# Crop adaptation through gene flow

immediate

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# Outline

### Introduction

"Whenever sympatric populations of crops and their wild relatives show a similar trait, there will be several possible explanations: (1) retention of an ancestral trait, (2) convergent or parallel evolution, and (3) introgression. Perhaps even more difficult is establishing the direction of introgression; does gene flow go from crop into wild species, the reverse, or in both directions?" and "For this reason, introgression may be best studied between crops and their more distant wild relatives where the probability of finding taxon-specific genetic markers is increased." (1)

# Maize

Supplement Li's work with the following ideas:

Maize is a classical model in crop species to study the issue of adaptive introgression from wild relatives, as it still grows in sympatry with many of its wild relatives (?).

Maize was domesticated about 10,000 years ago in Mexico (2) (3) in the Balsas River valley (4) from teosinte.

Mexican farmers have long been aware of maize-teosinte hybrids, and in some places, such as the Nobogame valley, farmers have observed desirable traits passed to their crops through the hybrid plants (5) (6) (7).

In particular, Zea mays ssp. mexicana (referred to mexicana hereafter) and Mexican highland maize have become a well-studied pair (although ssp. parvaglumis has also been shown to hybridize with maize, (5)).

Introgression between mexicana and Mexican highland maize has been reported based on evidence from both morphological data (???) and molecular analysis (?????). [[Li's and my bib tags might be in conflict, e.g. wilkes1977 is likely my wilkes1977hybridization.]]. Doebley ((8)) found that north-central high-elevation Mexican landraces (Apachito, Arrocillo Amarillo, Azul, Celaya, Chaloqueno, Conico, Conico Norteno, Gordo, Harinoso de Ocho Occidentales)[[special characters]] show evidence of introgression of two alleles from mexicana, and suspected that they might have fitness advantages for adapting to highland environmental conditions.

Population structure analyses by (4) estimate that *mexicana* gene flow has contributed to the gene pools of the Cacahuacintle, Palomero de Jalisco, and Palomero Taloqueno landraces, but also into (non-landrace) Mexican maize at the same elevation.

However, (?) for the first time revealed the evidence of adaptive introgression from *mexicana* to Mexican highland maize. The authors identified nine genomic regions, which showed evidence

of introgression from mexicana to maize in both the HAPMIX and the linkage model of STRUC-TURE analyses with over seven sympatric population pairs among the total nine pairs sampled. Among the nine regions, three spans the centromeres of chromosomes 5, 6, and 10, and one is located in the inversion polymorphism on chromosome 4, suggesting a significant role of genome structures restricting recombination in adaptive introgression. By further characterizing the nine introgression regions, it is found that most regions contain long tracts of zero diversity, enriched with QTL linked with anthocyanin content and leaf macrohairs (?) and over-represented with the SNPs demonstrating high association with temperature seasonality. Growth chamber experiments with maize populations with introgression from mexicana on chromosome 4 (associated with QTL controlling pigment density and macrohairs) and 9 (overlapped with QTL for macrohairs) exhibited more macrohairs and greater pigmentation under the highland environmental settings than the populations with absence of introgression from mexicana.

Gene flow between mexicana and maize is low enough that only fitness-neutral or advantageous alleles are likely to be passed (9).

From (10) Gene flow is probably greater from teosinte to maize. Certain morphological and flower timing characterists promote genetic isolation between maize and teosinte and slow rates of introgression. Gene flow from maize to teosinte occurs most easily when teosinte pollinates maize [[what paper can i cite to mention the gene tb1 and its role in hybridization, viz a viz restricting directionality of teosinte pollen into maize

From (11), bitocchi2009introgression Maize-maize introgression Research on introgression between highland and lowland maize shows adaptive traits being transferred

(12) has a good review of the history of domestication, introduction to Europe, and further development. (13) cites an example in Italy of one domesticated maize adapting to coastal environment by introgressions from another domesticated maize.

#### wheat

The first domesticated wheat was einkorn wheat ( $Tritium\ monococcum$ ), domesticated from wild einkorn ( $T.\ m.\ boeoticum$ ) in or around the Karacadag mountains of southeast Turkey (14; 15; 16) Subsequently, emmer wheat ( $T.\ dicoccum$ ) was domesticated from hybrids of diverse populations of wild emmer wheat ( $T.\ dicoccides$ ), also in the Fertile Crescent (17; 18; 19; 16) Two distinct taxa of  $T.\ dicoccides$  exist (a western one and a central-eastern one), but the central-eastern one was the progenitor species from which modern wheat was domesticated (20) The tetraploid durum wheat ( $T.\ turgidum$ ) is the result of hybridization between red wild einkorn wheat ( $T.\ urartu$ ) and wild goatgrass ( $Aegilops\ speltoides$ ) (21; 22) and a further hybridization with Tausch's goatgrass ( $A.\ tauschii$ ) produced the allohexaploid common bread wheat ( $T.\ aestivum$ ), the most agronomically important race of wheat (23; 24; 25) This hexaploidization event probably occurred twice, once in southeast Turkey/northern Syria and, more recently, in Iran (26) Each of the three genomes that comprise bread wheat shows genomic signatures of ancestral hybridizations as well (27).

The convoluted domestication of wheat is marked by many instances of hybridization between morphologically-distinct species within a restrained geographical range. Throughout wheat domestication, there has been gene flow between species and between wild and domesticated wheats. Emmer wheat, for example, has high genetic diversity, likely due to gene flow from wild emmer (19; 25) The a, b, c, and f alleles of mildew resistance gene Pm3 may have been introgressed into wheat from wild emmer wheat shortly after domestication (although the e, d, and g alleles were probably formed by de novo mutations) (28) Furthermore, wild wheat relatives are can be found in a much broader swath than the domesication center in the Fertile Crescent (29) Modern breeding efforts between wild and domesticated wheat have shown some success. For example, some

researchers have experimentally bred resistance traits into durum wheat (stem rust resistance from wild einkorn, (30) and powdery mildew resistance from *T. turgidum var. dicoccoides*, (31).

# barley

Domesticated and wild barleys belong to the same species, *Hordeum vulgare*, and are biologically capable of producing viable offpsring via hybridizaion (32). Barley (*Hordeum vulgare subsp. vulgare*) is believed to have been domesticated at least twice, once from wild *subsp. spontaneum* in the Fertile Crescent and once by *subsp. spontaneum var. agriocrithon* in Tibet roughly 10,000 years ago (33; 34; 35; 36; 37) but many of the details of barley domestication are still disputed.

