

Crop adaptation through gene flow

immediate

July 6, 2016

Outline

Sugarcane

The history of sugarcane is marked with phases of concerted interspecific breeding efforts, primarily but not exclusively between *S. officinarum* and *S. spontaneum* (1)

From (2)33—*daniels1987taxonomy*”*geneticallybasedtraits*”*evidenceforhybridoriginof*”*manycultivars*””*chro*
roach1995sugar”*spontaneoushybridizationbetweencultivatedandwildSaccharuminAustralasiaandislandsofthe*

Maize

rice

Several resistance genes (grassy stunt virus, bacterial blight, brown planthopper, blast) have been introgressed from wild relatives into *O. sativa* by researchers (3; 4). Beyond investigative experiments, gene flow from wild relatives has been used to produce agronomic rice varieties. Yatsen No. 1, for example, showed resistance to pests and diseases and adapted well to environmental conditions (5) Several lines were derived from Yatsen No. 1, and went on to be utilized extensively in parts of China.

wheat

The convoluted domestication of wheat is marked by many instances of hybridization between morphologically-distinct species within a restrained geographical range. Throughout wheat domestication, there has been gene flow between species and between wild and domesticated wheats. Emmer wheat, for example, has high genetic diversity, likely due to gene flow from wild emmer (6; 7) The a, b, c, and f alleles of mildew resistance gene Pm3 may have been introgressed into wheat from wild emmer wheat shortly after domestication (although the e, d, and g alleles were probably formed by de novo mutations) (8) Furthermore, wild wheat relatives are can be found in a much broader swath than the domestication center in the Fertile Crescent (9)

potato

Introgressive hybridization is widespread in potatoes (10) Andean potatoes exhibit high ecological versatility, due in part either to allelic diversity in polyploids or introgression of desirable alleles from wild relatives in diploids (11) Cultivated diploid potatoes have been shown introgressed with genes from *S. sparsipilum* (12) and *S. megistacrolobum* (13; 14). Drought- and frost-resistance genes introgressed from wild relatives expedited the spread of cultivated potatoes into the central Andean Altiplano (15; 16; 17) Resistance genes have been experimentally introgressed from

wild potato relatives (*S. tuberosum* subsp. *andigena*, (18), *S. bulbocastanum*, (19)) into cultivated potato, although this process is hindered by compatibility issues including ploidy levels and Endosperm Balance Number (20).

Cassava

Although cultivated cassava (*Manihot esculenta*) is interfertile with some of its wild relatives (the putative progenitor species *M. flabellifolia*, for example, (21), comparisons of single-copy nuclear gene *G3pdh* sequences provide little evidence to suggest that natural introgressive hybridization occurs, and if it does, it would appear to be in the direction of cultivar to wild relative (22) Clonal propagation and allopatry are listed among the barriers to gene flow between cassava and its wild relatives [[but see tovar2015diversity for discussion on elevated genetic diversity in *M. esculenta*, gene flow between species, occasional sexual reproduction, and trade between farmers]]. Breeders have introgressed genes for protein and disease resistance genes into cassava from *M. flabellifolia* (23)

Soybean

Soybeans (*Glycine max*) were domesticated in southern China (?) Soybeans (*Glycine max*) primarily self-pollinate, outcrossing at a rate of 1-8% Soybeans readily cross with their wild progenitor, *G. soja* (24) to the degree that some researchers (?) consider the two to be conspecific. Plants of intermediate morphology appear around fields when *S. soja* is also present, indicating spontaneous hybridization (25) The accession "*G. gracilis*" shows allelic contribution from both *G. max* and *G. soja* (?) Presently, however, there is little or no evidence that natural wild-to-crop introgressive hybridization has had an adaptive impact on cultivated soybeans.

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Palm

Oil palm (*Elaeis guineensis*) of western Africa has been crossed with the New-World species *E. oleifera*, the only other species in the genus. The two species differ in vegetative and reproductive morphology and in ecological adaptations. The hybrid appears to have partial resistance to pathogenic fungal *Phytophthora palmivora*, native to Latin America (?) The F1 hybrids have moderately diminished viability and fertility, but the reproductive barrier is not fully developed (?) Since there is only one wild relative in the same genus, and has not been sympatric with *E. guineensis* for a long time it seems unlikely that adaptive introgression has played a significant role in the domestication of this plant.

