# Reminder on hypothesis testing

Statistical test and p-value

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BIS2.0 Back2Basic, Mai 2025

# Guiding thread

#### Differential gene expression

Goal: Identify genes which show a **difference** in expression between two experimental conditions (i.e. greater than expected just due to natural random variation).

You are Mathéo Lode, and you have acces to preclinical data. Tumoral cells are given to mice to see the efficiency of a treatment. Each condition (control and treatment) have 3 observations available of RNAseq data.

# Null and alternative hypothesis

We have two hypothesis, which one is the most likely?

 $H_0$ : the effect can't be observed at population scale  $H_1$ : the effect is observed at population scale

We test the expression of one gene:

 $H_0$ : We don't see a greater difference in expression than expected for this gene  $H_1$ : We find out a significant difference in expression for this gene

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The **null hypothesis**  $H_0$  is the presumption of innocence. We reject  $H_0$  only if it is obvious that observations are incoherent with it.

The null hypothesis is never proved or established, but is possibly disproved, in the course of experimentation [Fisher et al., 1966], Absence of evidence is not evidence of absence [Altman and Bland, 1995].

### Three keys of a statistical test

**Test** of  $H_0$ : rule to reject or not  $H_0$  from data.

- Test statisitic T, it measure the effect: the more evident the effect, the greater the value of T.
- **Distribution** of T under  $H_0$ . If the probability that T being superior than 2 is below 5%, then observing  $t_{gene1} = 3$  must lead to reject  $H_0$ .
- Test's **p-value**: the probability, under  $H_0$ , that the test statistic T is greater than the observed value  $T_{obs}$ .

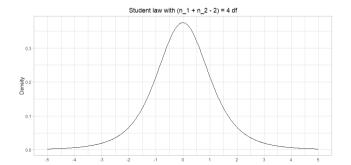
If the **p-value** is smaller than a level  $\alpha$  (usually  $\alpha = 0.05$ ), then the effect is significative at the level  $\alpha$ .

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### Reject zone on the distribution

We consider that the expression of the gene of interest have the same variance in the two experimental conditions, with respectively  $n_1 = n_2 = 3$  observations.

We test the expression of this gene: 
$$\begin{cases} H_0: \mu_A = \mu_B \\ H_1: \mu_A \neq \mu_B \end{cases}, \quad T = \frac{\hat{\mu}_A - \hat{\mu}_B}{\sqrt{\frac{\hat{\sigma}^2}{n_1} + \frac{\hat{\sigma}^2}{n_2}}}$$

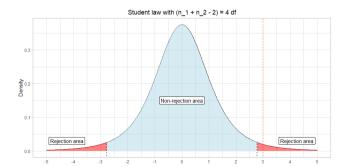


Parametric statistical test

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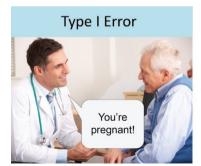
Parametric statistical test

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### Risks in statistical test

#### TRUTH

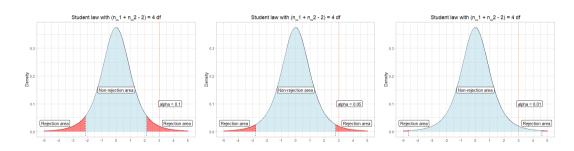
		There is no effect	There is an effect
		H <sub>0</sub> is true	$H_1$ is true
DECISION	Fail to reject H <sub>0</sub>	Good decision	$\beta$ (Type II error)
	Reject H <sub>0</sub>	$\alpha$ (Type I error)	Good decision
			(power $1-\beta$ )





### Influence of risk $\alpha$

→ a priori fixed risk: Probability we accept to be wrong when the truth is a absence of effect.



 $\rightarrow$  **Not a priori fixed risk**: At  $\alpha$  fixed, a good test try to minimise  $\beta$  (i.e. maximise the statistical power).

