

Intramolecular C-H--O interaction between lactam oxygen and N-alkyl protons

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We report evidence of an unusual C-H--O interaction between an α -methylene hydrogen of the alkylamine chain of substituted (N,N-dimethylamino)propyl-azetidinones, substituted (N,N-dimethylamino)propyl-thiazolidinones and substituted (N,N-dimethylamino)propyl-thiazinone and the lactam carbonyl oxygen. NMR analysis results, supported by molecular mechanic predictions, were in agreement with ab initio calculations. The observed interaction shorting the nitrogen—nitrogen distance in the H_1 -histamine antagonist, 2-(4-methylphenyl)-3-[3-(N,N-dimethylamino)propyl]-1,3-thiazolidin-4-one (1) could explain its fitting with the H_1 -antihistaminic pharmacophoric model and the high antihistaminic activity. © 2001 by Elsevier Science Inc.

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INTRODUCTION

Hydrogen bonding, especially the intramolecular variety, often plays a significant role in influencing the molecule conformation. Although research has discovered hydrogen bonding by the C-H group, few types of C-H bonds have been found that are polarized enough to form hydrogen bonds ^{1,2}. During a study on a recently synthesized series of 2-(substituted-phenyl)-3-[3-(N,N-dimethylamino) propyl]- 1,3-thiazolidin-4-ones³⁻⁵ active as H₁-histamine antagonists, ¹H NMR data showed that the geminal protons of the three methylene groups, α , β , and γ , belonging to the alkylaminic side chains of series are not equivalent and are coupled to each other in a gaucheanti conformation (1, Figures 1 and 2). The chemical shift

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difference (\sim 0.9 ppm) between the geminal α -CH₂ protons is very large and could be ascribed to an unusual intramolecular interaction, such as a hydrogen bond, between the amidic oxygen of the thiazolidinone ring and a hydrogen of the α -CH₂.

RESULTS

Because of the importance of the stabilizing role of such a CH--O interaction, a study was performed to explain and generalize what was observed in the thiazolidinone derivatives. Compounds 2, 3 and 5-12 with different lactam rings, chain length, and substituents were synthesized to investigate the effects of these differences on the CH--O bond strength (Figure 1). The experimental section reports general synthetic procedures and characterization of all new compounds on the basis of physicochemical and spectral data and elemental analysis. Table 1 lists the chemical shift values of alkylaminic chain protons and the Ha-Hb differences and shows that methylene protons are equivalent in the achiral compounds, whereas they are diastereotopic in the chiral ones. Moreover, the H_a-H_b difference appears depending on cycle width. ¹³C-¹H heteronuclear long-range coupling investigation performed on chiral and nonchiral thiazolidinones showed strong coupling between the carbonyl carbon and the α -CH₂ signals. COSY and NOESY experiments on the chiral thiazolidinones showed the interactions of the low field hydrogen (H_a) of α -CH₂ with the low field hydrogen of 5-CH₂ AB system of the thiazolidinone ring (H_{Δ}) and with the hydrogens of γ -CH₂. Furthermore the heterocyclic C₂-H showed a strong interaction with the low field hydrogen of β-CH₂ system (H_d). These results suggest that N-alkyl chain of chiral thiazolidinones stays in a α - β gauche β - γ anti-conformation where the dihedral angle between carbonyl group and one of α -CH₂ hydrogens is different from the other one. In the nonchiral compounds, the N-alkyl chain lies in the same conformation of the chiral compounds, but the

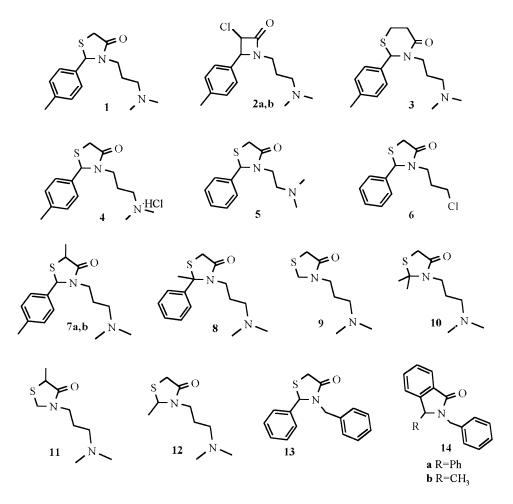


Figure 1. Thiazolidinone, azetidinone and thiazinone structures synthesized to study C-H--O interaction.

