

# Review: Adaptive Introgression Expanded the Genetic Base of Crops during post-Domestication Spread

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The process of domestication is often conceptualized as geographically constrained, with crops originating from a wild progenitor within one or more defined centers followed by expansion to the modern-day extent of cultivation [1]. However, archaeological and genetic evidence are beginning to reveal that, in many cases, domestication has been temporally protracted and geographically diffuse [2, 3, 4, 5, 6]. An additional important aspect of the emerging complexity of domestication is beneficial gene flow (*i.e.*, adaptive introgression) from locally adapted wild relatives during crop expansion following initial domestication.

Adaptive introgression has three components: hybridization between differentiated taxa, backcrossing to one of the parents, and selection on recombinant genotypes with progressively diminished linkage drag [7, 8]. In domesticated species, adaptive introgression would consist of crop-wild hybrids backcrossing to a crop followed by increase in frequency of adaptive wild haplotypes in the crop and selection against undesirable wild background. To date, the literature on crop-wild gene flow has largely focused on the risk of transgene introgression from domesticated crops into wild relatives (for a review, [9]) and on modern plant breeding efforts to introgress desired traits from wild relatives (for a review, [10]). The history of natural and potentially adaptive introgression of wild alleles into domesticated crops over evolutionary timescales has received considerably less attention. However, new tools and methods have recently been employed to detect genome-wide patterns of introgression, granting new insights into the prevalence of adaptive introgression in crop histories. Preliminary results suggest there is a need to expand our conception of domestication to include a broadening of the genetic base of crops that occurred through adaptive gene flow with newly encountered wild relatives during post-domestication expansion.

In this review, we will: 1) briefly describe recently developed methods for detecting introgression, 2) present case studies suggesting wild-to-crop introgression has conferred local adaptation, 3) consider how introgression bears upon fundamental questions of domestication, and 4) describe future advances in both basic and applied genetics that can be made through the study of introgression in agroecosystems.

## Introgression methods and their application

The decreasing cost of genome-wide resequencing and availability of reduced-representation genotyping (*e.g.*, GBS and RAD-Seq), combined with new analytical methods, has facilitated comprehensive study of introgression across a number of species (**Table 1**). High-density marker data can be used with recently developed methods to assign specific genomic regions to a taxon of origin

and identify introgression across taxa [11, 12, 13, 14, 15, 16]. The methods reviewed here do not include those marginally estimating introgression/migration rate as a component of demographic history (*e.g.*, Approximate Bayesian Computation (ABC) [17], diffusion approximations for demographic inference (*δaδi*) [18], isolation with migration models [19], and a series of methods utilizing the sequentially Markovian coalescent (PSMC, MSMC and SMC++) [20, 21, 22]). Rather, we focus on methods that explicitly identify introgressed genomic segments based on the extent of differentiation, patterns of nucleotide/haplotype sharing, and phylogenetic relationships.

First, introgressed segments are expected to show low differentiation from their source population. The  $F_{st}$  and  $d_{XY}$  statistics and their derivatives including  $G_{min}$  [16] and  $RND_{min}$  [15] gauge differentiation. The former two statistics are insensitive to rare migrants and therefore lack power to detect recent introgression, while the latter two overcome this limitation. Additionally,  $RND_{min}$  accounts for variable mutation rate, which is detected based on branch length to an outgroup [15]. These statistics have been further developed by adding differentiation between both non-admixed ( $A$ ) and admixed populations ( $B$ ) and a source population ( $C$ ) [23]. For example, the  $U_{A,B,C}(w,x,y)$  statistic summarizes number of sites where an allele at frequency  $y$  in the source population ( $C$ ) has a frequency higher than  $x$  in the admixed population ( $B$ ) and lower than  $w$  in the non-admixed population ( $A$ ). A similar statistic,  $Q95_{A,B,C}(w,y)$ , sets a hard cutoff at the 95<sup>th</sup> percentile of allele frequency in the admixed population ( $B$ ) [23]. Further modifications have allowed specification of more than one source population (see details in [23]).

Second, local ancestry deconvolution (also known as chromosome painting) assigns genomic regions to various source populations based on patterns of allele/haplotype sharing [24]. One form of chromosome painting utilizes a hidden Markov models to evaluate ancestry across admixed genomes through comparison to reference, non-admixed individuals (*e.g.*, HAPMIX [12]). Another clusters admixed populations with reference samples using a sliding-window approach (*e.g.*, PCAdmix [25] and LAMP [26]). And finally, introgression can be detected through chromosome painting by using a Bayesian model [27] in which deviations from Hardy-Weinberg equilibrium are minimized through creation of genetic groups (*e.g.*, fineSTRUCTURE [13]).

