

# SporeWeb: an interactive journey through the complete sporulation cycle of *Bacillus subtilis*

Robyn T. Eijlander<sup>1,2,\*</sup>, Anne de Jong<sup>1,2</sup>, Antonina O. Krawczyk<sup>1,2</sup>, Siger Holsappel<sup>2</sup> and Oscar P. Kuipers<sup>2,\*</sup>

<sup>1</sup>Top Institute Food and Nutrition (TIFN), Nieuwe Kanaal 9A, 6709 PA Wageningen, The Netherlands and

<sup>2</sup>Department of Molecular Genetics, Groningen Biomolecular Sciences and Biotechnology Institute, University of Groningen, 9747 AG Groningen, The Netherlands

Received July 26, 2013; Revised and Accepted October 4, 2013

## ABSTRACT

**Bacterial spores are a continuous problem for both food-based and health-related industries. Decades of scientific research dedicated towards understanding molecular and gene regulatory aspects of sporulation, spore germination and spore properties have resulted in a wealth of data and information. To facilitate obtaining a complete overview as well as new insights concerning this complex and tightly regulated process, we have developed a database-driven knowledge platform called SporeWeb (<http://sporeweb.molgenrug.nl>) that focuses on gene regulatory networks during sporulation in the Gram-positive bacterium *Bacillus subtilis*. Dynamic features allow the user to navigate through all stages of sporulation with review-like descriptions, schematic overviews on transcriptional regulation and detailed information on all regulators and the genes under their control. The Web site supports data acquisition on sporulation genes and their expression, regulon network interactions and direct links to other knowledge platforms or relevant literature. The information found on SporeWeb (including figures and tables) can and will be updated as new information becomes available in the literature. In this way, SporeWeb offers a novel, convenient and timely reference, an information source and a data acquisition tool that will aid in the general understanding of the dynamics of the complete sporulation cycle.**

## INTRODUCTION

During adverse environmental conditions, bacterial cells adopt developmental strategies, such as endospore

formation, to ensure their survival. Bacterial spores are a continuous problem in food- and health-related industries because of their persistence after treatments and their ability to revert to vegetative cells through the process of germination (1,2). For instance, the disease of anthrax can persevere through the ingestion of spores that are able to survive the gastrointestinal tract and germinate to vegetative cells that produce lethal toxins (3). On the other hand, use of bacterial spores in the form of bioinsecticides (4), antigen delivery systems and vaccines (5) or probiotics (6,7) are upcoming fields that offer attractive applications. Therefore, a better understanding of the sporulation and germination processes, the level of heterogeneity therein, all genes and proteins involved, as well as influential effects of environmental factors have formed important fields of study for the past decades (8) and have provided a wealth of knowledge (9–12).

Most of the work on sporulation has been performed using the Gram-positive non-pathogenic organism *Bacillus subtilis*. The obtained data are extremely valuable and are often used as a reference model in sporulation research concerning other (pathogenic) bacteria, including *Bacillus anthracis* (13,14), *Bacillus cereus* (15,16) and various *Clostridium* species and strains that are of both medical and industrial importance (including *Clostridium difficile*, *Clostridium perfringens* and *Clostridium botulinum*) (17–22). This results in even more data and information, with various theories and speculations on molecular mechanisms, conservation of ‘core’ sporulation genes and emergence of evolutionary foundations (23,24). Sporulation of *B. subtilis* is an extremely complex cellular developmental process (11,25,26). New technological advances such as RNA sequencing, identification of small non-coding RNAs and increased understanding of processes through mathematical modelling allow us to answer questions beyond previous expectations (27–31), but simultaneously add to the complexity. Newly sequenced bacterial genomes of other

\*To whom correspondence should be addressed. Tel: +31 50 3632190; Fax: +31 50 363 2348; Email: R.T.Eijlander@rug.nl  
Correspondence may also be addressed to Oscar P. Kuipers. Tel: +31 50 3632093; Fax: +31 50 363 2348; Email: O.P.Kuipers@rug.nl

spore-formers are increasingly available due to faster and cheaper methodologies and demand efficient analyses and readily available databases for comparison purposes (32). Therefore, a general overview on how spore formation is established (including which genes and regulatory pathways are involved) is very valuable to the field, but due to the complexity and dynamics increasingly difficult to obtain.

