



## Docking-based virtual screening for ligands of G protein-coupled receptors: Not only crystal structures but also *in silico* models

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### ABSTRACT

G protein-coupled receptors (GPCRs) regulate a wide range of physiological functions and hold great pharmaceutical interest. Using the  $\beta_2$ -adrenergic receptor as a case study, this article explores the applicability of docking-based virtual screening to the discovery of GPCR ligands and defines methods intended to improve the screening performance. Our controlled computational experiments were performed on a compound dataset containing known agonists and blockers of the receptor as well as a large number of decoys. The screening based on the structure of the receptor crystallized in complex with its inverse agonist carazolol yielded excellent results, with a clearly delineated prioritization of ligands over decoys. Blockers generally were preferred over agonists; however, agonists were also well distinguished from decoys. A method was devised to increase the screening yields by generating an ensemble of alternative conformations of the receptor that accounts for its flexibility. Moreover, a method was devised to improve the retrieval of agonists, based on the optimization of the receptor around a known agonist. Finally, the applicability of docking-based virtual screening also to homology models endowed with different levels of accuracy was proved. This last point is of uttermost importance, since crystal structures are available only for a limited number of GPCRs, and extends our conclusions to the entire superfamily. The outcome of this analysis definitely supports the application of computer-aided techniques to the discovery of novel GPCR ligands, especially in light of the fact that, in the near future, experimental structures are expected to be solved and become available for an ever increasing number of GPCRs.

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### 1. Introduction

G protein-coupled receptors (GPCRs) constitute the largest superfamily of human membrane signaling proteins. They are implicated in a vast array of physiological functions and pathological conditions and, thus, are highly pursued as targets for pharmacological intervention [1,2].

Due to difficulties inherent the crystallization of GPCRs, for years bovine rhodopsin has been the only member of the superfamily with an experimentally elucidated three-dimensional structure, and has been employed as a template for the construction of three-dimensional (3D) homology models. More recently, however, scientific breakthroughs yielded to the solution of the crystal structures of a few additional receptors, including the  $\beta_2$ -adrenergic receptor ( $\beta_2$ -AR), while structures of additional receptors are expected to be solved in the near future [2,3].

In this work, using the  $\beta_2$ -AR as a case study, we investigated the applicability of crystal structures and homology models endowed with different levels of accuracy to the identification of GPCR ligands, and defined methods intended to improve screening performance. We added a pool of known binders of the receptor to a large set of decoy compounds, and subsequently analyzed the ability of a series of controlled docking-based virtual screening experiments to prioritize ligands over decoys.

In particular, we: (a) assessed the excellent results attainable with the crystal structure of the receptor in complex with the inverse agonist carazolol [4,5]; (b) devised a method that managed to further improve the results through the generation of an ensemble of alternative conformations of the receptor that accounts for its flexibility – general importance: very rarely multiple crystal structures of a GPCR are available and can be used to account for its flexibility; (c) defined a method to invert the tendency of virtual screening to prioritize blockers over agonists, by optimizing the structure of the receptor around bound agonists – general importance: to date, no GPCR has been crystallized in complex with an agonist; (d) assessed the feasibility of the use of homology models in lieu of experimental structures, in the absence of the latter –

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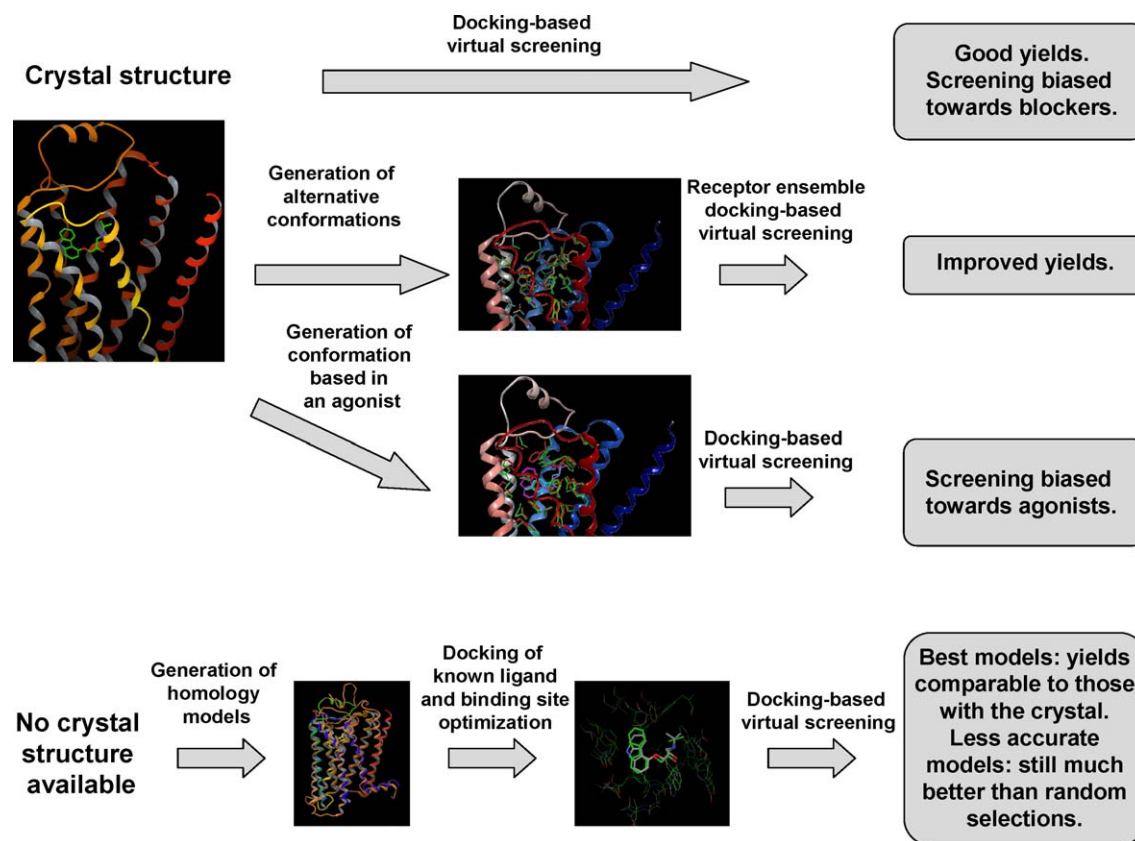


Fig. 1. Flowchart representation of the different procedures presented in this study.

general importance: for the vast majority of GPCRs crystal structures are not available. For this last point, we employed three rhodopsin-based  $\beta_2$ -AR homology models, endowed with different levels of structural accuracy, that we have recently published [6]. A flowchart illustrating the different points of the study is shown in Fig. 1.