Sugar Beet

Sugar beet was domesticated relatively recently Crop-wild weedy hybrids (wild alleles introgressing into crop populations) (26) Most introgression into sugar beet from wild relatives has been the

intentional product of breeding efforts for the purpose of disease resistance (a review of these efforts and outcomes can be found at (?))

barley

Wild-domesticate breeding experiments have shown that wild barleys have alleles for several important agronomic phenotypes, including brittleness, flowering time, plant height, lodging, and yield, (27; 28) Although the conditions of barley domestication would seem to allow (if not promote) natural adaptive introgression between barley and its wild relatives, there is little evidence of this genetic interaction at present. Barley/*spontaneum* hybrids are fertile, and morphologically intermediate (putatively hybrid) barleys are found when wild and domesticated barleys are grown in sympatry, but hybrids of other wild relatives generally exhibit greatly diminished fertility (26; 29)

0.1 Sweet Potato

Sweet potato (*Ipomoea batatas*) likely arose from multiple domestications, in Central America and in South America, which were secondarily hybridized upon introduction (30). [[see austin1988taxonomy for a summary of the genus *Ipomoea*, wild relatives and such.]]

Cotton

Cited (indirectly) in (?): 45 diploid species, 5 tetraploid monophyletic all tetraploids from New World two domestications, *G. hirsutum* (Mesoamerica/Caribbean) and *G. barbadense* (S. America/Caribbean)

Cited (indirectly) in (2) Evidence of introgression from domesticated cottons into wild.

Directly found by

Bidirectional interspecific introgression between cultivated and wild cottons account for a "significant" amount

Rapeseed

Domesticated rapeseed (*Brassica napus*) has many sympatric wild relatives, but reproductive barriers limit the degree of hybridization that occurs. (?) experimentally tested fertility rates between domesticated rapeseed and wild relatives. Also see (2)

Coconut

There are two types of cultivated coconut (Pacific and Indian/Atlantic), corresponding to two separate domestications, with a history of human-facilitated admixture between them (31) [[See cited references in intro of "Population Genetics, Lethal Yellowing Disease, and Relationships among Mexican and Imported Coconut Ecotypes"]]

Sorghum

(32) states that natural introgression has been an important force in sorghum evolution. Domesticated-weedy-wild complexes are common when sorghum cultivars are grown in sympatry with wild members of the *Eu-Sorghum* subgenus (33; 34; 35) Rates of outcrossing within sorghum range from 0-30 Natural introgression between wild and domesticated sorghum has been documented in both directions (36; 37; 38; 39; 40) These hybrids express diminished fertility. Members of *Sorghum bicolor* have adapted to a broad range of altitudinal, precipitation, and temperature clines across Africa and around the world (32; 9) Breeding efforts have focused on incorporating exotic germplasm

(wild sorghums as well as relatives from other genera (41) for its adaptations to biotic and abiotic stresses (42; 32; 43). Resistance to greenbug has been introgressed into cultivated sorghum through modern breeding efforts (43).

