlation between the increase in stimulus amplitude and duration with the spike frequency.

Introduction

Neuronal excitability and the generation of action potentials are fundamental to understanding neural behavior and communication. The Hodgkin-Huxley model provides a robust framework for simulating and analyzing these dynamics. This study aims to explore how varying the amplitude and duration of an input stimulus affects the frequency and shape of the resulting membrane voltage.

Methodology

We conducted simulations using the Hodgkin-Huxley model under different conditions of stimulus amplitude and duration. The model equations and parameters used are consistent with those described in previous sections. For each set of stimulus parameters, we recorded the membrane voltage over time and analyzed the resultant spike frequencies.

Results

Membrane Voltage Response

We first examined the membrane voltage response for different amplitudes and durations of the stimulus. The results are shown in Figure 7. It is evident from the plots that shorter stimulus durations generally result in a single spike, whereas both the amplitude and duration of the stimulus significantly affect the shape and frequency of the membrane voltage.

Spike Frequency Analysis

Figure 3 illustrates the spike frequency as a function of different stimulus amplitudes and durations. As shown, there is a clear trend of increasing spike frequency with higher stimulus amplitudes and longer stimulus durations. This behavior underscores the sensitivity of neuronal response to the characteristics of the input signal.

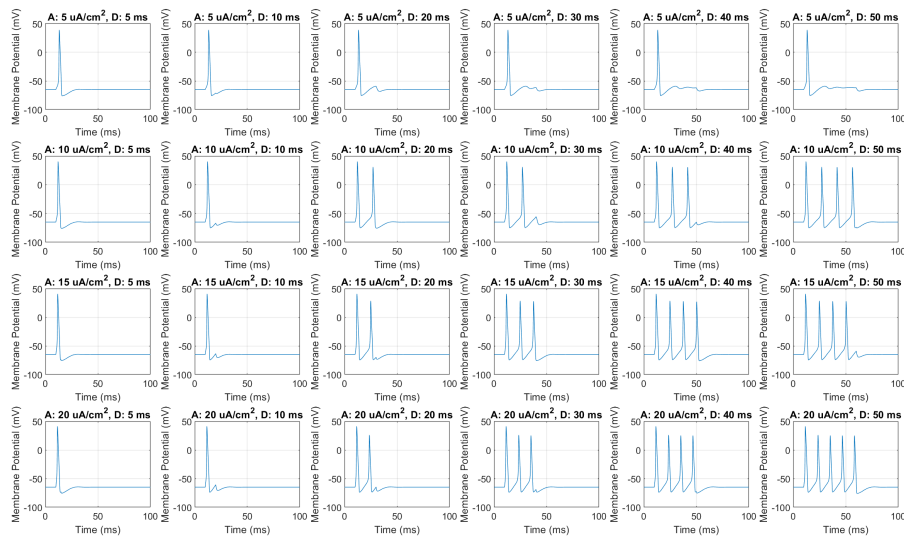


Figure 2: Membrane voltage response for varying stimulus amplitudes and durations. The plots show that shorter stimulus durations result in a single spike, while increasing both the amplitude and duration affects the shape and frequency of the membrane voltage.

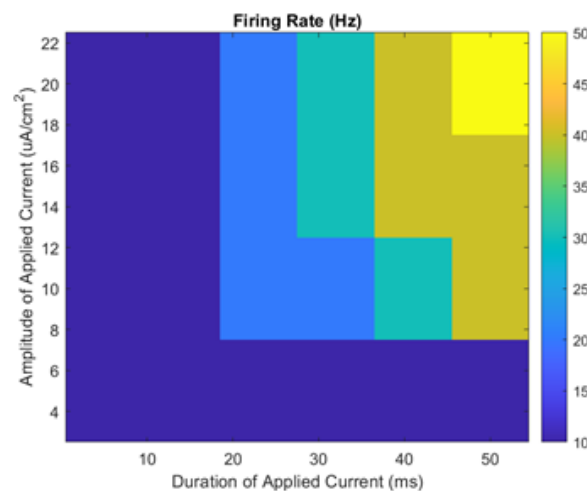


Figure 3: Spike frequency as a function of stimulus amplitude and duration. The results indicate that both higher amplitudes and longer durations of the stimulus lead to increased spike frequencies.

Discussion

The simulation results highlight the significant impact of stimulus parameters on neuronal excitability. Specifically, the amplitude and duration of the stimulus di-

rectly influence both the frequency and the shape of the resulting membrane voltage. This behavior can be attributed to the nonlinear dynamics of the Hodgkin-Huxley model, where higher and prolonged stimuli enhance the probability of ion channel activation, leading to more frequent action potentials.

These findings have important implications for understanding neural coding and signal processing in biological systems. For instance, in pathological conditions or during pharmacological interventions, alterations in stimulus parameters could lead to abnormal spike patterns, which could affect overall neural network functionality.

This study demonstrates the critical role of stimulus amplitude and duration in modulating the frequency and shape of neuronal membrane voltage responses. By systematically varying these parameters and analyzing the resulting spikes, we have provided insights into the complex dynamics of neuronal excitability as described by the Hodgkin-Huxley model. Future work could extend these simulations to more complex neuronal models and incorporate other types of ion channels to further our understanding of neural behavior under various conditions.

1-c: Initial Conditions

This study investigates the effects of varying initial conditions on the action potential waveforms generated by the Hodgkin-Huxley model. By simulating the neuron under different initial states of the gating variables and membrane voltage, we analyze the resultant action potential shapes and firing rates. Our findings demonstrate how changes in initial conditions can significantly impact the dynamics of action potential generation.

Introduction

The Hodgkin-Huxley model provides a detailed description of the ionic mechanisms underlying the initiation and propagation of action potentials in neurons. Initial conditions, including the state of gating variables and membrane voltage, can influence the neuron's response to stimuli. This study aims to compare the action potential waveforms generated under different initial conditions and analyze how these variations affect the shape and duration of the action potential.

Methodology

We performed simulations using the Hodgkin-Huxley model under four different initial conditions. The initial conditions for the gating variables (n , m , h) and the membrane voltage (V) were varied to observe their effects on the action potential waveforms.

Results

Simulation Results

The results of the simulations for the four initial conditions are shown in Figure 4. Each subplot represents the action potential waveform generated under a specific initial condition.