## 2. Materials and methods

### 2.1. Compound dataset

The compound dataset used for the virtual screening experiments was composed of a set of 60  $\beta_2$ -AR ligands (29 agonists and 31 blockers) with  $pK_i$  values above 5, which we had collected from the literature in a previous work [7], and 55,806 decoys, presumably inactive at the  $\beta_2$ -AR, extracted from the ZINC Database [8,9]. After downloading the subset of lead-like compounds (1,830,871 molecules), a diverse pool of it was selected with the help of the QuaSAR-Cluster module of MOE [10]. In particular, the clustering was performed through the following operations: (a) all compounds were sorted in alphabetical order, according to their names; (b) 184 2D descriptors were calculated with the QuaSAR-Descriptor function; (c) a Principal Components Analysis (PCA) was performed to reduce the number of variables to five components; (d) finally, the database was clustered into 55,806 groups with the QuaSAR-Cluster function and the first listed compound of each cluster was selected and added to the diverse subset of decoys.

Ligands and decoys were then subjected to an automatic preparation process, performed with the LigPrep tool of the Schrödinger package [11], generating all protonation and tautomeric states available within a pH range of  $7.0 \pm 2.0$ .

### 2.2. Protein preparation

Crystal structure (2rh1) [4,5] and the *in silico* models of the  $\beta_2$ -AR were subjected to the Protein Preparation Wizard workflow implemented in the Schrödinger package [11]. This added hydrogens, which were subsequently minimized using the OPLS.2005 force field and the Impact molecular mechanics engine, while heavy atoms were constrained. Furthermore, it optimized the protonation state of His residues and the orientation of hydroxyl groups, Asn residues, and Gln residues. More details on the protein preparation protocol can be found in a previously published work [6].

### 2.3. Molecular docking

Molecular docking experiments were carried out by means of the Glide [12], as implemented in the Schrödinger package, considering the ligands as flexible but treating the receptor as a rigid structure. A cubing docking grid was centered on Val114, and was given a dimension sufficient to accommodate compounds with a length  $\leq 15$  Å. The ligand-midpoint box was given a side of 10 Å. No scaling factors were applied to the van der Waals (vdW) radii of the receptor atoms, while a scaling factor of 0.8 was applied to the non-polar atoms of the ligands, defined as those with a partial charge lower than 0.15e. The HTVS (High Throughput Virtual Screening) and SP (Standard Precision) scoring functions of Glide were used, granting full flexibility to the ligands. A post-docking minimization, in which only the ligands were flexible, was performed on the output complexes in order to reduce the initially collected 25 poses per ligand to 5. A rescoring of the top ranking SP pose of each compound was then performed with the XP scoring function of Glide and with the London dG scoring function as implemented in the MOE [10]. Rescoring was always performed on the receptor–ligand complexes

just as resulting from the SP calculation, without minimizations or relaxations.

#### 2.4. Generation of alternative binding site conformations for receptor ensemble docking

The crystal structure of  $\beta_2$ -AR (2rh1) was optimized in the presence of three different ligands: carazolol (the co-crystallized ligand), carvedilol and ritodrine, through a stochastic global energy minimization in the torsional coordinate space, in a similar manner to what has been already described in detail [13–16]. The system

$$EF(\%SD) = \frac{\text{number of active compounds up to a given \%SD}}{\text{total number of active compounds in screened database}} \bigg/ \frac{\text{number of compounds up to a given \%SD}}{\text{total number of compounds in screened database}}$$

was represented through the ECEPP/3 force field, within the ICM platform [17]. The three positional and the three orientational coordinates of the ligand, together with the torsional coordinates of ligand and side-chains within 6 Å radius were considered free. The initial conformations of carvedilol and ritodrine chosen were those that showed maximum overlap with the co-crystallized carazolol. A quadratic restraint was imposed between the ligand charged amine and Asp113. No backbone relaxation was performed during the optimization process.

#### 2.5. Induced-fit docking procedure. Protein conformation adapted to the agonist isoproterenol

A conformation of the receptor optimized in complex with the agonist isoproterenol was obtained with the induced fit docking procedure, as implemented in the Schrödinger package [11,12,18]. Specifically, the induced fit procedure involved three consecutive steps: (a) a Glide-based docking of isoproterenol at the receptor, starting from the ligand pose extracted from the SP docking run (see Section 2.3), with the SP scoring function and a vdW scaling factor of 0.5 applied to non-polar atoms of receptor ligands, defined as those with partial charge lower than 0.25e for the receptor and lower than 0.15e for the ligands; (b) a Prime-based optimization of the ligand-binding pocket for all the complexes obtained in step (a), granting flexibility to the ligand and all residues within an 8 Å radius from the ligand; (c) a Glide-based docking of isoproterenol at each of the optimized ligand binding pockets obtained in step (b), with a vdW scaling factor of 0.8 for the non-polar atoms of the ligand only (only the complexes within 30 kcal/mol from the best were subjected to this final docking run). For steps (a) and (c) the docking grid was centered on Val114, and was given a size of 26 Å.

### 3. Results

As explained in Section 2, a virtual compound dataset composed of 60  $\beta_2$ -AR ligands (29 agonists and 31 blockers) and 55,806 decoys extracted from the ZINC Database, was docked to the receptor considering the flexibility of the ligands, but treating the protein as rigid [11,12]. The docking procedures and scoring functions that we applied are endowed with different computational time requirements, expected to be inversely correlated with their accuracy. The fastest and coarsest ones are intended to be used in the initial stages of virtual screening campaigns, in which datasets of millions of compounds are evaluated; in contrast, the slowest and most accurate ones are intended to be used in the subsequent stages of the campaigns, in which the datasets have been already considerably reduced in size. In our case, the average total CPU time required for the processing of 55,866 compounds, was 13 h for the HTVS dock-

ing, 94 h for the SP docking, and 18 h for the XP rescoring and 3 h for the London dG rescoring – these performances were obtained using a single processor of a Quad Core Xeon X5482 system (3.20 GHz, 2X 6MB L2Cache, 1600 MHz). Notably, the actual time needed to complete the calculations can be considerably reduced using more than one processor in parallel.

The results were analyzed through two-dimensional Cartesian graphs in which the percentage of the screened dataset (%SD) and the yield – defined as the percentage of known ligands retrieved – were plotted on the axes X and Y, respectively (see Figs. 2, 5, 7 and 8), and through the calculation of enrichment factors (see Figs. 3 and 9), defined according to the following formula:

