

CaFE Manual Release 1.0

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CHAPTER

ONE

INSTALLATION

1.1 Prerequisites

• VMD 1.9.2 or later

Note: There is a bug in the topotools1.5 plugin, which is included in VMD 1.9.2 and invoked by CaFE. You should change line 43 in:

Linux/MacOSX

\$VMDDIR/lib/plugins/noarch/tcl/topotools1.5/topoatoms.tcl

Windows

\$VMDDIR/plugins/noarch/tcl/topotools1.5/topoatoms.tcl

from:

```
$s set element [lindex $elements [ptefrommass $idx]]
```

to:

```
$s set element [lindex $elements [ptefrommass $a]]
```

Alternatively, you can copy the corrected topoatoms.tcl file in the patch folder, which is distributed with the CaFE source code, and replace the associated file in the topotools1.5 plugin folder in your installed VMD distribution.

- NAMD 2.9 or later
- APBS 1.3 or later

Optionally, for MM/PBSA only

• DelPhi 5.1 or later

Optionally, for MM/PBSA only

All these tools are commonly used in molecular simulations, please download the installation files and follow the corresponding instructions on their websites. Make sure that all the folders containing the binary files are appended to the PATH environment variable.

1.2 Download and install

After installing the above required programs, you could download the CaFE source code: CaFE_Plugin-master.zip from https://github.com/HuiLiuCode/CaFE_Plugin.

If you have root access for installation, just extract the compressed contents and rename the src folder to cafe1.0 and copy it to \$VMDDIR/lib/plugins/noarch/tcl (Linux/MacOSX) or \$VMDDIR/plugins/noarch/tcl (Windows), where \$VMDDIR is the directory in which VMD installed.

If you don't have root access, try to extract the contents and rename the src folder to cafe1.0 and copy it to somewhere in your \$HOME directory. Then, add the following to the VMD start-up file \$HOME/.vmdrc (Linux/MacOSX) or vmd.rc (Windows):

set auto_path [linsert \$auto_path 0 {/PATH/TO/YOUR/FOLDER}]

CHAPTER

TWO

APPLICATIONS

CaFE is an automatic pipeline tool for post-processing and energetic analysis, which is powered by VMD [HW96] (page 14). Prior to the calculations, topology and trajectory files are needed to be generated by common molecular dynamics (MD) simulation software. Currently, AMBER [CD05] (page 14) and NAMD [PJ05] (page 14) are generally supported. Other simulation packages using the same topology files are supported, too.

Note: If AMBER is used, you'd better set "iwrap=1" in the input file or use *cpptraj* to "image" the generated trajectory. If NAMD is used, you'd better set "wrapAll on" in the configuration file.

CaFE is implemented as a set of Tcl scripts. When using it, you should import the *cafe* package by appending the following line in the analysis scripts first:

```
package require cafe 1.0
```

After that, two major procedures can be invoked for binding free energy calculations.

2.1 The mmpbsa procedure

The **mmpbsa** procedure automates post-processing and energetic analysis using the molecular mechanics Poisson-Boltzmann surface area (MM/PBSA) method [KP00] (page 14). A trajectory file containing the binding complex is required, and then the receptor and ligand will be extracted from that according to the one-trajectory protocol [KP00] (page 14). For MM/PBSA calculations, either APBS [BN01] (page 14) or DelPhi [LL12] (page 14) is needed to be installed.

2.1.1 Usage

```
mmpbsa -top file -trj file [-args ...]
```

Mandatory arguments

• -top: topology file name

available values: a valid file name

default value: none-trj: trajectory file name

available values: a valid file name

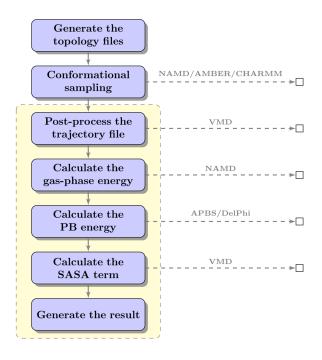


Fig. 2.1: **Work flow for MM/PBSA calculations**. The procedures in yellow background are automated by CaFE.