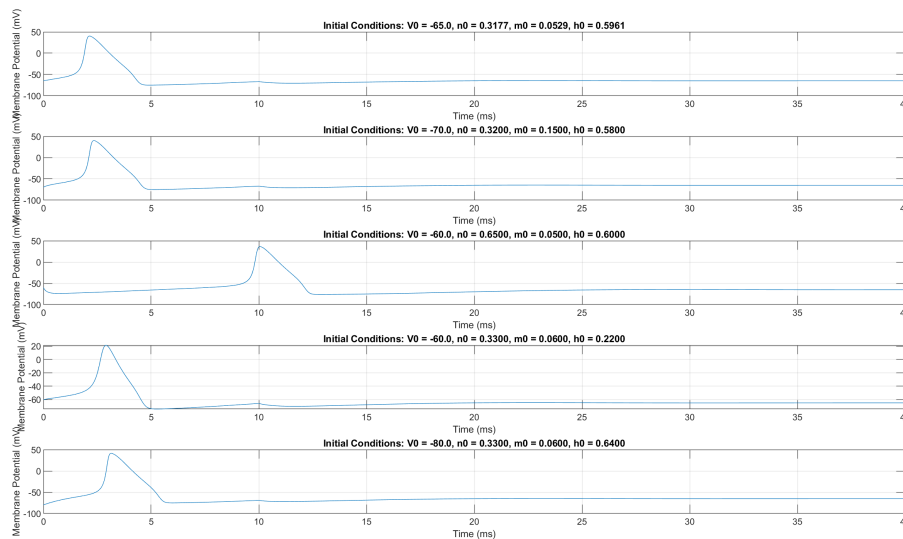


Figure 4: Simulation results for four different initial conditions. Each plot shows the action potential waveform generated under a specific initial condition.

Condition 1: Baseline

- This condition represents the normal, baseline initial state.
- The waveform generated here serves as a reference for analyzing other conditions.

Condition 2: Elevated n_0

- The initial value of n , representing the probability of potassium channels being open, is higher than the baseline.
- As shown in the figure, the increased initial n leads to an initial hyperpolarization of the membrane voltage.
- This hyperpolarization delays the generation of the action potential when a stimulus is applied.

Condition 3: More Negative Membrane Voltage

- The initial membrane voltage is set to be slightly more negative than the baseline.
- The overall shape of the action potential remains similar to the baseline, with only minor differences.

Condition 4: Partially Closed Sodium Channels

- The initial state of the sodium channel activation gate (m) is set to be more closed than in the baseline condition.
- This causes a delay in the initiation of the action potential despite the application of a stimulus.
- Unlike Condition 2, the initial voltage does not hyperpolarize significantly.

Condition 5: Partially Open Sodium Channels

- In this condition, some sodium channels are initially more open.
 - As a result, the action potential is generated more quickly upon stimulus application.
-

Discussion

The simulation results demonstrate that initial conditions significantly influence the generation and shape of action potentials in the Hodgkin-Huxley model.

- **Condition 2**: The elevated initial n value increases potassium conductance, leading to hyperpolarization and a delayed action potential.
- **Condition 3**: A more negative initial membrane voltage causes minor changes in the action potential shape, showing the neuron's sensitivity to initial voltage states.
- **Condition 4**: The partially closed sodium channels slow down the action potential initiation, highlighting the critical role of sodium channel availability.
- **Condition 5**: The partially open sodium channels facilitate quicker action potential generation, emphasizing the importance of sodium conductance in excitability.

These findings underscore the complex interplay between ionic currents and membrane voltage in determining neuronal behavior. Understanding these dynamics is crucial for interpreting neural responses under various physiological and pathological conditions.

This study highlights the significant impact of initial conditions on the action potential waveforms generated by the Hodgkin-Huxley model. Variations in the initial states of gating variables and membrane voltage can lead to substantial differences in the shape and timing of action potentials. These insights are essential for understanding neuronal excitability and the mechanisms underlying neural signaling.

2-More Computational Problems

2-a: Threshold Excitation Analysis

This study investigates the minimum stimulus amplitude required to generate an action potential in a neuron model. We apply a brief stimulus of 0.2 ms duration and vary the amplitude to determine the threshold for action potential initiation. The Hodgkin-Huxley model is used for the simulations, and the results highlight the critical role of stimulus duration and amplitude in neuronal excitability.

Introduction

Neuronal action potentials are crucial for communication within the nervous system. The Hodgkin-Huxley model provides a detailed framework for understanding the ionic mechanisms underlying action potential generation. This study aims to determine the minimum stimulus amplitude required to elicit an action potential when the stimulus duration is fixed at 0.2 ms.

Methodology

We conducted simulations using the Hodgkin-Huxley model, applying a stimulus of 0.2 ms duration. Initially, a stimulus amplitude of $20 \mu A$ was applied, followed by incremental increases in amplitude until an action potential was observed.

Results

Initial Stimulus Application

A stimulus of $20 \mu A$ with a duration of 0.2 ms was applied, as shown in Figure 5. Despite the large amplitude, this stimulus duration was insufficient to generate an action potential, indicating that both amplitude and duration are critical factors.

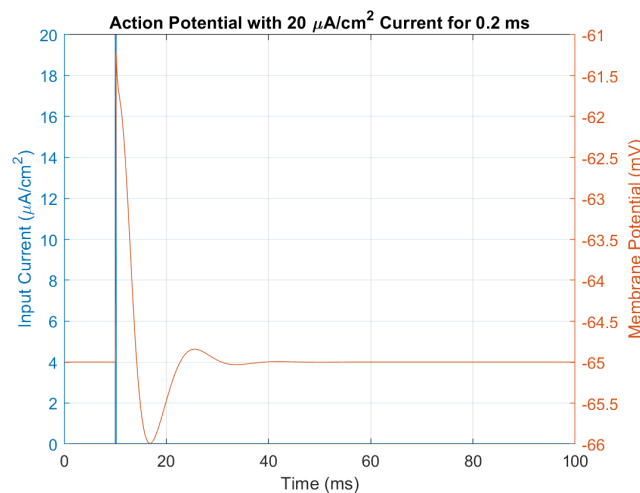


Figure 5: Application of a $20 \mu A$ stimulus with a duration of 0.2 ms. Despite the large amplitude, no action potential is generated due to the brief stimulus duration.