In this work, we describe a novel knowledge and data acquisition platform called SporeWeb (<http://sporeweb.molgenrug.nl>), which focuses on all developmental stages of sporulation of *B. subtilis* from a gene regulatory point of view. Through an interactive web interface querying the SporeWeb database (details available in Supplementary Material), it offers both a textual description and a graphical representation of the sequence of events throughout spore development (Figure 1). Importantly, it easily links to more catalogued information present on other knowledge Web sites, such as SubtiWiki (<http://subtiwiki.uni-goettingen.de/>) (33). This allows the reader to grasp what happens inside the cell on the regulatory level, with additional detailed information on key regulatory proteins involved. The database-driven SporeWeb Web site is dynamic and will be updated and extended when novel scientific data become available in the future. We believe that SporeWeb will be a continuous valuable asset to the research field of bacterial sporulation and will aid in our

overall understanding of this complex developmental process.

### An interactive journey through all stages of the bacterial sporulation process

Commitment to sporulation is characterized by asymmetric cell division and expression of dedicated gene sets (34). This expression is tightly regulated in various sequential developmental stages and governed by complex biochemical communication between the two compartments of the cell (11). SporeWeb offers an interactive review of this complete process, which is the result of an extensive literature study. The homepage serves as a starting point for any sporulation stage of interest, which can be accessed by clicking on the homepage image or by selecting the 'State' in the menu bar. Subsequent pages offer both detailed descriptions and schematic representations of development. The figures are interactive and dynamic: they contain clickable items of interest and ensure updated information as genes are added to or edited in the database. Legends to the figures are described in the vertical grey bar on the right of the web page, whereas a review-like description of the sporulation state is shown below the figure, with direct links to relevant literature references.

Sporulation-specific regulators and their regulons are described on individual pages, which can be accessed by



**Figure 1.** Graphical representation of the different levels in the SporeWeb Web site.

**SPOREWEB**  
BACILLUS SUBTILIS

HOME STATE CYTOSCAPE HEATMAP REGULONS LINKS REFERENCES

Search... **Search...**

**SpoIID Commitment to sporulation and engulfment (Stages II - III)**

**Sporulation cycle of *Bacillus subtilis***

**SpoIID**

- ↳ [SpoIID Commitment](#)
- ↳ [SpoIID Engulfment](#)
- ↳ [SubtiWiki SpoIID](#)

**SpoIID DURING COMMITMENT TO SPORULATION**

The gene encoding SpoIID is part of the  $\sigma^E$  regulon and encodes a DNA binding protein that acts both positively and negatively on a subset of  $\sigma^E$ -regulated genes. It plays an important role in the progression from stage III into stage IV of sporulation by enhancing the expression of genes required for the synthesis of pro- $\sigma^K$  <sup>109</sup>. Nevertheless, the majority of the  $\sigma^E$ -dependent genes co-controlled by SpoIID are repressed by this secondary regulator <sup>95</sup>, thereby fine-tuning timing of protein synthesis during sporulation stages II and III in the mother cell. Through the action of SpoIID, genes that are involved in the process of engulfment, spore cortex synthesis and the appearance of  $\sigma^K$  and  $\sigma^C$  are expressed in pulses that can time-wise be correlated and clustered together according to functionality <sup>95</sup>. For instance, short expression pulses of genes encoding some spore outer membrane proteins early in sporulation are necessary to ensure their proper localization before the engulfment process is complete. Also the production of some coat proteins not needed until the final stages of spore coat assembly is delayed by the action of SpoIID <sup>110, 111</sup>.

**Figure 2.** The ‘SpoIID during commitment’ page on SporeWeb. Detailed information on the role of a sporulation-specific regulator during any stage in sporulation can be found on such pages. The schematic representation depicts its own regulation as well as the genes under its control; positively regulated in a blue box, negatively regulated in a red box. A detailed list containing all genes within the regulon can be accessed using the SubtiWiki link, or downloaded as an Excel file by clicking on the coloured boxes or the XLXS icon. To see what happens to this regulator during other stages in sporulation, simply click on the link provided in the grey bar on the right.