default value: none

Optional arguments

• -top_type: topology file type
available values: parm7 (AMBER format), psf (CHARMM/NAMD format), auto

default value: auto

-trj_type: trajectory file type
 available values: all of the coordinate and trajectory file types supported by VMD
 default value: auto

• -par: CHARMM-formatted force field parameter file, required only if the PSF topology type is used available values: a valid file name

default value: \$CAFEDIR/par_all22_prot.inp

• -out: output file name

available values: a valid file name

default value: result.log

• -debug: debug level

available values: 0 (remove all of the intermediate files), 1 (remove a part of them), 2 (keep all)

default value: 0

```
• -first: the first frame index, starting from 0
  available values: an integer >= 0
  default value: 0
• -last: the last frame index, starting from 0
  available values: an integer >= 0 or -1 (stands for the last frame of the trajectory)
  default value: -1
• -stride: stride
  available values: an integer >= 1
  default value: 1
• -com: complex selection
  available values: a valid selection string used by VMD
  default value: ""
• -rec: receptor selection
  available values: a valid selection string used by VMD
  default value: ""
• -lig: ligand selection
  available values: a valid selection string used by VMD
  default value: ""
• -mm: do gas-phase calculations or not
  available values: 0 (do not) or 1 (do)
  default value: 0
• -mm_exe: path to the NAMD binary
  available values: a valid path
  default value: "namd2"
• -mm diel: dielectric constant
  available values: a float
  default value: 1.0
• -pb: do PB calculations or not
  available values: 0 (do not), 1 (use DelPhi), 2 (use APBS)
  default value: 0
• -pb_exe: path to the DelPhi/APBS binary
  available values: a valid path
  default value: "delphi77"
• -pb_siz: radii parameter file, not a necessity (DelPhi-only)
  available values: a valid file name
  default value: ""
```

```
• -pb_crg: charge parameter file, not a necessity (DelPhi-only)
  available values: a valid file name
  default value: ""
• -pb_rad: type of PB radii
  available values: bondi, rowland, mparse, parse, charmm, roux, parm7 (valid only if the topol-
  ogy is an AMBER parm7 file, using the radii included in it)
  default value: bondi
• -pb_indi: internal dielectric constant
  available values: a float
  default value: 1.0
• -pb_exdi: external dielectric constant
  available values: a float
  default value: 80.0
• -pb_scale: the reciprocal of grid spacing
  available values: a float
  default value: 2.0
• -pb_perfil: percentage of filling
  available values: a float
  default value: 80.0
• -pb_prbrad: probe radius
  available values: a float
  default value: 1.4
• -pb_linit: max number for linear iterations
  available values: an integer > 3
  default value: 1000
• -pb_maxc: convergence threshold
  available values: a float > 0.0
  default value: 0.0001
• -pb_bndcon: boundary condition (DelPhi-only)
  available values: 1 (zero boundary condition), 2 (dipolar), 4 (coulombic)
  default value: 4
• -pb_bcfl: boundary condition (APBS-only)
  available values: zero (zero boundary condition), sdh (single Debye-Huckel), mdh (multiple Debye-
  Huckel)
  default value: sdh
```

```
• -pb_chgm: the method that charges are mapped to the grids (APBS-only)
  available values: spl0 (linear splines), spl2 (cubic B-spline), spl4 (quintic B-spline)
  default value: spl0
• -pb_srfm: the model used to construct the dielectric and ion-accessibility coefficients (APBS-only)
  available values: mol, smol, spl2, spl4
  default value: smol
• -pb_swin: spline window width (APBS-only)
  available values: a float > 0.0
  default value: 0.3
• -pb_sdens: density of grids (APBS-only)
  available values: a float > 0.0
  default value: 10.0
• -sa: do SA calculations or not
  available values: 0 (do not), 1 (use VMD)
  default value: 0
• -sa rad: type of SA radii
  available values: bondi, rowland, mparse, parse, charmm, roux, parm7 (valid only if the topol-
  ogy is an AMBER parm7 file, using the radii included in it)
  default value: bondi
• -sa_gamma: surface tension
  available values: a float
  default value: 0.005
• -sa_beta: surface offset
  available values: a float
  default value: 0.0
• -sa_prbrad: probe radius
  available values: a float
  default value: 1.4
• -sa_samples: number of samples
  available values: an integer > 0
  default value: 500
```

For more details about the parameters, please see the manuals of VMD (supported file types, selection string, SA calculations) and APBS/DelPhi (PB calculations).