It is easier to conclude a significative difference between two mean if:

Means are largely different,

Parametric statistical test

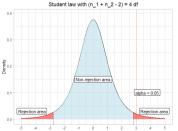
- The variability is low in both population
- We have access to a lot of data

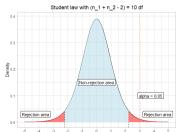
# Influence of risk $\beta$

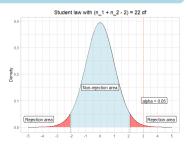
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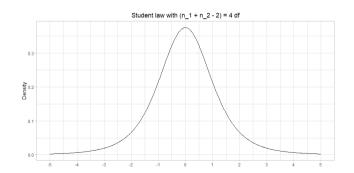


Parametric statistical test

p-value = 
$$\mathbb{P}(T = t_{gene1}|\mathsf{H}_0)$$
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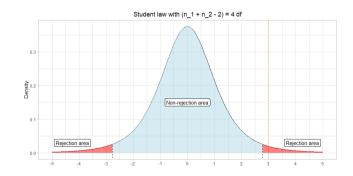
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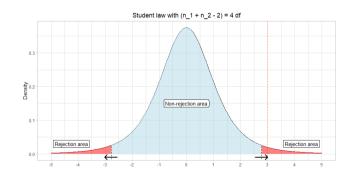
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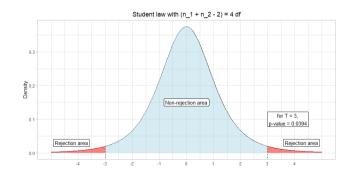
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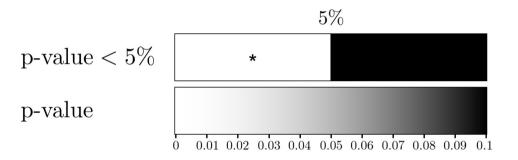
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$$t_{gene1} = 3$$



### A p-value can be interpretted



There is a reason that the speedometer in your car doesn't just read "slow" and "fast". (F. Harrel, 'warning about the use of cutoffs after logistic regression' in R-help, 2011)

### Non-parametric statistical tests

Assumptions on data distribution → Parametric tests (normality, homogeneity of variance, ...) No assumptions of a specific distribution  $\rightarrow$  Non-parametric tests  $\implies$ More reliable when data samples have a small size

### Non-parametric doesn't mean no assumptions

	parametric test	non-parametric test
use	row data	row data, but a lot use ranks  • Supp details
assume	identically distributed and independent data, homogenous variance,	less needed, at least independent data
power:	optimal if assumptions are respected, drop quickly if not	stable/robust but less than parametric in the best case

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#### Non-parametric test still have assumption:

- Wilcoxon-Mann-Whitney assume independent observations.
- Kruskal-Wallis is a non-parametric version of ANOVA, but assume all groups have an identically shaped and scaled distribution [Kruskal and Wallis, 1952]
   [Wikipedia, ].

Multiple statistical tests

### Differential gene expression

Goal: Identify genes which show a **difference** in expression between two experimental conditions (i.e. greater than expected just due to natural random variation).

### Testing all the genes

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Multiple statistical tests oooooooo

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⇒ How much false positive discovered gene can we expect? Since we fixed  $\alpha = 0.05$ , we expect 5% of the 20061 gene tested to be false positive, so 1003 genes or 31.1% of our list of gene differentially expressed.

### Multiple hypothesis

A null hypothesis collection,  $H_0^{(k)}$  k = 1, ..., m, with  $m_0$  true null hypothesis.

m boxes out of which  $m_0$  are empty



For the  $k^{th}$  box,  $H_0^{(k)}$ : box is empty

### Multiple p-values

For the  $k^{th}$  test, the associated p-value is  $p_k = \mathbb{P}_{\mathsf{H}_0^{(k)}} \big( \text{reject } \mathsf{H}_0^{(k)} \big).$ 

I weight each box and evaluate its probability  $p_k$  of being empty

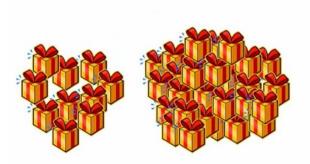


Take these ones ...

 $p_k \leq \alpha$ 

### Multiple p-values

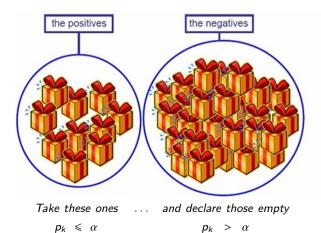
For the  $k^{th}$  test, the associated p-value is  $p_k = \mathbb{P}_{H_0^{(k)}}$  (reject  $H_0^{(k)}$ ).



and declare those empty

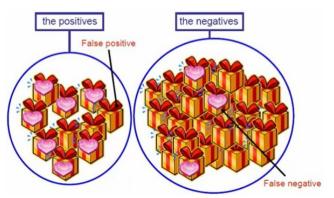
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Take these ones ... and declare those empty

$$p_k \leq \alpha$$

$$p_k > a$$

# An efficient procedure

#### What is an **efficient procedure**?

- Controlling the risk of false positive; as few disappointing discovery as possible.
- Great proportion of true positives within the positives; as much discovery as possible.
- No cofounding effects; knowing what we discover.