Variety agriocrithon (*H. vulgare ssp. vulgare f. agriocrithon*) is genetically diverse, and is found throughout much of the range of Barley. Some have suggested that agriocrithon may be the progenitor of six-rowed barley, the product of a hybridization between eastern and western cultivated barley, or a wild-domesticate hybrid (38; 39; 40) but (36) dispute these theories, suggesting instead that it arose from *ssp. spontaneum*, perhaps more than once (in Israel as well as in Tibet). Presently, the distribution of *spontaneum* consists of the Mediterranean, the Middle-East, and West-Central Asia, while other barley wild relatives inhabit Asiatic regions (including Tibet) more broadly (41; 42; 29) These wild relatives inhabit regions spanning such abiotic clines as temperature, precipitation, soil type, and altitude, as well as biotic clines (41).

The barley domestication process has reduced the number of alleles in the domesticate to only 40Wild-domesticate breeding experiments have shown that wild barleys have alleles for several important agronomic phenotypes, including brittleness, flowering time, plant height, lodging, and yield, (43; 44)Ålthough the conditions of barley domestication would seem to allow (if not promote) natural adaptive introgression between barley and its wild relatives, there is little evidence of this genetic interaction at present. Owing to this sparcity of evidence is the low rate of outcrossing of spontaneum, estimated by (45) to be 1.6There has been little genetic investigation into spontaneous barley/spontaneum hybrids (46)Barley/spontaneum hybrids are fertile, and morphologically intermediate (putatively hybrid) barleys are found when wild and domesticated barleys are grown in sympatry, but hybrids of other wild relatives generally exhibit greatly diminished fertility (46; 42)Even when the two are not grown immediately adjacent to one another, introgression from wild to domesticate has been shown to happen over distances of more than a kilometer (47).

### sunflower

The common sunflower (Helianthus annuus) shows evidence of domestication in eastern United States (48; 49) with additional evidence of a possible second origin of domestication in Mexico (50)The species H. annuus is a versatile species, capable of adapting to a wide range of environments across the Americas, Europe, Asia, and Australia (51). This versatility may be due in part either to phenotypic plasticity or genetic adaptability (52)Domesticated sunflower lives in sympatry with wild relatives like H. petiolaris and H. bolanderi and forms stable hybrid populations (53; 54; 55)Wild sunflowers are locally adapted, and weedy hybrid populations share these adaptations (51)The species H. annuus shows signs of persistent introgressive hybridization with H. petiolaris with evidence of positive selection driving some of the genetic differentiation between the two species (56)The species H. annuus adapted to the environment in Texas by hybridizing with cucumberleaf sunflower (H. debilis ssp. cucumerifolius), gaining advantageous alleles (these hybrids are now called H. annuus ssp. texanus, (57; 58; 59; 60).

Helianthus has several genes for downy mildew resistance. Each imparts resistance to one or more races of Plasmopara halstedii, one of the most agronomically important diseases in sunflower

cultivation (61) Some of these downy mildew resistence genes were found in wild relatives (including H. argophyllus, H. tuberosus, and H. praecox) and have been successfully bred into modern H. annus (62) PlArg, an allele found in wild silverleaf sunflowers (H. argophyllus, inbred line Arg1575-2), confers resistance to all known (20 or more) races of downey mildew (63) while others (Pl1-Pl11) are effective for one or more types (64) Silverleaf sunflower has also been the focus of drought resistance breeding efforts (65) and Phomopsis resistance breeding efforts (66).

#### rice

There are two main cultigens of rice; Asian rice (Oryza sativa) and African rice (O. glaberrima), which are easily distinguish by ligule length, number of secondary panicle branches, panicle axis thickness, and differences in life cycles (35). The centers of rice domestication are not known with complete confidence, but genetic and archaeobotanical evidence points towards both the Yangzee Basin in China and the Ganges plains in India for O. sativa, 8,200-13,500 years ago, from wild O. rufipogon (35; 67; 68; 69) or "the Asian form of O. perennis complex" (35) and the Upper Niger River delta in Mali, Africa for O. glaberrima, 2-3,000 years ago, from a wild ancestor, perhaps O. barthii (68) or O. breviligulata (35) Asian rice has two main subspecies, Indica (with subpopulations indica and aus) and Japonica (with subpopulations temperate japonica, tropical japonica known as javanica, and aromatic) (70; 71; 68). Indica and Japonica are likely the result of independent domestications from separate O. rufipogon populations in India/Indochina and southern China, respectively (72) These subspecies display adaptations to the environmental coniditions corresponding to the differentiated geographical locations they inhabit (73).

Both Asian and African rice naturally hybridize with other domesticated subspecies and with wild relatives (of which there are about twenty (68) and introgression is common (35; 74; 75) The greater genetic diversity within *O. sativa* is likely due to introgression with wild relatives both during domestication and upon the dispersal of *O. sativa* into new environments and sympatry with new relatives (74). Several resistance genes (grassy stunt virus, bacterial blight, brown planthopper, blast) have been introgressed from wild relatives into *O. sativa* by researchers (76; 77). Beyond investigative experiments, gene flow from wild relatives has been used to produce agronomic rice varieties. Yatsen No. 1, for example, showed resistance to pests and diseases and adapted well to environmental conditions (78) Several lines were derived from Yatsen No. 1, and went on to be utilized extensively in parts of China.

### rye

Rye domestication has received comparatively little academic interest. Although there is little evidence for or against the role of natural introgression in rye domestication, rye (*Secale cereale*) has wild relatives outside its center of domestication in or around the Fertile Crescent (79) and the species contains domesticated, weedy, and wild rye subspecies (80).

#### potato

Modern potato (Solanum tuberosum) is believed to have been domesticated in southern Peru in sympatry with a multitude of wild relatives about 6000 years ago, although the exact location and formal classification and phylogenetic relationships between these taxa have long been disputed (81; 82; 83; 84) The northern members of the polyphyletic S. brevicaule wild potato complex have been identified as likely progenitors of modern potatoes (85), but determination of a single progenitor species is unlikely, either because widespread gene flow in the complex will mask signs of this phylogeny or because s. tuberosum has a polyphyletic origin. Also, these northern members of the

S. breviaule complex are not clearly defined, and may in fact be one singular species (in which case the species name would be S. bukasovii (82).

