

PhEVER: a database for the global exploration of virus–host evolutionary relationships

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ABSTRACT

Fast viral adaptation and the implication of this rapid evolution in the emergence of several new infectious diseases have turned this issue into a major challenge for various research domains. Indeed, viruses are involved in the development of a wide range of pathologies and understanding how viruses and host cells interact in the context of adaptation remains an open question. In order to provide insights into the complex interactions between viruses and their host organisms and namely in the acquisition of novel functions through exchanges of genetic material, we developed the PhEVER database. This database aims at providing accurate evolutionary and phylogenetic information to analyse the nature of virus–virus and virus–host lateral gene transfers. PhEVER (<http://pbil.univ-lyon1.fr/databases/phever>) is a unique database of homologous families both (i) between sequences from different viruses and (ii) between viral sequences and sequences from cellular organisms. PhEVER integrates extensive data from up-to-date completely sequenced genomes (2426 non-redundant viral genomes, 1007 non-redundant prokaryotic genomes, 43 eukaryotic genomes ranging from plants to vertebrates) and offers a clustering of proteins into homologous families containing at least one viral sequences, as well as alignments and phylogenies for each of these families. Public access to PhEVER is available through its webpage and through all dedicated ACNUC retrieval systems.

INTRODUCTION

Viruses are responsible for a large number of infectious diseases and cancers. Recently, new viral diseases have emerged leading to severe consequences on human activities. The emergence of many of these new viruses can be attributed to recombining viruses as well as to host species jump (1–3). Therefore, understanding how viruses interact with their hosts and more specifically how the complex interactions between viruses and their host organisms are acquired and maintained throughout evolution, remains a major challenge (4–7). In order to assess this question, it is of prime importance to be able to detect and quantify the occurrence of lateral gene transfer events, and the impact of these events on viral–host co-evolution. Indeed, the mechanisms behind fast viral adaptation are far from being elucidated. Thus, we developed a global approach aimed at providing accurate evolutionary and phylogenetic information to tackle these questions.

The major drawback of currently available databases of homologous families to the study of viral homologies and lateral gene transfer in viruses is their taxonomic compartmentalization. Indeed, current databases present families of homologies either restricted to viruses only [Protein Clusters (8), GeneTree (9)] or to viral taxonomic groups [Viral Orthologous Cluster (10)], some also not presenting viral information [HomoloGene (11)]. The few databases that do present viral and non-viral sequences, such as Pfam (12) or the Conserved Domain Database (13) do not provide complete phylogenetic trees. This translates into the fact that it is not currently possible to have a global view on viral–host lateral gene transfers due to the difficulty of obtaining global information on cross-taxon transfers at the viral level.

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We present the first public release of PhEVER, a unique database of homologous gene families containing sequences (i) of all completely sequenced viruses and (ii) from fully sequenced cellular organisms. The protein sequences are clustered—without *a priori* and according to similarity criteria—into families containing either only viral sequences or both viral and cellular sequences. PhEVER integrates extensive data from up-to-date completely sequenced genomes spanning a wide taxonomic range (2426 non-redundant viral genomes, 1007 non-redundant prokaryotic genomes, 43 eukaryotic genomes ranging from plants to vertebrates). To our knowledge, this is the most complete database of families of homologous viral sequences. Indeed, it not only spans all known viral groups but it also has the unique feature of presenting homologies with eukaryotic and prokaryotic sequences. The database offers a clustering of proteins into homologous families containing at least one viral sequence, as well as pre-computed alignments and phylogenies for each of these families. Alignments and phylogenies are built according to state-of-the-art phylogeny procedures and we provide tools to edit them and recompute them on the fly (14). We also provide the possibility for users to assign their sequence of interest to a family and to re-build the phylogeny accordingly through the HoSeqI tool (15). PhEVER thus constitutes a comprehensive working tool to detect sequence homologies and possible gene transfer events. Public access and documentation is available through the database webpage and through all dedicated ACNUC retrieval systems (16).

FAMILIES OF VIRAL PROTEINS

We developed a genome-wide cross-taxon approach to build a database of families of homologous sequences and to provide accurate alignments and phylogenies for the constructed families. The layout of the implementation of the PhEVER database is represented in Figure 1 and the details concerning its sequence content are available in Table 1. In order to avoid redundancy due to the availability of numerous genomes of similar bacterial and viral strains in public databases, we collected all completely sequenced viral and bacterial genomes from RefSeq Viral (17) and Genome Reviews (18), two non-redundant and curated databases of completely sequenced genomes. To this high-quality curated data composed of Archaea, Bacteria and Eukarya, we added nine eukaryotic genomes from Ensembl (*Aedes aegypti*, *Anopheles gambiae*, *Bos taurus*, *Caenorhabditis elegans*, *Danio rerio*, *Drosophila melanogaster*, *Gallus gallus*, *Homo sapiens*, *Mus musculus*) (19) to allow for a large representation of species from the different domains of life. The PhEVER database was structured under the ACNUC system (20) allowing it to be queried using a web interface and a large number of tools specifically developed for this database management system (14).