2.1.2 Example

Now, suppose that we have performed the conformational sampling by using MD simulations, there are two folders in the working folder workdir:

- 1. md, which contains topology file com.psf, and generated trajectory com.dcd;
- 2. toppar, which contains force field parameter files.

In the PSF file, the receptor and ligand have been put into segments named "PRO" and "LIG", respectively. The trajectory file is totally 1 ns long and saved every 100 ps. We want to perform MM/PBSA calculations from 0 to 1 ns and produce snapshots every 100 ps. APBS is used in the PB calculations.

A typical analysis script is something like this:

```
package require cafe 1.0
                 ../md/com.psf \
mmpbsa -top
      -trj
                 ../md/com.dcd \
      -out
               mmpbsa.log \
      -par
                 ../toppar/par_all22_prot.prm \
      -par
                 ../toppar/par_all36_cgenff.prm \
      -par
                ../toppar/toppar_water_ions_namd.str \
                 ../toppar/lig.str \
      -par
      -com
                 "segname PRO LIG" \
                 "segname PRO" \
      -rec
      -lig
                 "segname LIG" \
      -first 0 \
      -last
                -1 \
      -stride 1 \setminus
                1 \
                2 \
      -pb
      -pb_exe apbs \
      -pb_rad mparse \
      -pb_bcfl mdh \
      -pb_chgm spl2 \
       -sa
                 1 \
      -sa_rad mparse \
      -sa_gamma 0.00542 \
      -sa_beta 0.92
quit
```

Create a new folder mmpbsa in workdir and save the above contents to a file named mmpbsa.vmd in it.

Note: Don't forget the backslash!

Then run commands as follows in the terminal, which will perform the MM/PBSA calculations and generate results in plain text:

```
$ cd workdir/mmpbsa
$ vmd -dispdev text -eofexit < mmpbsa.vmd > vmd.log
```

The output file mmpbsa.log, which contains the calculated binding free energy, is self-explanatory. Files that are required to perform the exemplified calculations are in the examples folder distributed alongside the source code.

2.2 The lie procedure

The **lie** procedure automates post-processing and energetic analysis using the linear interaction energy (LIE) method [AJ94] (page 14). Two topology as well as MD trajectory files containing the solvated complex and ligand are required.

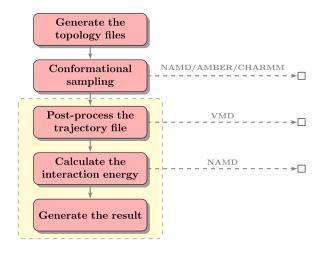


Fig. 2.2: Work flow for LIE calculations. The procedures in yellow background are automated by CaFE.

2.2.1 **Usage**

```
lie -top_bound file -trj_bound file -top_free file -trj_free file [-args ...]
```