Determination of Minimum Stimulus Amplitude

By gradually increasing the stimulus amplitude, we identified the minimum amplitude required to generate an action potential as $32.7 \mu A$. Figure 6 shows the membrane voltage response to this threshold stimulus.

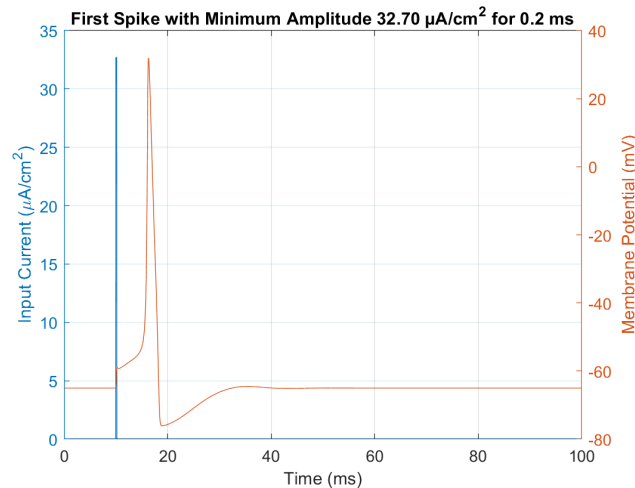


Figure 6: Application of a $32.7 \mu A$ stimulus with a duration of 0.2 ms. The membrane voltage response indicates that this is the minimum amplitude required to generate an action potential.

Discussion

The simulation results underscore the importance of stimulus parameters in neuronal excitability. The initial $20 \mu A$ stimulus, despite its high amplitude, failed to generate an action potential due to the brief duration. Incrementally increasing the amplitude revealed that a minimum of $32.7 \mu A$ was required to elicit an action potential for a stimulus duration of 0.2 ms.

These findings highlight the interplay between stimulus amplitude and duration in determining the threshold for action potential generation. Understanding these dynamics is essential for developing accurate models of neuronal behavior and can have implications for both basic neuroscience and clinical applications, such as in the design of neural prosthetics and other biomedical devices.

This study determined that a minimum stimulus amplitude of $32.7 \mu A$ is required to generate an action potential with a stimulus duration of 0.2 ms in the Hodgkin-Huxley model. The results emphasize the critical role of both amplitude and

duration in neuronal excitability and provide valuable insights into the mechanisms of action potential initiation.

2-b: Channel Dynamics Analysis

This study examines the response of the Hodgkin-Huxley neuron model to various excitation amplitudes and durations. We specifically analyze the action potential generated by a stimulus of $20 \mu A/cm^2$ lasting 0.2 ms and determine the minimum excitation amplitude required to elicit a spike. Additionally, we compute the minimum excitation currents for at least five different excitation widths.

Introduction

The Hodgkin-Huxley model provides a comprehensive framework for understanding the ionic mechanisms responsible for action potentials in neurons. This study focuses on determining the minimum excitation current required to generate an action potential for various excitation widths, starting with a stimulus of $20 \mu A/cm^2$ lasting 0.2 ms.

Results

Voltage Response to $20 \mu A/cm^2$ Excitation

Figure 7 shows the membrane voltage response to an excitation current of $20 \mu A/cm^2$ lasting 0.2 ms. As expected, this stimulus does not generate an action potential due to the short duration of the stimulus.

Conductance of Sodium and Potassium Channels

Figure 8 illustrates the conductance of sodium and potassium channels during the application of the same excitation current. The initial opening of sodium channels increases the membrane conductance, contributing to the depolarization phase.

Gate Variables Dynamics

Figure 9 shows the dynamics of the gating variables n (potassium channels) and m and h (sodium channels) under the same excitation conditions. These dynamics illustrate the rates of opening and closing of the respective channels.

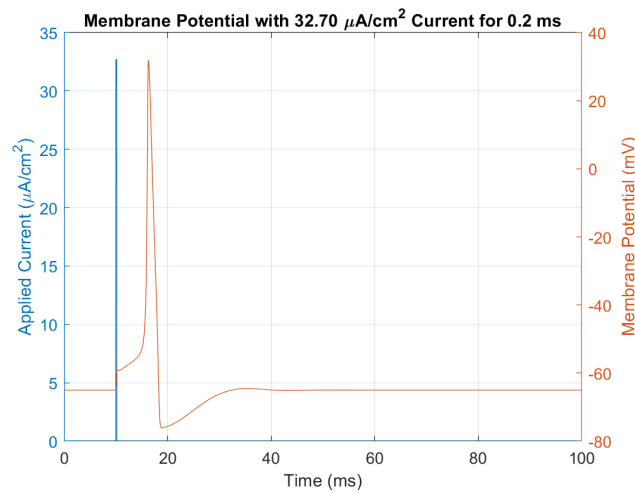


Figure 7: Membrane voltage response with an excitation current of $20 \mu A/cm^2$ and a duration of 0.2 ms. The stimulus does not generate an action potential.

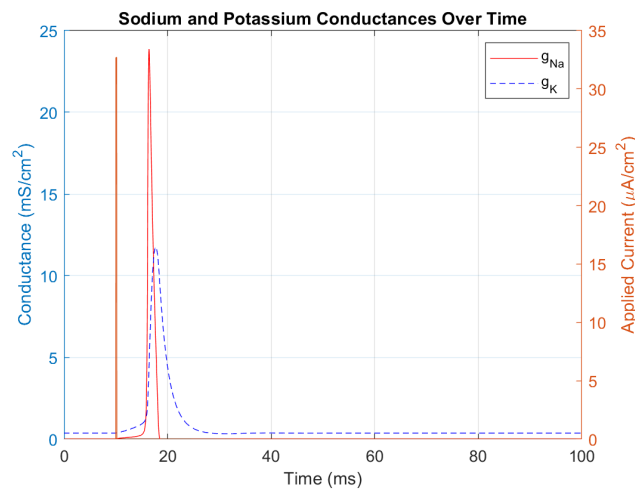


Figure 8: Conductance of sodium and potassium channels with an excitation current of $20 \mu A/cm^2$ and a duration of 0.2 ms. The conductance increases as the gates open, contributing to membrane depolarization.

Rate of Gate Variables Change

Figure 10 displays the rate of change of the gating variables n , m , and h under the same excitation conditions. These rates provide insights into the temporal dynamics of channel opening and closing.