From the flat files containing the genomes and their annotations, two databases were built under the ACNUC database management system (20), which is specifically aimed at building, storing and querying biological sequence data. One of them contains the nucleic sequences, the other contains the proteins generated by translating all CDS of the complete genomes—using the

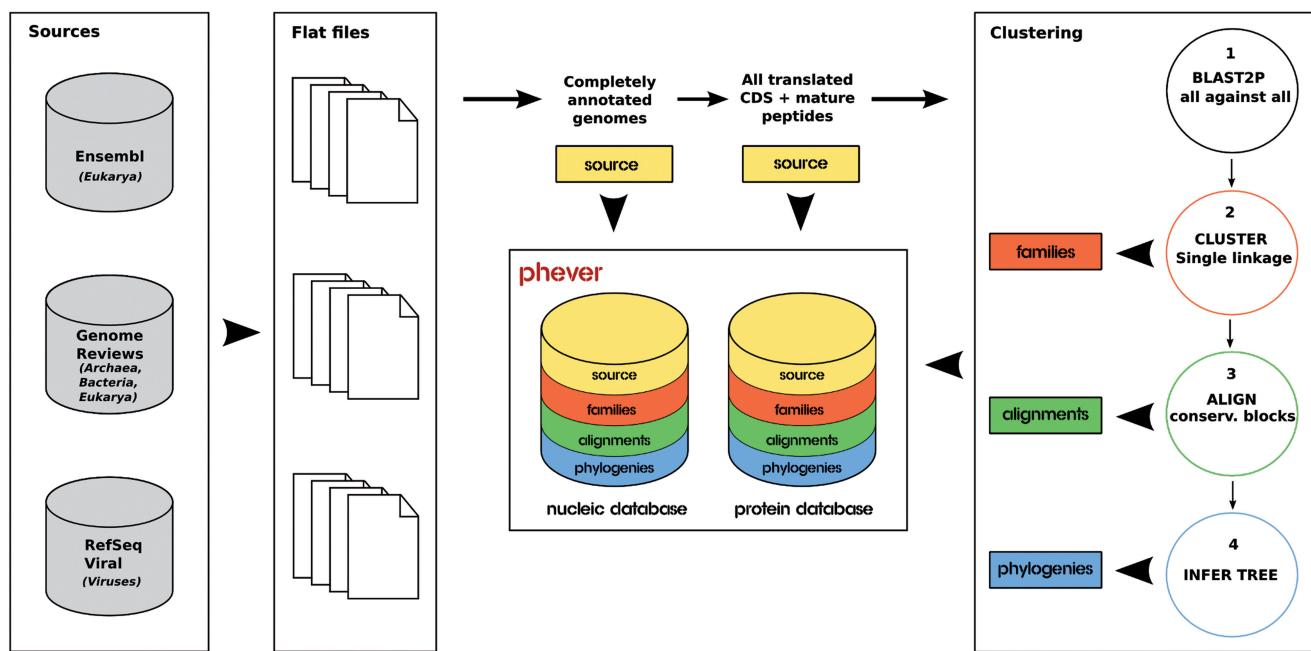


Figure 1. Flow chart of the PhEVER building process. Complete genomes and their annotations were retrieved from three external public databases (Ensembl, Genome Reviews and RefSeq Viral) to provide high-quality non-redundant data for Eukarya, Archaea, Bacteria and Viruses. Two databases (nucleic acids, proteins) were constructed from this data to form PhEVER. All annotated CDS and mature peptides were translated and used for the clustering procedure. The homologous families thus produced were annotated in PhEVER, and alignments and phylogenies were built for each family and incorporated in the databases.

Table 1. Global species and sequence content in PhEVER

	No. of species	No. of proteins ^a	No. of proteins in fam. ^b	Data source
All	3476	4 515 271	333 618 (1%)	—
Viruses	2426	82 929 (2718 m.p. ^c)	82 784 (100%)	RefSeq
Bacteria	937	3 207 914	232 066 (7%)	Genome Reviews
Archaea	70	158 919	6702 (4%)	Genome Reviews
Eukarya	43	1 065 509	12 066 (1%)	Ensembl + Genome Reviews
Eukarya (excl. <i>Anopheles</i>) ^d	—	580 567	11 201 (2%)	—

^aProteins correspond to translated annotated CDS. The number of proteins in the protein database is therefore equal to the number of CDS in the nucleic database.