Third, the ABBA-BABA statistic (also known as the D-statistic) and related statistics are widely applied to introgression detection. These statistics make inferences regarding introgression based on genomic patterns of derived variants that are shared between populations or species. Patterns of allele sharing are interpreted in a phylogenetic context and the method is best suited to detection of introgression at the genome level. Elaborations of the D-statistic capable of localizing introgression to specific genomic regions include  $\hat{f}_d$  [11] and the five-taxon D-statistic [14]. The former is quite similar to the D-statistic but uses allele frequencies from each population/species, and the latter detects introgression based on the localized phylogenetic pattern and is capable of determining introgression directionality.

Application of these approaches, in combination with population genetic statistics to detect selection, suggest introgression can play an adaptive role. For example, based on sequence divergence methods, introgression has been detected across *Mimulus* (*i.e.*, monkeyflower) species and appears to play a role in both adaptation to pollinator preference and speciation [28]. Likewise, the HAPMIX chromosome painting method was applied by Jeong et al. [29] to detect introgression from Nepalese Sherpa to Tibetan humans at loci controlling high altitude adaptation (reviewed in [30]). Finally, the phylogenetic ABBA-BABA statistic has revealed introgression of wing coloration loci conferring Müllerian mimicry across butterfly species [31].

# Crop adaptation through introgression

Genome-wide data from extensive samples of crops and their wild relatives, in combination with the new methods described above, have recently allowed detailed analysis of wild-to-crop introgression in some of the world’s most important crops (**Table 2**). Below we present a summary of findings from maize, barley, rice, and potato, four promising systems in which introgression from wild relatives appears to have played an adaptive role.

## 1. Maize:

The relationship between maize (*Zea mays* ssp. *mays*) and the teosinte *Zea mays* ssp. *mexicana* (hereafter, *mexicana*) offers a prime case study of adaptive wild-to-crop introgression. Maize was domesticated from *Zea mays* ssp. *parviglumis* (hereafter, *parviglumis*) approximately 9,000 BP in the lowlands of the Balsas River Valley in Mexico [32]. From this domestication center, maize spread into the highlands of the Mexican Central Plateau, where it came into sympatry with *mexicana*. Introgression from *mexicana* to maize in the Central Plateau has been reported based on both morphological [33] and molecular [34, 35] data. However, Hufford et al. [36] first localized *mexicana* introgression to chromosomal regions and provided evidence that it was likely adaptive. The authors identified nine genomic regions in several maize populations which consistently showed evidence of *mexicana* introgression based on chromosome painting methods including HAPMIX (Figure 1). These introgressed segments overlapped with QTL that had previously been found to control anthocyanin content and leaf macrohairs [37], traits known to be adaptive at high elevation. In a growth chamber experiment, the authors demonstrated that maize populations with *mexicana* introgression showed greater plant height (a proxy for fitness) under highland environmental conditions than populations that lacked introgression. Height differences were not detected under lowland conditions, providing further evidence of local adaptation.

Populations of *mexicana* cannot be found outside the highlands of Mexico, yet maize has colonized and adapted to high elevation in a number of other regions. A recent study [4] employed the ABBA-BABA and  $\hat{f}_d$  statistics to evaluate if maize with *mexicana* introgression was transferred to other highland regions or whether highland adaptation was obtained *de novo* outside of Mexico. Overall, analyses revealed that *mexicana* introgression was pervasive in maize from Mesoamerican high elevation regions (the highlands of Mexico, Guatemala, and the southwestern United States), but more distant high elevation regions (*e.g.*, the Andes) showed no *mexicana* ancestry. Maize from the lowlands of northern South America appears to have been derived from Mesoamerican lowland maize and high elevation adaptation in Andean maize likely occurred *de novo* from this genetic base.

## 2. Barley:

Barley (*Hordeum vulgare* subsp. *vulgare*) was domesticated at least twice roughly 8,000 to 10,000 BP: once from the wild subsp. *spontaneum* in the Fertile Crescent and once from subsp. *spontaneum* var. *agriocrithon* in Tibet [38, 39, 40, 41, 42, 43]. Presently, the distribution of subsp. *spontaneum* stretches from the eastern Mediterranean through the Middle-East to west-central Asia, spanning clines in temperature, precipitation, soil type, and altitude [44]. Cultivated barley is found throughout much of the *spontaneum* distribution and barley-*spontaneum* hybrids are fertile and commonly found when these taxa co-occur. In fact, signatures of contemporary gene flow between wild and domesticated barley have been detected across distances greater than a kilometer [45].

Poets and co-authors [46] recently investigated the range-wide contribution of wild barley to landraces, assessing both genome-wide and geographical patterns of introgression. This study identified several lines of evidence consistent with wild introgression aiding the expansion and adaptation of domesticated barley. Using chromosome painting methods, the authors identified genomic regions of shared ancestry linking particular landraces to numerous wild relative populations. These results suggested landraces may have received wild introgression on a continual basis during post-domestication expansion. However, barley landraces also showed an excess of ancestry from nearby wild relatives, indicating a prevalence of local and potentially adaptive gene flow. Limited admixture linkage disequilibrium and small tracts of identity by state suggested even locally introgressed chromosomal regions are ancient, perhaps dating to the early expansion of barley post-domestication. While these results are consistent with adaptive introgression, wild barley haplotypes have yet to be definitively linked to specific adaptations in landraces.