# An efficient procedure

#### What is an efficient procedure?

- Controlling the risk of false positive; as few disappointing discovery as possible.
- Great proportion of true positives within the positives; as much discovery as possible.
- No cofounding effects; knowing what we discover.

#### How to build an efficient procedure?

- 1. A powerfull Design of Experiment; if the gift is much heavier than the box, it's easier.
- 2. A good choice for the threshold  $\alpha$ .

While a good design does not guarantee a successful experiment, a suitably bad design guarantees a failed experiment—no results or incorrect results. (K.M. Kerr, 'Experimental design to make the most of microarray studies', 2003)





Day 2

8 Control samples

8 Treatment samples

The global mean of samples for our 2 conditions are significatively differents,

### 1. Design of Experiment (DoE)





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Day 2

8 Treatment samples

The global mean of samples for our 2 conditions are significatively differents, but there are unidentifiability between effect of the treatment s and the day s.

### 1. Design of Experiment (DoE)

For example, we study our 2 conditions (Control/Treatment) with 8 samples each:





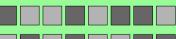
8 Control samples

Day 2

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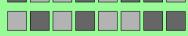
The global mean of samples for our 2 conditions are significatively differents, but there are **unidentifiability** between effect of the treatment and the day

Day 1



8 Control samples

Day 2



8 Treatment samples

Randomisation

#### 1. Design of Experiment (DoE): Fractional plan



Especially if money is an issue, you can test multiple factors at the same time with a minimum of combination [Husson, ].

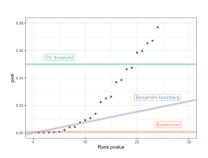
#### 2. A good choice for the threshold $\alpha$

We want to have a decision rule on the m p-values  $p_k$ ,  $1, \ldots, m$ :

- At which threshold  $\alpha$  do we reject  $H_0^{(k)}$  ?
- If p-values are ordered, at which rank  $\hat{k}$  p-values  $p_{(1)}$  to  $p_{(\hat{k})}$  lead to reject the null hypothesis, while p-values  $p_{(\hat{k}+1)}$  to  $p_{(m)}$  doesn't ?

#### For a threshold $\alpha$ :

- $P_{\alpha}$ : numbers of gene (gift box) which reject the null hypothesis (known).
- FP<sub>α</sub>: numbers of gene (gift box) which wrongly reject the null hypothesis (unknown).



### Family Wise error rate (FWER)

To define the new threshold  $\alpha_{FWER}$ , we set:

$$\mathsf{FWER}_{\alpha_{\mathsf{FWFR}}} \leq \mathbb{P}(\mathsf{FP}_{\alpha_{\mathsf{FWFR}}} > 0) = \alpha, \quad \mathsf{equivalently} \ \mathbb{P}(\mathsf{FP}_{\alpha_{\mathsf{FWFR}}} = 0) \geq 1 - \alpha$$

#### Bonferroni procedure

If the number of expected false positive discovery is  $FP_{\alpha} = m \times \alpha$  we only have to take

$$\alpha_{FWER} = \frac{\alpha}{m}.$$

The same result is obtain if we adjust p-values:  $p_k \leqslant \frac{\alpha}{m} \Leftrightarrow \tilde{p}_k = mp_k \leqslant \alpha$ . Supp details

⇒ FWER procedures tends to be very restrictive with few false positive.

In our case,  $\alpha_{FWER} = \frac{0.05}{20061} = 2.492 \times 10^{-6}$  which lead to 38 positive genes.

### False discovery rate (FDR)

To define the new threshold  $\alpha_{FDR}$ , we set:

$$\mathsf{FDR}_{\alpha_{FDR}} = \mathbb{E}\left[\frac{\mathit{FP}_{\alpha_{FDR}}}{\mathit{P}_{\alpha_{FDR}}}\right] \leqslant \alpha$$

#### Benjamini-Hochberg procedure

The number of false positive is unknown, so we control the number of positive and estimate the esperance of fale positive. For that, we order p-values and for the  $k^{th}$  p-value we have

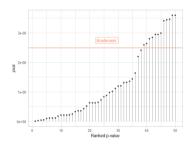
$$\alpha_{FDR} = k \frac{\alpha}{m}$$
.

