

Package ‘QCpipeline’

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Type Package

Title Utilities for the QC pipeline

Description Configuration and plotting code

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Suggests MSBVAR, GWASdata

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QCpipeline-package	<i>QC utility functions</i>
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Description

This package contains functions for the QC pipeline.

Author(s)

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bestLegendPos	<i>Best legend position</i>
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Description

Find the section of a plot with the fewest number of points for placing a legend

Usage

```
bestLegendPos(x, y)
```

Arguments

x	Points on the x axis
y	Points on the y axis

Value

Returns one of the legend placement strings from the list "bottomright", "bottom", "bottomleft", "left", "topleft", "top", "topright", "right" and "center".

Author(s)

Stephanie Gogarten

Examples

```
x <- sample(1:100, 50, replace=TRUE)
y <- sample(1:100, 50, replace=TRUE)
plot(x,y)
legend(bestLegendPos(x,y), "points", pch=1)
```

boxplotMeanSD	<i>Boxplot with mean and SD</i>
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Description

Boxplot with mean and SD

Usage

```
boxplotMeanSD(x, y, data=NULL, xlab=NULL, ylab=NULL, nSD=1, ...)
```

Arguments

x	vector or character string denoting column in data
y	vector or character string denoting column in data
data	data.frame
xlab	title for x axis (defaults to x if data is not NULL)
ylab	title for y axis (defaults to y if data is not NULL)
nSD	number of standard deviations to plot
...	additional plotting arguments

Author(s)

Jess Shen

Examples

```
age <- sample(25:55, 100, replace=TRUE)
sex <- sample(c("M", "F"), 100, replace=TRUE)
boxplotMeanSD(sex, age)

data <- data.frame(age, sex)
boxplotMeanSD("sex", "age", data)
```

dbgapAnnotation	<i>Write annotation files for dbGaP</i>
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Description

dbgapScanAnnotation and dbgapSnpAnnotation create text files appropriate for posting on dbGaP.

Usage

```
dbgapScanAnnotation(scanAnnot, dir=".",
  consentVar="consent", subjVar="subj.plink", dupVar="dup.post", omitVar="no.post",
  annotationCol="annotation", analysisCol="analysis")

dbgapSnpAnnotation(snpAnnot, dir=".",
  annotationCol="annotation", analysisCol="analysis")
```

Arguments

scanAnnot	A ScanAnnotationDataFrame .
snpAnnot	A SnpAnnotationDataFrame .
dir	A character string with the directory for file output.
consentVar	The variable in scanAnnot containing consent levels.
subjVar	The logical variable in scanAnnot indicating unique subjects to post.
dupVar	The logical variable in scanAnnot indicating duplicate scans to post.
omitVar	The logical variable in scanAnnot indicating scans to be omitted from posting.
annotationCol	The logical column in the metadata indicating which variables should be included in the annotation files.
analysisCol	The logical column in the metadata indicating which variables should be included in the analysis files.

Details

dbgapScanAnnotation writes the following files to dir:

- Sample_annotation.csv
- Sample_annotation_consent_*.csv
- Sample_annotation_duplicates.csv
- Sample_annotation_duplicates_consent_*.csv
- Sample_annotation_DD.txt
- Sample_analysis.csv
- Sample_analysis_duplicates.csv
- Sample_analysis_DD.txt

dbgapSnpAnnotation writes the following files to dir:

- SNP_annotation.csv
- SNP_annotation_DD.txt
- SNP_analysis.csv
- SNP_analysis_DD.txt

Which variables should be written to the annotation and analysis files are indicated in the metadata columns `annotationCol` and `analysisCol`.

The data dictionary files are populated from the metadata. The "type" column is automatically generated from the classes of the variables in `scanAnnot` and `snpAnnot`.

Author(s)

Stephanie Gogarten

make.allele.mappings *Make allele mapping file*

Description

Function to take Illumina build 37 SNP annotation data file and make an Allele Mappings table

Usage

```
make.allele.mappings(snp.dat)
```

```
make.allele.annotation(map, alleles=c("top", "design", "fwd", "plus"))
```

Arguments

snp.dat	a data frame made from SNP Illumina annotation file (i.e., "HumanOmni2.5-4v1_D.csv"), with following fields: "IlmnID", "Name", "IlmnStrand", "SNP", "GenomeBuild", "SourceStrand", "SourceSeq", "TopGenomicSeq", "RefStrand"
map	a data frame with allele mappings (output of make.allele.mappings), with following fields: "snp", "alle.AB", "alle.design", "alle.top", "alle.fwd", "alle.plus"
alleles	character string with type of alleles to return in the annotation table ("top", "design", "fwd", or "plus")

Details

In the case of indels, the make.allele.mappings codes A or B as "-" and the other allele with the insertion/deletion sequence. make.allele.annotation codes indels as "D" and "I" (useful for making PLINK files).