Although potatoes are usually propogated clonally, farmers also promote sexual hybridization at times to improve disease resistance and develop new cultivars (86) Farmers continue to grow potatoes in close proximity to wild relatives, resulting in domesticate-weedy-wild hybrid complexes which promote introgressive hybridizatiion (87; 88; 89) These complexes, combined with a diverse range of biotic and environmental selective pressures and local farming practices (human-mediated migration, isolated farmsteads in fertile valleys, clonal propogation, and intentional maintenance of a variety of landraces), have fostered expansion of genetic diversity within potatoes subsequent to domestication (90) However, as farmers tend to abandon fields after being used for potato cultivation, it is less likely that hybrids have an opportunity to form stable populations for maintained introgressive gene flow into the domesticated potato gene pool (90) Also, natural selection may be subverted by farmer preference and artificial selection (91).

Introgressive hybridization is widespread in potatoes (92) The various cultivars of Andean potatoes are interfertile, forming one large plastic gene pool (86) Andean potatoes exhibit high ecological versatility, due in part either to alleleic diversity in polyploids or introgression of desirable alleles from wild relatives in diploids (93) Cultivated diploid potatoes have been shown introgressed with genes from S. sparsipilum (87) and S. megistacrolobum (88; 94). Drought- and frost-resistance genes introgressed from wild relatives expediated the spread of cultivated potatoes into the central Andean Altiplano (95; 96; 97) Resistance genes have been experimentally introgressed from wild poptato relatives (S. tuberosum subsp. andigena, (98), S. bulbocastanum, (99)) into cultivated potato, although this process is hindered by compatability issues including ploidy levels and Endosperm Balance Number (100).

#### tomato

The domestication center for tomato (Solanum lycopersicum) is not known with certainty, but (101) quotes Dr. Charles Rick (University of California, August 1978) stating that tomato was domesticated from Solanum lycopersicum cerasiforme in the New World (most likely Mexico) [[http://www.landscapeimagery.com/tomato.html]]. (102; 103) and [[http://www.landscapeimagery.com/tomato.h agree with this hypothesis. (104), however, conclude that v. cerasiformae is not the direct ancestor of the domesticated tomato, but rather a hybrid between wild tomatoes L. esculentum and L. pimpinellifolium. Widescale recent introgression between wild relatives confound attempts at elucidating phylogenies.

Since the work of Charles Rick in 1940s and 50s (105) breeding efforts have focused on incorporating and utilizing the relatively diverse genetic germplasm of wild tomato relatives (106; 107; 108) Wild tomato relatives have been shown to harbor resistance genes for more than 40 agronomically important diseases, and breeders are working to introgress them into cultivated tomato (?)  $\dot{A}$ lso, (109) mapped a tomato yellow leaf curl virus resistance gene in an accession of L. hirsutum f. glabratum (110) to an introgression on chromosome 11, which originated from the wild relative L. hirsutum. Similar resistance traits has also been shown to have been imparted from L. chilense, L. pimpinellifolium, and L. cheesmanii (109).

### Common Bean

Common bean (*Phaseolus vulgaris*) was domesticated from a wild common bean at least twice, once in the Lerma-Santiago Basin of Mesoamerica and once in the southern Andes (111) The domesticated common bean progenitor species (*P. vulgaris var. aborigineus*) and other wild relative species can

be found across the breadth of this region and beyond, from Mexico to Argentina, inhabiting a broad range of environments (112; 29)Because common bean is generally self-pollinating (outcrossing at an estimated rate under 3These expectations are supported by evidence provided by (113)However, hybrid populations can still be found in sympatry with wild and domesticated common bean, forming domesticate-hybrid-weedy complexes.

Some evidence for introgression exists in the direction of domesit to wild common bean, leading to increased seed size, an adaptation that permitted their spread to higher elevations with cooler climates (114) Wild common beans with this introgression are also occasionally harvested along with cultivated common bean and consumed by subsistence farmers (113) Also, (115) found via STRUCTURE analysis that certain Andean wild common bean accessions were possibly the result of hybridizations with domesticated common bean. The disparity in gene flow directionality may be due in part to the large population size of common beans in a cultivated field relative to the population size of wild common beans within pollen dispersal range (a distance that may be quite small, (113). Farmer practices may also be diminishing introgressive gene flow. Though some hybrids show heterosis, others (especially F1 hybrids) do not, expressing smaller plant size, smaller seed size, and increased mortality (116; 117) Because F1 hybrids are intermediate in seed size and color, farmers may effectively identify and avoid hybrid stock.

Though limited, there is also some evidence of natural introgression of wild genes into domesticated common bean. (113) was unable to determine how much of the genetic variability of domesticated common bean landraces originated from weedy or wild types, but point to the allele for 'L' phaseolin as a possible example of an allele that has been introgressed from wild common beans into domesticated. Experimentally, common leaf blight and white mold resistances have been bred into common bean from scarlet runner bean (*P. coccineus*) (118; 119) as was resistance to weevils (*Zabrotes subfasciatus* and *Acanthoscelides obtectus*) from Mexican wild bean species (120) but it may take careful, larger-scale breeding programs to exploit heterotic hybrids for agronomic gain (117).

## Sorghum

Anthropological evidence points to a domestication event of sorghum (Sorghum bicolor subsp. bicolor) some 5-6,000 years ago in Ethiopian Sudan (121)However, comparative isozyme analyses (122) have suggested a possible secondary domestication event (sorghum race kafir) in the southeastern Savanna. The progenitor species was S. arundinaceum (123; 124)The genus Sorghum contains 25 species, but these are split between five subgenera: Eu-Sorghum, Chaetosorghum, Heterosorghum, Para-Sorghum, and Stiposorghum, and it is Eu-Sorghum that includes all domesticated, weedy, and closely-related wild relatives (125; 126). Within Eu-Sorghum, there are currently nine races, five cultivated (bicolor, guinea, kafir, caudatum, and durra) and four wild (arundinaceum, virgatum, aethiopicum, and verticilliflorum) (121)Members of Sorghum bicolor are interfertile, though outcrossing rates and fertility can be low due to ploidy incompatability (123; 127).

(128) states that natural introgression has been an important force in sorghum evolution. Domesticate-weedy-wild complexes are common when sorghum cultivars are grown in sympatry with wild members of the Eu-Sorghum subgenus (129; 130; 131) Rates of outcrossing within sorghum range from 0-30Natural introgression between wild and domesticated sorghum has been documented in both directions (132; 133; 134; 123; 135) These hybrids express diminished fertility. Members of Sorghum bicolor have adapted to a broad range of altitudinal, precipitation, and temperature clines across Africa and around the world (128; 29) Breeding efforts have focused on incorporating exotic germplasm (wild sorghums as well as relatives from other genera (136) for its adaptations to biotic

and abiotic stresses (137; 128; 138). Resistance to greenbug has been introgressed into cultivated sorghum through modern breeding efforts (138).