 α -CH $_2$ hydrogens give rise only to a signal owing to the equivalence of two sides of the thiazolidinone plane. In this situation the 1 H NMR signals will be the average of signals arising from equivalent populations of enantiotopic conformers. Figure 2 shows the best conformation of 1 that agrees with the NMR results.

DISCUSSION

To investigate these results and to justify the origin of the large H_a-H_b nonequivalence, the geometries of thiazolidinones 1 and 5-12 were generated within SYBYL⁶ starting from the relevant internal coordinates determined for 2-(4-nitrophenyl)-3-[3-(N,N-dimethylamino) propyl]-1,3-thiazolidin-4-one hydrochloride by X-ray diffraction. $^{\scriptscriptstyle 1}$ X-ray data showed that the H-C $_{\alpha^-}$ N_3 - C_4 =O atoms lie on the same plane, that the C_4 =O--H- C_α distance is 2.33 Å, shorter than van der Waals radii, and that the C₄=O--H angle is 110°. Molecular mechanics calculations of the low-energy conformations of all derivatives were carried out by TRIPOS force field6 of SYBYL. Geometry optimizations were realized with the semiempirical quantummechanical method AM1,7 available in the MOPAC program8 (default settings; keyword: "MMOK"). Two low energy conformers (I and II) of 1 and 5-12, showed a gauche-anti conformation of the alkylamine chain and the $H-C_{\alpha}-N_3-C_4$ \longrightarrow O

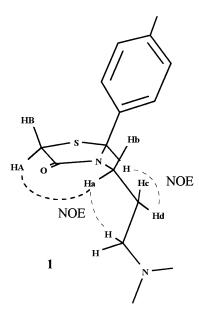


Figure 2. NOE interaction detected by COSY and NOESY experiments on 1.

Table 1. ¹H NMR data of α,β and γ -CH₂ protons of 1-3 and 5-13 in C₆D₆ and 25°C (multiplet center) and of 4 in D₂O.

H _a -H _b difference	cpd	$lpha ext{-CH}_2$		$\beta\text{-CH}_2$		γ -CH $_2$		
		$\overline{\mathrm{H_{a}}}$	H_{b}	$\overline{\mathrm{H_{c}}}$	$H_{\rm d}$	$H_{\rm e}$	H_{f}	
0.96	1	3.70	2.74	1.54	1.41	2.03	1.91	
0.59	2a	3.48	2.89	1.65		2.24		
0.55	2b	3.46	2.89	1.59		2.16		
1.63	3	4.31	2.68	1.88	1.69	2.35	2.02	
0.36	4	3.43	3.07	1.69		2.94		
1.25	5	3.80	2.55	2.21	1.97			
0.85	6	3.44	2.59	1.54	1.44	2.97	2.91	
0.85	7a	3.60	2.75	1.	57	2	18	
0.90	7b	3.65	2.75	1.	64	2.1	18	
0.64	8	3.55	2.91	1.86	1.70	2.0	05	
0.00	9	3.	13	1.41		2.01		
0.00	10	3.	3.14		1.68		2.09	
0.12	11	3.20	3.08	1.	40	1.9	99	
0.69	12	3.53	2.84	1.52	1.37	2.04	1.96	
1.84	13	4.66	2.82					
1.75	$14a^a$	5.42	3.67					
1.05	$14b^a$	5.20	4.15					

