

IA, database of known ligands of aminoacyl-tRNA synthetases

Mieczyslaw Torchala · Marcin Hoffmann

Received: 3 July 2007 / Accepted: 3 September 2007 / Published online: 20 September 2007
© Springer Science+Business Media B.V. 2007

Abstract The IA database contains 240 structures of known inhibitors of aminoacyl-tRNA synthetases. Structures can be downloaded in different file formats (mol, sdf, smile, png). The search engine offers possibility of searching for the ligands with a given functional group. Additionally, one can search for ligands that act on selected synthetases and from particular references. The data include information which synthetase a given ligand inhibits together with the inhibition constant (IC_{50}) if known. Database is freely available at <http://ia.bioinfo.pl/>

Keywords Aminoacyl-tRNA synthetases · Aminoacyl-tRNA synthetases inhibitors · Molecular docking · Virtual high throughput screening

Introduction

Aminoacyl-tRNA synthetases (AA-RS) are a group of enzymes that ensure the fidelity of transfer of genetic information from the DNA into the protein [1]. They are found in all living organisms and catalyze the esterification

of a particular tRNA with its cognate amino acid. Their key mode of action is graphically depicted in Fig. 1.

AA-RS fulfill many criteria indispensable for good antibacterial agents [2], among others they are crucial for viability, different for procariotic and eucariotic organism and share a common catalytic site. These and other advantages cause that AA-RS constitute an appealing molecular target for drug design for compounds active against pathogenic bacteria. Moreover it seems possible to create selective drugs which do not act on host aminoacyl-tRNA synthetases.

Inhibition of these enzymes is possible in every stage [3]. In the first stage, substrate-binding can be interrupted by analogues of amino acids. In the second stage it is possible to generate mimetic of the enzyme-bound reaction intermediate (AA-AMP). As the result no AA-tRNA is synthesized so finally it will lead to the interruption of a polypeptide chain elongation and inhibition of cell growth.

Currently, rational drug design often proceeds via structure based virtual screening of possible inhibitors. Sometimes the process is facilitated by a feedback from experimental measurements. In such case the collection of compounds that are known to act as inhibitors facilitates the discovery of agents of a desired activity. Our database of molecules known to inhibit enzymatic activities of aminoacyl-tRNA synthetases gathers experimental data from various sources and can be used to find other molecules which can be potential inhibitors and test their usefulness using methods like virtual screening or molecular docking.

Methods and database overview

We searched for known inhibitors of aminoacyl-tRNA synthetases in articles published from 2001 to 2006, found by PubMed search engine. Reviews published before 2001

M. Torchala · M. Hoffmann (✉)
BioInfoBank Institute, ul. Limanowskiego 24A,
Poznan 60-744, Poland
e-mail: mmh@bioinfo.pl

M. Torchala
Faculty of Physics, Adam Mickiewicz University,
Umultowska 85, Poznan 61-614, Poland

M. Hoffmann
Faculty of Chemistry, Adam Mickiewicz University,
Grunwaldzka 6, Poznan 60-780, Poland

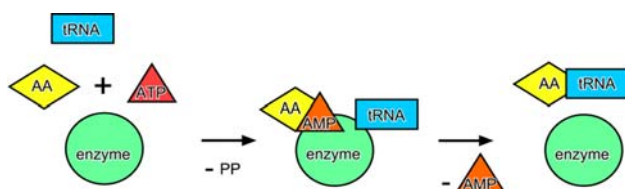


Fig. 1 In the first ATP dependent stage amino acid (AA) is recognized by the enzyme and carboxyl group of amino acid is activated as aminoacyl-AMP (AA-AMP) is formed. In the second stage amino acid moiety is transferred to its cognate tRNA creating aminoacyl-tRNA (AA-tRNA), a substrate for synthesis of proteins

were also taken into account in creation of the database of inhibitors of aminoacyl-tRNA synthetases.

Molecules 0000–0239 were obtained from 22 papers [2–23]. Next Marvin applets [24] were used to create files with 3D structures of these compounds in ‘mol’ format and to convert structures between different file types. Two conformations were chosen for each molecule. The conformation A is the one of the lowest energy in Marvin’s optimization with Dreiding force field [25]. The conformation B is the conformation whose RMSD (root mean square deviation) from the conformer A is the largest in the set of conformations generated by Marvin’s molconvert utility.

The presented database is composed of 240 molecules known from literature to inhibit enzymatic activity of aminoacyl-tRNA synthetases. The geometries of all these molecules are available in two different conformations. The MySQL database is equipped with a search engine to search for molecules having desired functional groups, i.e. possessing a given smile’s substring. It is also possible to define up to two substrings and use %, AND or OR operators. Additionally, one can search for ligands that act on selected synthetases and from particular references. The data include information which synthetase a given ligand inhibits together with the inhibition constant (IC_{50}) if known.

Conclusions

We presented here the database containing the structures of 240 known inhibitors of aminoacyl-tRNA synthetases ready to search and download, as well as experimental information on these inhibitors. The 3D geometries of ligands, after downloading, can be easily used for various computational tests like molecular docking [26–28] or virtual high throughput screening [29–31] experiments. Furthermore, the IA database can be easily expanded and updated to include more data on aminoacyl-tRNA synthetases inhibitors.

Acknowledgements Financial support by European Commission and Polish Ministry for Education and Science is acknowledged (BioSapiens project grant number LSHG-CT-2003-503265, and grant number PBZ-MNiI-2/1/2005). MH thanks the Foundation for Polish Science for support via FOCUS program. The authors thank the anonymous referee for the suggestions on improving the IA database.