#### 3.1. Virtual screening based on the crystal structure of the $\beta_2$ -AR

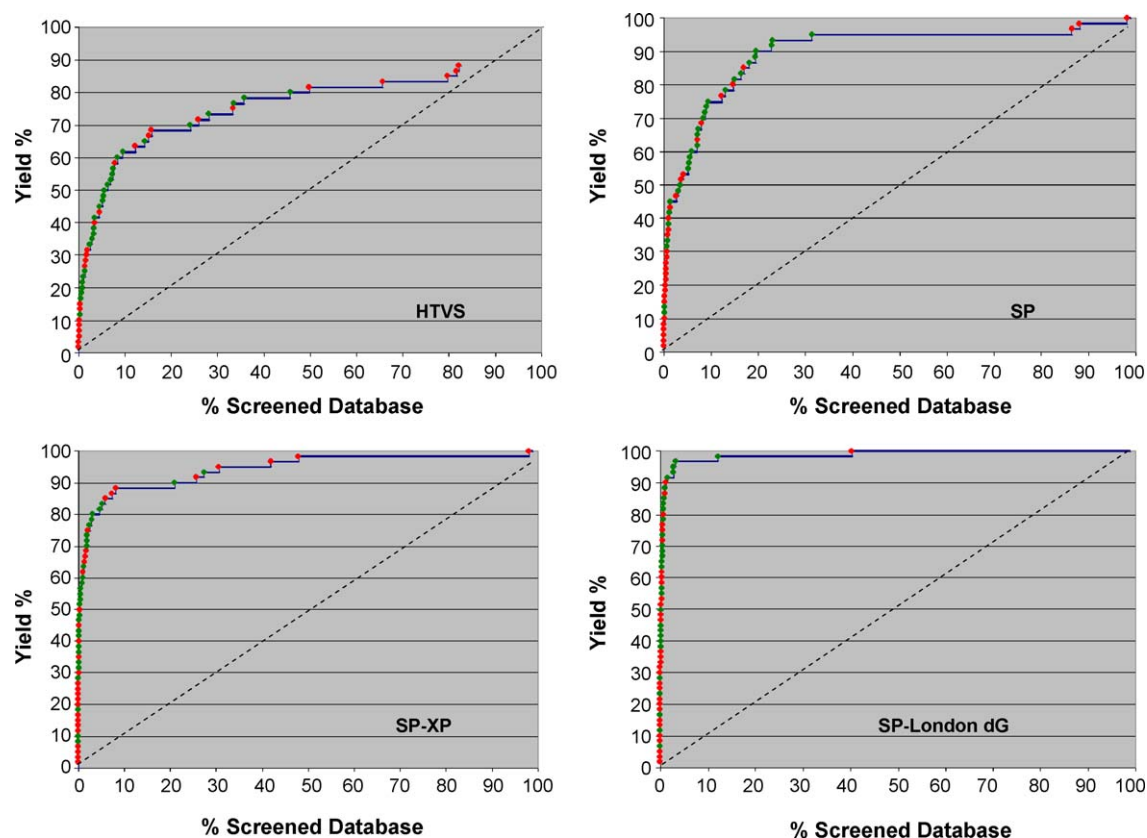
The results obtained when basing the screening on the  $\beta_2$ -AR crystal structure were remarkable (Figs. 2 and 3 and Table 1). Even the fast and coarse HTVS docking mode produced outstanding results, with a clear prioritization of ligands over decoys. As expected, the best results were obtained rescoring the SP results with the more accurate XP and London dG scoring functions, which both showed very high yields in ligand retrieval already in the top ranking portions of the dataset. For example, with the London dG scoring function, the yield reached 50% within the top 0.21% of the dataset, while, within the top 2.5% of the screened database, it reached 91.67% (see Table 1). Similar results were obtained with the XP scoring function. Notably, with the same two scoring functions, we achieved an EF above 300 within the top 10 compounds (i.e. the top 0.02% of the dataset). The EF then gradually declines when considering increasingly large portions of the screened dataset (see Fig. 3).

#### 3.2. Improving virtual screening performance through receptor ensemble docking

Crystal structures offer only a static representation of the receptors, which, in fact, are intrinsically flexible entities. On the basis of our previous experience [19–21], we reasoned that generating alternative conformations of the ligand binding pocket, hence accounting for its flexibility, could improve virtual screening. Thus, through stochastic energy minimizations of the side chains, alternative conformations of the  $\beta_2$ -AR in complex with three representative ligands were generated, starting from the docking poses obtained as described in the previous section (Fig. 4). To thoroughly explore different scenarios, carazolol, which is the co-crystallized ligand, and two additional ligands were selected. Namely, these were ritodrine, which was well prioritized over the decoys when using the crystal structure, and carvedilol, which, instead, was not.

Following the receptor ensemble docking (RED) approach [19,21], the compound dataset was independently docked at each of the three alternative conformations. The results from these three docking runs were then combined with those obtained with the crystal structure and, for each ligand, the pose that obtained the maximum score was selected, thus giving to each docked ligand the possibility to choose its preferred receptor conformation.

The data clearly indicated that combining the results obtained with the three models and the crystal structures were always significantly improved over those obtained using the crystal structure alone (Fig. 5). For all the scoring functions, but especially for HTVS, there was a noticeable improvement since the very early stages of the screening: within the top 0.5% of the screened database, the yield reached 15.0%, 26.67%, 55.0% and 73.33% for HTVS, SP,



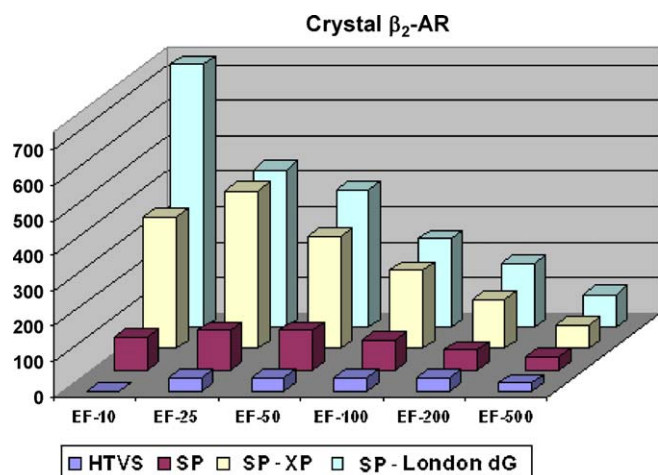
**Fig. 2.** Percentage yield of ligand retrieval plotted versus percentage of screened compounds, sorted according to their docking rank, for the  $\beta_2$ -AR crystal structure using the HTVS, SP, SP-XP and SP-London dG scoring functions. Red dots represent blockers and green dots represent agonists. (For interpretation of the references to color in this figure caption, the reader is referred to the web version of the article.)

XP and London dG, when using crystal structure, while, with the same scoring functions, it reached 30.0%, 33.33%, 56.67% and 76.67%, when combining multiple structures. Similar conclusions remained still valid also when only one of the alternative conformations was used in addition to the crystal structure, as shown in Figs. S1–S3 of the supplementary data for carazolol, carvedilol, and ritodrine, respectively. The structure that worked the best, both alone and in combination with the crystal structure, was the one optimized around the docked carazolol. On the other hand, the

structure that, when taken alone, yielded the poorest results with all the scoring functions, with the exception of London dG, was the one optimized around the docked carvedilol, a ligand that, as we mentioned, received a poor score in the initial SP docking.