# References

- [1] J. Doebley, "Molecular evidence for gene flow among zea species," *BioScience*, pp. 443–448, 1990.
- [2] B. D. Smith, "The initial domestication of cucurbita pepo in the americas 10,000 years ago," *Science*, vol. 276, no. 5314, pp. 932–934, 1997.
- [3] M. B. Hufford, X. Xu, J. Van Heerwaarden, T. Pyhäjärvi, J.-M. Chia, R. A. Cartwright, R. J. Elshire, J. C. Glaubitz, K. E. Guill, S. M. Kaeppler, et al., "Comparative population genomics of maize domestication and improvement," Nature genetics, vol. 44, no. 7, pp. 808–811, 2012.
- [4] Y. Matsuoka, Y. Vigouroux, M. M. Goodman, J. Sanchez, E. Buckler, and J. Doebley, "A single domestication for maize shown by multilocus microsatellite genotyping," *Proceedings of the National Academy of Sciences*, vol. 99, no. 9, pp. 6080–6084, 2002.
- [5] H. Wilkes, "Hybridization of maize and teosinte, in mexico and guatemala and the improvement of maize," *Economic Botany*, pp. 254–293, 1977.
- [6] C. Lumholtz, Unknown Mexico: a record of five years' exploration among the tribes of the western Sierra Madre; in the Tierra Caliente of Tepic and Jalisco; and among the Tarascos of Michoacan, vol. 2. C. Scribner's sons, 1902.
- [7] H. G. Wilkes, "Teosinte introgression in the maize of the nobogame valley," *Botanical Museum Leaflets, Harvard University*, pp. 297–311, 1970.
- [8] J. Doebley, M. M. Goodman, and C. W. Stuber, "Patterns of isozyme variation between maize and mexican annual teosinte," *Economic Botany*, vol. 41, no. 2, pp. 234–246, 1987.
- [9] M. Slatkin, "Gene flow and the geographic structure of natural populations," *Science*, vol. 236, no. 4803, pp. 787–792, 1987.
- [10] B. M. Baltazar, J. de Jesus Sanchez-Gonzalez, L. de la Cruz-Larios, and J. B. Schoper, "Pollination between maize and teosinte: an important determinant of gene flow in mexico," *Theoretical and Applied Genetics*, vol. 110, no. 3, pp. 519–526, 2005.
- [11] C. Jiang, G. Edmeades, I. Armstead, H. Lafitte, M. Hayward, and D. Hoisington, "Genetic analysis of adaptation differences between highland and lowland tropical maize using molecular markers," *Theoretical and Applied Genetics*, vol. 99, no. 7-8, pp. 1106–1119, 1999.
- [12] E. Bitocchi, L. Nanni, M. Rossi, D. Rau, E. Bellucci, A. Giardini, A. Buonamici, G. G. Vendramin, and R. Papa, "Introgression from modern hybrid varieties into landrace populations of maize (zea mays ssp. mays l.) in central italy," *Molecular Ecology*, vol. 18, no. 4, pp. 603–621, 2009.
- [13] K. Hammer, P. Perrino, and D. Pignone, "Collection of plant genetic resources in south italy, 1986," *Die Kulturpflanze*, vol. 35, no. 2, pp. 389–399, 1987.

- [14] M. Heun, R. Schäfer-Pregl, D. Klawan, R. Castagna, M. Accerbi, B. Borghi, and F. Salamini, "Site of einkorn wheat domestication identified by dna fingerprinting," *Science*, vol. 278, no. 5341, pp. 1312–1314, 1997.
- [15] J. R. Harlan and D. Zohary, "Distribution of wild wheats and barley," Science, vol. 153, no. 3740, pp. 1074–1080, 1966.
- [16] H. Özkan, A. Brandolini, R. Schäfer-Pregl, and F. Salamini, "Aflp analysis of a collection of tetraploid wheats indicates the origin of emmer and hard wheat domestication in southeast turkey," *Molecular Biology and Evolution*, vol. 19, no. 10, pp. 1797–1801, 2002.
- [17] S. Lev-Yadun, A. Gopher, and S. Abbo, "The cradle of agriculture," *Science*, vol. 288, no. 5471, p. 1602, 2000.
- [18] P. Civáň, Z. Ivaničová, and T. A. Brown, "Reticulated origin of domesticated emmer wheat supports a dynamic model for the emergence of agriculture in the fertile crescent," 2013.
- [19] M.-C. Luo, Z.-L. Yang, F. M. You, T. Kawahara, J. G. Waines, and J. Dvorak, "The structure of wild and domesticated emmer wheat populations, gene flow between them, and the site of emmer domestication," *Theoretical and Applied Genetics*, vol. 114, no. 6, pp. 947–959, 2007.
- [20] H. Ozkan, A. Brandolini, C. Pozzi, S. Effgen, J. Wunder, and F. Salamini, "A reconsideration of the domestication geography of tetraploid wheats," *Theoretical and Applied Genetics*, vol. 110, no. 6, pp. 1052–1060, 2005.
- [21] E. McFadden and E. Sears, "The origin of triticum spelta and its free-threshing hexaploid relatives," *Journal of Heredity*, vol. 37, no. 4, pp. 107–116, 1946.
- [22] G. Petersen, O. Seberg, M. Yde, and K. Berthelsen, "Phylogenetic relationships of triticum and aegilops and evidence for the origin of the a, b, and d genomes of common wheat (triticum aestivum)," *Molecular phylogenetics and evolution*, vol. 39, no. 1, pp. 70–82, 2006.
- [23] F. Salamini, H. Özkan, A. Brandolini, R. Schäfer-Pregl, and W. Martin, "Genetics and geography of wild cereal domestication in the near east," *Nature Reviews Genetics*, vol. 3, no. 6, pp. 429–441, 2002.
- [24] J. F. Hancock, Plant evolution and the origin of crop species. CABI, 2012.
- [25] J. Dvorak, E. D. Akhunov, A. R. Akhunov, K. R. Deal, and M.-C. Luo, "Molecular characterization of a diagnostic dna marker for domesticated tetraploid wheat provides evidence for gene flow from wild tetraploid wheat to hexaploid wheat," *Molecular biology and evolution*, vol. 23, no. 7, pp. 1386–1396, 2006.
- [26] R. J. Giles and T. A. Brown, "Gludy allele variations in aegilops tauschii and triticum aestivum: implications for the origins of hexaploid wheats," *Theoretical and applied genetics*, vol. 112, no. 8, pp. 1563–1572, 2006.
- [27] T. Marcussen, S. R. Sandve, L. Heier, M. Spannagl, M. Pfeifer, K. S. Jakobsen, B. B. Wulff, B. Steuernagel, K. F. Mayer, O.-A. Olsen, et al., "Ancient hybridizations among the ancestral genomes of bread wheat," Science, vol. 345, no. 6194, p. 1250092, 2014.
- [28] N. Yahiaoui, S. Brunner, and B. Keller, "Rapid generation of new powdery mildew resistance genes after wheat domestication," *The Plant Journal*, vol. 47, no. 1, pp. 85–98, 2006.