^aSpectrum recorded in carbon tetrachloride.⁹

atoms lying on the same plane in agreement with an attractive interaction between the carbonyl oxygen and an α-CH₂ hydrogen (Figure 3). When X = Y, the two thiazolidinone planes are equal, I and II have the same energy and are equally probable, and ¹H NMR spectrum shows only one α -CH₂ signal. When $X \neq Y$, $H_a \neq H_b$, the two low-energy minimized conformers, I and II, contribute differently to the H_a and H_b shifts, and ¹H NMR spectrum shows two α -CH $_2$ signals. According to the energy difference in 1 ($\Delta E = 1 \text{ kcal/mol}$; I constitutes approximately 80%, and II 20%), the dominance of I agrees with the NOESY results (Figure 2) and X-ray data.3 NMR experiments performed in a temperature range of 25-50 °C showed no temperature-dependent chemical shift, reflecting a stable population distribution. Moreover, nonprotic solvents, such as benzene, seem to increase the reported conformational preference, whereas water weakens it, pushing the two α -CH₂ signals (1 H NMR data of 4) very close. All data are in agreement with the reported C-H--O interaction.² In conclusion, the hydrogenoxygen interaction, resulting in a low field H_a, plays the same role in chiral and achiral thiazolidinones stabilizing low-energy conformers. In the chiral compounds these two conformers are pseudo-diastereoisomers, whereas in nonchiral compounds they are enantiotopic forms. This attractive interaction is not affected by substituent effects, steric hindrance, or shieldingdeshielding effects, so it appears similar to an electrostatic interaction as in a hydrogen bond. In fact, in 9 as in 10, the α -CH₂ signals lie at 3.13 and at 3.14 δ , whereas in **8** and **12** the C-H_a signals lie at 3.55 and 3.53 δ excluding any inductive and steric effect on the alkyl chain chemical shifts.8 2-Substituted and unsubstituted thiazolidinones as azetidinones 2a, b and thiazinone 3, showed the same N-alkylaminic chain conformation. Figure 4 shows a generic lactam and the optimized structures of 1, 2, and 3 obtained from the SYBYL program. In the azetidinones 2 as in the thiazinone 3 the α -CH₂ hydrogens are diastereotopic and, the calculated angles C_{α} - N_3 - C_4 of 1, C_{α} - N_1 - C_2 of **2(a, b)** and C_{α} - N_3 - C_4 of **3** were, respectively, 123°, 130°, and 119° (Figure 4). The hydrogen-oxygen distances were 2.3, 2.8, and 2.2 Å showing that the thiazinone geometry could allow the best hydrogen interaction. According to this, the chemical shift difference between the H_a and H_b in 3 is larger (1.65 δ) than in the corresponding five and four member rings of 1 and 2 (0.96 and 0.65 δ), respectively. On the basis of these results, it seems reasonable that azetidinone, thiazolidinone, and thiazinone systems sufficiently influence the polar-

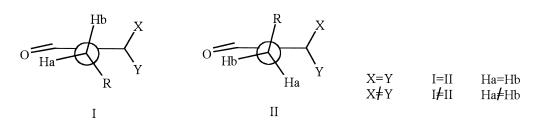


Figure 3. Two low energy thiazolidinone conformers contributing differently to the α -CH₂ shifts.

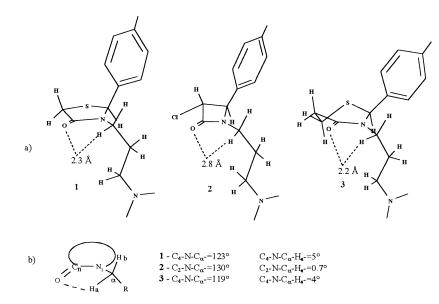


Figure 4. (a) 1, 2 and 3 structures represent the minimum energy conformers obtained from SYBYL package. (b) Generic N-alkyl lactam structure. C_4 -N- C_{α} and C_4 -N- C_{α} -H_a didehral angles are reported.

ization of the C-H bond¹⁰⁻¹² acting as electron-withdrawing group on the side N-alkylic chain and allow the development of an attractive intramolecular interaction between C_{\alpha}-H_{\alpha} and the carbonyl oxygen. To gain further insight into the electronic origin of the behavior of these molecules, we performed a number of quantum mechanical computations by the Gaussian 98 package¹³ using the very reliable PBE0 density functional¹⁴ and the 6-311G(d,p) basis set,15 which provide essentially converged results at the DFT level.¹⁶ In a first step, we have fully optimized the geometries of 1 and of the two model compounds 3-methyl-1,3-thiazolidin-4-one (15a) and 3-ethyl-1,3-thiazolidin-4-one (15b) as shown in Figure 5. In all cases, the most stable conformer has one of the α -CH₂ hydrogens in the plane of the thiazolidinone ring and a short O-H distance (2.333, 2.328, and 2.306 Å, respectively), which is quite close to the molecular mechanics result. Analogous computations for 15b in aqueous solution by the C-PCM model¹⁷ show that environmental effects do not alter the structure of these compounds significantly. Next, the net atomic charges of 15b have been obtained by a number of different procedures, including Mulliken^{18,19} and natural bond orbital (NBO)²⁰ population analyses, fitting of the molecular electrostatic potential (MEP)^{21,22} and atomic polar tensor (APT)(Table 2).²³ Whereas the different procedures give widely different charges, all the methods agree in indicating that the α -methylene unit is more positive than the β -methyl group. Moreover, a significant Coulombic interaction is operative between the oxygen atom and the α-CH_a hydrogen. The NBO analysis also allows a dissec-