### 3.3. Prioritizing agonists or blockers

The  $\beta_2$ -AR has been crystallized in complex with carazolol, an inverse agonist. Thus, it was not surprising to note that, although the virtual screening experiments based on the crystal structure effectively differentiate both agonists and blockers from non-binders, they generally prioritize blockers over agonists. This tendency can be seen in Fig. 2, where red and green dots represent blockers and agonists, respectively, and confirms what we reported in a recent publication [7].



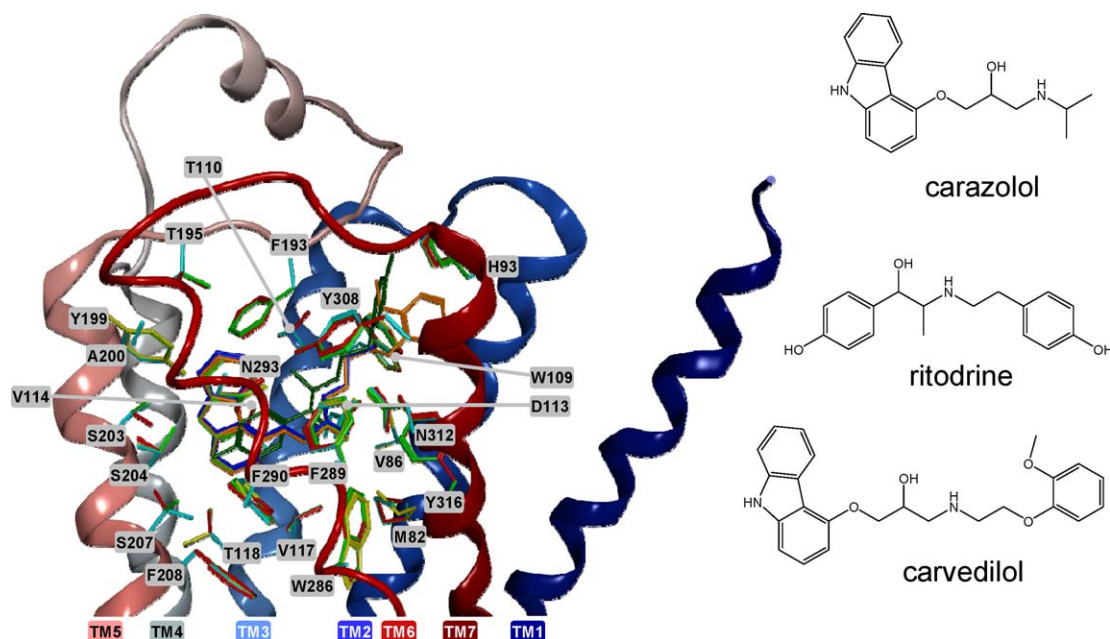
**Fig. 3.** Enrichment of true ligands concentration within the top scoring 10–500 compounds when compared to their concentration in the whole database, for the screening based on the crystal structure. Values expressed as enrichment factors (EF). Numerical values can be found in Table S1 of the supplementary data.

**Table 1**

Percentage yield of ligand retrieval and enrichment factor (EF) within various percentages of top scoring compounds in the screening based on the  $\beta_2$ -AR crystal structure, applying the London dG and XP scoring functions on the poses selected with the SP docking.

SP-London dG			SP-XP		
Screened database (%)	Yield (%)	EF	Screened database (%)	Yield (%)	EF
0.10	35.00	349.16	0.10	30.00	299.28
0.20	46.67	232.79	0.20	41.67	207.85
0.50	73.33	146.83	0.50	55.00	110.13
1.00	85.00	84.95	1.00	61.67	61.63
2.50	91.67	36.66	2.50	76.67	30.66
5.00	96.67	19.34	5.00	81.67	16.34
10.00	96.67	9.67	10.00	88.33	8.83