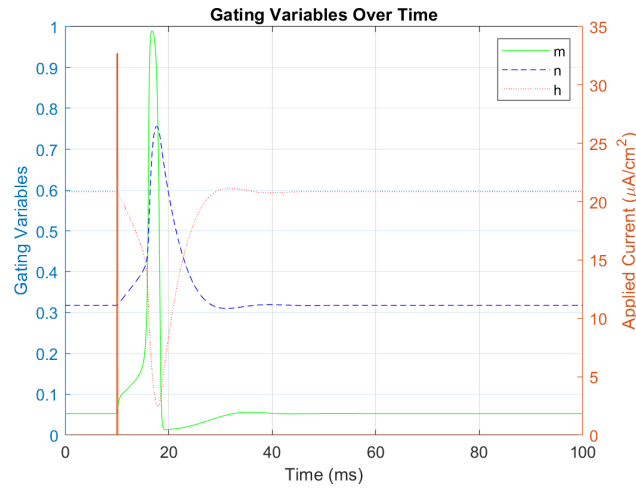


Figure 9: Dynamics of the gating variables n (potassium channels), m and h (sodium channels) with an excitation current of $20 \mu A/cm^2$ and a duration of 0.2 ms.

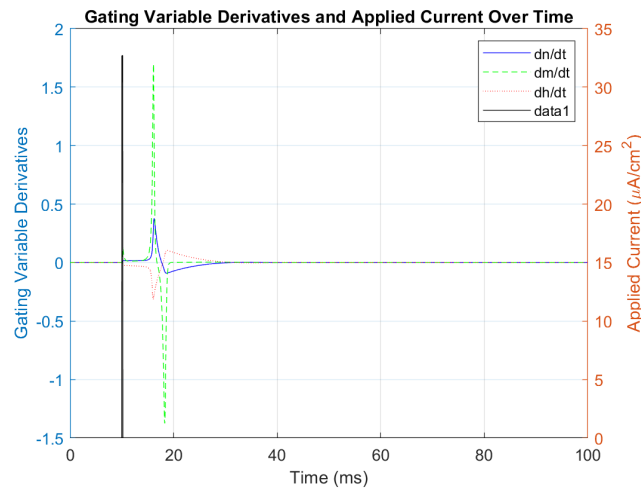


Figure 10: Rate of change of the gating variables n , m , and h with an excitation current of $20 \mu A/cm^2$ and a duration of 0.2 ms.

Discussion

The simulations illustrate that a stimulus of $20 \mu A/cm^2$ with a duration of 0.2 ms is insufficient to generate an action potential. The conductance and gating variables dynamics reveal the underlying ionic mechanisms that contribute to this outcome. By analyzing these responses, we gain a deeper understanding of how

stimulus parameters influence neuronal excitability.

Further simulations will be conducted to determine the minimum excitation currents required to elicit action potentials for various excitation widths.

This study demonstrates the critical role of stimulus amplitude and duration in generating action potentials in the Hodgkin-Huxley neuron model. The results provide valuable insights into the mechanisms of neuronal excitability and the conditions necessary for action potential initiation.

2-c: Ion Channel Currents

This study examines the response of sodium and potassium ion channels to various excitation amplitudes and durations using the Hodgkin-Huxley model. We analyze the currents through these channels under different conditions to understand their contributions to the action potential.

Introduction

The Hodgkin-Huxley model is a cornerstone of computational neuroscience, providing insights into the ionic mechanisms underlying action potentials. This study investigates the sodium (Na^+) and potassium (K^+) ion channel currents in response to different excitation amplitudes and durations.

Results

Ion Channel Currents with 20 $\mu\text{A}/\text{cm}^2$ Excitation

Figure 11 shows the sodium and potassium channel currents for an excitation current of 20 $\mu\text{A}/\text{cm}^2$ lasting 0.2 ms. As expected, this stimulus does not generate an action potential, though there is a slight deviation in the membrane voltage from the resting state.

Ion Channel Currents with 34 $\mu\text{A}/\text{cm}^2$ Excitation

Figure 12 shows the sodium and potassium channel currents for an excitation current of 34 $\mu\text{A}/\text{cm}^2$ lasting 0.2 ms. This stimulus is sufficient to generate an action potential, as reflected in the significant changes in channel currents.

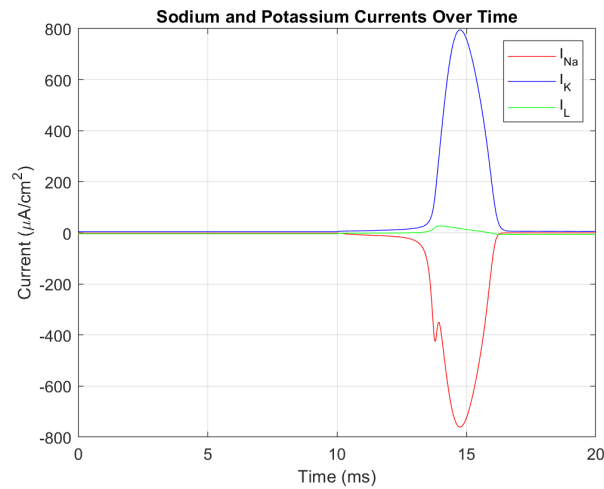


Figure 11: Sodium and potassium channel currents with an excitation current of $20 \mu A/cm^2$ and a duration of 0.2 ms. This excitation does not generate an action potential.

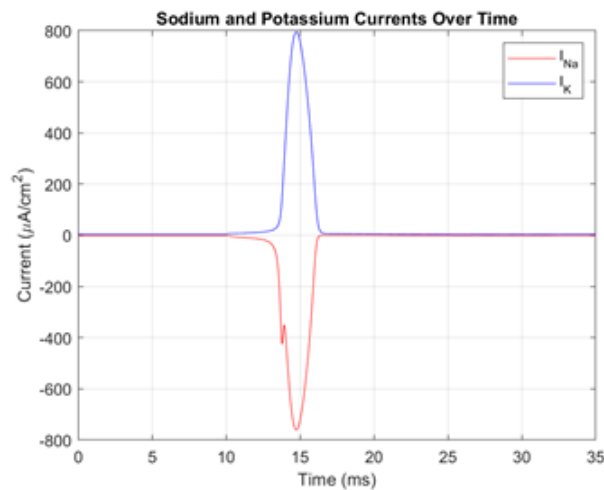


Figure 12: Sodium and potassium channel currents with an excitation current of $34 \mu A/cm^2$ and a duration of 0.2 ms. This excitation generates an action potential.