 $\begin{array}{ccc}
S \nearrow O & S \nearrow O \\
CH_3 & CH_3
\end{array}$ 15a 15b

Figure 5. 1,3-thiazolidinones used for quantum mechanical computations.

tion of the interactions between different localized orbitals into physical significant terms. In the present case, the most significant (and conformation-dependent) interaction occurs between an oxygen lone-pair and the C-Ha σ^* bond orbital. This interaction thus contributes to further stabilize conformers with the C-H_a bond lying in the ring plane. Further studies will be performed on this topic. This C-H--O interaction, which is similar to a hydrogen bond, stabilizes a gauche-anti spatial arrangement of the alkylaminic chain of 2-(substituted-phenyl)-3-[3-(N,N-dimethylamino)propyl]-1,3-thiazolidin-4-one series, thereby reducing the distance between the two nitrogens; this geometry can dramatically affect H₁-histamine antagonism.^{24,25} H₁-histamine antagonist thiazolidinones,3,5,26 as the majority of potent antihistamines, are tertiary amines and exist at physiological pH with the nitrogen of the side chain protonated. Nevertheless, when they interact with a receptor, 26,27 it can be a nonaqueous milieu where the hydrogen-bond constrained conformation is reasonable. Better fitting with the receptor site can be responsible for the high H₁-histamine receptor affinity.

MATERIALS AND METHODS

The IR spectra were taken on a Perkin-Elmer 1760-X IFT (Perkin Elmer Corp., Norwalk, CT USA) spectrophotometer in potassium bromide. The ¹H (500 MHz) and ¹³C (125 MHz) NMR spectra were recorded on a Fourier-transform Bruker

Table 2. Net atomic charges of 15b obtained by different procedures

	Mulliken	ESP	NBO	APT
qO	-0.388	-0.514	-0.602	-0.794
qN	-0.420	-0.249	-0.474	-0.802
qH_{a}	0.162	0.056	0.234	0.162
qCH ₂	0.141	0.254	0.226	0.351
qCH ₃	0.066	-0.062	0.018	-0.002

spectrometer AMX 500 equipped with a Bruker X-32 computer, using the UXNMR software package (Unix Software (ver. 950901) developed by Bruker Analytische Messtechnik GmbH, Rheinstetten, Germany). Standard pulse sequences were employed for DEPT and COSY. Phase sensitive NOESY spectra were measured with a mixing time $t_{\rm m}$ 800 ms, while HMQC and HMBC were optimized for $^1J_{\rm C-H}=135$ Hz and $^{2,3}J_{\rm C-H}=8$ and 10 Hz, respectively. To enhance the results, nitrogen was bubbled through the sample to remove oxygen. Electron impact (ei) mass spectra were obtained at 70 eV on a VG ZAB 2F spectrometer. The purity of the compounds was checked by ascending tlc on Merck's precoated silica-gel plates (0.25 mm) (Merck Eurolab, Milano, Italy).

Preparation of 1,3-thiazolidin-4-ones (1, 5–6, 8–13) and of 1,3-thiazin-4-one (3).