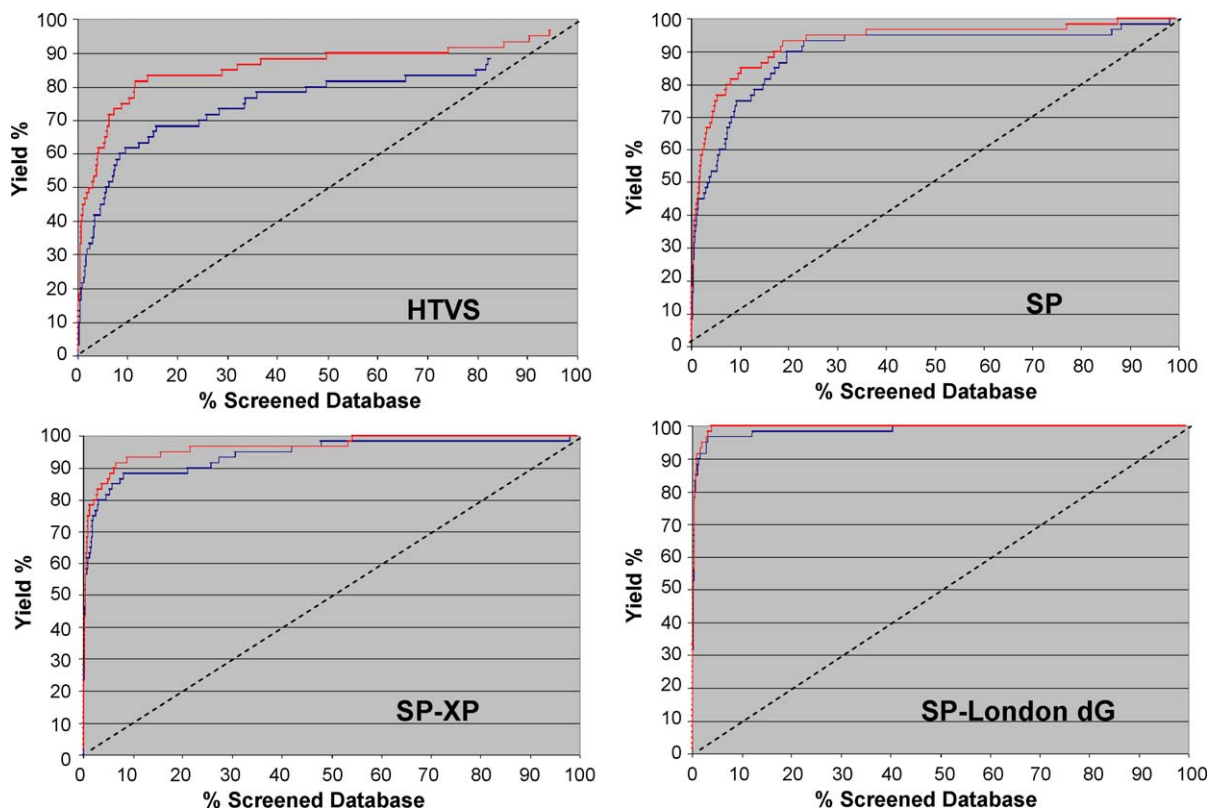




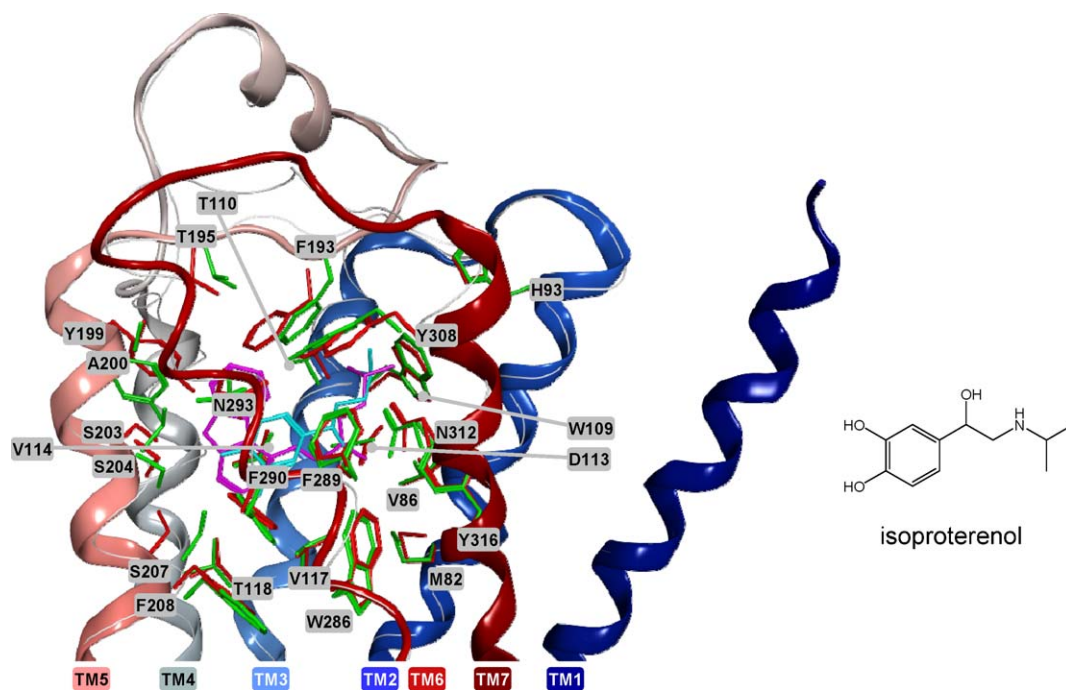
**Fig. 4.** Alternative conformations of the  $\beta_2$ -AR in complex with three representative ligands (red crystal, yellow carvedilol protein, orange carvedilol ligand, light green ritodrine protein, dark green ritodrine ligand, cyan carazolol protein, blue carazolol ligand). The ribbon representation of the backbone, identical for all the structures since the optimizations did not allow movements of the backbone, is colored with a smooth transition from red to blue moving from the N-terminus to the C-terminus. (For interpretation of the references to color in this figure caption, the reader is referred to the web version of the article.)

Here, this tendency was successfully reversed, prioritizing agonists over blockers, by generating an alternative protein conformation optimized with the full agonist isoproterenol (Fig. 6). Unlike for the generation of the alternative conformations described in

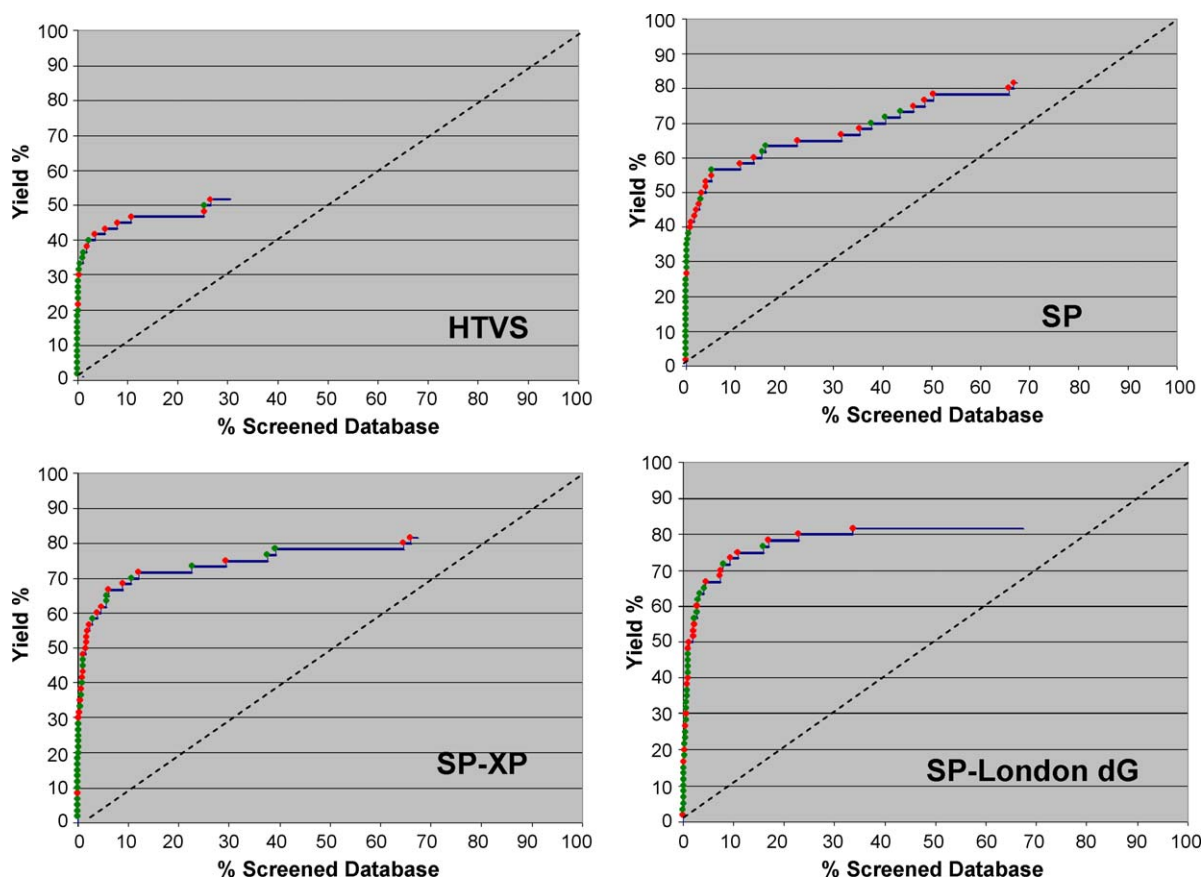
Section 2, for this purpose a more drastic approach was employed, based on the Schrödinger's induced fit protocol, which, for the residues around the ligand, samples the degrees of freedom of the side chains in the dihedral space and grants flexibility to the back-



**Fig. 5.** Comparison of the virtual screening results obtained when using the  $\beta_2$ -AR crystal structure alone (blue) and when combining it with three alternative conformations according to the receptor ensemble docking approach (red). The percentage yield of ligand retrieval is plotted versus percentage of screened compounds, sorted according to their docking rank, using the HTVS, SP, SP-XP and SP-London dG scoring functions. (For interpretation of the references to color in this figure caption, the reader is referred to the web version of the article.)