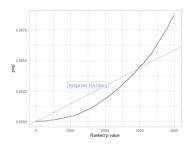
Again we can rather look at adjust p-values:  $\tilde{p}_k = \frac{mp_k}{k} \leqslant \alpha$ . Supp details

⇒ FDR procedures are less demanding than FWER, and provides much more positives.

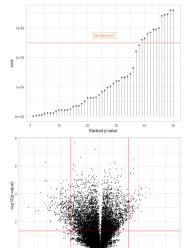
In our case,  $\alpha_{FDR} = k \frac{0.05}{20061}$  which lead to 1958 positive genes ( $\alpha_{FDR} = 0.00488$ ).

## Ranked p-values



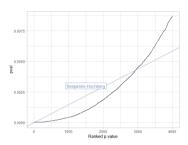


# Ranked p-values

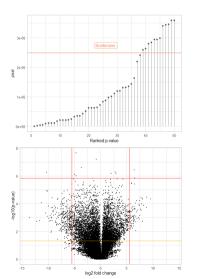


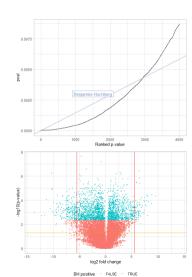
log2 fold change

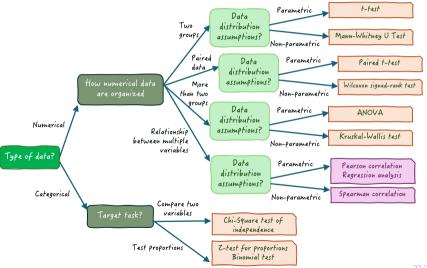
-15

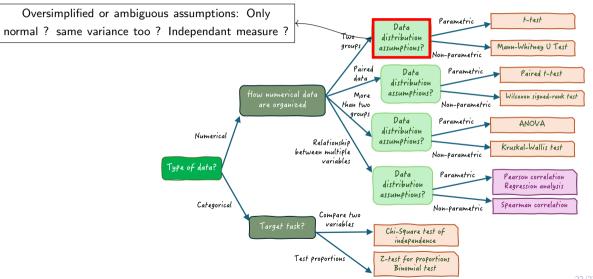


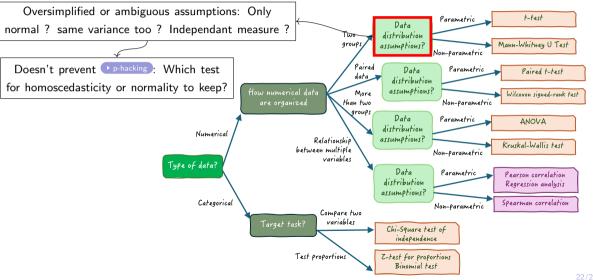
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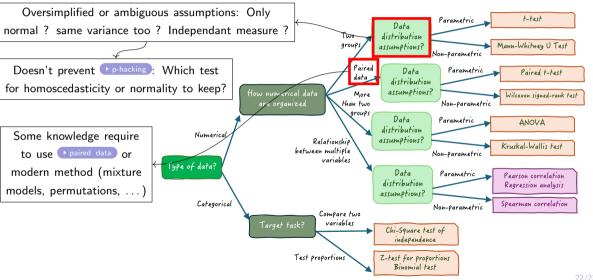


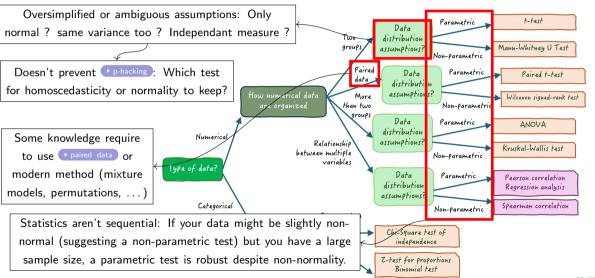












You knew 90% of all tests used, but now you better understand what you do with them: how they work and what they assume. With these new insights, you precise your limits of understanding and when you reach it.

#### Take home message

- → Use decision trees as a starting point rather than a definitive guide [MacFarland et al., 2016], consult statistical literature for complex cases.
- → With great power comes great responsability [Stan Lee, ], you make arbitrary choices (and it's ok). Always understand why each decision point matters.