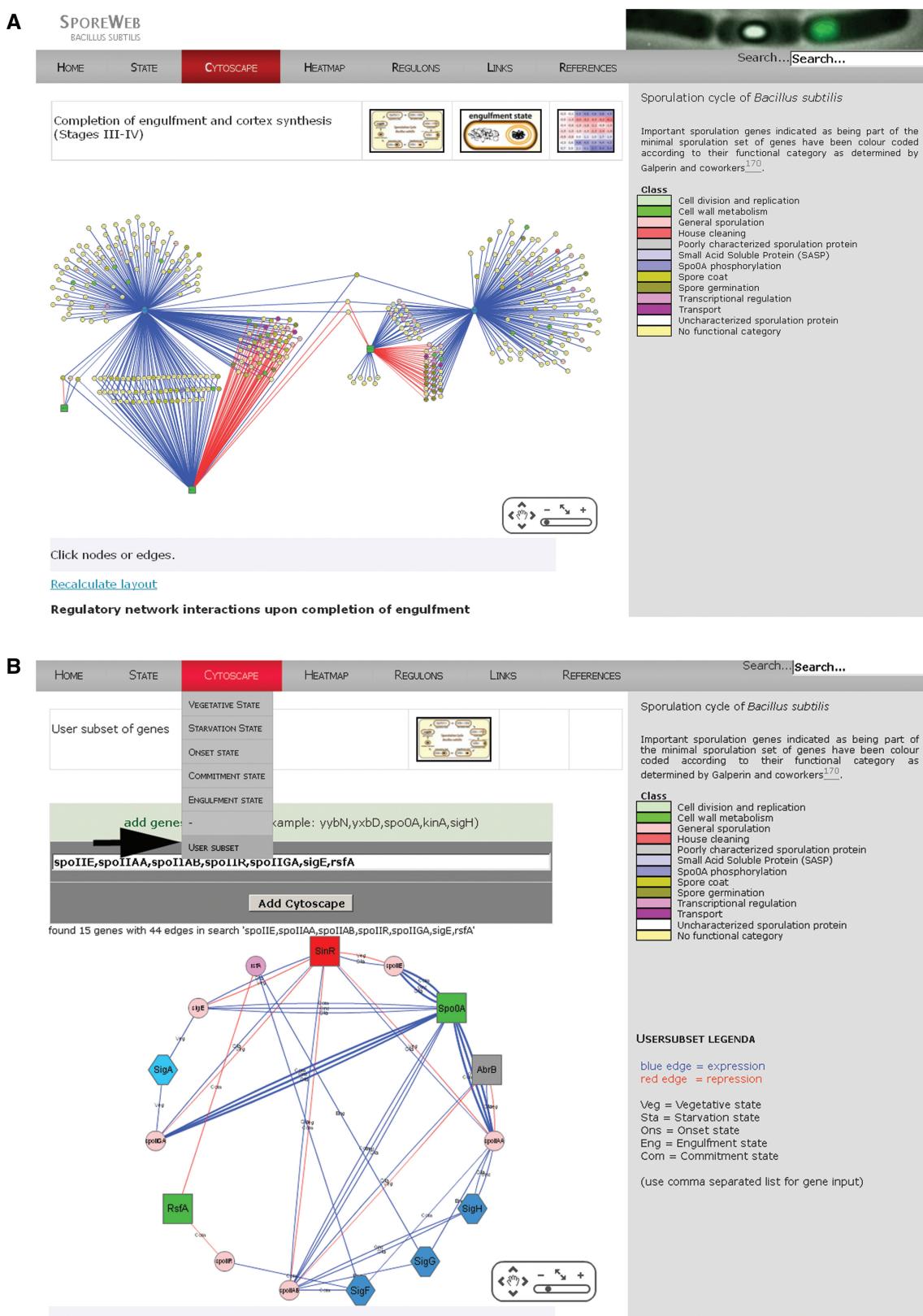
clicking on the item in the schematic figures or via the menu bar. An example of such a page is shown in Figure 2. Genes under transcriptional control of a specific regulator have been indicated in blue or red boxes for transcriptional activation or repression, respectively. Additionally, there is a direct link to the SubtiWiki list of regulon members. Lists and descriptions of all genes within the regulon can be downloaded in the form of updatable Excel sheets by clicking on the coloured boxes or via the Excel icon in the top right corner (Figure 2).