Mandatory arguments

• -top_bound: topology file name for the bound state

available values: a valid file name

default value: none

• -trj_bound: trajectory file name for the bound state

available values: a valid file name

default value: none

• -top_free: topology file name for the free state

available values: a valid file name

default value: none

• **-trj_free**: trajectory file name for the free state

available values: a valid file name

default value: none

Optional arguments

```
• -top_type: topology file type
  available values: parm7 (AMBER format), psf (CHARMM/NAMD format), auto
  default value: auto
• -trj_type: trajectory file type
  available values: all of the coordinate and trajectory file types supported by VMD
  default value: auto
• -par: CHARMM-formatted force field parameter file, required only if the PSF topology type is used
  available values: a valid file name
  default value: $CAFEDIR/par_all22_prot.inp
• -out: output file name
  available values: a valid file name
  default value: result.log
• -debug: debug level
  available values: 0 (remove all of the intermediate files), 1 (remove a part of them), 2 (keep all)
  default value: 0
• -first_bound: the first frame index for the bound state, starting from 0
  available values: an integer >= 0
  default value: 0
• -last_bound: the last frame index for the bound state, starting from 0
  available values: an integer \ge 0 or -1 (stands for the last frame of the trajectory)
  default value: -1
• -stride bound: stride for the bound state
  available values: an integer >= 1
  default value: 1
• -lig_bound: ligand selection for the bound state
  available values: a valid selection string used by VMD
  default value: ""
• -first_free: the first frame index for the free state, starting from 0
  available values: an integer >= 0
  default value: 0
• -last_free: the last frame index for the free state, starting from 0
  available values: an integer >= 0 or -1 (stands for the last frame of the trajectory)
  default value: -1
```

-stride_free: stride for the free state
 available values: an integer >= 1
 default value: 1

• -lig_free: ligand selection for the free state

available values: a valid selection string used by VMD

default value: ""

• -mm_exe: path to the NAMD binary

available values: a valid path

default value: "namd2"

• -alpha: van der Waals coefficient

available values: a float **default value**: 0.18

• -beta: electrostatic coefficient

available values: a float default value: 0.33

• -gamma: offset

available values: a float default value: 0.0

For more details about the parameters, please see the manual of VMD (supported file types, selection string).

2.2.2 Example

Now, suppose that we have performed the conformational sampling by using MD simulations, there are two folders in the working folder workdir:

- 1. md, which contains topology files com.psf and lig.psf, and generated trajectories com.dcd and lig.dcd;
- 2. toppar, which contains force field parameter files.

In both the PSF files, the ligand has been put into a segment named "LIG". Each of the two trajectory files is 1 ns long and saved every 100 ps. We want to perform LIE calculations from 0 to 1 ns and produce snapshots every 100 ps.

A typical analysis script is something like this:

```
package require cafe 1.0
lie -top_bound
                 ../md/com.psf \
   -trj_bound
                 ../md/com.dcd \
               ../md/lig.psf \
    -top_free
    -trj_free
                 ../md/lig.dcd \
                 lie.log \
    -out
   -par
                 ../toppar/par_all22_prot.prm \
                 ../toppar/par_all36_cgenff.prm \
    -par
                  ../toppar/toppar_water_ions_namd.str \
    -par
```

```
../toppar/lig.str \
   -lig_bound "segname LIG" \
               "segname LIG" \
   -lig_free
   -first_bound 0 \setminus
   -last_bound
                 -1 \
   -stride_bound 1 \
   -first_free 0 \
   -last_free
                -1 \
   -stride_free 1 \
   -alpha 0.18 \
                0.5 \
   -beta
   -gamma
                 0.0
quit
```

Note: The values of alpha, beta and gamma are dependent on your simulated systems. Please see related references for the choice of values.

Create a new folder lie in workdir and save the above contents to a file named lie.vmd in it.

Note: Don't forget the backslash!

Then run commands as follows in the terminal, which will perform the LIE calculations and generate results in plain text:

```
$ cd workdir/lie
$ vmd -dispdev text -eofexit < lie.vmd > vmd.log
```

The output file lie.log, which contains the calculated binding free energy, is self-explanatory. Files that are required to perform the exemplified calculations are in the examples folder distributed alongside the source code.

CHAPTER

THREE

ABOUT CAFE

CaFE is a free and open-source VMD plugin for binding affinity prediction using end-point free energy methods. It is being developed in Hou Lab at Zhejiang University.

3.1 Citation

If CaFE is utilized in scientific work that results in a publication, it is expected that the following reference is cited:

Liu H, Hou T.

CaFE: a tool for binding affinity prediction using end-point free energy methods.

In submission

3.2 License

CaFE is freely available under the GPL License:

CaFE: Calculation of Free Energy

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