- [30] Z. Gerechter-Amitai, I. Wahl, A. Vardi, and D. Zohary, "Transfer of stem rust seedling resistance from wild diploid einkorn to tetraploid durum wheat by means of a triploid hybrid bridge," *Euphytica*, vol. 20, no. 2, pp. 281–285, 1971.
- [31] A. Blanco, A. Gadaleta, A. Cenci, A. Carluccio, A. Abdelbacki, and R. Simeone, "Molecular mapping of the novel powdery mildew resistance gene pm36 introgressed from triticum turgidum var. dicoccoides in durum wheat," *Theoretical and Applied Genetics*, vol. 117, no. 1, pp. 135–142, 2008.
- [32] R. Von Bothmer, N. Jacobsen, C. Baden, R. Jørgensen, and I. Linde-Laursen, "An ecographical study of genus hordeum," 1995.
- [33] R. Takahashi, "The origin and evolution of cultivated barley," *Advances in genetics*, vol. 7, pp. 227–266, 1955.
- [34] A. Badr, R. Sch, H. El Rabey, S. Effgen, H. Ibrahim, C. Pozzi, W. Rohde, F. Salamini, et al., "On the origin and domestication history of barley (hordeum vulgare)," *Molecular Biology and Evolution*, vol. 17, no. 4, pp. 499–510, 2000.
- [35] H.-I. Oka, Origin of cultivated rice. Elsevier, 2012.
- [36] P. Azhaguvel and T. Komatsuda, "A phylogenetic analysis based on nucleotide sequence of a marker linked to the brittle rachis locus indicates a diphyletic origin of barley," *Annals of botany*, vol. 100, no. 5, pp. 1009–1015, 2007.
- [37] G. Haberer and K. F. Mayer, "Barley: From brittle to stable harvest," *Cell*, vol. 162, no. 3, pp. 469–471, 2015.
- [38] G. Staudt, "The origin of cultivated barleys: a discussion," *Economic Botany*, vol. 15, no. 3, pp. 205–212, 1961.
- [39] D. Zohary, "Is hordeum a griocrithon the ancestor of six-rowed cultivated barley?," *Evolution*, vol. 13, no. 2, pp. 279–280, 1959.
- [40] P. Murphy, J. Witcombe, P. Shewry, and B. Miflin, "The origin of six-rowed wildbarley from the western himalaya," *Euphytica*, vol. 31, no. 1, pp. 183–192, 1982.
- [41] E. Nevo and G. Chen, "Drought and salt tolerances in wild relatives for wheat and barley improvement," *Plant, cell & environment*, vol. 33, no. 4, pp. 670–685, 2010.
- [42] J. R. Harlan, "The living fields," Our agricultural heritage.-271 S, 1995.
- [43] M. Von Korff, H. Wang, J. Léon, and K. Pillen, "Ab-qtl analysis in spring barley: Ii. detection of favourable exotic alleles for agronomic traits introgressed from wild barley (h. vulgare ssp. spontaneum)," *Theoretical and Applied Genetics*, vol. 112, no. 7, pp. 1221–1231, 2006.
- [44] L. L. Handley, E. Nevo, J. A. Raven, R. MartInez-Carrasco, C. M. Scrimgeour, H. Pakniyat, and B. P. Forster, "Chromosome 4 controls potential water use efficiency (δ13c) in barley," Journal of Experimental Botany, vol. 45, no. 11, pp. 1661–1663, 1994.
- [45] A. Brown, D. Zohary, and E. Nevo, "Outcrossing rates and heterozygosity in natural populations of hordeum spontaneum koch in israel," *Heredity*, vol. 41, no. 1, pp. 49–62, 1978.

- [46] N. C. Ellstrand, Dangerous liaisons?: when cultivated plants mate with their wild relatives. JHU Press, 2003.
- [47] G. Hillman, R. Hedges, A. Moore, S. Colledge, and P. Pettitt, "New evidence of lateglacial cereal cultivation at abu hureyra on the euphrates," *The Holocene*, vol. 11, no. 4, pp. 383–393, 2001.
- [48] A. V. Harter, K. A. Gardner, D. Falush, D. L. Lentz, R. A. Bye, and L. H. Rieseberg, "Origin of extant domesticated sunflowers in eastern north america," *Nature*, vol. 430, no. 6996, pp. 201–205, 2004.
- [49] D. M. Wills and J. M. Burke, "Chloroplast dna variation confirms a single origin of domesticated sunflower (helianthus annuus l.)," *Journal of Heredity*, vol. 97, no. 4, pp. 403–408, 2006.
- [50] D. L. Lentz, M. D. Pohl, J. L. Alvarado, S. Tarighat, and R. Bye, "Sunflower (helianthus annuus l.) as a pre-columbian domesticate in mexico," *Proceedings of the National Academy of Sciences*, vol. 105, no. 17, pp. 6232–6237, 2008.
- [51] N. C. Kane and L. H. Rieseberg, "Genetics and evolution of weedy helianthus annuus populations: adaptation of an agricultural weed," *Molecular Ecology*, vol. 17, no. 1, 2008.
- [52] J. L. Maron, M. Vilà, R. Bommarco, S. Elmendorf, and P. Beardsley, "Rapid evolution of an invasive plant," *Ecological Monographs*, vol. 74, no. 2, pp. 261–280, 2004.
- [53] A. Schwarzbach and L. Rieseberg, "Likely multiple origins of a diploid hybrid sunflower species," *Molecular Ecology*, vol. 11, no. 9, pp. 1703–1715, 2002.
- [54] L. H. Rieseberg, D. E. Soltis, and J. D. Palmer, "A molecular reexamination of introgression between helianthus annuus and h bolanderi (compositae)," *Evolution*, pp. 227–238, 1988.
- [55] M. E. Welch and L. H. Rieseberg, "Patterns of genetic variation suggest a single, ancient origin for the diploid hybrid species helianthus paradoxus," *Evolution*, vol. 56, no. 11, pp. 2126–2137, 2002.
- [56] Y. Yatabe, N. C. Kane, C. Scotti-Saintagne, and L. H. Rieseberg, "Rampant gene exchange across a strong reproductive barrier between the annual sunflowers, helianthus annuus and h. petiolaris," *Genetics*, vol. 175, no. 4, pp. 1883–1893, 2007.
- [57] S.-C. Kim and L. H. Rieseberg, "Genetic architecture of species differences in annual sunflowers: implications for adaptive trait introgression," *Genetics*, vol. 153, no. 2, pp. 965–977, 1999.
- [58] C. B. Heiser Jr, "Hybridization in the annual sunflowers: Helianthus annuus× h. debilis var. cucumerifolius," *Evolution*, pp. 42–51, 1951.
- [59] L. H. Rieseberg, J. Whitton, and K. Gardner, "Hybrid zones and the genetic architecture of a barrier to gene flow between two sunflower species," *Genetics*, vol. 152, no. 2, pp. 713–727, 1999.
- [60] L. H. Rieseberg, S. Beckstrom-Sternberg, and K. Doan, "Helianthus annuus ssp. texanus has chloroplast dna and nuclear ribosomal rna genes of helianthus debilis ssp. cucumerifolius," *Proceedings of the National Academy of Sciences*, vol. 87, no. 2, pp. 593–597, 1990.