General Procedure

An equimolar mixture (0.01 mol) of aldehyde or ketone and appropriate alkylamine (0.01 mol) in dry benzene (50 ml) was refluxed until no more water was collected in a Dean-Stark water separator. α -Mercaptoacetic acid (0.01 mol) (β -mercaptopropionic acid for compound 3) was added, dropwise, to this crude mixture, and the reaction was carried out at reflux temperature until stoichiometric water was collected. The mixtures, cooled and evaporated *in vacuo*, afforded the crude compounds as pale yellow oils that were purified by TLC (Silica gel, Merck 0.5 mm; eluent: n-exane:ethanol:TEA, 10: 3:2, vv). The compound 13 was collected as white powder after purification.

2-(4-Methylphenyl)-3-[2-(*N*,*N***-dimethylamino**)**propyl]-1, 3-thiazolidin-4-one** (1): 1 H NMR: 6.98 (2H, d; J = 8.1 Hz), 6.87 (2H, d; J = 8.1 Hz), 5.35 (1H, d; J = 1.9 Hz), 3.70 (1H, dd; J = 14.2, 6.4 Hz), 3.48 (1H, dd; J = 15.1, 1.9 Hz), 3.35 (1H, d; J = 15.1 Hz), 2.74 (1H, ddd; J = 14.2, 6.4 Hz), 2.03 (1H, ddd; 12.5, 5.3, 6.4 Hz), 2.01 (3H, s), 1.91 (1H, ddd; 12.5, 5.3, 6.4 Hz), 1.89 (6H, s), 1.54 (1H, m), 1.41 (1H, m); 13 C NMR: 178.8 (s), 38.7 (s), 137.5 (s), 129.6 (2C, d), 127.4 (2C, d), 67.0 (d), 63.5 (t), 56.8 (t), 45.1 (2C, q), 41.4 (t), 32.7 (t), 25.2 (t), 20.9 (q); mass spectrum, m/e M+ 278. *Anal.* Calcd. for $C_{15}H_{22}N_2OS$: C, 64.71; H, 7.96; N, 10.06. Found: C, 64.69; H, 7.94; N, 10.07.

2-(4-Methylphenyl)-3-[3-(*N,N***-dimethylamino)propyl]-1, 3-thiazin-4-one** (3): 1 H NMR: 7.11 (2H, d; J = 8.0 Hz), 6.95 (2H, d; J = 8.0Hz), 5.65 (1H, s), 4.31 (1H, ddd; J = 13.1, 8.2, 7.9), 2.68 (1H, dt; J = 13.1, 7.5 Hz), 2.65 (1H, ddd; J = 17.5, 10.9, 6.35 Hz), 2.56 (1H, ddd; J = 17.5, 6.3, 6.4 Hz), 2.47 (1H, ddd; J = 13.6, 10.9, 6.1), 2.35 (1H, ddd; J = 12.1, 8.4, 6.5), 2.10 (3H, s), 2.02 (7H, s+m), 1.95 (1H, m), 1.88 (1H, m), 1.69 (1H, ddt; J = 13.4, 6.7, 6.5); 13 C NMR: 170.77 (s), 140.6 (d), 129.7 (2C, d), 129.6 (2C, d), 61.01 (d), 54.87 (t), 44.82 (t), 43.25 (2C, q), 33.15 (t), 24.13 (t), 20.39 (t), 19.11 (q); mass spectrum, m/e M+ 292. *Anal.* Calcd. for $C_{16}H_{24}N_2OS$: C, 65.71; H, 8.27; N, 9.58. Found: C, 65.90; H, 8.24; N, 9.61.

2-Phenyl-3-[2-(*N*,*N***-dimethylamino**)**ethyl]-1,3-thiazolidin-4-one** (5): 1 H NMR: 7.02 (5H, m), 5.67 (1H, d; J = 1.7 Hz), 3.80 (1H, dt; J = 14.2, 6.4 Hz), 3.45 (1H, dd; J = 15.1, 1.7 Hz), 3.36 (1H, d; J = 15.1 Hz), 2.55 (1H, ddd; J = 14.2, 6.4 Hz), 2.21 (1H, ddd; 12.5, 5.3, 6.4 Hz), 1.97 (1H, ddd; 12.5, 5.3, 6.4 Hz), 1.90 (6H, s); 13 C NMR: 170.77 (s), 140.6 (s), 129.0 (2C,

d), 128.8 (d), 127.3 (2C, d), 63.7 (d), 57.0 (t), 45.3 (2C, q), 40.4 (t), 32.6 (t) mass spectrum, m/e M+ 250. *Anal.* Calcd. for $C_{13}H_{18}N_2OS$: C, 62.37; H, 7.25; N, 11.14. Found: C, 64.20; H, 7.23; N, 11.16.