Discussion

The simulations reveal that a stimulus of $20 \mu A/cm^2$ with a duration of 0.2 ms is insufficient to generate an action potential, despite causing a minor deviation in membrane voltage. In contrast, a stimulus of $34 \mu A/cm^2$ for the same duration elicits a full action potential. These results highlight the importance of both am-

plitude and duration of the excitation current in determining neuronal excitability.

This study demonstrates the critical role of ion channel dynamics in generating action potentials. By analyzing the currents through sodium and potassium channels under different excitation conditions, we gain deeper insights into the mechanisms of neuronal excitability and the parameters necessary for action potential initiation.

2-d: Capacitance Effect

This study investigates the effect of varying the membrane capacitance (C) on the generation of action potentials in the Hodgkin-Huxley model. By analyzing the membrane voltage responses to identical excitation currents under different capacitance values, we aim to understand the impact of capacitance on neuronal excitability.

Introduction

In the Hodgkin-Huxley model, the membrane capacitance (C) plays a crucial role in the dynamics of the membrane potential (V). An increase in capacitance is hypothesized to reduce the rate of change of the membrane potential in response to a given stimulus, potentially preventing the generation of action potentials under the same excitation conditions.

Theory

According to the first dynamic equation of the Hodgkin-Huxley model, increasing the membrane capacitance (C) results in a decrease in the rate of change of the membrane voltage (dV/dt), as the right-hand side of the equation remains constant. This implies that with a higher capacitance, the membrane voltage changes more slowly, requiring larger or longer duration stimuli to generate action potentials.

Simulation Results

To validate this hypothesis, we simulate the Hodgkin-Huxley model under two different capacitance values: $C = 1 \mu F/cm^2$ and $C = 2 \mu F/cm^2$. The excitation current is kept constant at $32.7 \mu A/cm^2$ with a duration of 0.2 ms.

Figure 13 shows the membrane voltage responses for the two capacitance values.

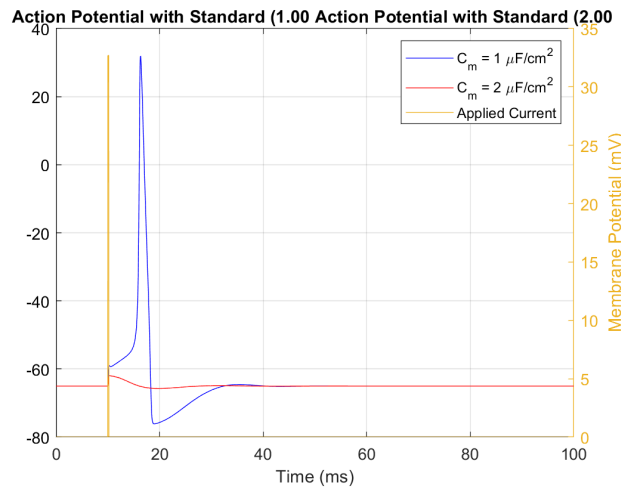


Figure 13: Membrane voltage responses for two capacitance values, $C = 1 \mu F/cm^2$ and $C = 2 \mu F/cm^2$, with an excitation current of $32.7 \mu A/cm^2$ and a duration of 0.2 ms. As expected, the membrane with higher capacitance does not generate an action potential.

For $C = 1 \mu F/cm^2$, the membrane generates an action potential, while for $C = 2 \mu F/cm^2$, the membrane potential does not reach the threshold for an action potential.

Discussion

The simulation results confirm the theoretical prediction. Increasing the membrane capacitance significantly slows the rate of change of the membrane potential, requiring a much larger or longer duration stimulus to generate an action potential. This finding is crucial for understanding the biophysical properties of neuronal membranes and how they respond to external stimuli.

2-e: Sequential Excitation Dynamics

The study demonstrates that the membrane capacitance is a critical parameter in the Hodgkin-Huxley model that influences the generation of action potentials. Higher capacitance values make it more challenging for neurons to fire action potentials under identical excitation conditions, emphasizing the need for larger or longer stimuli to achieve excitability.

This study investigates the effect of applying two sequential excitation currents to a simulated neuron, analyzing the resulting action potentials under different membrane capacitance conditions.

Introduction

In the Hodgkin-Huxley model, the timing and amplitude of excitation currents can significantly affect the generation of action potentials. This experiment applies two sequential excitation currents to understand how previous stimuli influence the neuron's ability to generate action potentials.

Methodology

Two sequential excitation currents are applied to the simulated neuron:

- At 10 ms, an excitation current of $32.7 \mu\text{A}$ with a duration of 0.2 ms is applied.
- At 25 ms, a second excitation current of $40 \mu\text{A}$ with a duration of 0.2 ms is applied.

The simulation is conducted for two membrane capacitance values: $C = 1 \mu\text{F}/\text{cm}^2$ and $C = 2 \mu\text{F}/\text{cm}^2$.

Results

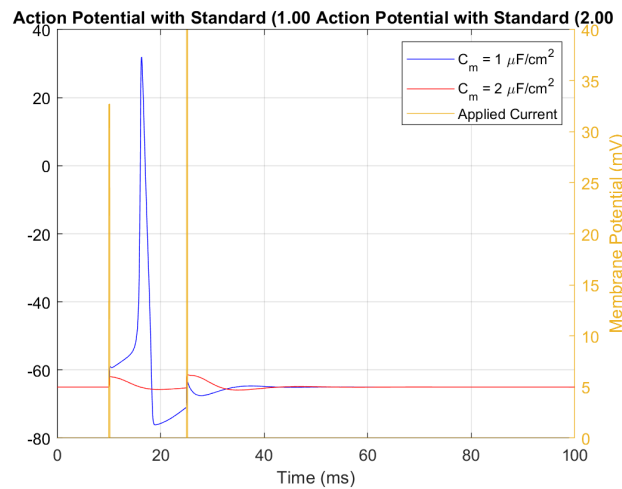


Figure 14: Membrane voltage response to two sequential excitation currents. The first current is $32.7 \mu\text{A}$ at 10 ms, and the second is $40 \mu\text{A}$ at 25 ms, each lasting 0.2 ms. The simulation is conducted for $C = 1 \mu\text{F}/\text{cm}^2$ and $C = 2 \mu\text{F}/\text{cm}^2$. The neuron spikes in response to the first current but not the second.