- [61] Y. Cohen and W. Sackston, "Factors affecting infection of sunflowers by plasmopara halstedii," *Canadian Journal of Botany*, vol. 51, no. 1, pp. 15–22, 1973.
- [62] J. Miller and T. Gulya, "Inheritance of resistance to race 4 of downy mildew derived from interspecific crosses in sunflower," *Crop science*, vol. 31, no. 1, pp. 40–43, 1991.
- [63] C. Dussle, V. Hahn, S. Knapp, and E. Bauer, "Pl arg from helianthus argophyllus is unlinked to other known downy mildew resistance genes in sunflower," *Theoretical and applied genetics*, vol. 109, no. 5, pp. 1083–1086, 2004.
- [64] M. Rahim, C. Jan, and T. Gulya, "Inheritance of resistance to sunflower downy mildew races 1, 2 and 3 in cultivated sunflower," *Plant Breeding*, vol. 121, no. 1, pp. 57–60, 2002.
- [65] F. Sauca *et al.*, "Introgression of drought-resistance gene (s) from helianthus argophyllus to helianthus annuus specie, using embryo rescue techniques.," *Romanian Agricultural Research*, no. 27, pp. 47–51, 2010.
- [66] G. Besnard, Y. Griveau, M. Quillet, H. Serieys, P. Lambert, D. Vares, and A. Bervillé, "Specifying the introgressed regions from h. argophyllus in cultivated sunflower (helianthus annuus l.) to mark phomopsis resistance genes," *Theoretical and applied genetics*, vol. 94, no. 1, pp. 131–138, 1997.
- [67] D. Q. Fuller, Y.-I. Sato, C. Castillo, L. Qin, A. R. Weisskopf, E. J. Kingwell-Banham, J. Song, S.-M. Ahn, and J. Van Etten, "Consilience of genetics and archaeobotany in the entangled history of rice," Archaeological and Anthropological Sciences, vol. 2, no. 2, pp. 115–131, 2010.

[68]

- [69] J. Molina, M. Sikora, N. Garud, J. M. Flowers, S. Rubinstein, A. Reynolds, P. Huang, S. Jackson, B. A. Schaal, C. D. Bustamante, et al., "Molecular evidence for a single evolutionary origin of domesticated rice," Proceedings of the National Academy of Sciences, vol. 108, no. 20, pp. 8351–8356, 2011.
- [70] T.-T. Chang, "Origin, domestication, and diversification," Rice: origin, history, technology, and production. New Jersey USA: J. Wiley and Sons, Inc, pp. 3–25, 2003.
- [71] J. Glaszmann, "Isozymes and classification of asian rice varieties," *Theoretical and Applied Genetics*, vol. 74, no. 1, pp. 21–30, 1987.
- [72] J. P. Londo, Y.-C. Chiang, K.-H. Hung, T.-Y. Chiang, and B. A. Schaal, "Phylogeography of asian wild rice, oryza rufipogon, reveals multiple independent domestications of cultivated rice, oryza sativa," *Proceedings of the National Academy of Sciences*, vol. 103, no. 25, pp. 9578–9583, 2006.
- [73] G. Khush, D. Brar, P. Virk, S. Tang, S. Malik, G. Busto, Y. Lee, R. McNally, L. Trinh, Y. Jiang, et al., "Classifying rice germplasm by isozyme polymorphism and origin of cultivated rice," IRRI Discuss Pap, vol. 46, p. 279, 2003.
- [74] G. Second, "Origin of the genic diversity of cultivated rice (oryza spp.): study of the polymorphism scored at 40 isozyme loci.,", vol. 57, no. 1, pp. 25–57, 1982.