^bNumber of proteins associated to a family, followed by the proportion of proteins associated to a family in the taxonomic group.

^c2718 mature peptides are added to the 80 210 proteins translated from all CDS.

^dThe genome of *A. gambiae* contains data from different haplotypes and presents therefore a high level of redundancy.

appropriate genetic codes. For viral genomes presenting polypeptides which further mature in vivo into mature peptides, the mature peptides were added to the set of translated proteins according to the annotations specified in the given genome (Figure 1). Table 1 lists the global content of the databases as well as their original sources. Annotations were extracted from UniProtKB via the cross-references found in the CDS (21). Subsequently, sequences in the nucleic and protein PhEVER database were clustered into families and were assigned a family accession number. Alignments and phylogenies were built for each of these families according to state-of-the-art procedures (Figure 1). The classification of all organisms present in the database was retrieved from the taxonomy database at National Centre for Biotechnology Information (11) and is available on the PHEVER web interface.

CLUSTERING INTO FAMILIES

The clustering of proteins into homologous families was constructed using an automated procedure similar to the one described in (14) and implemented within a parallel framework in a software package called SiLiX. Briefly, sequences were assigned to protein families by simple transitive link using the following criteria. A similarity search of all translated proteins and mature peptides against all was performed using BLASTP2 similarity search with the BLOSUM62 substitution matrix, a 10^{-4} e-value threshold and the ‘m S’ filter option (22,23). For each pair of sequences, HSPs which were not compatible with a global alignment were removed. Two sequences were included in the same family if the sum of the remaining HSPs covered >80% of the proteins length (and at least 100 amino acids) and if their identity was $\geq 35\%$. These two criteria were previously shown to provide a good trade-off between the ability of clustering sequences from divergent organisms and the quality of resulting alignments for subsequent phylogenetic analyses (14). Finally, only families containing at least one viral sequence were integrated in the database. For each of the families, a small description built from the gene annotations ordered by frequency is available on the web interface. Figure 2 presents the distribution of viral species, proteins and families according to each viral group (A) as well as a Venn diagram

representing the families content (B). Figure 2A presents PhEVER’s broad taxonomical distribution covering all Baltimore groups (24). This distribution is naturally biased towards dsDNA, ssDNA and positive-sense ssRNA viruses reflecting the bias in genomic sequencing efforts. Indeed these groups contain long-studied viral families—either for their medical interest or for their economical impacts—such as Caudovirales, Poxviridae, Herpesvirales, Flaviviridae, Picornavirales or Parvoviridae. Figure 2 (B) shows a large number of orphan families indicating that a significant proportion of viral proteins (32%) do not contain any homologs with proteins from known genomes. These proteins, among which some might possibly be caused by annotation errors, are unfit for comparative functional analysis and should be the focus of future experimental studies to validate them and to provide with crucial information on viral mechanisms. Figure 2B also shows the small number of families sharing sequences from both viruses and eukaryotes compared to the relatively high number of families sharing sequences from both viruses and bacteria. This observation may be due to different underlying biological mechanisms but might also be an indicator of a still low coverage of the Eukarya domain.

DETECTION OF HORIZONTAL GENE TRANSFER

One of the applications of PhEVER is the detection of horizontal gene transfer events by comparing a gene family tree with the expected species tree. The discrepancies between the gene family history and the species phylogeny can then be an indication of possible events including a gene duplication, a gene loss or a horizontal gene transfer. The quality of the gene phylogenies is therefore essential and in this perspective, we implemented a procedure based on a rigorous methodology. First, the clustering into families was built with criteria leading to conservative families. Second, maximum likelihood phylogenetic trees were inferred for all families based on conserved aligned blocks. In our databases, for all families containing at least three sequences, pre-computed alignments and phylogenies are therefore already available. This allows for a simple and accurate overview of any family without the need of heavy computations and