### 3. Asian Rice:

The details of Asian rice (*Oryza sativa*) domestication are still debated. Certain genetic and archaeobotanical evidence point toward independent domestications of the two prominent varietal groups *japonica* and *indica* from the wild species *Oryza rufipogon* (*rufipogon* hereafter) in China and the Indian Ganges plain, respectively [47], with a potential third domestication event giving rise to *aus* in Bangladesh or central India [48]. Other studies support a single domestication in China, with later divergence of *japonica* and *indica* [49, 50] during crop expansion. For example, Huang and colleagues [50] developed a genetic map of rice variation, which they used to measure genetic distance between a range-wide sample of wild and domesticated rice, finding *japonica* was domesticated near the Pearl River in Guangxi province, China, and that *indica* was likely the result of hybridization between *japonica* and local *rufipogon* populations in Southern and South-eastern Asia. In a re-examination of these same data, [48] found evidence supporting independent domestications of *japonica*, *indica*, and *aus*, as well as a hybrid origin (*japonica* x *aus*) of *aromatic* rice. A recent third analysis by Choi and co-authors [51] compared these two disparate results and the methods utilized therein, concluding that domestication alleles (including *LABA1*, *PROG1*, and *sh1*) arose during a single domestication event of *japonica*, and that these alleles introgressed into several wild *rufipogon* subpopulations which thereby became the progenitors of other Asian rice varieties.

The findings of [51] bear similarities to a hypothesis posited by Vaughan and colleagues [52]. According to this hypothesis, domestication alleles arose in a single cultivated rice population and subsequently introgressed into diverse cultivated populations (some *japonica*-like, some *indica*-like). As these now fully domesticated populations spread further into new environments, they potentially received introgression from locally adapted wild relatives, retaining alleles that improved fitness. While the precise history of domesticated rice remains in question, multiple lines of evidence indicate diverse wild populations have contributed to domesticated germplasm and suggest adaptive introgression may have played a role during the expansion of this important crop.

### 4. Potato

Modern potato (*Solanum tuberosum*) was likely domesticated in southern Peru in sympatry with several wild relatives approximately 6,000-10,000 BP. The exact progenitor has remained in question for some time [53, 54, 55, 56], but a distance-based phylogeny constructed using genotypic data from a *Solanum* diversity panel recently identified *S. candolleianum* as the most probable progenitor [57]. The lack of clarity regarding a progenitor has been due, in

part, to extensive post-domestication hybridization between potato and a number of related species.

While potatoes are primarily propagated clonally, farmers do at times promote sexual reproduction for improvement of the crop and development of new cultivars [58]. Close proximity of domesticated potatoes and wild relatives, active hybridization, and local selection pressure favoring wild haplotypes across a diverse range of biotic and environmental conditions have likely fostered an expansion of genetic diversity within potatoes subsequent to domestication [59]. The prevalence of wild introgression was recently clarified in a broad survey of potato diversity by Hardigan and colleagues [60]. These authors discovered that both diploid and tetraploid domesticates had received extensive introgression from more than ten species of wild *Solanum*, with a continued broadening of the genetic base of potato as it spread away from its Peruvian origin. In certain cultivars, wild ancestry was estimated at upwards of 30%. Genes located within these introgressed regions were more likely to be highly-expressed and stress-inducible, and contained loci related to disease resistance, drought tolerance, and heat tolerance, suggesting introgression conferred adaptations critical to survival, possibly facilitating tolerance for new environmental pressures during range expansion [60].

## Re-evaluating domestication

A framework in which crops are domesticated from a single wild population or even a single species is an oversimplification when introgression is extensive during crop geographic expansion. The addition of ongoing gene flow to our understanding of crop demography could therefore bear importantly on fundamental questions of crop domestication:

### What is the progenitor of a crop?

Depending on the extent of post-domestication gene flow with new wild relatives, identification of a crop's progenitor can be complicated or confounded entirely. Introgression between a crop and newly-encountered taxa decreases divergence of the crop from these donors. This signal could be mistaken for origin rather than gene flow. For example, when determining a single origin of maize from *parviglumis*, Matsuoka and colleagues [32] identified a paradox: while *parviglumis* is found exclusively in the lowlands of southwest Mexico, maize with allele frequencies most similar to *parviglumis* was found in the highlands of the Mexican Central Plateau. Several years later, van Heerwaarden *et al.* [34] resolved the paradox by determining that widespread introgression in the highlands from *mexicana*, which is closely related to *parviglumis*, has caused maize from this region to appear ancestral. Similarly, extensive post-domestication adaptive introgression from potato wild relatives long obscured this crop's origin. Recent work has shown that, following the original domestication event of *Solanum tuberosum* in the central Andes, potato received introgression from as many as four additional species during colonization of the highest elevations of the Andes and the lowlands of the Chilean coast [88, 89, 90].