References

- [1] B. Roach, "Origin and improvement of the genetic base of sugarcane," in *Proc Aust Soc Sugar Cane Technol*, vol. 11, pp. 34–47, 1989.
- [2] N. C. Ellstrand, H. C. Prentice, and J. F. Hancock, "Gene flow and introgression from domesticated plants into their wild relatives," *Annual review of Ecology and Systematics*, pp. 539–563, 1999.
- [3] D. Brar and G. Khush, "Alien introgression in rice," in *Oryza: From Molecule to Plant*, pp. 35–47, Springer, 1997.
- [4] G. S. KHUSH and K. Ling, "Inheritance of resistance to grassy stunt virus and its vector in rice," *Journal of Heredity*, vol. 65, no. 3, pp. 135–136, 1974.
- [5] Y. Ting, "Wild rice of kwangtung and new variety bred from the hybrids of wild rice with cultivated rice," *Coll. Agric. Sun Yatsen Univ., Agron. Bull*, vol. 3, pp. 1–22, 1933.
- [6] M.-C. Luo, Z.-L. Yang, F. M. You, T. Kawahara, J. G. Waines, and J. Dvorak, "The structure of wild and domesticated emmer wheat populations, gene flow between them, and the site of emmer domestication," *Theoretical and Applied Genetics*, vol. 114, no. 6, pp. 947–959, 2007.
- [7] J. Dvorak, E. D. Akhunov, A. R. Akhunov, K. R. Deal, and M.-C. Luo, "Molecular characterization of a diagnostic dna marker for domesticated tetraploid wheat provides evidence for gene flow from wild tetraploid wheat to hexaploid wheat," *Molecular biology and evolution*, vol. 23, no. 7, pp. 1386–1396, 2006.
- [8] N. Yahiaoui, S. Brunner, and B. Keller, "Rapid generation of new powdery mildew resistance genes after wheat domestication," *The Plant Journal*, vol. 47, no. 1, pp. 85–98, 2006.
- [9]
- [10] P. Grun, "The evolution of cultivated potatoes," *Economic Botany*, vol. 44, no. 3, pp. 39–55, 1990.
- [11] K. S. Zimmerer, "The ecogeography of andean potatoes," *BioScience*, pp. 445–454, 1998.
- [12] D. Rabinowitz, C. Linder, R. Ortega, D. Begazo, H. Murguia, D. Douches, and C. Quiros, "High levels of interspecific hybridization between *Solanum sparsipilum* and *S. stenotomum* in experimental plots in the andes," *American Potato Journal*, vol. 67, no. 2, pp. 73–81, 1990.
- [13] T. Johns, Z. Huaman, C. Ochoa, and P. E. Schmiediche, "Relationships among wild, weed, and cultivated potatoes in the *Solanum x ajanhuiri* complex," *Systematic Botany*, pp. 541–552, 1987.
- [14] Z. Huamán, J. Hawkes, and P. Rowe, "*Solanum ajanhuiri*: an important diploid potato cultivated in the andean altiplano," *Economic Botany*, vol. 34, no. 4, pp. 335–343, 1980.

- [15] T. Johns and S. L. Keen, "Ongoing evolution of the potato on the altiplano of western bolivia," *Economic Botany*, vol. 40, no. 4, pp. 409–424, 1986.
- [16] J. G. Hawkes, "Origin of solanum juzepczukii buk and s curtilobum juz et buk," *ZEITSCHRIFT FUR PFLANZENZUCHTUNG-JOURNAL OF PLANT BREEDING*, vol. 47, no. 1, p. 1, 1962.
- [17] P. Schmiediche, J. Hawkes, and C. Ochoa, "Breeding of the cultivated potato species solanum x juzepczukii buk. and solanum x curtilobum juz. etbuk.," *Euphytica*, vol. 29, no. 3, pp. 685–704, 1980.
- [18] J. R. van der Voort, K. Kanyuka, E. van der Vossen, A. Bendahmane, P. Mooijman, R. Klein-Lankhorst, W. Stiekema, D. Baulcombe, and J. Bakker, "Tight physical linkage of the nematode resistance gene gpa2 and the virus resistance gene rx on a single segment introgressed from the wild species solanum tuberosum subsp. andigena cpc 1673 into cultivated potato," *Molecular plant-microbe interactions*, vol. 12, no. 3, pp. 197–206, 1999.
- [19] E. Van Der Vossen, A. Sikkema, B. t. L. Hekkert, J. Gros, P. Stevens, M. Muskens, D. Wouters, A. Pereira, W. Stiekema, and S. Allefs, "An ancient r gene from the wild potato species solanum bulbocastanum confers broad-spectrum resistance to phytophthora infestans in cultivated potato and tomato," *The Plant Journal*, vol. 36, no. 6, pp. 867–882, 2003.
- [20] S. Johnston, T. Den Nijs, S. Peloquin, and R. Hanneman Jr, "The significance of genic balance to endosperm development in interspecific crosses," *Theoretical and applied genetics*, vol. 57, no. 1, pp. 5–9, 1980.
- [21] A. C. Roa, M. M. Maya, M. Duque, J. Tohme, A. C. Allem, and M. W. Bonierbale, "Aflp analysis of relationships among cassava and other manihot species," *Theoretical and applied Genetics*, vol. 95, no. 5-6, pp. 741–750, 1997.
- [22] K. M. Olsen and B. A. Schaal, "Evidence on the origin of cassava: phylogeography of manihot esculenta," *Proceedings of the National Academy of Sciences*, vol. 96, no. 10, pp. 5586–5591, 1999.
- [23] O. A. Akinbo, *Introgression of high protein and pest resistance genes from inter-specific hybrids of Manihot esculenta ssp flabellifolia into cassava (Manihot esculenta Crantz)*. PhD thesis, University of the Free State, 2008.
- [24] R. Singh and T. Hymowitz, "The genomic relationship between glycine max (l.) merr. and g. soja sieb. and zucc. as revealed by pachytene chromosome analysis," *Theoretical and Applied Genetics*, vol. 76, no. 5, pp. 705–711, 1988.
- [25] S. Kwon, K. Im, and J. Kim, "Studies on the diversity of seed weight in the korean soybean land races and wild soybean," *Korean J Breed*, vol. 4, no. 1, pp. 70–74, 1972.
- [26] N. C. Ellstrand, *Dangerous liaisons?: when cultivated plants mate with their wild relatives*. JHU Press, 2003.
- [27] M. Von Korff, H. Wang, J. Léon, and K. Pillen, "Ab-ql analysis in spring barley: Ii. detection of favourable exotic alleles for agronomic traits introgressed from wild barley (h. vulgare ssp. spontaneum)," *Theoretical and Applied Genetics*, vol. 112, no. 7, pp. 1221–1231, 2006.