Note that "RefStrand" in build 37 annotations from Illumina is result of BLAST search of DESIGN strand. In previous strand annotations from Illumina for build 36 arrays, "BlastStrand" held +/- result of BLAST search of the SOURCE sequence.

Value

make.allele.mappings returns a data frame object with columns ("snp", "alle.AB", "alle.design", "alle.top", "alle.fwd", "alle.plus").

make.allele.annotation returns a data frame with columns "snp", "alleleA.*", "alleleB.*" where "*" = alleles.

Author(s)

Sarah Nelson

Examples

```
## Not run:
## Load in Illumina annotation
column.select <- rep("NULL", times=21)
column.select[c(1:4, 9:11, 16:18, 21)] <- NA ## only read in select columns
snp.dat <- read.csv(file="/projects/geneva/geneva_sata/SNP_annotation/Illumina/HumanOmni2.5_4v1/HumanOmni2.
skip=7, colClasses=column.select, nrow=2450000) ## last rows are for control probes
```

```

map.final <- make.allele.mappings(snp.dat)
write.csv(map.final, file="/projects/geneva/geneva_sata/SNP_annotation/Illumina/HumanOmni2.5_4v1/testfn.csv",
          row.names=FALSE,quote=FALSE)

## Create a data frame with one row per SNP, TOP alleles, and indels coded as D/I
snp.annot <- make.allele.annotation(map.final, alleles="top")

## Example allele mappings table:
#
#      snp alle.AB alle.design alle.top alle.fwd alle.plus
# rs1000000      A      T      A      T      A
# rs1000000      B      C      G      C      G
# rs1000002      A      A      A      A      T
# rs1000002      B      G      G      G      C
# rs10000023     A      T      A      T      T
# rs10000023     B      G      C      G      G

## With the following data dictionary:
# variable      type      description
# snp           text      rs id and other comparable identifiers for a snp
# alle.AB       text      A or B, per the Illumina genotyping system nomenclature
# alle.design    text      nucleotide(s) corresponding to A or B allele for design strand
# alle.top       text      nucleotide(s) corresponding to A or B allele for Illumina TOP strand
# alle.fwd       text      nucleotide(s) corresponding to A or B allele for FORWARD strand, with respect to dbSNP reference
# alle.plus      text      nucleotide(s) corresponding to A or B allele
# for PLUS(+) strand, relative to the forward direction in the human
# reference genome sequence

## End(Not run)

```

plot2DwithHist	<i>Scatterplot with density</i>
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Description

plot2DwithHist produces a scatterplot of y vs x, along with histograms of the marginal distributions of x and y.

Usage

```
plot2DwithHist(x, y, xlab=NULL, ylab=NULL, xlim=NULL, ylim=NULL,
               sublab=NULL, mn=NULL, sd=NULL, ...)
```

Arguments

x	vector of x coordinates
y	vector of y coordinates
xlab	x-axis label (defaults to variable name)
ylab	y-axis label (defaults to variable name)
xlim	x-axis limits (defaults to [min,max] of X, plus a bit of space)
ylim	y-axis limits (defaults to [min,max] of Y, plus a bit of space)
sublab	sub-label (instead of main, since there's no room)

mn	2-element vector with mean of x and y
sd	2-element vector with sd of x and y
...	additional arguments to pass to points

Author(s)

Leila Zelnick

Examples

```
library(MSBVAR)
# generate some multivariate normal example data
n <- 5000
mu <- c(0, 2)
vmat <- matrix(c(1, 0.7, 0.7, 1), nrow=2)

dat <- rmultnorm(n, mu, vmat) # generates n multivariate normal obs.
x <- dat[,1]
y <- dat[,2]

plot2DwithHist(x, y, xlab="This is the X variable", ylab="This is the Y variable.",
  sub="Example Plot!")
# defining axis limits
plot2DwithHist(x, y, xlab="This is the X variable", ylab="This is the Y variable.",
  sub="Example Plot!", xlim=c(0,4), ylim=c(-2,2))
```

readConfig	<i>Read a configuration file</i>
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Description

Read a configuration file

Usage

```
readConfig(file, ...)

setConfigDefaults(config, required, optional, default)
```

Arguments

file	file where column 1 is parameter name and column 2 is value.
...	additional arguments to read.table
config	named character vector
required	character vector of required parameter names
optional	character vector of optional parameter names
default	character vector of default values for parameters in optional

Value

`readConfig` returns a named character vector of parameter values.

`setConfigDefaults` takes a named character vector returned by `readConfig` and adds additional parameters in `optional` with values in `default`. An error will result if a parameter in `required` is missing from `config`.

Author(s)

Stephanie Gogarten

Examples

```
f <- tempfile()
write.table(cbind(letters[1:10], 1:10), file=f, quote=FALSE,
            row.names=FALSE, col.names=FALSE)
config <- readConfig(f)

required <- letters[1:5]
optional <- letters[11:15]
default <- 11:15
config <- setConfigDefaults(config, required, optional, default)

unlink(f)
```


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