#### Graphic representations of regulon interactions define subgroups of co-regulated genes

As sporulation progresses, sporulation-specific sigma factors are expressed and activated in a spatial and temporal manner (11,26). Together with secondary regulator proteins, they control the timing, sequence and level of gene expression that are necessary for formation, maturation and release of the endospore. Various transcriptomic studies in *B. subtilis* have led to the identification of genes controlled by these sigma factors and

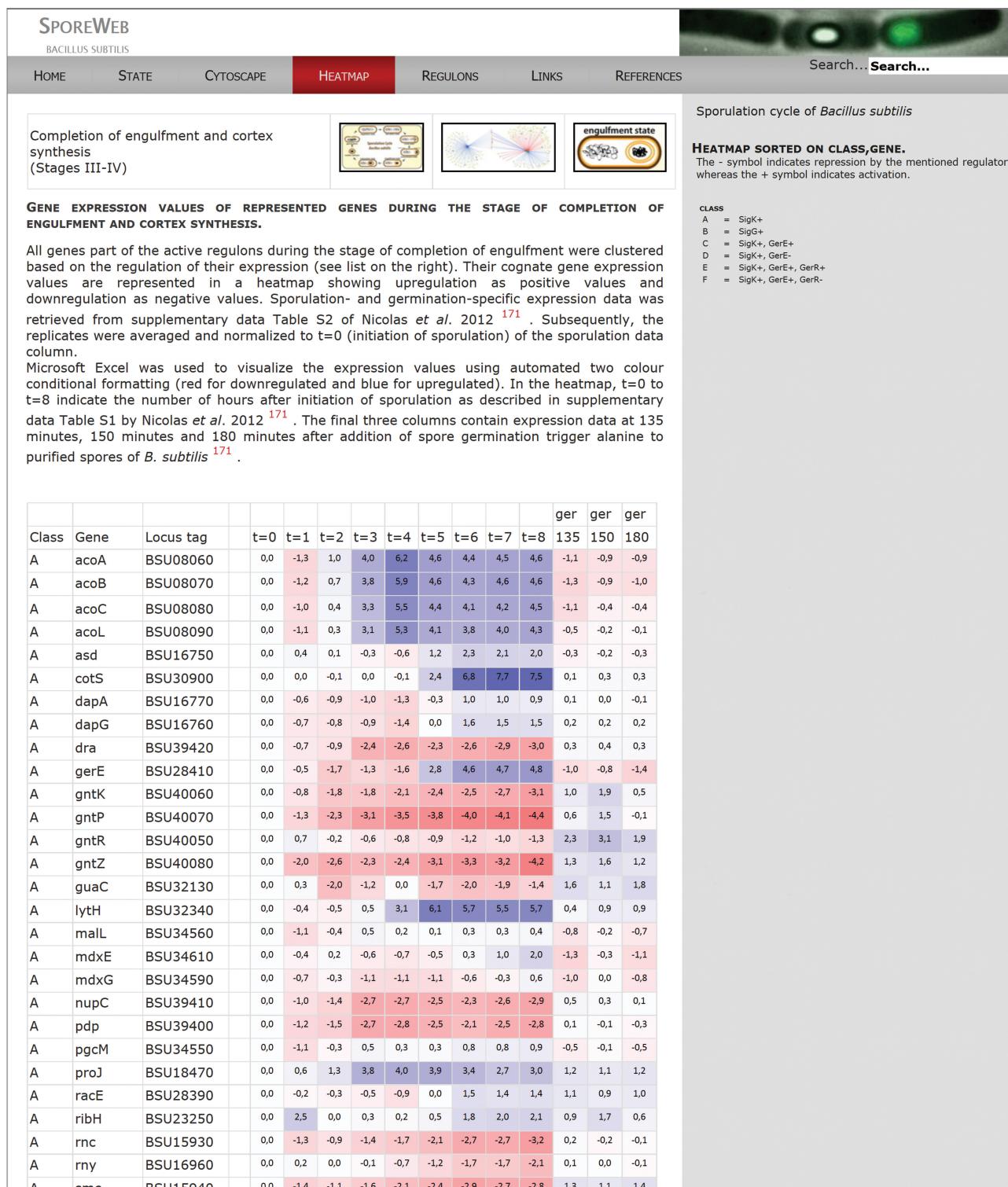
other regulators to map the sporulation gene regulatory networks (35–42). In SporeWeb, we have visualized these networks using Cytoscape-generated layouts for every sporulation-specific stage (details available in Supplementary Material) (43). These layouts can be accessed via the ‘Cytoscape’ option in the menu bar, or the Cytoscape icon on the top of every State page.