Beyond confounding detection of progenitor taxa, extensive introgression may necessitate a reevaluation of crop origins. In cases like maize and potato it is important to recognize the substantial contributions of introgressing taxa to the genetic base of modern crops. Broad recognition of the role these wild relatives have played in crop adaptation could further their use in breeding and elevate their conservation status.

## When was a crop domesticated?

Estimates of the timing of initial domestication are often based on levels of sequence divergence between a crop and populations of its presumed progenitor (*e.g.*, [32, 49]). In highly introgressed domesticates, these estimates will be based on comparison of both crop and introgressant haplotypes to those of the presumed progenitor. In such cases, divergence time is a mixture of time since domestication and time since split of the progenitor and the introgressing taxon. This phenomenon, in combination with divergence of crop samples from true ancestral crop populations, ongoing evolution of crop progenitors, and problems with assuming evolution under a molecular clock [91], may help explain discrepancies between domestication dates based on genetic and archaeological data. More accurate estimates of the timing of domestication may be obtained from genetic data by excluding loci that show signatures of introgression or by explicitly including estimates of introgression when modeling a crop’s demographic history.

## How was genome-wide diversity shaped by domestication?

Measurement of the strength of the initial domestication bottleneck may also be impacted by adaptive introgression during the spread of crops. Crop wild relatives have distinct demographies when compared to domesticates and may therefore have contrasting effective population sizes ( $N_e$ ). The influence of wild relative introgression on estimates of the domestication bottleneck will depend on a number of factors including the magnitude of gene flow, the  $N_e$  of the introgressing taxon, and the strength of selection on haplotypes following introgression. For example, substantial introgression at neutral loci from a wild taxon with a historically higher  $N_e$  will lead to underestimates of the overall strength of the domestication bottleneck.

## What candidate genes were targeted by selection during domestication?

Loci targeted by selection during domestication can be identified through so-called “bottom-up” approaches based on population genetic signatures [92]. Ideally, candidate loci will be identified by first constructing a demographic model representing the history of the domesticate. In this approach, diversity data from neutral loci are fit to potential models of a crop’s demography and then statistical tests of selection are used to identify candidate domestication genes under the most likely model. Due to the difficulty of this approach and the uncertainty associated with any given demography, many studies identify domestication loci using a strict outlier approach in which loci showing, for example, the greatest reduction in nucleotide diversity or the highest allele frequency differentiation in the domesticate relative to the wild progenitor are identified as candidates. Introgression during crop expansion may influence candidate gene detection using both demographic-modeling and strict-outlier approaches. For example, *mexicana* introgression into maize described above accounts for approximately 20% of the genome of maize in the highlands of Mexico [34]. Takuno and co-authors [93] have shown that a demographic model incorporating this introgression is a significantly better fit to empirical data than a model lacking introgression. Failure to account for introgression in maize would therefore compromise domestication candidate detection, particularly if a study contained maize samples from the Mexican highlands. Likewise, introgression that increased nucleotide diversity in the domesticate or decreased differentiation at domestication loci would confound a strict outlier approach. However, previous work, also in maize, has shown that known domestication loci are particularly resistant to introgression [36], likely due to ongoing selection favoring the domesticated phenotype.

## Future studies in crop-wild introgression

### Basic:

Research has so far shown that adaptive crop-wild introgression has played a significant role in the domestication and dispersal of many agronomically-important crops. However, the dynamics of this process are not yet fully understood, especially in the context of individual case examples, and many questions remain.

What is the genomic architecture of this introgression, and does the architecture suggest that it has been adaptive? At what geographic scale is introgression adaptive? To what extent does this depend on the slope of environmental gradients such as temperature, precipitation, and elevation across these ranges? We can look at conservation of genomic architecture across landscapes and between populations, and make predictions about introgressions and their relationship with local adaptation. If the genomic architecture of an introgressed region is conserved across a broad ecogeographical region, this suggests that the introgression imparts adaptation to general environmental or climatic variables. On the other hand, if the genomic architecture is conserved within populations but not between nearby populations in the region, this suggests that the introgressed regions offer adaptations to more local selective pressures. If the genomic architecture of an introgressed region is not conserved within a population, there is little evidence that the introgression is adaptive. Also, identification of environmental and/or disease tolerance loci within introgressed regions suggest adaptive significance.

After hybridization events that lead to introgression, how long might the detectable genomic signals of introgression persist? Introgressed regions are easier to detect when there has been limited recombination to break them apart. Therefore, introgressions are easiest to detect when they are either recent (few generations means few recombination events) or involve structural variation (which diminishes recombination rate). Because recombination progressively breaks apart LD in introgressed regions, measurements of LD can be used to date introgression events (as in [46]). Mutation rate, effective population size, and strength of selection also may influence the rate of breakup of introgressed regions.