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#### Common non-parametric tests Inon-parametric tests assumptions

Test	Data Type Used	Note
Rank-based tests		
Mann-Whitney U (Wilcoxon rank-sum)	Ranks	Compares ranks between two independent groups.
Wilcoxon signed-rank test	Ranks of differences	Based on the ranks of paired differences.
Kruskal–Wallis	Ranks	Extension of Mann–Whitney to $k$ independent groups.
Friedman	Ranks within blocks	For $k$ repeated measures or blocks, ranks computed within each block.
Spearman's $ ho$	Ranks	Monotonic rank correlation.
Kendall's $ au$	Pair orderings	Measures concordance/discordance.
Tests using raw data (or categories)		·
Sign test	Signs (±)	Uses only the sign of differences, not their magnitude or rank.
McNemar's test	Binary categories	Counts discordances in a $2\times2$ before/after contingency table.
Chi-squared test of independence	Counts	Works on cell frequencies in a contingency table.
Kolmogorov-Smirnov test	Raw values	Compares empirical distribution functions.
Lilliefors test	Raw values	Adaptation of KS for normality testing without fixed parameters.
Log-rank test	Survival times	Compares survival curves based on exact event times.

#### Bonferroni procedure FWER

$$\begin{aligned} \mathsf{FWER} &= & \mathbb{P}\big([p_1 \leqslant \alpha_{\mathit{FWER}}] \text{ or } \dots \text{ or } [p_m \leqslant \alpha_{\mathit{FWER}}]\big) \\ &\leqslant & \mathbb{P}\big([p_1 \leqslant \alpha_{\mathit{FWER}}]\big) \, + \, \dots \, + \, \mathbb{P}\big([p_m \leqslant \alpha_{\mathit{FWER}}]\big) \\ &\leqslant & m_0 \times \alpha_{\mathit{FWER}} \end{aligned}$$
 If  $\alpha_{\mathit{FWER}} = \frac{\alpha}{m}$ , then  $\mathsf{FWER} = \frac{m_0}{m} \alpha \leqslant \alpha$ .

#### 

$$\begin{aligned} \mathsf{FDR} &= & \mathbb{E}\left[\frac{\mathit{FP}_{\alpha_{\mathit{FDR}}}}{\mathit{P}_{\alpha_{\mathit{FDR}}}}\right] = \frac{\mathbb{E}\left[\mathit{FP}_{\alpha_{\mathit{FDR}}}\right]}{\mathit{P}_{\alpha_{\mathit{FDR}}}} \\ &= & \frac{\mathit{m}_0 \; \alpha_{\mathit{FDR}}}{\mathit{P}_{\alpha_{\mathit{FDR}}}} = \frac{\mathit{m}_0}{\mathit{m}} \frac{\mathit{m} \; \alpha_{\mathit{FDR}}}{\mathit{P}_{\alpha_{\mathit{FDR}}}} \leq \frac{\mathit{m} \; \alpha_{\mathit{FDR}}}{\mathit{P}_{\alpha_{\mathit{FDR}}}}. \end{aligned}$$

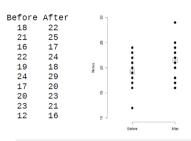
If, for the  $k^{th}$  p-value,  $\alpha_{FDR} = k \frac{\alpha}{m}$ , then FDR  $\leq \alpha$ .

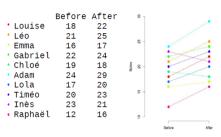
### 

Decision tree doesn't prevent p-hacking [Head et al., 2015]: Which normal test will you keep, qq-plot,  $\chi^2$  adequation, Kolmogorov-Smirnov, Lilliefors, Anderson-Darling, Shapiro-Wilks,... Same question for homoscedasticity.

A comparison of Shapiro-Wilk, Kolmogorov-Smirnov, Lilliefors and Anderson-Darling test concluded that Shapiro-Wilk have the best power for the same significance level, followed by Anderson-Darling [Razali et al., 2011].

#### 





R> wilcox.test(x, y, paired = FALSE)
Wilcoxon rank sum test with continuity
correction

W = 35, **p-value = 0.2716**alternative hypothesis: true location shift is not equal to 0

R> t.test(x, y, paired = FALSE)
Two Sample t-test

t = -1.3529, df = 18, p-value = 0.1928 alternative hypothesis: true difference in means is not equal to  $\theta$ 

R> wilcox.test(x, y, paired = TRUE) Wilcoxon rank sum test with continuity correction

V = 5, **p-value = 0.02428** alternative hypothesis: true location shift is not equal to  $\theta$ 

R> t.test(x, y, paired = TRUE)
Two Sample t-test

t = -3.1461, df = 9, **p-value = 0.01181** alternative hypothesis: true difference in means is not equal to 0