- [75] K. Zhao, M. Wright, J. Kimball, G. Eizenga, A. McClung, M. Kovach, W. Tyagi, M. L. Ali, C.-W. Tung, A. Reynolds, et al., "Genomic diversity and introgression in o. sativa reveal the impact of domestication and breeding on the rice genome," PloS one, vol. 5, no. 5, p. e10780, 2010.
- [76] D. Brar and G. Khush, "Alien introgression in rice," in *Oryza: From Molecule to Plant*, pp. 35–47, Springer, 1997.
- [77] G. S. KHUSH and K. Ling, "Inheritance of resistance to grassy stunt virus and its vector in rice," *Journal of Heredity*, vol. 65, no. 3, pp. 135–136, 1974.
- [78] Y. Ting, "Wild rice of kwangtung and new variety bred from the hybrids of wild rice with cultivated rice," Coll. Agric. Sun Yatsen Univ., Agron. Bull, vol. 3, pp. 1–22, 1933.
- [79] N. Vavilov, "Geographical centers of our cultivated plants," in *Proceedings of V international congress on genetics*, New York, USA, pp. 342–369, 1928.
- [80] G. S. Khush and G. L. Stebbins, "Cytogenetic and evolutionary studies in secale. i. some new data on the ancestry of s. cereale," *American Journal of Botany*, pp. 723–730, 1961.
- [81] Z. Huamán and D. M. Spooner, "Reclassification of landrace populations of cultivated potatoes (solanum sect. petota)," *American Journal of Botany*, vol. 89, no. 6, pp. 947–965, 2002.
- [82] D. M. Spooner, K. McLean, G. Ramsay, R. Waugh, and G. J. Bryan, "A single domestication for potato based on multilocus amplified fragment length polymorphism genotyping," Proceedings of the National Academy of Sciences of the United States of America, vol. 102, no. 41, pp. 14694–14699, 2005.
- [83] B. Pickersgill and C. B. Heiser Jr, "Origins and distribution of plants domesticated in the new world tropics," *Origins of agriculture*, 1977.
- [84] J. G. Hawkes, "The evolution of cultivated potatoes and their tuber-bearing wild relatives," *Die Kulturpflanze*, vol. 36, no. 1, pp. 189–208, 1988.
- [85] D. S. Correll *et al.*, "The potato and its wild relatives.," *The potato and its wild relatives.*, 1962.
- [86] C. Quiros, R. Ortega, L. Van Raamsdonk, M. Herrera-Montoya, P. Cisneros, E. Schmidt, and S. Brush, "Increase of potato genetic resources in their center of diversity: the role of natural outcrossing and selection by the andean farmer," *Genetic Resources and Crop Evolution*, vol. 39, no. 2, pp. 107–113, 1992.
- [87] D. Rabinowitz, C. Linder, R. Ortega, D. Begazo, H. Murguia, D. Douches, and C. Quiros, "High levels of interspecific hybridization between solanum sparsipilum ands. stenotomum in experimental plots in the andes," *American Potato Journal*, vol. 67, no. 2, pp. 73–81, 1990.
- [88] T. Johns, Z. Huaman, C. Ochoa, and P. E. Schmiediche, "Relationships among wild, weed, and cultivated potatoes in the solanum x ajanhuiri complex," *Systematic Botany*, pp. 541–552, 1987.
- [89] C. R. Linder, Diversity within traditional potato agriculture in the Peruvian Andes and its relevance to crop evolution. Cornell University, Aug., 1987.

- [90] S. Brush, R. Kesseli, R. Ortega, P. Cisneros, K. Zimmerer, and C. Quiros, "Potato diversity in the andean center of crop domestication," *Conservation Biology*, vol. 9, no. 5, pp. 1189–1198, 1995.
- [91] S. B. Brush, H. J. Carney, and Z. Humán, "Dynamics of andean potato agriculture," *Economic Botany*, vol. 35, no. 1, pp. 70–88, 1981.
- [92] P. Grun, "The evolution of cultivated potatoes," *Economic Botany*, vol. 44, no. 3, pp. 39–55, 1990.
- [93] K. S. Zimmerer, "The ecogeography of andean potatoes," BioScience, pp. 445–454, 1998.
- [94] Z. Huamán, J. Hawkes, and P. Rowe, "Solanum ajanhuiri: an important diploid potato cultivated in the andean altiplano," *Economic Botany*, vol. 34, no. 4, pp. 335–343, 1980.
- [95] T. Johns and S. L. Keen, "Ongoing evolution of the potato on the altiplano of western bolivia," *Economic Botany*, vol. 40, no. 4, pp. 409–424, 1986.
- [96] J. G. Hawkes, "Origin of solanum juzepczukii buk and s curtilobum juz et buk," ZEITSCHRIFT FUR PFLANZENZUCHTUNG-JOURNAL OF PLANT BREEDING, vol. 47, no. 1, p. 1, 1962.
- [97] P. Schmiediche, J. Hawkes, and C. Ochoa, "Breeding of the cultivated potato species solanum x juzepczukii buk. and solanum x curtilobum juz. etbuk.," *Euphytica*, vol. 29, no. 3, pp. 685–704, 1980.
- [98] J. R. van der Voort, K. Kanyuka, E. van der Vossen, A. Bendahmane, P. Mooijman, R. Klein-Lankhorst, W. Stiekema, D. Baulcombe, and J. Bakker, "Tight physical linkage of the nematode resistance gene gpa2 and the virus resistance gene rx on a single segment introgressed from the wild species solanum tuberosum subsp. andigena cpc 1673 into cultivated potato," Molecular plant-microbe interactions, vol. 12, no. 3, pp. 197–206, 1999.
- [99] E. Van Der Vossen, A. Sikkema, B. t. L. Hekkert, J. Gros, P. Stevens, M. Muskens, D. Wouters, A. Pereira, W. Stiekema, and S. Allefs, "An ancient r gene from the wild potato species solanum bulbocastanum confers broad-spectrum resistance to phytophthora infestans in cultivated potato and tomato," The Plant Journal, vol. 36, no. 6, pp. 867–882, 2003.
- [100] S. Johnston, T. Den Nijs, S. Peloquin, and R. Hanneman Jr, "The significance of genic balance to endosperm development in interspecific crosses," *Theoretical and applied genetics*, vol. 57, no. 1, pp. 5–9, 1980.
- [101] W. Sims, "History of tomato production for industry around the world," in *Symposium on Production of Tomatoes for Processing 100*, pp. 25–26, 1979.
- [102] L. D. Robertson, J. A. Labate, M. Razdan, and A. Mattoo, "Genetic resources of tomato (lycopersicon esculentum mill.) and wild relatives," *Genetic Improvement of Solanaceous Crops. Tomato*, vol. 2, pp. 25–75, 2007.
- [103] Y. Bai and P. Lindhout, "Domestication and breeding of tomatoes: what have we gained and what can we gain in the future?," *Annals of Botany*, vol. 100, no. 5, pp. 1085–1094, 2007.
- [104] T. C. Nesbitt and S. D. Tanksley, "Comparative sequencing in the genus lycopersicon: implications for the evolution of fruit size in the domestication of cultivated tomatoes," *Genetics*, vol. 162, no. 1, pp. 365–379, 2002.