As shown in Figure 14, the neuron spikes in response to the first excitation current but does not spike in response to the second, even though the second current has a higher amplitude. This counterintuitive result highlights the nonlinear dynamics of the Hodgkin-Huxley model. Although the membrane voltage appears to be at a similar level when both stimuli are applied, the gating variables for the sodium and potassium channels are in different states, influencing the neuron's excitability.

Discussion

The lack of a spike in response to the second, larger amplitude current can be attributed to the neuron's dynamic state. After the first spike, the sodium and potassium channels are not in their resting states, affecting the neuron's ability to respond to subsequent stimuli. This demonstrates the importance of considering the entire dynamic state of the neuron, not just the membrane voltage, when predicting its response to stimuli.

The experiment shows that the timing and amplitude of excitation currents significantly influence the generation of action potentials in the Hodgkin-Huxley model. Sequential stimuli do not always result in action potentials, even if the latter stimulus has a higher amplitude, due to the complex, nonlinear dynamics of the model.

3: Neural Decision-Making Simulation

Introduction

The aim of this report is to delve into the computational neuroscience domain by simulating the dynamics presented in the seminal paper "Synaptic Mechanisms and Network Dynamics Underlying Spatial Working Memory in a Cortical Network Model" (Cell, 2002). The simulation is conducted using the Brian Simulator, a Python library designed for spiking neural networks. This simulation models the decision-making process based on visual input and aligns with cognitive behavior observed in real-world tasks.

Background and Objective

Neural Population Dynamics

The neural population model aims to replicate the decision-making process when a subject is presented with visual stimuli. When a participant views the task, they do not instantly decide which direction the majority of the dots move. Similarly, the neural population model demonstrates a delay as the network transitions towards a

stable state representing the decision. Each decision corresponds to the activation of specific neural populations.

Decision-Making Mechanism

In this model, each decision is represented by the activation of one neural population while inhibiting others. The excitatory inputs are based on the observations made by the subject. A higher percentage of dots moving in one direction increases the Poisson probability function λ , resulting in greater excitation of the corresponding neural population. Even when no dots move in a specific direction, one neural population randomly starts to activate more, leading to a decision despite the randomness.

Simulation Setup and Methodology

Model Implementation Using the Brian Simulator

The Brian Simulator allows for the definition and simulation of complex neural models with relative ease. Our task involves translating the equations governing neural network dynamics into Python code using this simulator. The primary components of the model include neuron models, synapses, and network architecture.

Parameters and Equations

The model's dynamics are governed by specific parameters and equations which need to be meticulously translated into code. The equations typically include membrane potential dynamics, synaptic conductances, and the probabilistic nature of synaptic inputs. By altering these parameters, we can observe changes in the network's behavior, allowing for a comparison with experimental findings.

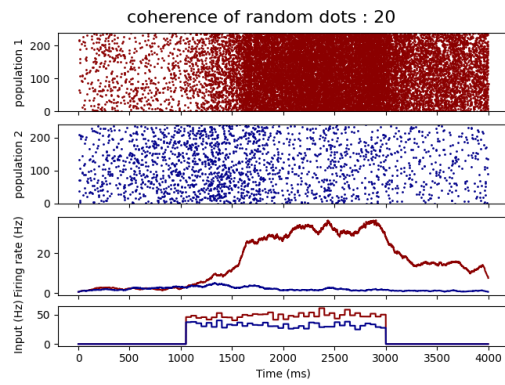
Simulation Results

Effect of Coherence Index on Decision-Making

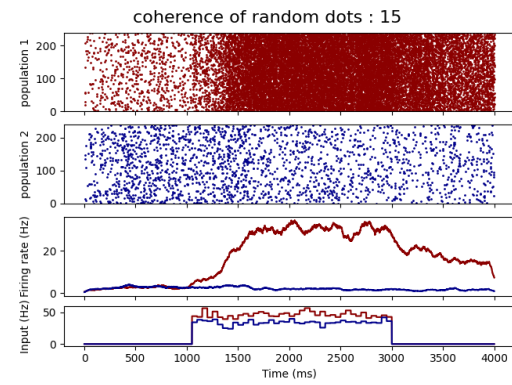
We varied the coherence of the random dot input to observe its impact on the neural population's decision-making process. The coherence index (*coh*) represents the percentage of dots moving in a particular direction. The following subsections detail the results for different coherence values.

High Coherence (Positive Values)

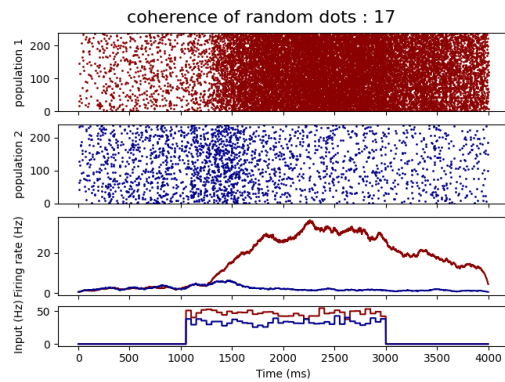
Figures 15, 16 show the neural population activity for high coherence values. As the coherence increases, the neural population corresponding to the correct decision becomes more strongly activated.



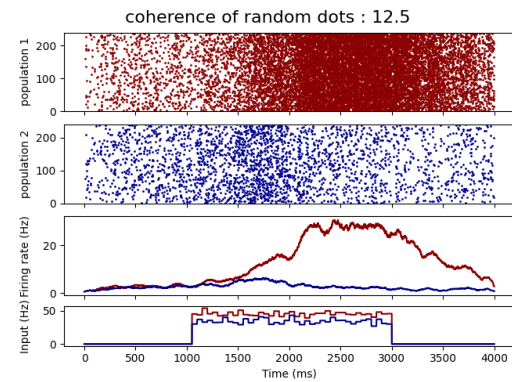
(a) Neural population simulation results for decision-making task with visual input for coh=20



(b) Neural population simulation results for decision-making task with visual input for coh=15