An example of such a layout is shown in Figure 3A. This representation immediately shows which genes are under single, dual or even triple or quadruple control and which genes are co-regulated during a specific sporulation stage. There is a zoom-in function that allows the user to identify genes or regulators of interest. The name and direct links to the gene SubtiWiki and MicroScope MaGe pages ([http://www.genoscope.cns.fr/agc/micro\\_scope/home/](http://www.genoscope.cns.fr/agc/micro_scope/home/)) are provided when the node is clicked. Furthermore, recently published spore-specific gene classification of the minimal sporulation gene set by Galperin *et al.* (24) has been integrated to immediately appreciate similarities and differences in regulation of functional classes of genes. Additionally, a visualization tool called ‘User Subset’ has been implemented in the Cytoscape



**Figure 3.** Cytoscape-generated layouts on gene regulatory networks. (A) Active regulators during completion of engulfment are indicated by green squares (proteins) or blue hexagons (sigma factors). Positive or negative effects on the transcription of genes (coloured circles) are indicated by connecting blue and red lines, respectively. Important sporulation genes indicated as being part of the minimal sporulation gene set have been colour-coded according to their functional category as determined by Galperin *et al.* (24). Genes unassigned to these functional categories are indicated as yellow circles. These layouts are available on SporeWeb for five different stages during spore formation. (B) A personal Cytoscape layout on specific genes of interest can be generated using the 'User Subset' option in the 'Cytoscape' menu (indicated by a black arrow). Genes (separated by a comma

(continued)



**Figure 4.** Heatmap representation of gene expression values during the engulfment state of sporulation. Tables like these are available on SporeWeb for five different stages in spore formation. Expression value data were derived from Nicolas *et al.* (45). Activation of gene expression is indicated as positive values in blue boxes, whereas downregulation is indicated as negative values in red boxes. Genes are categorized in classes listed A–L according to their documented regulation. Expression values are shown throughout the complete sporulation process ( $t = 0$ – $t = 8$ ) and for three time points taken during spore germination (ger).

**Figure 3.** Continued

only) should be typed in the white bar and will be organized in a graphical network via the ‘Add Cytoscape’ option. Coloured nodes (shapes) and edges (lines) represent genes and connections as described for Figure 3A. Three-letter abbreviations at the edges indicate during which stage in sporulation the particular regulation is relevant. For Spo0A regulation, thicker edges represent high-threshold genes, whereas thin edges represent low-threshold genes (44).

menu bar that will allow the users to generate their own Cytoscape interaction figure of specific sporulation genes of interest (Figure 3B).

#### Gene expression values during sporulation are visualized in informative heatmaps

A large group of important sporulation genes has already been identified and characterized, although many remain whose function and/or regulation is still unknown. To further visualize timing and co-regulation of gene expression during sporulation, we have displayed a recent sporulation-specific transcriptional dataset from Nicolas *et al.* (45) in colour-coded heatmaps. These heatmaps can be accessed via the ‘Heatmap’ option on the menu bar, or via the heatmap icon at the top of every State page. Genes are categorized in classes based on their previously documented regulation. Their expression values during the complete course of sporulation as well as during three time points of germination are shown in colour-coded boxes (Figure 4). In this way, differences and similarities in expression between co-regulated genes are visible and can provide clues about possible function and/or regulation of uncharacterized genes. The heatmaps can be downloaded via the Excel icon on the top right of the page.

#### Concluding remarks and perspectives

Knowledge on bacterial sporulation is rapidly growing, partly due to novel technological developments. This progress also reveals additional levels of complexity and makes it increasingly difficult to obtain a general understanding, especially for non-specialists in the field. Furthermore, rapid advances in DNA and RNA sequencing technologies have enabled faster and cheaper access to genomic- and transcriptional data of a large number of bacterial species. This leads to an expansion of our knowledge from laboratory-adapted model bacteria, such as *B. subtilis*, to more industrially or medically relevant species and strains. The data reveal high levels of conservation of certain genes or regulatory modules on the one hand and highlight important differences in gene presence/absence and regulatory events on the other, which have significant implications for the overall process of spore formation in specific groups of bacteria (23,24). The wealth of information that has been generated over decades by research on bacterial model organisms is extremely useful and usable as reference material for those bacterial species for which genetic manipulation and *in vivo* validation is less straightforward.