**Fig. 6.** Alternative protein conformation optimized with the full agonist isoproterenol (red: receptor in the crystal structure; pink: carazolol; green: isoproterenol-adapted receptor; cyan: isoproterenol). The backbone of the crystal structure is shown in ribbon representation, and is colored with a smooth transition from red to blue moving from the N-terminus to the C-terminus. The backbone of the isoproterenol-adapted receptor is shown as a thin white line.



**Fig. 7.** Percentage yield of ligand retrieval plotted versus percentage of screened compounds, sorted according to their docking rank, for the isoproterenol-adjusted structure of the  $\beta_2$ -AR using the HTVS, SP, SP-XP and SP-London dG scoring functions. Red dots represent blockers and green dots represent agonists. A significant migration of the agonists towards the top scoring positions can be noticed when comparing these results with those shown in Fig. 2. (For interpretation of the references to color in this figure caption, the reader is referred to the web version of the article.)

**Table 2**

Blockers and agonists retrieved within the top scoring compounds using the crystal structure and the isoproterenol-adjusted structure.

	HTVS		SP	
	Crystal structure	Isoproterenol-adjusted structure	Crystal structure	Isoproterenol-adjusted structure
TOP-50				
Blockers	2	0	6	1
Agonists	0	11	0	14
Total	2	11	6	15
TOP-100				
Blockers	4	1	7	2
Agonists	0	14	2	18
Total	4	15	9	20
TOP-250				
Blockers	8	2	13	2
Agonists	1	17	2	20
Total	9	19	15	22
TOP-500				
Blockers	8	2	17	3
Agonists	5	18	4	21
Total	13	20	21	24
	SP-XP		SP-London dG	
	Crystal structure	Isoproterenol-adjusted structure	Crystal structure	Isoproterenol-adjusted structure
TOP-50				
Blockers	13	1	16	1
Agonists	4	11	5	2
Total	17	12	21	3
TOP-100				
Blockers	16	1	17	1
Agonists	8	16	10	6
Total	24	17	27	7
TOP-250				
Blockers	18	3	24	3
Agonists	13	17	18	11
Total	31	20	42	14
TOP-500				
Blockers	18	7	28	6
Agonists	18	20	23	17
Total	36	27	51	23

bone [22]. The analysis of the virtual screening conducted using this alternative structure, shown in Fig. 7 and Table 2, clearly showed that with the HTVS, SP, and XP scoring functions agonists effectively migrated towards the top scoring positions of the database. As an example, using the XP scoring function, 11 agonists and 1 blocker were retrieved within the 50 top scoring compounds with the isoproterenol-adjusted structure, as opposed to the 4 agonists and 13 blockers retrieved with the plain crystal structure. Also the London dG scoring function tended to prioritize agonists over blockers with the isoproterenol-adjusted structure. However, the global retrieval yields fell significantly when using this structure, thus resulting in a lower absolute number of agonists found within the top scoring compounds, contrary to what detected with the other functions (see Table 2).

#### 3.4. Virtual screening based on homology models of the $\beta_2$ -AR

Crystal structures exist only for very limited numbers of GPCRs [2,3]. However, these can be used as templates for the construction of homology models of the remaining members of the superfamily. Thus, it is of extreme importance to establish the applicability of such homology models to docking-based virtual screenings. For this purpose, here, the same docking-based virtual screening was also applied using three homology models recently published by Costanzi [6], dubbed model 1, model 2 and model 2-F290g+. From

**Table 3**

Percentage yield of ligand retrieval and enrichment factor (EF) within various percentages of top scoring compounds in the screening based on the  $\beta_2$ -AR homology models, applying the London dG and XP scoring functions on the poses selected with the SP docking.

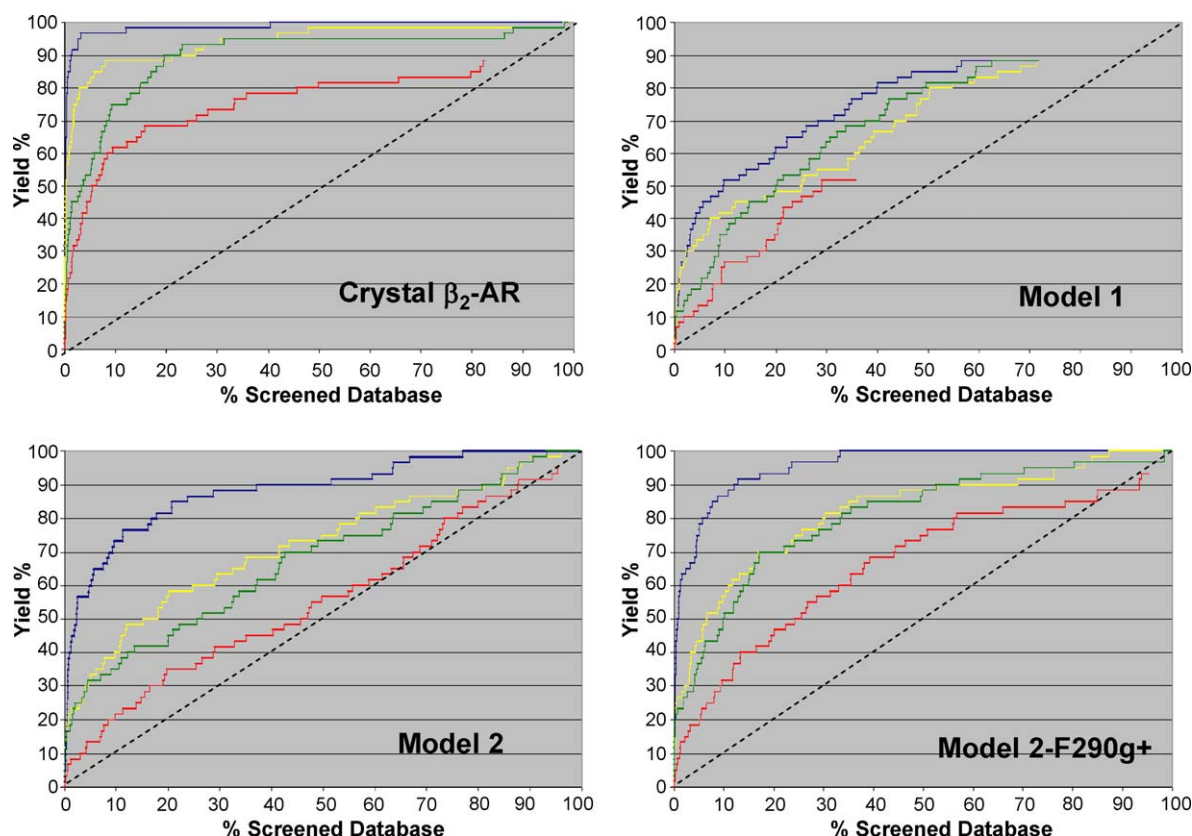
SP-London dG			SP-XP		
Screened database (%)	Yield (%)	EF	Screened database (%)	Yield (%)	EF
Model 1					
0.10	3.33	33.22	0.10	3.33	33.22
0.20	5.00	24.94	0.20	5.00	24.94
0.50	11.67	23.37	0.50	13.33	26.69
1.00	21.67	21.66	1.00	20.00	19.99
2.50	30.00	12.00	2.50	30.00	12.00
5.00	41.67	8.33	5.00	33.33	6.67
10.00	51.67	5.17	10.00	41.67	4.17
Model 2					
0.10	5.00	49.88	0.10	10.00	99.76
0.20	8.33	41.55	0.20	11.67	58.21
0.50	16.67	33.38	0.50	16.67	33.38
1.00	40.00	39.98	1.00	21.67	21.66
2.50	53.33	21.33	2.50	23.33	9.33
5.00	60.00	12.00	5.00	31.67	6.33
10.00	73.33	7.33	10.00	40.00	4.00
Model 2-F290g+					
0.10	20.00	199.52	0.10	11.67	116.42
0.20	25.00	124.70	0.20	20.00	99.76
0.50	45.00	90.11	0.50	25.00	50.06
1.00	56.67	56.64	1.00	26.67	26.65
2.50	65.00	25.99	2.50	30.00	12.00
5.00	75.00	15.00	5.00	43.33	8.67
10.00	86.67	8.67	10.00	56.67	5.67