References

1. Szymanski M, Deniziak M, Barciszewski J (2000) *Acta Biochim Pol* 47:821
2. Pohlmann J, Brotz-Oesterhelt H (2004) *Curr Drug Targets Infect Disord* 4:261
3. Kim S, Lee SW, Choi EC, Choi SY (2003) *Appl Microbiol Biotechnol* 61:278
4. Sukuru SCK, Crepin T, Milev Y, Marsh LC, Hill JB, Anderson RJ, Morris JC, Rohatgi A, O’Mahony G, Grøtli M, Danel F, Page MGP, Härtlein M, Cusack S, Kron MA, Kuhn LA (2006) *J Comput Aided Mol Des* 20:159
5. Kim SE, Kim SY, Kim S, Kang T, Lee J (2005) *Bioorg Med Chem Lett* 15:3389
6. Farhanullah, Kim SY, Yoon E-J, Choi E-C, Kim S, Kang T, Samrin F, Purie S, Lee J (2006) *Bioorg Med Chem* 14:7154
7. Jarvest RL, Erskine SG, Forrest AK, Fosberry AP, Hibbs MJ, Jones JJ, O’Hanlon PJ, Sheppard RJ, Worby A (2005) *Bioorg Med Chem Lett* 15:2305
8. Critchley IA, Young CL, Stone KC, Ochsner UA, Guiles J, Tarasow T, Janjic N (2005) *Antimicrob Agents Chemother* 49:4247
9. Petraitis V, Petraitiene R, Kelaher AM, Sarafandi AA, Sein T, Mickiene D, Bacher J, Groll AH, Walsh TJ (2004) *Antimicrob Agents Chemother* 48:3959
10. Kanamaru T, Nakano Y, Toyoda Y, Miyagawa KI, Tada M, Kaisho T, Nakao M (2001) *Antimicrob Agents Chemother* 45:2455
11. Winum JY, Scozzafava A, Montero JL, Supuran CT (2005) *Med Res Rev* 25:186
12. Schimmel P, Tao J, Hill J (1998) *FASEB J* 12:1599
13. Yu XY, Hill JM, Yu G, Yang Y, Kluge AF, Keith D, Finn J, Gallant P, Silverman J, Lim A (2001) *Bioorg Med Chem Lett* 11:541
14. Banwell MG, Crasto CF, Easton CJ, Forrest AK, Karoli T, March DR, Mensah L, Nairn MR, O’Hanlon PJ, Oldham MD, Yue W (2000) *Bioorg Med Chem Lett* 10:2263
15. Jarvest RL, Berge JM, Houge-Frydrych CS, Mensah LM, O’Hanlon PJ, Pope AJ (2001) *Bioorg Med Chem Lett* 11:2499
16. Yu XY, Finn J, Hill JM, Wang ZG, Keith D, Silverman J, Oliver N (2004) *Bioorg Med Chem Lett* 14:1343
17. Tandon M, Coffen DL, Gallant P, Keith D, Ashwell MA (2004) *Bioorg Med Chem Lett* 14:1909
18. Hurdle JG, O’Neill AJ, Chopra I (2005) *Antimicrob Agents Chemother* 49:4821
19. Jarvest RL, Armstrong SA, Berge JM, Brown P, Elder JS, Brown MJ, Copley RC, Forrest AK, Hamprecht DW, O’Hanlon PJ, Mitchell DJ, Rittenhouse S, Witty DR (2004) *Bioorg Med Chem Lett* 14:3937
20. Jarvest RL, Berge JM, Brown P, Houge-Frydrych CS, O’Hanlon PJ, McNair DJ, Pope AJ, Rittenhouse S (2003) *Bioorg Med Chem Lett* 13:1265
21. Qiu X, Janson CA, Smith WW, Green SM, McDevitt P, Johanson K, Carter P, Hibbs M, Lewis C, Chalker A, Fosberry A, Lalonde J, Berge J, Brown P, Houge-Frydrych CS, Jarvest RL (2001) *Protein Sci* 10:2008

22. Finn J, Mattia K, Morytko M, Ram S, Yang Y, Wu X, Mak E, Gallant P, Keith D (2003) *Bioorg Med Chem Lett* 13:2231
23. Lee J, Kim SE, Lee JY, Kim SY, Kang SU, Seo SH, Chun MW, Kang T, Choi SY, Kim HO (2003) *Bioorg Med Chem Lett* 13:1087
24. Marvin applets. Retrieved from <http://www.chemaxon.com/6/13/2007>
25. Mayo SL, Olafson BD, Goddard WA III (1990) *J Phys Chem* 94:8897
26. Moustakas DT, Lang PT, Pegg S, Pettersen E, Kuntz ID, Brooijmans N, Rizzo RC (2006) *J Comput Aided Mol Des* 20:601
27. Bursulaya BD, Totrov M, Abagyan R, Brooks CL (2003) *J Comput Aided Mol Des* 17:755
28. Eitner K, Gaweda T, Hoffmann M, Jura M, Rychlewski L, Barciszewski J (2007) *J Chem Inf Model* 47:695
29. Zavodszky MI, Sanschagrin PC, Kuhn LA, Korde RS (2002) *J Comput Aided Mol Des* 16:883
30. Sippl W (2002) *J Comput Aided Mol Des* 16:825
31. Plewczynski D, Spieser SAH, Koch U (2006) *J Chem Inf Model* 46:1098