To provide researchers from all disciplines and expertise levels an accessible overview of the current state of knowledge on *B. subtilis* sporulation, we have constructed an interactive and graphical knowledge platform called SporeWeb. We believe that the potential of SporeWeb lies in the combination of a source of information on bacterial sporulation, an accessible starting point for further detailed investigation and a dynamic platform that can be adjusted and supplemented as new relevant data become available in the future. Moreover, the user-friendly interface and intuitive organization provide comprehensible data acquisition for both specialists and non-specialists

in the field. Future perspectives for SporeWeb can include expansion to ‘sister’ Web sites. These can contain similar content on sporulation from a proteomics point of view or of (for instance) members of the *B. cereus* group (including *B. anthracis* and *Bacillus thuringiensis*) and/or *Clostridium* species, which would be a constructive addition to the applications of SporeWeb and a valuable contribution of knowledge to the entire field of sporulation research.

#### SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

#### ACKNOWLEDGEMENTS

The authors are grateful for the time and dedication of Dr Adam Driks in thorough previewing of SporeWeb and they also thank many others in the microbiological and bacterial sporulation community who have come forward with useful suggestions and/or contributions.

#### FUNDING

TI Food and Nutrition, Wageningen, The Netherlands. Funding for open access charge: TI Food and Nutrition.

*Conflict of interest statement.* None declared.

#### REFERENCES

- Eijlander,R.T., Abee,T. and Kuipers,O.P. (2011) Bacterial spores in food: how phenotypic variability complicates prediction of spore properties and bacterial behavior. *Curr. Opin. Biotechnol.*, **22**, 180–186.
- Augustin,J.C. (2011) Challenges in risk assessment and predictive microbiology of foodborne spore-forming bacteria. *Food Microbiol.*, **28**, 209–213.
- Mock,M. and Fouet,A. (2001) Anthrax. *Annu. Rev. Microbiol.*, **55**, 647–671.
- Schnepf,E., Crickmore,N., Van Rie,J., Lereclus,D., Baum,J., Feitelson,J., Zeigler,D.R. and Dean,D.H. (1998) *Bacillus thuringiensis* and its pesticidal crystal proteins. *Microbiol. Mol. Biol. Rev.*, **62**, 775–806.
- Amuguni,H. and Tzipori,S. (2012) *Bacillus subtilis*: a temperature resistant and needle free delivery system of immunogens. *Hum. Vaccin. Immunother.*, **8**, 979–986.
- Bader,J., Albin,A. and Stahl,U. (2012) Spore-forming bacteria and their utilisation as probiotics. *Benef. Microbes.*, **3**, 67–75.
- Permpoonpattana,P., Hong,H.A., Khanuja,R. and Cutting,S.M. (2012) Evaluation of *Bacillus subtilis* strains as probiotics and their potential as a food ingredient. *Benef. Microbes.*, **3**, 127–135.
- Gould,G.W. (2006) History of science-spores. *J. Appl. Microbiol.*, **101**, 507–513.
- Higgins,D. and Dworkin,J. (2012) Recent progress in *Bacillus subtilis* sporulation. *FEMS Microbiol. Rev.*, **36**, 131–148.
- Errington,J. (2010) From spores to antibiotics via the cell cycle. *Microbiology*, **156**, 1–13.
- Hilbert,D.W. and Piggot,P.J. (2004) Compartmentalization of gene expression during *Bacillus subtilis* spore formation. *Microbiol. Mol. Biol. Rev.*, **68**, 234–262.
- Moir,A. (2006) How do spores germinate? *J. Appl. Microbiol.*, **101**, 526–530.
- Liu,H., Bergman,N.H., Thomason,B., Shallom,S., Hazen,A., Crossno,J., Rasko,D.A., Ravel,J., Read,T.D., Peterson,S.N. *et al.* (2004) Formation and composition of the *Bacillus anthracis* endospore. *J. Bacteriol.*, **186**, 164–178.