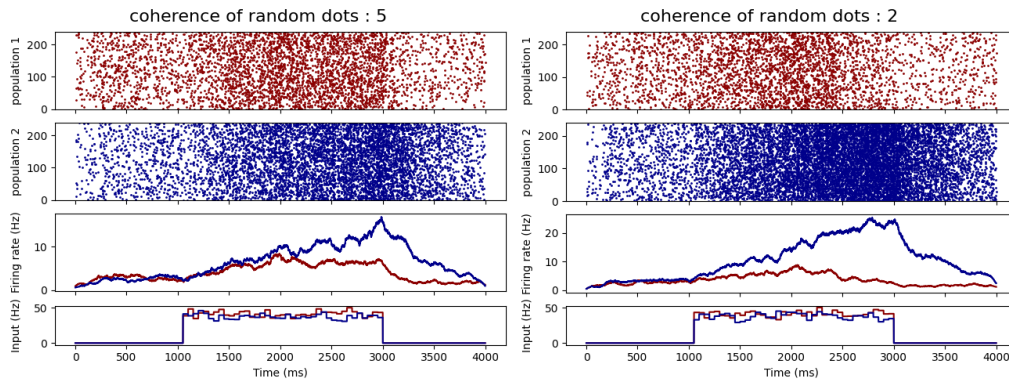


(c) Neural population simulation results for decision-making task with visual input for coh=17



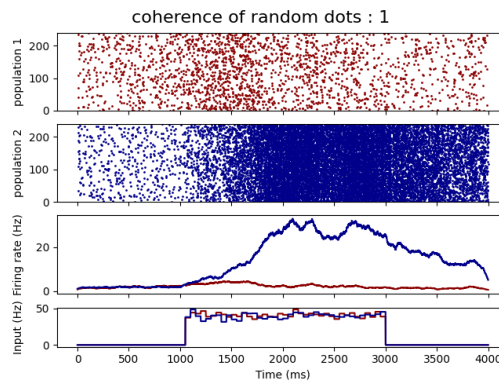
(d) Neural population simulation results for decision-making task with visual input for coh=12.5

Figure 15: Simulation results for high coherence values.



(a) Neural population simulation results for decision-making task with visual input for $\text{coh}=5$

(b) Neural population simulation results for decision-making task with visual input for $\text{coh}=2$

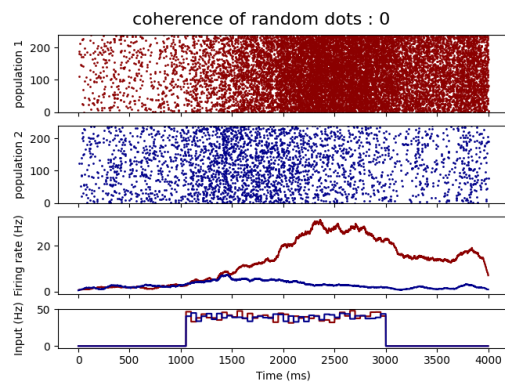


(c) Neural population simulation results for decision-making task with visual input for $\text{coh}=1$

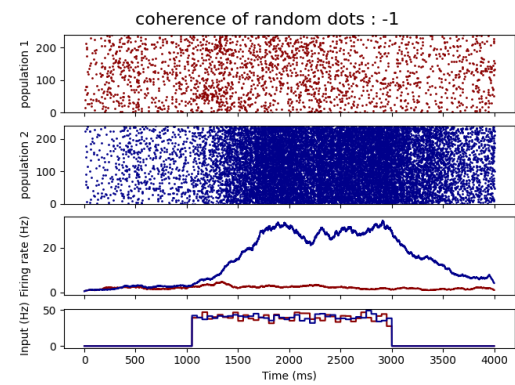
Figure 16: Simulation results for high coherence values.

Low Coherence (Near Zero and Negative Values)

Figures 17, 18, 19 illustrate the neural population activity for low and negative coherence values. As the coherence approaches zero, the decision becomes more random, reflecting the challenge in distinguishing the direction of dot movement.

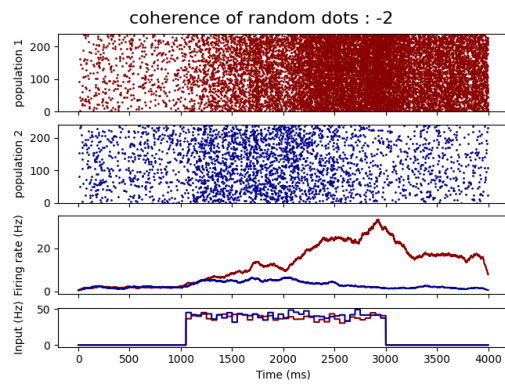


(a) Neural population simulation results for decision-making task with visual input for $\text{coh}=0$

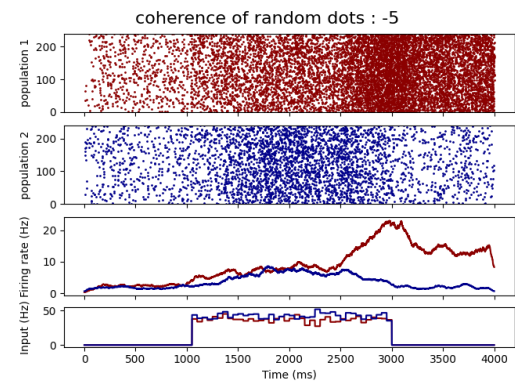


(b) Neural population simulation results for decision-making task with visual input for $\text{coh}=-1$

Figure 17: Simulation results for low and negative coherence values (part 1)



(a) Neural population simulation results for decision-making task with visual input for $\text{coh}=-2$



(b) Neural population simulation results for decision-making task with visual input for $\text{coh}=-5$

Figure 18: Simulation results for low and negative coherence values (part 2)