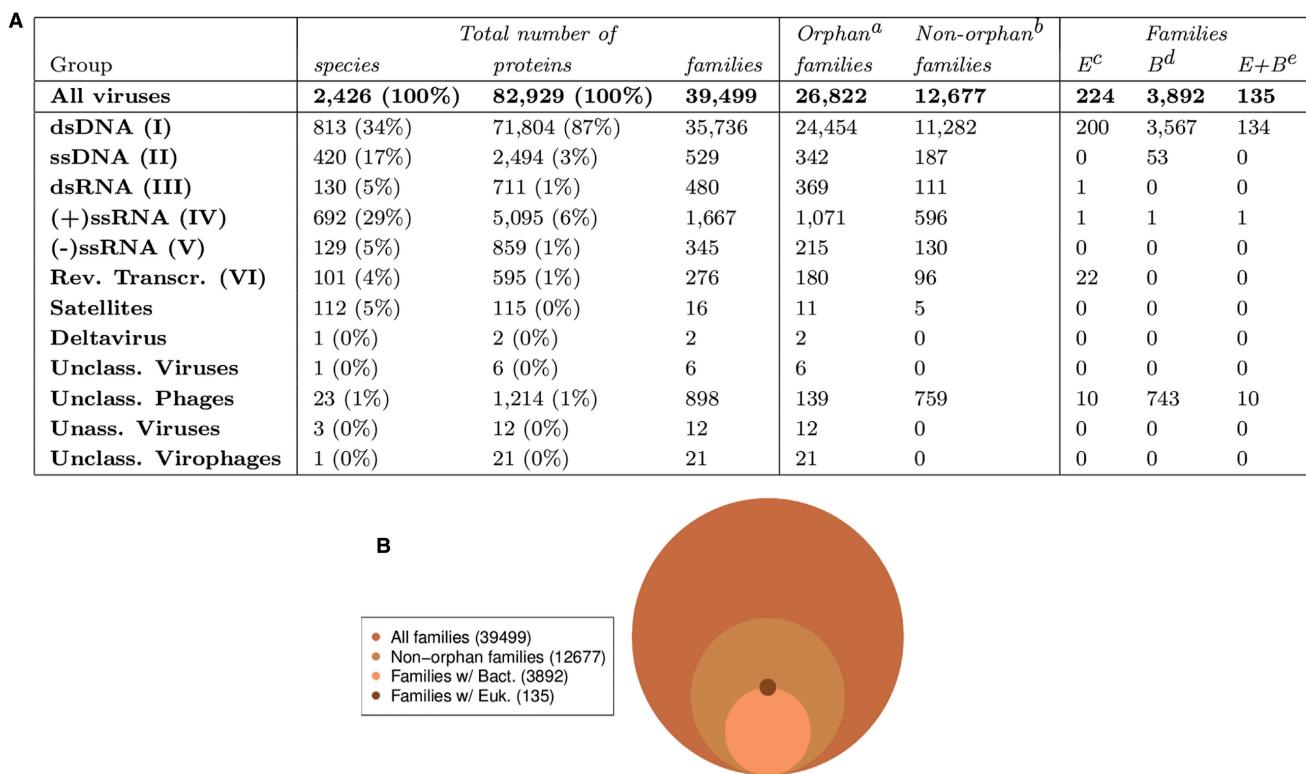


Figure 2. Global information on PhEVER content. (A) Number of species, proteins, families in each viral group [groups correspond to the NCBI Taxonomy following the ICTV nomenclature (36); groups corresponding to the Baltimore classification are additionally indicated in Roman notation (24)]. (B) Venn diagram of the number of families, non-orphan families and families containing Bacteria and Eukarya. ^aOrphan families are families containing only one viral sequence and no other sequence. This sequence presents no sufficient similarity with any sequence of the database, ^bNon-orphan families are all families containing at least two sequences, ^cFamilies containing at least one viral sequence and one eukaryotic (E) sequence, ^dFamilies containing at least one viral sequence and one bacterial (B) sequence and ^eFamilies containing at least one viral sequence, one eukaryotic sequence and one bacterial sequence (E+B).

can be a useful tool to search for lateral gene transfers in viral genes. For each family containing at least three sequences, alignments were estimated using MUSCLE with default parameters (25). All alignments were treated with Gblocks (26) to select conserved blocks. Phylogenetic trees were inferred by maximum likelihood using PhyML (27) with a JTT evolutionary model (28). Branch support was inferred using the Shimodaira–Hasegawa-like non-parametric procedure implemented in PhyML (27,29). To accommodate for weak phylogenetic signal, a thorough exploration of the tree space was made through topological rearrangements using the Nearest Neighbor Interchange topology search method. Finally, for visualization purposes, the trees were then rooted using midpoint rooting. This procedure allowed us to build accurate phylogenies sustained by branches of high support values. Indeed, Figure 3 shows that ~80% of all branch supports have a value higher than 75 and one-third are above 95. Global statistics are biased by the presence of low branch supports. These are mostly due to few small families of less than 10 leaves, indicating that most families present robust phylogenies (Figure 3B). Finally, Figure 3C indicates that the low branch support values are attributable to very similar sequences with small branch lengths and might be linked to unresolved topologies. By contrast, all branch lengths longer than 0.15 subst/site

display a high branch support which reveals the accuracy of our phylogenetic inference procedure.