At what taxonomic scale does introgression occur? When do species become so diverged that introgression is either maladaptive or impossible (due to Dobzhansky-Muller incompatibilities or other mechanisms)? Theory indicates that the most significant limiting factor to gene flow between progenitor and domesticate is divergence time. Over time, diverged populations drift and become increasingly incompatible. Small effective population size, and correspondingly high genetic load, of the introgressive population also limits gene flow. Although perhaps less applicable in crop systems, this effect is seen in other well-documented cases of introgression (for example, Neanderthal introgression into humans, [94]). *iiiiiii HEAD I tracked down the sources that indicate that introgression is suppressed around genes. The case example was Neanderthal alleles into humans, the paper titled "The genetic history of Ice Age Europe" [? ], cited by the Graham Coop paper. However, I'm unclear if this is a general rule, or simply a consequence of Neanderthal being high in deleterious genetic load. Wouldn't crop systems, with wild relatives harboring putatively adaptive alleles with low load, show a different pattern? I could also talk about cross-incompatibility factors in maize/teosinte hybridization vs. highly compatible asian rice and its relatives. I probably wouldn't need a new source for the rice, but I might need to cite a paper about tcb1, ga1, and ga2.*

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### Applied:

Our identification and understanding of introgression in agroecosystems would be augmented by the development of wild relative genomic resources, such as annotated genomic sequence assemblies

and functional genomic data sets [50]. Additional study of introgression in agroecosystems could lead to advances in both basic and applied genetics, and specifically the continued improvement of modern crops. Loci underlying the domesticated phenotype can be more clearly identified by removing the confounding population genetic signals of introgression. These loci are potentially beneficial targets for crop improvement. Furthermore, adaptive introgression that is demonstrably tied to a specific environment may include beneficial alleles that can be utilized in crop breeding. Identification of wild relative species or populations that harbor putatively adaptive genotypes may facilitate greater prioritization of conservation efforts.

## Conclusions

In the same way that Darwin explored natural selection by studying domesticated species (going so far as to feature domestic pigeons as his "special group" at the beginning of his *On the Origin of Species* [95]), we frequently find that modern crop systems, in tandem with their extant wild relatives, offer unparalleled opportunities for the study of several evolutionary questions, including the role of adaptive introgression in dispersal and range expansion. The advent and application of genomics tools has lead to mounting evidence which underscores the prevalence of genetic admixture between crops and wild relatives during and well beyond the initial stages of domestication. Likewise, population genetic theory reinforces the proclivity of wild relatives to provide advantageous, locally-adapted alleles to crops as they disperse beyond their domestication centers into new geographies with new ecological pressures and niches. Within this new conception, the process of crop range expansion is a concurrent element of domestication, rather than a distinct event subsequent to it. This is a revolutionary departure from the prevailing model, and as such, offers new questions and new opportunities for investigation. These questions take on practical importance as well as theoretical. Understanding a crop's history of domestication in terms of selection, adaptation, and admixture empowers efforts at crop development and improvement. As the historic role of wild relatives in the development of many crops is increasingly clarified, it is likely that we may reevaluate the importance of conservation of wild populations (in particular or in general) that harbor agronomically-valuable alleles that may be targeted by pre-breeding efforts. These efforts may be of particular value in the face of future climate volatility and change.

Domesticated crops are among humanity's first and greatest inventions. Interwoven in the genetic history of these crops is the story of our ancestors as they transitioned through the Agronomic Revolution. In this more nuanced and dynamic understanding of crop domestication, we gain insight into the evolutionary journey that crops and humanity took, and continue to take, together.