the structural point of view, the three models, when compared with the crystal structure showed different levels of accuracy: (a) model 1 featured a second extracellular loop (EL2) built by homology to rhodopsin, buried into the opening of the interhelical cavity in a manner unnatural for the  $\beta_2$ -AR, and showed six out of the twenty residues in the binding pocket in the wrong rotameric state; (b) model 2 featured an EL2 built *de novo*, with a more native-like conformation, at least in the segment that lines the binding pocket, and only one residue, namely Phe290, in the wrong rotameric state; (c) model 2-F290g+ was a modified version of model 2, featuring, as the only difference with the latter, residue Phe290 in the gauche+ conformation, to match the crystal structure.

As evident from Fig. 8, the virtual screening experiments conducted at the three models resulted in a significant prioritization of ligands versus decoys, especially when using more sophisticated scoring functions. Not surprisingly, the homology models performed accordingly to their level of accuracy. With the London dG rescoring of the SP docking, 50% of the true ligands were recovered within the top 0.72%, 2.25%, and 9.55% of the screened database, with model 1, model 2, and model 2-F290g+, respectively. Moreover, in the top 2.5% of the screened database, the yield was 65.0%, 53.33%, and 30.0% with model 1, model 2, and model 2-F290g+, respectively (see Table 3). The same trend was evident from the analysis of the EF (see Fig. 9).

#### 4. Discussion

As mentioned, rhodopsin has been for years the only receptor with an experimentally elucidated 3D structure. However, recent breakthroughs led to the solution of the crystal structure of a few additional receptors, including the  $\beta_2$ -AR, while more structures are expected to be solved in the near future. In this context, our analysis of the applicability of the crystal structure of a GPCR to virtual screening is indeed very timely. The positive results that we obtained in this case study encourage the application of computer-aided techniques to the rational identification of GPCR ligands, and



**Fig. 8.** Percentage yield of ligand retrieval plotted versus percentage of screened compounds, sorted according to their docking rank, for the  $\beta_2$ -AR crystal structure and three homology models using the HTVS (red), SP (green), SP-XP (yellow) and SP-London dG (blue) scoring functions. (For interpretation of the references to color in this figure caption, the reader is referred to the web version of the article.)

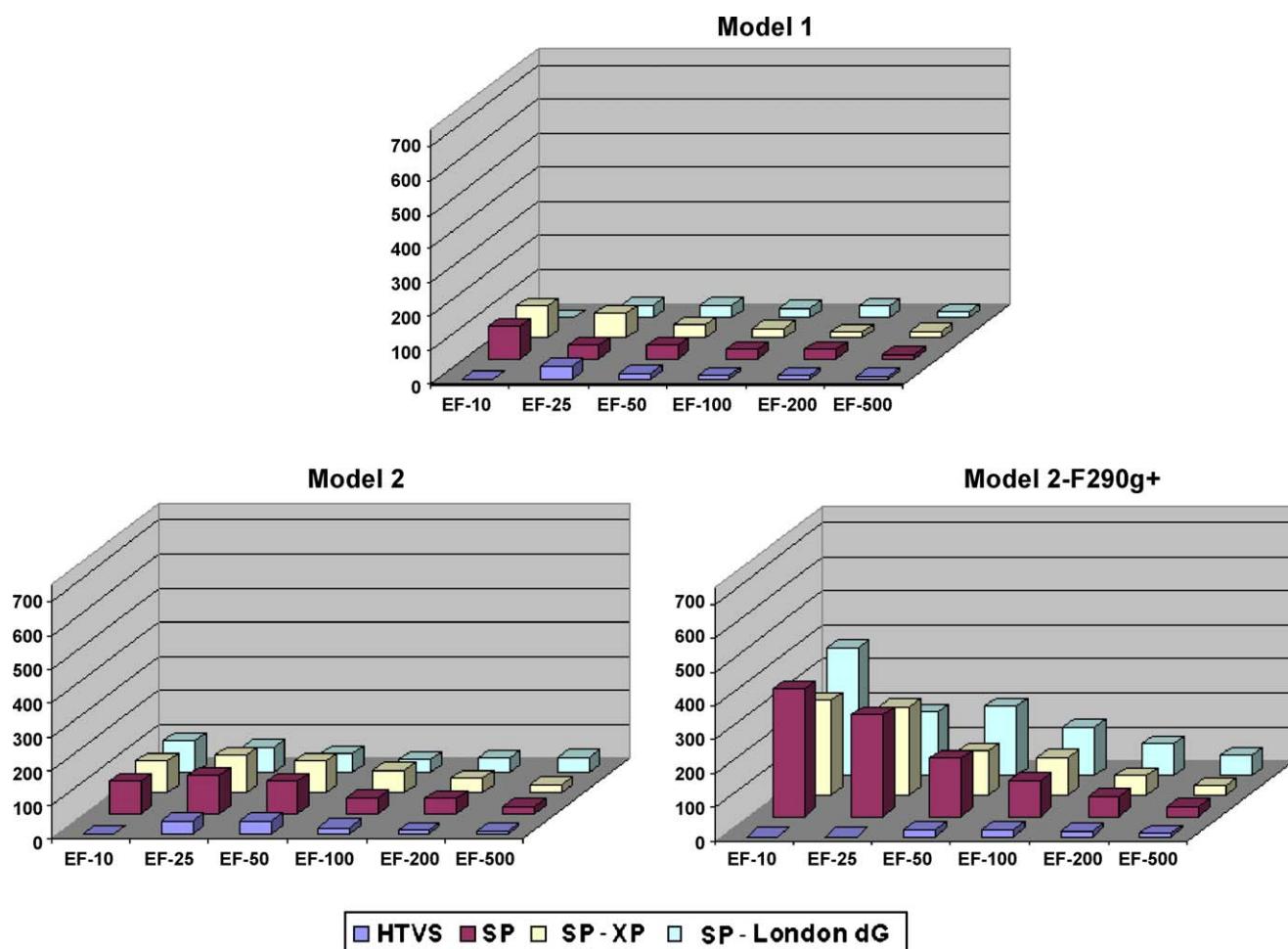
are very much in line with the outcome of actual virtual screening campaigns based on the crystal structures of the  $\beta_2$  adrenergic and the adenosine  $A_{2A}$  receptors [23–25]. Notably, our virtual screening experiments did not necessitate the consideration of explicit water molecules in the binding pocket. However, for other systems, the inclusion of one or more water molecules crucial for the mediation of receptor ligand-interactions may be essential [26].