QUERY THE DATABASE

PhEVER is structured under the ACNUC sequence database management system and a large number of tools have been developed around this database management system [see (16) for an overview]. PhEVER can therefore be queried using (i) web applications, (ii) standalone software (iii) or embedded within Python, R or C code. The PhEVER web interface is available at <http://pbil.univ-lyon1.fr/databases/phever> and allows to search for sequences or families by combining several criteria (including species, gene names, annotation terms) as well as by crossing taxa. The graphical user interface QUERY_WIN and the terminal-based interface Raa_query (16) implement more features than the web interface and allow for remote ACNUC access and query as well as for the automatization of querying processes through standalone software. Finally the C language API, Python language API (16) and the seqinR package for R (30) implement tools for integrating queries in user designed code. Note that the PhEVER database can also be installed on a local machine or server for fastest data access. All files necessary for the PhEVER

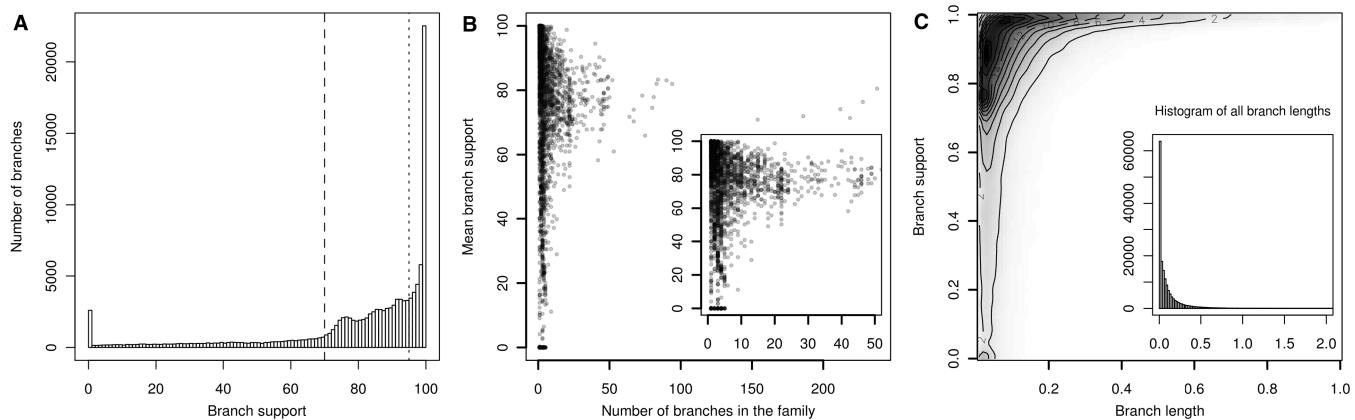


Figure 3. Branch support in PhEVER trees. (A) Histogram of node supports in all phylogenetic trees. The dashed line indicates the support value of 70 (80% of the distribution is above this threshold), the dotted line indicates the support value of 95 (one-third of the distribution is above this threshold). (B) Relation between the mean node support in a family and the size of this family, measured by the number of nodes in the family. The inner figure shows the mean node support of the smaller families. (C) Relation between branch support and branch length. Only branch lengths inferior to 1 are plotted on this figure. The inner figure shows the distribution of branch lengths justifying the focus on branch lengths inferior to 1. For all figures, only trees with more than four leaves are presented here. Branches with length smaller than 10^{-5} were considered unresolved multifurcations and were discarded.