14. Fisher,N. and Hanna,P. (2005) Characterization of *Bacillus anthracis* germinant receptors *in vitro*. *J. Bacteriol.*, **187**, 8055–8062.
15. van der Voort,M., García,D., Moezelaar,R. and Abee,T. (2010) Germinant receptor diversity and germination responses of four strains of the *Bacillus cereus* group. *Int. J. Food Microbiol.*, **139**, 108–115.
16. Vries de,Y.P., Hornstra,L.M., de Vos,W.M. and Abee,T. (2004) Growth and sporulation of *Bacillus cereus* ATCC 14579 under defined conditions: temporal expression of genes for key sigma factors. *Appl. Environ. Microbiol.*, **70**, 2514–2519.
17. Xiao,Y., Francke,C., Abee,T. and Wells-Bennik,M.H. (2011) Clostridial spore germination versus bacilli: genome mining and current insights. *Food Microbiol.*, **28**, 266–274.
18. Paredes-Sabja,D., Setlow,P. and Sarker,M.R. (2011) Germination of spores of *Bacillales* and *Clostridiales* species: mechanisms and proteins involved. *Trends Microbiol.*, **19**, 85–94.
19. Steiner,E., Dago,A.E., Young,D.I., Heap,J.T., Minton,N.P., Hoch,J.A. and Young,M. (2011) Multiple orphan histidine kinases interact directly with Spo0A to control the initiation of endospore formation in *Clostridium acetobutylicum*. *Mol. Microbiol.*, **80**, 641–654.
20. Burns,D.A. and Minton,N.P. (2011) Sporulation studies in *Clostridium difficile*. *J. Microbiol. Methods*, **87**, 133–138.
21. Rosenbusch,K.E., Bakker,D., Kuijper,E.J. and Smits,W.K. (2012) *C. difficile* 630Δerm Spo0A regulates sporulation, but does not contribute to toxin production, by direct high-affinity binding to target DNA. *PLoS One*, **7**, e48608.
22. Labbé,R.G. and Dürre,P. (2005) Sporulation of clostridia. In: Dürre,P. (ed.), *Handbook on Clostridia*. CRC Press, Taylor & Francis Group, FL, USA, pp. 647–669.
23. de Hoon,M.J.L., Eichenberger,P. and Vitkup,D. (2010) Hierarchical evolution of the bacterial sporulation network. *Curr. Biol.*, **20**, R735–R745.
24. Galperin,M.Y., Mekhedov,S.L., Puigbo,P., Smirnov,S., Wolf,Y.I. and Rigden,D.J. (2012) Genomic determinants of sporulation in *Bacilli* and *Clostridia*: towards the minimal set of sporulation-specific genes. *Environ. Microbiol.*, **11**, 2870–2890.
25. Stragier,P. and Losick,R. (1996) Molecular genetics of sporulation in *Bacillus subtilis*. *Annu. Rev. Genet.*, **30**, 297–241.
26. Pigott,P.J. and Hilbert,D.W. (2004) Sporulation of *Bacillus subtilis*. *Curr. Opin. Microbiol.*, **7**, 579–586.
27. Silvaggi,J.M., Perkins,J.B. and Losick,R. (2006) Genes for small, noncoding RNAs under sporulation control in *Bacillus subtilis*. *J. Bacteriol.*, **188**, 532–541.
28. Schmalisch,M., Maiques,E., Nikolov,L., Camp,A.H., Chevreux,B., Muffler,A., Rodriguez,S., Perkins,J. and Losick,R. (2010) Small genes under sporulation control in the *Bacillus subtilis* genome. *J. Bacteriol.*, **192**, 5402–5412.
29. Liebal,U.W., Millat,T., De Jong,I.G., Kuipers,O.P., Volker,U. and Wolkenhauer,O. (2010) How mathematical modelling elucidates signalling in *Bacillus subtilis*. *Mol. Microbiol.*, **77**, 1083–1095.
30. Jabbari,S., Heap,J.T. and King,J.R. (2011) Mathematical modelling of the sporulation-initiation network in *Bacillus subtilis* revealing the dual role of the putative quorum-sensing signal molecule PhrA. *Bull. Math. Biol.*, **73**, 181–211.