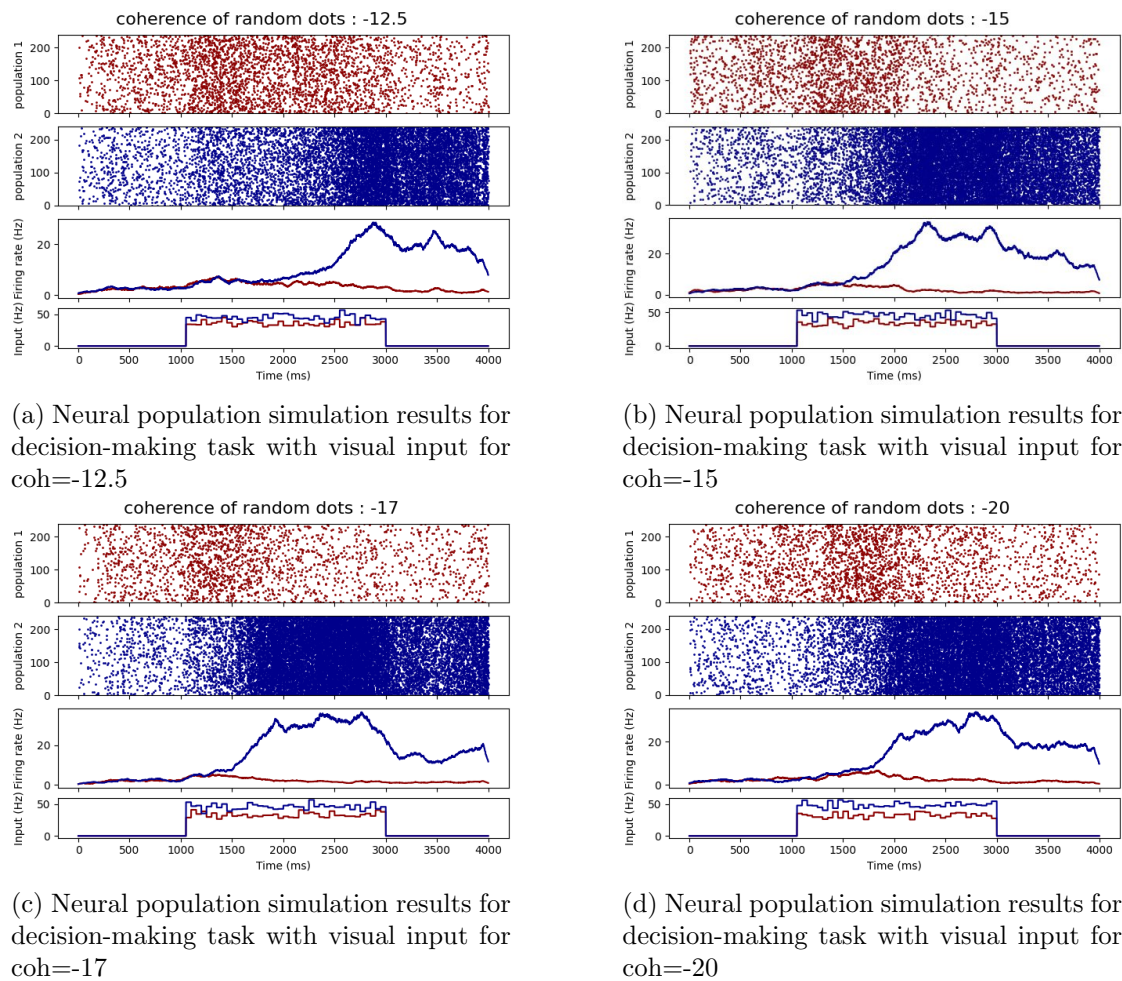


Figure 19: Simulation results for low and negative coherence values (Part 3).

Randomness in Decision-Making ($\text{coh}=0$)

As expected, when the coherence index is zero, the inputs to the two neural populations are very similar, leading to random activation of one population over the other. This randomness reflects real-world scenarios where participants struggle to make accurate decisions when visual inputs are ambiguous.

Behavioral Analysis

Figure 20 shows the percentage of correct responses when the visual input is presented for a fixed duration of 1 second across different coherence indices. As anticipated, the percentage of correct responses increases with higher coherence indices.

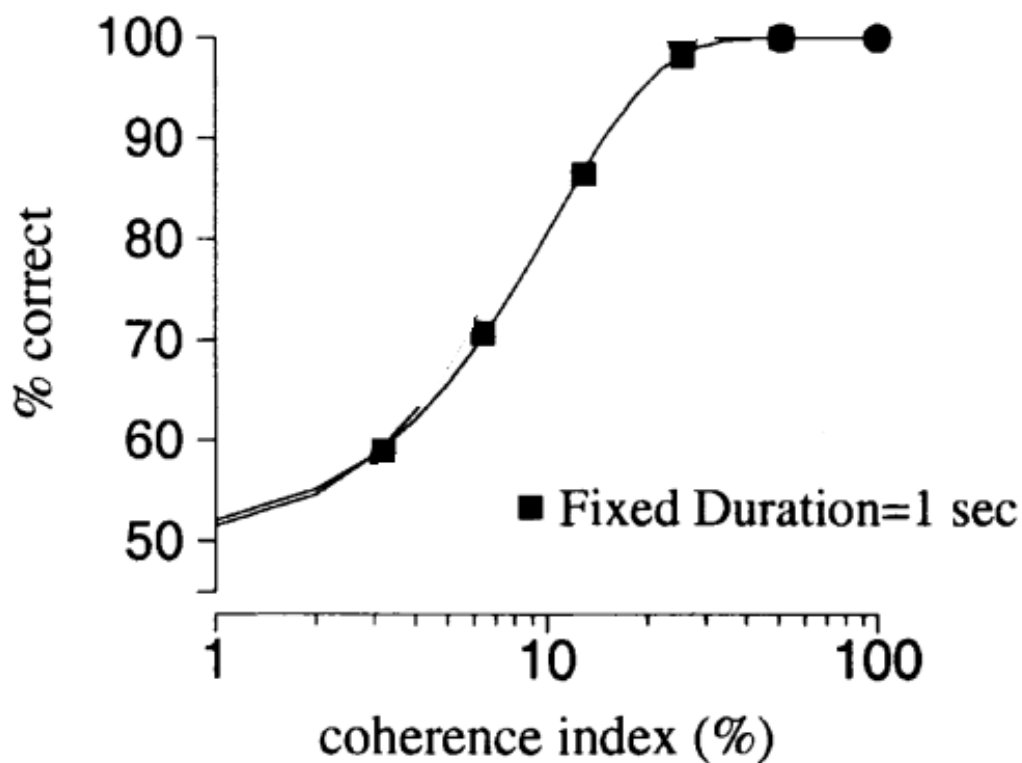


Figure 20: Percentage of correct responses for different coherence indices.

The simulation results demonstrate the correlation between the coherence index of visual stimuli and the decision-making accuracy in a neural population model. As the coherence increases, the neural population corresponding to the correct decision becomes more active, leading to higher accuracy in decision-making. This model effectively replicates cognitive behaviors observed in real-world tasks, providing valuable insights into the neural mechanisms underlying decision-making processes.