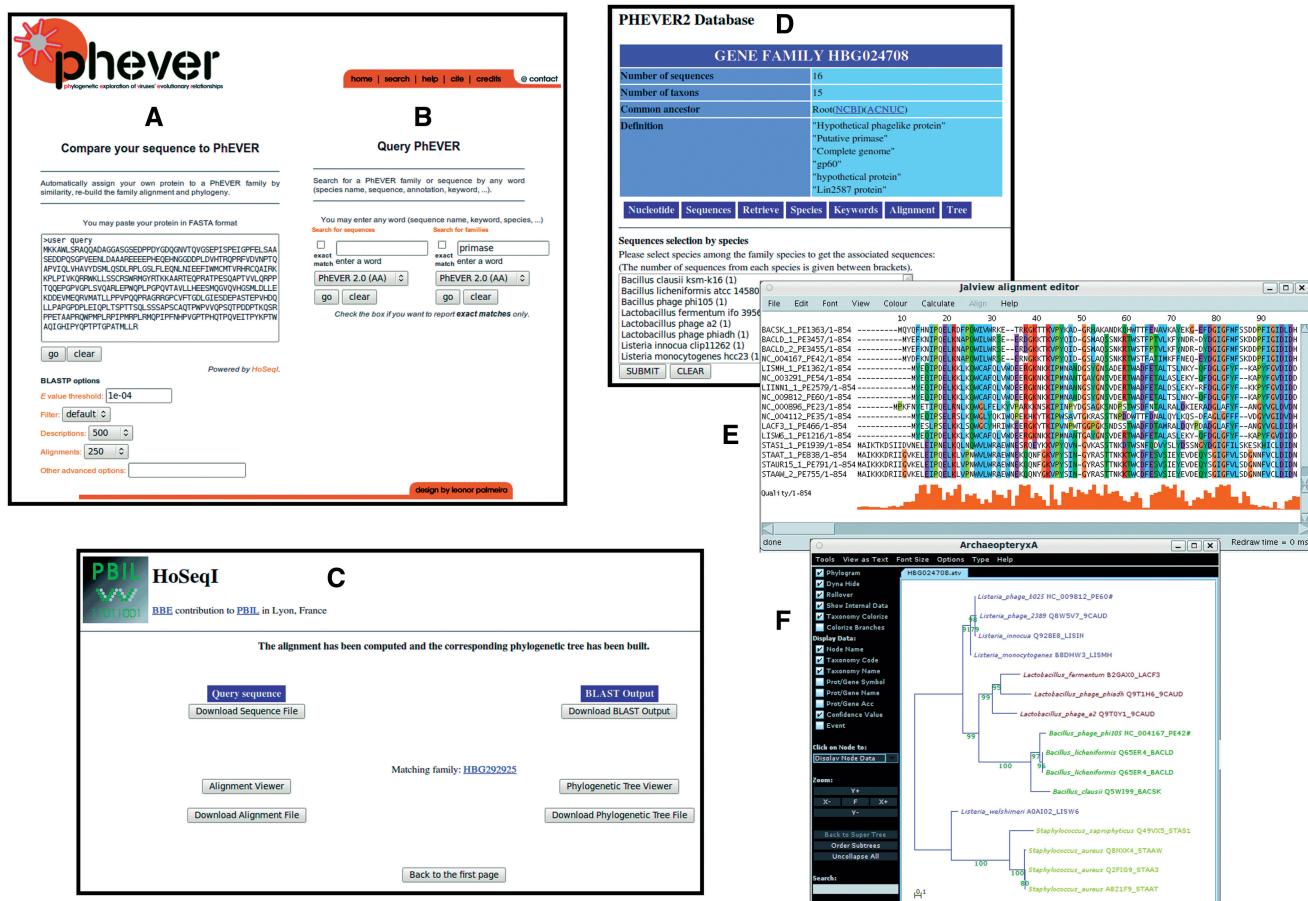


Figure 4. Overview of some of the possibilities of the PhEVER interface. (A) HoSeqI tool to query PhEVER families with a user provided sequence, (B) Query the database with terms including gene name, species name, annotation terms, (C) Alignments and phylogenies can be recomputed on the fly through the HoSeqI tool, (D) Visualization of a PhEVER family and the species and sequences composing it, (E) JalView applet to edit the pre-computed alignments and (F) ATV applet to edit the pre-computed phylogenies.

installation are provided through our ftp website or by simple request.

PHEVER WEB INTERFACE

The PhEVER web interface allows for two query forms represented in Figure 4A and B. On the one hand (Figure 4A), the HoSeqI tool allows to search in PhEVER families with a user provided query (15). This query is used to BLAST the proteins present in PhEVER and to match the most related family. Alignments and phylogenies for this new family containing the user provided sequence can be recomputed on the fly, visualized and manipulated with Java applets (Figure 4F) (31,32). On the other hand (Figure 4B), the query tool allows to directly query the database with a very diverse range of terms including gene name, annotation term, species name, protein accession number, genome accession number and family accession number. The species and sequences represented in each family detected by the query can then be viewed (Figure 4D) as well as the alignments and phylogenies which can be edited online via Java applets (Figure 4E) (31,32). More details on how to query PhEVER are presented in the Supplementary Data.

CONCLUSION AND PERSPECTIVES

PhEVER is the first open access database to provide information at the cross-taxa scale for the analysis of virus–virus and virus–host protein transfers. It compiles information from all kingdoms of life, and handles data from the genomes of all completely sequenced viruses and prokaryotes and of a large range of eukaryotes. It is the largest database of viral homologous families and offers highly accurate pre-computed alignments and phylogenies, making it a powerful tool for the analysis of horizontal gene transfer and more widely for the analysis of gene history.

Our objective is to continue the development of PhEVER around the analysis of protein evolution in the context of virus–virus and virus–host interactions. More specifically, the next step we have under development is the detection of evolutionary conserved modules in the proteins present in PhEVER. Indeed, there is strong evidence that proteins evolve in a modular way, where modules are defined as parts of proteins sharing a common evolutionary history. These modules act as small interchangeable blocks of sequences that may be combined into proteins and form novel functions (33–35). We are interested in providing a global tool allowing to analyse the weight of modular evolution in viral adaptation. We will therefore implement the detection of modules in PhEVER proteins to provide information concerning the exchanges of genetic information at the sub-protein level.