- [28] L. L. Handley, E. Nevo, J. A. Raven, R. MartInez-Carrasco, C. M. Scrimgeour, H. Pakniyat, and B. P. Forster, "Chromosome 4 controls potential water use efficiency ($\delta^{13}C$) in barley," *Journal of Experimental Botany*, vol. 45, no. 11, pp. 1661–1663, 1994.
- [29] J. R. Harlan, "The living fields," *Our agricultural heritage.-271 S*, 1995.
- [30] C. Roullier, A. Duputié, P. Wennekes, L. Benoit, V. M. F. Bringas, G. Rossel, D. Tay, D. McKey, and V. Lebot, "Disentangling the origins of cultivated sweet potato (*ipomoea batatas* (l.) lam.)," *PLoS One*, vol. 8, no. 5, p. e62707, 2013.
- [31] B. F. Gunn, L. Baudouin, and K. M. Olsen, "Independent origins of cultivated coconut (*cocos nucifera* l.) in the old world tropics," *Plos one*, vol. 6, no. 6, p. e21143, 2011.
- [32] I. P. PO, "Sorghum in the eighties," 1982.
- [33] J. De Wet, "Systematics and evolution of sorghum sect. sorghum (gramineae)," *American Journal of Botany*, pp. 477–484, 1978.
- [34] H. Doggett and B. Majisu, "Disruptive selection in crop development," *Heredity*, vol. 23, no. 1, pp. 1–23, 1968.
- [35] H. G. Baker, "Human influences on plant evolution," *Economic Botany*, vol. 26, no. 1, pp. 32–43, 1972.
- [36] L. Kuhlman, B. Burson, P. Klein, D. Stelly, and W. Rooney, "Interspecific sorghum breeding using *s. macrospermum*," in *Proceedings of the ASA-CSSA-SSA 2006 International Meetings, Indianapolis*, pp. 12–16, 2006.
- [37] P. Aldrich and J. Doebley, "Restriction fragment variation in the nuclear and chloroplast genomes of cultivated and wild sorghum bicolor," *Theoretical and Applied Genetics*, vol. 85, no. 2-3, pp. 293–302, 1992.
- [38] P. Aldrich, J. Doebley, K. Schertz, and A. Stec, "Patterns of allozyme variation in cultivated and wild sorghum bicolor," *Theoretical and Applied Genetics*, vol. 85, no. 4, pp. 451–460, 1992.
- [39] H. Doggett *et al.*, *Sorghum*. No. 2. ed., Longman Scientific and Technical, 1988.
- [40] H. G. Baker, "Migrations of weeds," *Valentine, D, H ed (s). Taxonomy phytogeography and evolution*, pp. 327–347, 1972.
- [41] J. De Wet, S. Gupta, J. Harlan, and C. Grassl, "Cytogenetics of introgression from *saccharum* into sorghum," *Crop Science*, vol. 16, no. 4, pp. 568–572, 1976.
- [42] V. G. Reddy, H. Upadhyaya, and C. Gowda, "Current status of sorghum genetic resources at icrisat: their sharing and impacts," *International Sorghum and Millets Newsletter*, vol. 47, pp. 9–13, 2006.
- [43] J. W. Johnson and G. L. Teetes, "Breeding for arthropod resistance in sorghum," *Harris, MK Biology and breeding for resistance to arthropods and pathogens in agricultural plants. Texas, USA*, pp. 168–180, 1979.