Crystal structures, however, are only static representations of proteins that, instead, are inherently flexible. Neglecting protein flexibility by using one rigid representation of the target has been shown to adversely affect virtual screening performance [27–29]. Receptor ensemble docking, a method where docking is performed on several distinct structures and the results are subsequently combined, has been successfully used to account for protein flexibility in virtual screening campaigns for ligands of protein kinases [19,21,30,31], GPCRs [20], and other protein targets [29,32]. Here, we implemented receptor ensemble docking to mimic the flexibility of the  $\beta_2$ -AR by generating and using more than one binding site conformation, and observed significantly improved virtual screening yields, in agreement with what was found for other receptors.

Moreover, all the known crystal structures of GPCRs, including that of the  $\beta_2$ -AR, have been obtained in complex with antagonists or inverse agonists, thus raising possible concerns on their applicability to the identification of agonists. According to mutagenesis and crystallographic data, blockers and agonists of the  $\beta_2$ -AR establish an ionic interaction Asp113 in TM3, through their protonated amino group, and a hydrogen-bond with Asn312 in TM7, through the  $\beta$ -hydroxyl group [33,34]. As shown by the crystal structure, the inverse agonist carazolol also establishes a direct interaction with Ser203 in TM5. The catechol hydroxyl groups of the agonists are also supposed to interact with this residue as well as two other

serines located in TM5, namely Ser204 and Ser207 [35]. However, such interactions cannot be detected when docking of agonists to a rigid representation of the receptor crystallized in its ground state. For these interactions to occur, a conformational change in the protein is necessary. Modeling studies suggested that agonists triggered a counterclockwise rotation of TM5 [36,37], leading to the formation of hydrogen bonds with the three above-mentioned Ser residues. In agreement with the need for a conformational change, our virtual screening based on the crystal structure generally prioritized blockers over agonists, although being clearly able to distinguish the latter from the decoys (Fig. 2). In this study, we defined a method that successfully inverted this tendency, by utilizing a model optimized around an agonist (see Fig. 5). Notably, de Graaf and Rognan have shown that a crystal structure modified to reflect the early conformational events in  $\beta_2$ -AR activation led to the selective retrieval of full and partial agonists, in a screen that combined docking and molecular interaction fingerprints [38]. Similarly, Abagyan and co-workers have improved the retrieval of agonists by changing the conformation of TM5, a domain which is thought to be related with the activation of the receptor [39]. However, the procedure utilized here for the generation of the agonist optimized receptor does not necessitate prior knowledge of the activation process. It just requires docking an agonist at the crystal structure and subjecting the resulting complex to automatic optimization according to the InducedFit protocol, as implemented in the Schrodinger package [11,12,18]. Confirming these theoretical findings, a recent virtual screening campaign for ligands of the free fatty receptor 1 (FFA1), based on a model optimized in complex with a potent agonist [40–42], prioritized the retrieval of activators over blockers [43]. It is worth noting that the protocol that we used for the generation of the isoproterenol-adapted confor-





**Fig. 9.** Enrichment of true ligands concentration within the top scoring 10–500 compounds when compared to their concentration in the whole database, for the screenings based on the three homology models. Values expressed as enrichment factors (EF). Numerical values can be found in [Table S2 of the supplementary data](#).

mation of the  $\beta_2$ -AR accounts only for local changes within the ligand-binding pocket, and is not intended to study the activation process. In fact, GPCR activation is a complex phenomenon that may imply relatively large conformational changes as well alterations of dimerization interfaces [33,34,44]. We have been able to follow the initial phases of the activation process by means of a modified induced fit protocol that granted flexibility to entire domains of the receptor (data still unpublished). Alternatively, computational hypotheses on the activation process have been inferred through molecular dynamics and coarse-grain simulations [45–48].

Lastly, we demonstrated that docking-based virtual screening techniques are not only applicable to crystal structures but also to homology models. This point is of extreme importance because it extends their area of applicability to the vast majority of receptors, for which crystal structures are not available. Not surprisingly, the three homology models that we tested performed according to their level of accuracy. Notably, the most accurate model, although obviously outperformed by the crystal structure, yielded excellent results. The conformation of the second extracellular loop, a domain heavily involved in ligand binding and receptor activation [47,49], affected substantially the accuracy of the binding site, and thus the virtual screening, which showed significantly improved performances when the loop was built *de novo*. Thus, accurately modeling the extracellular loops appears as one of the most crucial aspects to ensure the applicability of virtual screening to GPCR models – the use of mixed computational and experimental techniques in which the helical bundle is modeled *in silico* while the extracellular

regions are modeled through NMR may offer an attractive solution to this challenging problem [50]. It is worth noting that with the HTVS scoring function, which is the fastest but also the least sophisticated of those that we applied, model 1 and model 2 did not perform outstandingly, although yielding results by far better than a random selection (represented by the dotted diagonal in Fig. 6). This observation suggests that, when conducting virtual screening campaigns based on GPCR homology models, it might be convenient to use directly a scoring function with a higher level of accuracy, rather than subjecting the database to a preliminary screen with the HTVS scoring function.

## 5. Conclusion

This study shows the applicability of the  $\beta_2$ -AR crystal structure to docking-based virtual screening, defines a method to optimize the results by mimicking the flexibility of the protein through the use of generated alternative conformations of the binding pocket, defines a method to guide the screening towards the discovery of agonists, and provides evidence that also *in silico* models are powerful tools on which to base the virtual screenings. Our results definitely encourage the application of computer-aided techniques to the discovery of novel GPCR ligands. Moreover, the practical applicability of these computer-based drug discovery techniques promises to considerably expand and bloom in the near future, as experimental structures are expected to be produced and become available for an ever greater number of GPCRs.

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## Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.jm.2010.11.005

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