In conclusion, PhEVER aims at being a comprehensive tool for the analysis of virus–virus and virus–host relationships from an evolutionary point of view, namely through the analysis of genomic interchanges. It should become a

valuable tool for anyone working on viral evolution, but also to understand the general mechanisms behind protein evolution and functional innovation.

DATABASE ACCESS AND UPDATES

Public access and documentation is freely available through the database webpage (<http://pbil.univ-lyon1.fr/databases/phever/>) and through all dedicated ACNUC retrieval systems. More information on dedicated ACNUC retrieval systems, such as standalone query software (16), the seqinR package for R (30) or the C and Python APIs, can be obtained in the Supplementary Data or on the PhEVER webpage. The PhEVER flat files for local installation are available through our ftp server (<ftp://pbil.univ-lyon1.fr/pub/phever>) and instructions are available on the database webpage. The PhEVER database is updated every 6 months. This update frequency allows to follow the fast pace of viral and prokaryotic genome sequencing as well as to obtain updated genomic annotations for large eukaryotic genomes. Previous versions of the database remain available upon request.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

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REFERENCES

1. Sharp,P.M., Bailes,E., Chaudhuri,R.R., Rodenburg,C.M., Santiago,M.O. and Hahn,B.H. (2001) The origins of acquired immune deficiency syndrome viruses: where and when? *Philos. Trans. Roy. Soc. Lond. B Biol. Sci.*, **356**, 867–876.
2. Graham,R.L. and Baric,R.S. (2010) Recombination, reservoirs, and the modular spike: mechanisms of coronavirus cross-species transmission. *J. Virol.*, **84**, 3134–3146.
3. Holmes,E.C. (2010) The comparative genomics of viral emergence. *Proc. Natl Acad. Sci. USA*, **107(Suppl. 1)**, 1742–1746.
4. Shackelton,L.A. and Holmes,E.C. (2004) The evolution of large DNA viruses: combining genomic information of viruses and their hosts. *Trends Microbiol.*, **12**, 458–465.