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Domesticated Crop	Compatible Wild Relatives	Hybrids and/or Hybridization	Evidence of Crop Introgression	Evidence of Adaptativeness	Sources
Apple ( <i>Malus domestica</i> )	<i>M. sylvestris</i> , <i>M. orientalis</i> , <i>M. baccata</i> , <i>M. sieversii</i>	X	X		[87]
Asian Rice ( <i>Oryza sativa</i> )	<i>O. rufipogon</i>	X	X	X	[50]
Barley ( <i>Hordeum vulgare</i> )	<i>H. v.</i> subsp. <i>spontaneum</i>	X	X	X	[46]
Cassava ( <i>Manihot esculenta</i> )	<i>M. glaziovii</i>	X	X	X	[78]
Common Bean ( <i>Phaseolus vulgaris</i> )	<i>P. v.</i> var. <i>aborigineus</i> , <i>P. v.</i> var. <i>mexicanus</i> [[not in this source]]	X	X		[83]
Grapes ( <i>Vitis vinifera</i> subsp. <i>vinifera</i> )	<i>V. v.</i> subsp. <i>sylvestris</i>	X	X		[84]
Maize ( <i>Zea mays</i> subsp. <i>mays</i> )	<i>Z. m.</i> subsp. <i>mexicana</i> , <i>Z. m.</i> subsp. <i>parviglumis</i>	X	X	X	[36]
Olive ( <i>Olea europaea</i> ssp. <i>europaea</i> var. <i>sylvestris</i> subsp. <i>sativa</i> )	<i>O. e.</i> ssp. <i>europaea</i> var. <i>sylvestris</i>	X	X		[81]
Potato ( <i>Solanum tuberosum</i> )	many	X	X	X	[60]
Soybeans ( <i>Glycine max</i> )	<i>G. soja</i>	X	X		[82]
Sorghum ( <i>Sorghum bicolor</i> subsp. <i>bicolor</i> )	<i>S. b.</i> subsp. <i>arundinaceum</i> , <i>S. b.</i> subsp. <i>drummondii</i>	X	X		[85]
Sunflower ( <i>Helianthus annuus</i> )	<i>H. argophyllus</i> , <i>H. bolanderi</i> , <i>H. debilis</i> , <i>H. petiolaris</i>	X			[77]
Tomato ( <i>Solanum lycopersicum</i> )	<i>S. pimpinellifolium</i>	X	X	X	[80]
Wheat ( <i>Triticum monococcum</i> , <i>T. dicoccum</i> , <i>T. aestivum</i> )	<i>T. m. boeoticum</i> , <i>T. diocoides</i> , <i>T. urartu</i> , <i>Aegilops speltoides</i> , <i>A. tauschii</i>	X	X		[86]

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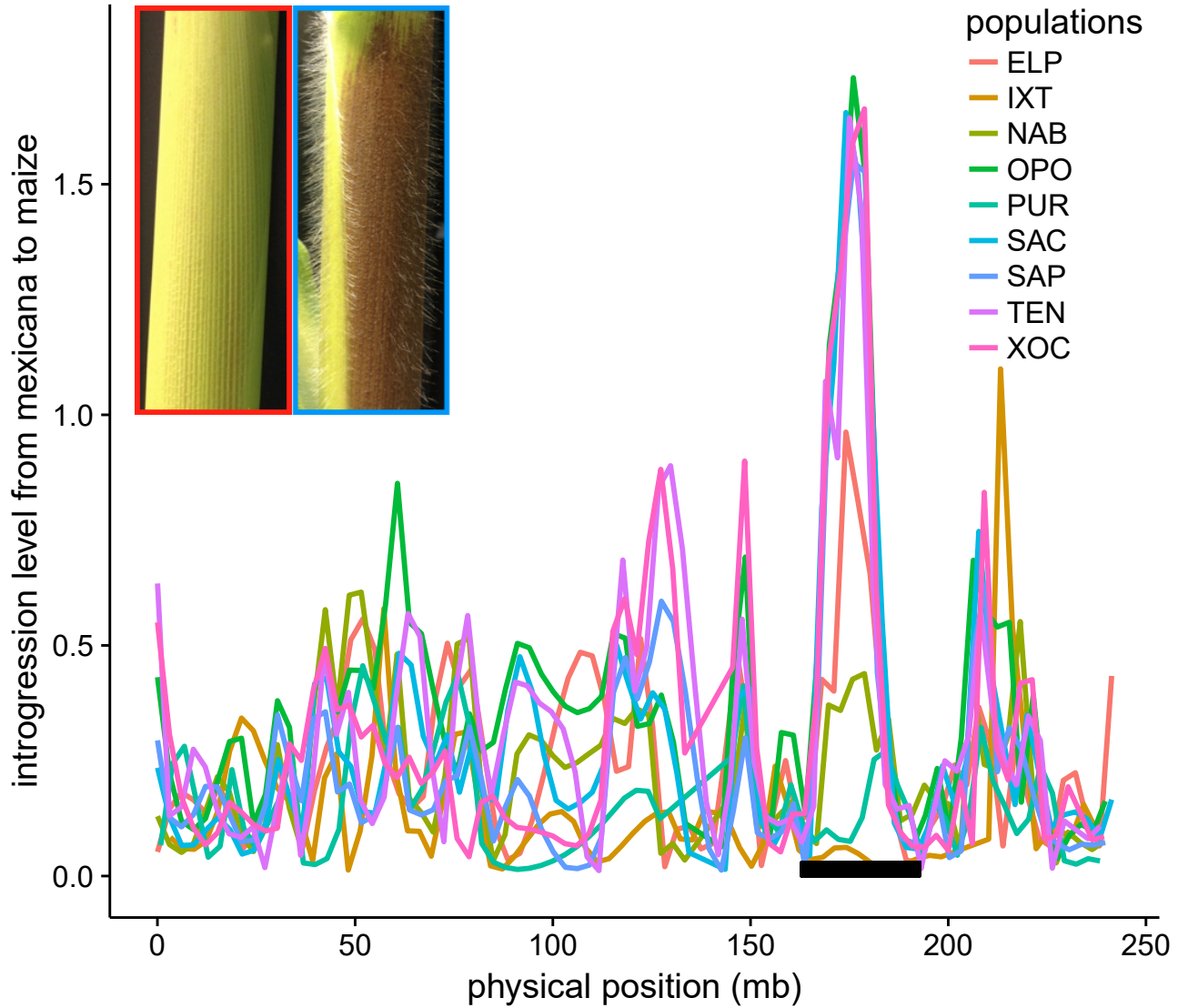