2-Phenyl-3-(3-chloropropyl)-1,3-thiazolidin-4-one (6): 1 H NMR: 7.01 (3H, d+t; J = 8.8 Hz), 6.94 (2H, d; J = 8.7 Hz), 5.12 (1H, d; J = 1.8 Hz), 3.44 (1H, ddd; J = 13.8, 7.3, 7.0 Hz), 3.35 (1H, dd; J = 15.7, 1.7 Hz), 3.26 (1H, d; J = 15.7 Hz), 2.97 (1H, ddd; J = 10.5, 6.3, 2.0 Hz), 2.91 (1H, ddd; J = J = 10.5, 6.3, 1.8 Hz), 2.59 (1H, dt; J = 13.8, 7.3, 7.0 Hz), 1.54 (1H, seven peak multiplet), 1.44 (1H, seven peak multiplet); 13 C NMR: 170.83 (s), 140.11 (s), 129.10 (3C, d), 127.26 (2C, d), 63.62 (d), 42.15 (t), 40.81 (t), 32.46 (t), 30.11 (t); mass spectrum, m/e M+ 255, (M+2) 257. *Anal.* Calcd. for $C_{12}H_{14}$ CINOS: C, 56.35; H, 5.52; N, 5.48. Found: C, 56.32; H, 5.50; N, 5.47.

3-(N,N-dimethylamino)propyl-1,3-thiazolidin-4-one (9): 1 H NMR: 3.75 (2H, bs), 3.21 (2H, bs), 3.13 (2H, t; J = 6.9 Hz), 2.01 (2H, t; J = 6.9 Hz), 1.99 (6H, s), 1.41 (2H, p; J = 6.9 Hz); 13 C NMR: 170.25 (s), 56.79 (t), 47.21 (t), 45.41 (q), 45.34 (q), 42.63 (t), 32.18 (t), 25.57 (t); mass spectrum, m/e M+ 188. *Anal.* Calcd. for $C_8H_{16}N_2OS$: C, 51.03; H, 8.57; N, 14.88. Found: C, 51.23; H, 8.55; N, 14.86.

2-Dimethyl-3-(*N*,*N***-dimethylamino**)**propyl-1,3-thiazolidin-4-one** (**10**) 1 H NMR: 3.29 (2H, s), 3.14 (2H, dd; J = 7.8 Hz), 2.09 (2H, t; J = 6.9 Hz), 2.03 (6H, s), 1.68 (2H, m), 1.20 (6H, s); 13 C NMR: 170.02 (s), 66.70 (s) 57.22 (t), 45.28 (2C, q), 40.09 (t), 32.11 (t), 30.23 (2C, q), 27.39 (t); mass spectrum, m/e M+ 216. *Anal.* Calcd. for $C_{10}H_{20}N_{2}OS$: C, 55.52; H, 9.32; N, 12.95. Found: C, 55.48; H, 9.30; N, 12.96.

2-Methyl-3-[3-(*N,N***-dimethylamino)propyl]-1,3-thiazolidin-4-one** (**12**): 1 H NMR: 4.21 (1H, dq; J = 5.9, 1.6 Hz), 3.53 (1H, ddd; J = 13.4, 7.3, 2.0 Hz), 3.27 (1H, dd; J = 15.9, 1.6 Hz), 3.16 (1H, d; J = 15.9 Hz), 2.84 (1H, ddd; J = ; J = 13.4, 7.3, 2.0 Hz), 2.04 (1H, m) 1.99 (6H, s), 1.96 (1H, m), 1.52 (1H, m), 1.37 (1H, m), 1.02 (3H, d; J = 5.9 Hz); 13 C NMR: 171.50 (s), 56.84 (t), 56.42 (d), 45.28 (2C, q), 40. 2 (t), 32.01 (t), 25.71

(t), 23.02 (q); mass spectrum, m/e M+ 202. *Anal.* Calcd. for $C_9H_{18}N_2OS$: C, 53.43; H, 8.97; N, 13.85. Found: C, 53.48; H, 8.99; N, 13.81.