5. Hamblly,E. and Suttle,C.A. (2005) The viriosphere, diversity, and genetic exchange within phage communities. *Curr. Opin. Microbiol.*, **8**, 444–450.
6. Filée,J. (2009) Lateral gene transfer, lineage-specific gene expansion and the evolution of Nucleo Cytoplasmic Large DNA viruses. *J. Invertebr. Pathol.*, **101**, 169–171.
7. Hughes,A.L., Irausquin,S. and Friedman,R. (2010) The evolutionary biology of poxviruses. *Infect. Genet. Evol.*, **10**, 50–59.
8. Klimke,W., Agarwala,R., Badretdin,A., Chetvernin,S., Ciuffo,S., Fedorov,B., Kiryutin,B., O'Neill,K., Resch,W., Resenchenk,S. et al. (2009) The National Center for Biotechnology Information's Protein Clusters Database. *Nucleic Acids Res.*, **37**(Database issue), D216–D223.
9. Tian,Y. and Dickerman,A.W. (2007) GeneTrees: a phylogenomics resource for prokaryotes. *Nucleic Acids Res.*, **35**(Database issue), D328–D331.
10. Ehlers,A., Osborne,J., Slack,S., Roper,R.L. and Upton,C. (2002) Poxvirus Orthologous Clusters (POCs). *Bioinformatics*, **18**, 1544–1545.
11. Sayers,E.W., Barrett,T., Benson,D.A., Bolton,E., Bryant,S.H., Canese,K., Chetvernin,V., Church,D.M., Dicuccio,M., Federhen,S. et al. (2010) Database resources of the National Center for Biotechnology Information. *Nucleic Acids Res.*, **38**(Database issue), D5–D16.
12. Finn,R.D., Mistry,J., Tate,J., Coggill,P., Heger,A., Pollington,J.E., Gavin,O.L., Gunasekaran,P., Ceric,G., Forslund,K. et al. (2010) The Pfam protein families database. *Nucleic Acids Res.*, **38**(Database issue), D211–D222.
13. Marchler-Bauer,A., Anderson,J.B., Chitsaz,F., Derbyshire,M.K., DeWeese-Scott,C., Fong,J.H., Geer,L.Y., Geer,R.C., Gonzales,N.R., Gwadz,M. et al. (2009) CDD: specific functional annotation with the Conserved Domain Database. *Nucleic Acids Res.*, **37**(Database issue), D205–D210.
14. Penel,S., Arigon,A.-M., Dufayard,J.-F., Sertier,A.-S., Daubin,V., Duret,L., Gouy,M. and Perrière,G. (2009) Databases of homologous gene families for comparative genomics. *BMC Bioinformatics*, **10**(Suppl. 6), S3.
15. Arigon,A.-M., Perrière,G. and Gouy,M. (2006) HoSeqI: automated homologous sequence identification in gene family databases. *Bioinformatics*, **22**, 1786–1787.
16. Gouy,M. and Delmotte,S. (2008) Remote access to ACNUC nucleotide and protein sequence databases at PBIL. *Biochimie*, **90**, 555–562.
17. Pruitt,K.D., Tatusova,T., Klimke,W. and Maglott,D.R. (2009) NCBI Reference Sequences: current status, policy and new initiatives. *Nucleic Acids Res.*, **37**(Database issue), D32–D36.
18. Sterk,P., Kulikova,T., Kersey,P. and Apweiler,R. (2007) The EMBL Nucleotide Sequence and Genome Reviews Databases. *Methods Mol. Biol.*, **406**, 1–21.
19. Hubbard,T.J.P., Aken,B.L., Ayling,S., Ballester,B., Beal,K., Bragin,E., Brent,S., Chen,Y., Clapham,P., Clarke,L. et al. (2009) Ensembl 2009. *Nucleic Acids Res.*, **37**, D690–D697.
20. Gouy,M., Gautier,C., Attimonelli,M., Lanave,C. and di Paola,G. (1985) ACNUC—a portable retrieval system for nucleic acid sequence databases: logical and physical designs and usage. *CABIOS*, **1**, 167–172.
21. UniProt Consortium. (2009) The Universal Protein Resource (UniProt) 2009. *Nucleic Acids Res.*, **37**(Database issue), D169–D174.
22. Altschul,S.F., Gish,W., Miller,W., Myers,E.W. and Lipman,D.J. (1990) Basic local alignment search tool. *J. Mol. Biol.*, **215**, 403–410.
23. Wootton,J. and Federhen,S. (1996) Analysis of compositionally biased regions in sequence databases. *Methods Enzymol.*, **266**, 554–571.
24. Baltimore,D. (1971) Expression of animal virus genomes. *Bacteriol Rev.*, **35**, 235–241.
25. Edgar,R.C. (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.*, **32**, 1792–1797.
26. Castresana,J. (2000) Selection of conserved blocks from multiple alignments for their use in phylogenetic analysis. *Mol. Biol. Evol.*, **17**, 540–552.
27. Guindon,S. and Gascuel,O. (2003) A simple, fast, and accurate algorithm to estimate large phylogenies by maximum likelihood. *Systematic Biol.*, **52**, 696–704.
28. Jones,D.T., Taylor,W.R. and Thornton,J.M. (1992) The rapid generation of mutation data matrices from protein sequences. *CABIOS*, **8**, 275–282.
29. Shimodaira,H. and Hasegawa,M. (1999) Multiple comparisons of log-likelihoods with applications to phylogenetic inference. *Mol. Biol. Evol.*, **16**, 1114–1116.
30. Charif,D. and Lobry,J. (2007) SeqinR 1.0-2: a contributed package to the R project for statistical computing devoted to biological sequences retrieval and analysis. In Bastolla,U., Porto,M., Roman,H.E. and Vendruscolo,M. (eds), *Structural Approaches to Sequence Evolution: Molecules, Networks, Populations*. Springer, NY, pp. 207–232.
31. Clamp,M., Cuff,J., Searle,S.M. and Barton,G.J. (2004) The Jalview Java alignment editor. *Bioinformatics*, **20**, 426–427.
32. Zmasek,C.M. and Eddy,S.R. (2001) ATVis: display and manipulation of annotated phylogenetic trees. *Bioinformatics*, **17**, 383–384.
33. Patthy,L. (2003) Modular assembly of genes and the evolution of new functions. *Genetica*, **118**, 217–231.
34. Moore,A.D., Björklund,A.K., Ekman,D., Bornberg-Bauer,E. and Elofsson,A. (2008) Arrangements in the modular evolution of proteins. *Trends Biochem. Sci.*, **33**, 444–451.
35. Trifonov,E.N. and Frenkel,Z.M. (2009) Evolution of protein modularity. *Curr. Opin. Struct. Biol.*, **19**, 335–340.
36. Fauquet,C., Mayo,M.A., Maniloff,J., Desselberger,U. and Ball,L.A. (eds), (2005) *Virus Taxonomy: VIIIth Report of the International Committee on Taxonomy of Viruses*. Elsevier Academic Press, Oxford, UK.