Figure 1: Hapmix vectors indicating introgression level from *mexicana* to Mexican highland maize on chromosome 4. Data adopted from Hufford et al. [36]. ELP: EL Porvenir; IXT: Ixtlan ; NAB: Nabogame; OPO: Opopeo; PUR: Puruandiro; SAC: Santa Clara; SAP: San Pedro; TEN: Tenango del Aire; XOC: Xochimilco. Insets showed the phenotypes of macrohairs on lowland (red) and highland (blue) maize stems. The black bar indicated one of the QTLs for macrohairs and pigment density in Lauter et al. [37].

Table 1: List and brief description of recently developed methods and examples of empirical studies employing these methods.

| Methods                                   | Data Type                                 | References | Applications |
|-------------------------------------------|-------------------------------------------|------------|--------------|
| <b>Divergence</b>                         |                                           |            |              |
| Gmin                                      | biallelic SNP                             | [16]       | [61]         |
| RNDmin                                    | phased haplotype                          | [15]       | [62]         |
| $U_{A,B,C(w,x,y)}$ and $Q95_{A,B,C(w,y)}$ | biallelic SNP                             | [30]       | [63]         |
| <b>Chromosome Painting</b>                |                                           |            |              |
| Hapmix                                    | phased haplotype; reference panel         | [12]       | [36, 64]     |
| RASPberry                                 | phased haplotype                          | [65]       | [66]         |
| MultiMix                                  | phased/unphased genotype; reference panel | [67]       | [68]         |
| PCAdmix                                   | phased haplotype                          | [25]       | [69, 70]     |
| LAMP                                      | phased haplotypes; reference panel        | [26]       | [71]         |
| <b>Phylogenetic Relationship</b>          |                                           |            |              |
| ABBA-BABA/D-statistics                    | biallelic SNP                             | [72]       | [31]         |
| fd statistic                              | biallelic SNP                             | [11]       | [73, 74]     |
| five taxon D statistics                   | biallelic SNP                             | [14]       | [75, 76]     |

| Crop                                                                           | Compatible Wild Relatives                                                                                         | Hybrids and/or Hybridization | Evidence of Crop Introgression | Evidence of Adaptiveness | Source     |
|--------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------|------------------------------|--------------------------------|--------------------------|------------|
| Maize ( <i>Zea mays</i> subsp. <i>mays</i> )                                   | <i>Z. m.</i> subsp. <i>mexicana</i> , <i>Z. m.</i> subsp. <i>parviglumis</i>                                      | X                            | X                              | X                        | [36]       |
| Asian Rice ( <i>Oryza sativa</i> )                                             | <i>O. rufipogon</i>                                                                                               | X                            | X                              | X                        | [50]       |
| Barley ( <i>Hordeum vulgare</i> )                                              | <i>H. v.</i> subsp. <i>spontaneum</i>                                                                             | X                            | X                              | X                        | [46]       |
| Sunflower ( <i>Helianthus annuus</i> )                                         | <i>H. argophyllus</i> , <i>H. bolanderi</i> , <i>H. debilis</i> , <i>H. petiolaris</i>                            | X                            |                                |                          | [77]       |
| Cassava ( <i>Manihot esculenta</i> )                                           | <i>M. glaziovii</i>                                                                                               | X                            | X                              | X                        | [78]       |
| Potato ( <i>Solanum tuberosum</i> )                                            | many                                                                                                              | X                            | X                              | X                        | [60, 79? ] |
| Tomato ( <i>Solanum lycopersicum</i> )                                         | <i>S. pimpinellifolium</i>                                                                                        | X                            | X                              | X                        | [80]       |
| Olive ( <i>Olea europaea</i> ssp. <i>europaea</i> var. <i>sativa</i> )         | <i>O. e.</i> ssp. <i>europaea</i> var. <i>sylvestris</i>                                                          | X                            | X                              |                          | [81]       |
| Soybeans ( <i>Glycine max</i> )                                                | <i>G. soja</i>                                                                                                    | X                            | X                              |                          | [82]       |
| Common Bean ( <i>Phaseolus vulgaris</i> )                                      | <i>P. v.</i> var. <i>aborigineus</i> , <i>P. v.</i> var. <i>mexicanus</i> [[not in this source]]                  | X                            | X                              |                          | [83]       |
| Grapes ( <i>Vitis vinifera</i> subsp. <i>vinifera</i> )                        | <i>V. v.</i> subsp. <i>sylvestris</i>                                                                             | X                            | X                              |                          | [84]       |
| Sorghum ( <i>Sorghum bicolor</i> subsp. <i>bicolor</i> )                       | <i>S. b.</i> subsp. <i>arundinaceum</i> , <i>S. b.</i> subsp. <i>drummondii</i>                                   | X                            | X                              |                          | [85]       |
| Wheat ( <i>Triticum monococcum</i> , <i>T. dicoccum</i> , <i>T. aestivum</i> ) | <i>T. m. boeoticum</i> , <i>T. diocoides</i> , <i>T. urartu</i> , <i>Aegilops speltoides</i> , <i>A. tauschii</i> | X                            | X                              |                          | [86]       |
| Apple ( <i>Malus domestica</i> )                                               | <i>M. sylvestris</i> , <i>M. orientalis</i> , <i>M. baccata</i> , <i>M. sieversii</i>                             | X                            | X                              |                          | [87]       |