31. Levine,J.H., Fontes,M.E., Dworkin,J. and Elowitz,M.B. (2012) Pulsed feedback defers cellular differentiation. *PLoS Biol.*, **10**, e1001252.
32. Rottger,R., Rückert,U., Taubert,J. and Baumbach,J. (2012) How little do we actually know? – on the size of gene regulatory networks. *IEEE/ACM Trans. Comput. Biol. Bioinform.*, **9**, 1293–1300.
33. Mäder,U., Schmeisky,A.G., Florez,L.A. and Stülke,J. (2012) SubtiWiki—a comprehensive community resource for the model organism *Bacillus subtilis*. *Nucleic Acids Res.*, **40**, D1278–D1287.
34. Parker,G.F., Daniel,R.A. and Errington,J. (1996) Timing and genetic regulation of commitment to sporulation in *Bacillus subtilis*. *Microbiology*, **142(Pt 12)**, 3445–3452.
35. Steil,L., Serrano,M., Henriques,A.O. and Völker,U. (2005) Genome-wide analysis of temporally regulated and compartment-specific gene expression in sporulating cells of *Bacillus subtilis*. *Microbiology*, **151**, 399–420.
36. Cangiano,G., Mazzone,A., Baccigalupi,L., Iстикато,R., Eichenberger,P., De Felice,M. and Ricca,E. (2010) Direct and indirect control of late sporulation genes by Gerr of *Bacillus subtilis*. *J. Bacteriol.*, **192**, 3406–3413.
37. Britton,R.A., Eichenberger,P., Gonzalez-Pastor,J.E., Fawcett,P., Monson,R., Losick,R. and Grossman,A.D. (2002) Genome-wide analysis of the stationary-phase sigma factor (sigma-H) regulon of *Bacillus subtilis*. *J. Bacteriol.*, **184**, 4881–4890.
38. Wang,S.T., Setlow,B., Conlon,E.M., Lyon,J.L., Imamura,D., Sato,T., Setlow,P., Losick,R. and Eichenberger,P. (2006) The forespore line of gene expression in *Bacillus subtilis*. *J. Mol. Biol.*, **358**, 16–37.
39. Eichenberger,P., Fujita,M., Jensen,S.T., Conlon,E.M., Rudner,D.Z., Wang,S.T., Ferguson,C., Haga,K., Sato,T., Liu,J.S. et al. (2004) The program of gene transcription for a single differentiating cell type during sporulation in *Bacillus subtilis*. *PLoS Biol.*, **2**, e328.
40. Eichenberger,P., Jensen,S.T., Conlon,E.M., van Ooij,C., Silvaggi,J., Gonzalez-Pastor,J.E., Fujita,M., Ben-Yehuda,S., Stragier,P., Liu,J.S. et al. (2003) The sigmaE regulon and the identification of additional sporulation genes in *Bacillus subtilis*. *J. Mol. Biol.*, **327**, 945–972.
41. Molle,V., Fujita,M., Jensen,S.T., Eichenberger,P., Gonzalez-Pastor,J.E., Liu,J.S. and Losick,R. (2003) The Spo0A regulon of *Bacillus subtilis*. *Mol. Microbiol.*, **50**, 1683–1701.
42. Fawcett,P., Eichenberger,P., Losick,R. and Youngman,P. (2000) The transcriptional profile of early to middle sporulation in *Bacillus subtilis*. *Proc. Natl Acad. Sci. USA*, **97**, 8063–8068.
43. Saito,R., Smoot,M.E., Ono,K., Ruscheinski,J., Wang,P.L., Lotia,S., Pico,A.R., Bader,G.D. and Ideker,T. (2012) A travel guide to Cytoscape plugins. *Nat. Methods*, **9**, 1069–1076.
44. Fujita,M., González-Pastor,J.E. and Losick,R. (2005) High- and low-threshold genes of the Spo0A regulon of *Bacillus subtilis*. *J. Bacteriol.*, **187**, 1357–1368.
45. Nicolas,P., Mäder,U., Dervyn,E., Rochat,T., Leduc,A., Pigeonneau,N., Bidnenko,E., Marchadier,E., Hoebeke,M., Aymerich,S. et al. (2012) Condition-dependent transcriptome reveals high-level regulatory architecture in *Bacillus subtilis*. *Science*, **335**, 1103–1106.