- [105] C. M. Rick and P. G. Smith, "Novel variation in tomato species hybrids," *American Naturalist*, pp. 359–373, 1953.
- [106] C. M. Rick, "Tomato-like nightshades: affinities, autoecology, and breeders opportunities," *Economic Botany*, vol. 42, no. 2, pp. 145–154, 1988.
- [107] J. Miller and S. Tanksley, "Rflp analysis of phylogenetic relationships and genetic variation in the genus lycopersicon," *Theoretical and applied genetics*, vol. 80, no. 4, pp. 437–448, 1990.
- [108] C. Rick, "The potential of exotic germplasm for tomato improvement," 1982.
- [109] P. M. Hanson, D. Bernacchi, S. Green, S. D. Tanksley, V. Muniyappa, A. S. Padmaja, H.-m. Chen, G. Kuo, D. Fang, and J.-t. Chen, "Mapping a wild tomato introgression associated with tomato yellow leaf curl virus resistance in a cultivated tomato line," *Journal of the American Society for Horticultural Science*, vol. 125, no. 1, pp. 15–20, 2000.
- [110] M. Banerjee *et al.*, "Transfer of tomato leaf curl virus resistance from lycopersicon hirsutum f. glabratum to l. esculentum," *Plant Breeding*, vol. 105, no. 2, pp. 156–159, 1990.
- [111] M. Kwak, J. A. Kami, and P. Gepts, "The putative mesoamerican domestication center of is located in the lerma–santiago basin of mexico," *Crop Science*, vol. 49, no. 2, pp. 554–563, 2009.
- [112] D. Debouck and S. THOME, "Implications for bean breeders of studies on the origin of common beans, phaseolus vulgaris l," in *THE BEAN breeders. Cali, Workshop at CIAT*, 1988.
- [113] R. Papa and P. Gepts, "Asymmetry of gene flow and differential geographical structure of molecular diversity in wild and domesticated common bean (phaseolus vulgaris l.) from mesoamerica," *Theoretical and Applied Genetics*, vol. 106, no. 2, pp. 239–250, 2003.
- [114] D. G. Debouck, O. Toro, O. M. Paredes, W. C. Johnson, and P. Gepts, "Genetic diversity and ecological distribution of phase olus vulgaris (fabaceae) in northwestern south america," *Economic botany*, vol. 47, no. 4, pp. 408–423, 1993.
- [115] M. Kwak and P. Gepts, "Structure of genetic diversity in the two major gene pools of common bean (phaseolus vulgaris l., fabaceae)," *Theoretical and Applied Genetics*, vol. 118, no. 5, pp. 979–992, 2009.
- [116] J. A. GUTIERREZ and S. P. SINGH, "Heterosis and inbreeding depression in dry bush beans, phaseolus vulgaris 1.," *Canadian journal of plant science*, vol. 65, no. 2, pp. 243–249, 1985.
- [117] O. M. Paredes and P. Gepts, "Extensive introgression of middle american germplasm into chilean common bean cultivars," *Genetic Resources and Crop Evolution*, vol. 42, no. 1, pp. 29–41, 1995.
- [118] S. Park and B. Dhanvantari, "Transfer of common blight (xanthomonas campestris pv phase-oli) resistance from phaseolus coccineus lam. to p. vulgaris l. through interspecific hybridization," Canadian journal of plant science, vol. 67, no. 3, pp. 685–695, 1987.
- [119] H. F. Schwartz, K. Otto, H. Terán, M. Lema, and S. P. Singh, "Inheritance of white mold resistance in phaseolus vulgaris× p. coccineus crosses," *Plant Disease*, vol. 90, no. 9, pp. 1167–1170, 2006.

- [120] J. L. Kornegay and C. Cardona, "Inheritance of resistance to acanthoscelides obtectus in a wild common bean accession crossed to commercial bean cultivars," *Euphytica*, vol. 52, no. 2, pp. 103–111, 1991.
- [121] C. W. Smith and R. A. Frederiksen, Sorghum: Origin, history, technology, and production, vol. 2. John Wiley & Sons, 2000.
- [122] Y. Shechter and J. De Wet, "Comparative electrophoresis and isozyme analysis of seed proteins from cultivated races of sorghum," *American Journal of Botany*, pp. 254–261, 1975.
- [123] H. Doggett et al., Sorghum. No. 2. ed., Longman Scientific and Technical, 1988.
- [124] J. R. Harlan and J. M. de Wet, "Toward a rational classification of cultivated plants," *Taxon*, pp. 509–517, 1971.
- [125]
- [126] E. D. Garber et al., "Cytotaxonomic studies in the genus sorghum.," Univiversity of California Publications in Botany, vol. 23, pp. 283–362, 1950.
- [127] P. E. Arriola and N. C. Ellstrand, "Crop-to-weed gene flow in the genus sorghum (poaceae): spontaneous interspecific hybridization between johnsongrass, sorghum halepense, and crop sorghum, s. bicolor," *American Journal of Botany*, pp. 1153–1159, 1996.
- [128] I. P. PO, "Sorghum in the eighties," 1982.
- [129] J. De Wet, "Systematics and evolution of sorghum sect. sorghum (gramineae)," American Journal of Botany, pp. 477–484, 1978.
- [130] H. Doggett and B. Majisu, "Disruptive selection in crop development," *Heredity*, vol. 23, no. 1, pp. 1–23, 1968.
- [131] H. G. Baker, "Human influences on plant evolution," *Economic Botany*, vol. 26, no. 1, pp. 32–43, 1972.
- [132] L. Kuhlman, B. Burson, P. Klein, D. Stelly, and W. Rooney, "Interspecific sorghum breeding using s. macrospermum," in *Proceedings of the ASA-CSSA-SSA 2006 International Meetings*, *Indianapolis*, pp. 12–16, 2006.
- [133] P. Aldrich and J. Doebley, "Restriction fragment variation in the nuclear and chloroplast genomes of cultivated and wild sorghum bicolor," *Theoretical and Applied Genetics*, vol. 85, no. 2-3, pp. 293–302, 1992.
- [134] P. Aldrich, J. Doebley, K. Schertz, and A. Stec, "Patterns of allozyme variation in cultivated and wild sorghum bicolor," *Theoretical and Applied Genetics*, vol. 85, no. 4, pp. 451–460, 1992.
- [135] H. G. Baker, "Migrations of weeds," Valentine, D, H ed (s). Taxonomy phytogeography and evolution, pp. 327–347, 1972.
- [136] J. De Wet, S. Gupta, J. Harlan, and C. Grassl, "Cytogenetics of introgression from saccharum into sorghum," *Crop Science*, vol. 16, no. 4, pp. 568–572, 1976.

- [137] V. G. Reddy, H. Upadhyaya, and C. Gowda, "Current status of sorghum genetic resources at icrisat: their sharing and impacts," *International Sorghum and Millets Newsletter*, vol. 47, pp. 9–13, 2006.
- [138] J. W. Johnson and G. L. Teetes, "Breeding for arthropod resistance in sorghum," *Harris, MK Biology and breeding for resistance to arthropods and pathogens in agricultural plants. Texas, USA*, pp. 168–180, 1979.