2-Phenyl-3-benzyl-1,3-thiazolidin-4-one (13): 1 H NMR: 6.50 (8H, m), 6.33 (2H, m), 4.66 (1H, d; J = 14.7 Hz), 4.61 (1H, d; J = 1.7 Hz), 2.95 (1H, dd; J = 15.4, 1.7 Hz), 2.82 (1H, d; J = 14.7 Hz), 2.79 (1H, d; J = ; J = 15.4); 13 C NMR: 170.71 (s), 140.20 (s), 136.33 (s), 129.05 (2C, d), 128.88 (d), 128.58 (2C, d), 128.40 (d), 128.00 (2C,d), 127.30 (2C,d), 62.53 (d), 46.20 (t), 32.56 (t); mass spectrum, m/e M+ 269; Mp 142-3 $^{\circ}$ C. *Anal.* Calcd. for C₁₆H₁₅NOS: C, 71.34 ; H, 5.61 ; N, 5.20: Found: C, 71.35 H, 5.59; N, 5.18.

Preparation of cis and trans 3-chloro-[3-(N,N-dimethylamino)propyl]-4-(4-methylphenyl)-azetidin-2-ones (2a, 2b)

An equimolar mixture (0.01 mol) of *p*-tolualdehyde and *N*,*N*-dimethyl-alkylamine (0.01 mol) in dry benzene (50 ml) was refluxed until no more water was collected in a Dean-Stark water separator. The crude mixture was extracted with a HCl 2N (100 ml). The aqueous line, alkalinized with NaOH and extracted with diethyl ether (50 ml), yielded a pale-yellow oil that was diluted with DMF (50 ml). Chloroacetyl chloride (0.01 mol) was added to the solution and the mixture, refluxed for 2 h, and evaporated *in vacuo*, which yielded the crude *trans* (2a) and *cis* (2b) azetidinones. 2a and 2b were separated and purified by TLC (Silica gel, Merck 0.5 mm; eluent: *n*-exane: ethanol:TEA, 10:3:2, vv)

3-Chloro-1-[3-(N,N-dimethylamino)propyl]-4-4-methylphenyl-azetidin-2-ones (2a. 2b): trans-2a- ¹H NMR: 7.20 (2H, d; J = 7.9), 7.16 (2H, d; J = 7.9), 4.50 (1H, d; J = 1.6),4.44 (1H, d; J = 1.6), 3.48 (1H, dt; J = 7.0, 14.0), 2.89 (1H, dt; J = 7.0, 14.0, 2.33 (3H, s), 2.24 (2H, m), 2.14 (6H, s), 1.65 (2H, m); ¹³C NMR: 163.62 (s), 139.44 (s), 132.01 (s), 129.83 (2C, d), 126.42 (2C, d), 66.03 (d), 63.01 (d), 56.64 (t), 45.15 (2C, q), 39.33 (t), 25.41 (t), 21.05 (q); mass spectrum, m/e M+ 280, (M+2) 282. Anal. Calcd. for $C_{15}H_{21}CIN_2O$: C, 64.16; H, 7.54; N, 9.98. Found: C, 64.61; H, 7.55; N, 9.93. *cis-2b* ¹H NMR: 7.14 (2H, d; J = 7.9), 7.08 (2H, d; J = 7.9), 5.01 (1H, d; J = 4.8), 4.86 (1H, d; J = 4.8), 3.46 (1H, dt; J = 7.0, 14.0), 2.91 (1H, dt; J = 7.0, 14.0), 2.29 (3H, s), 2.16 (2H, m), 2.07(6H, s), 1.59 (2H, m) ¹³C NMR: 160.51 (s), 140.32 (s), 128.13 (s), 127.81 (2C, d), 125.45 (2C, d), 66.01 (d), 58.02 (d), 50.93 (t), 44.63 (2C, q), 40.22 (t), 25.49 (t), 21.09 (q); mass spectrum, m/e M+ 280, (M+2) 282. Anal. Calcd. for C₁₅H₂₁ClN₂O: C, 64.16; H, 7.54; N, 9.98. Found: C, 64.62; H, 7.54; N, 9.91.

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