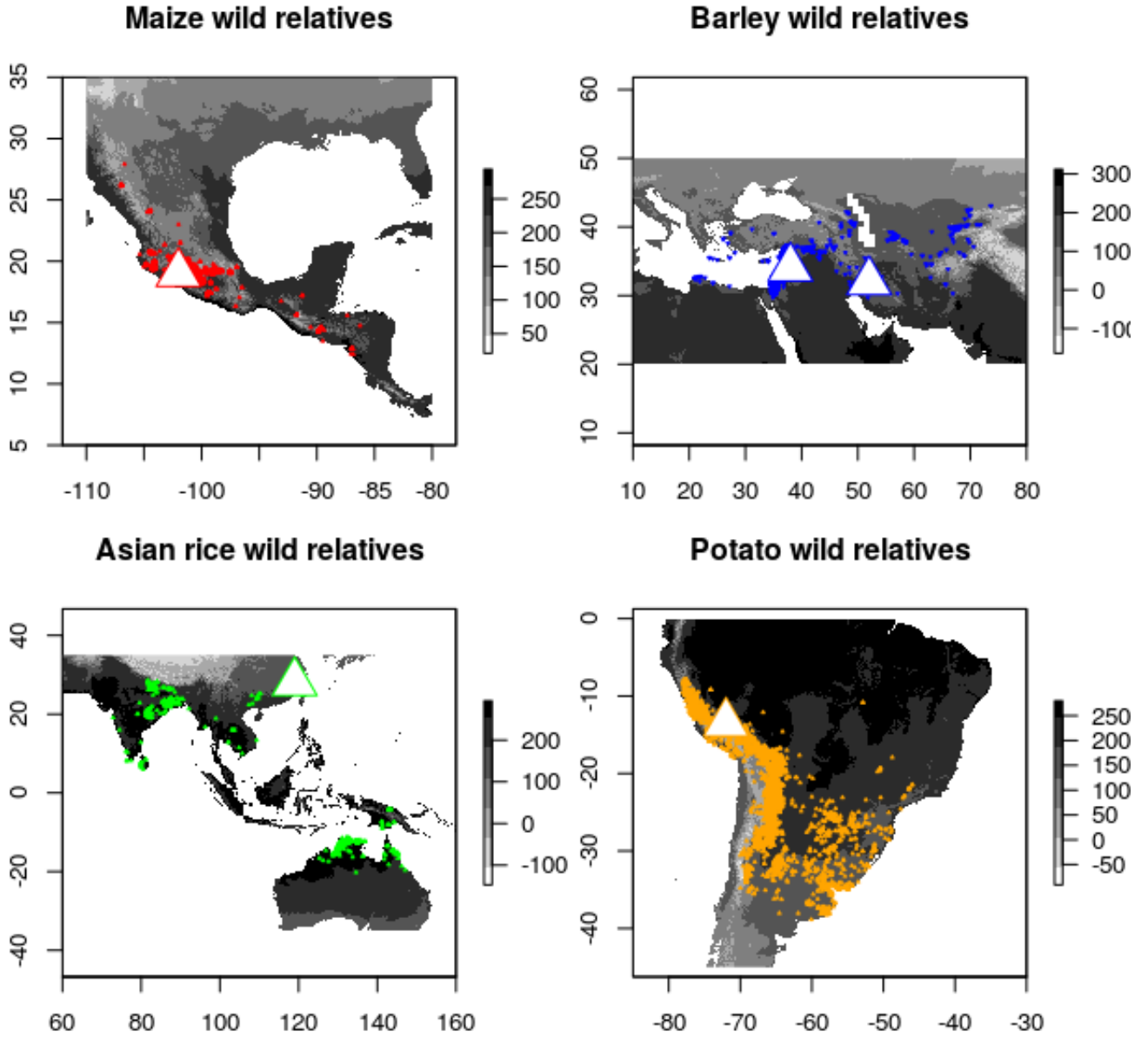


Figure 2: Map of the natural ranges of wild relatives of four domesticated crops, overlaid with average annual temperature. The distribution of average annual temperature experienced in the geographic home ranges of wild relatives interfertile with four crops